

International Journal of Medical Laboratory Shahid Sedoughi University of Medical Sciences

Original Article

Diversity and Distribution Patterns of Culturable Airborne Fungi in Jiroft City

Mohadeseh Kamali¹ M.D., Mehdi Taheri Sarvtin^{2*} Ph.D.

¹ Department of Internal Medicine, School of Medicine, Jiroft University of Medical Sciences, Jiroft, Iran
² Department of Medical Mycology and Parasitology, School of Medicine, Jiroft University of Medical Sciences, Jiroft, Iran

Article history Received: 18 Jun 2023 Accepted: 29 Nov 2023 Available online: 24 Feb 2024

Keywords Airborne Distribution Diversity Fungi



© This article is distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use and redistribution provided that the original author and source are credited.

Background and Aims: Fungal spores constitute a significant portion of primary biological aerosol particles, and large amounts have been identified in the air. They are present in the atmosphere throughout the year, and their concentration varies due to meteorological factors such as temperature, relative humidity, location, construction conditions, and vegetation. Exposure to airborne fungi has been linked with diseases such as allergic respiratory symptoms, rhinitis, asthma, and various infections. This study investigates the diversity and distribution of airborne fungal spores in Jiroft City, located in the southern region of Iran.

ABSTRACT

Materials and Methods: Airborne fungal composition was sampled from indoor, outdoor, and hospital air using the sedimentation plate method. Grown fungi were identified using standard mycological techniques, including determining macroscopic and microscopic characteristics.

Results: A total of 230 colonies belonging to 11 genera of fungi were isolated from all culture media. Fungi were isolated from 53 (80.3%) samples. The highest and lowest percent of positive samples were found in outdoor (100%) and hospital (64.3%) air samples, respectively. *Cladosporium* (43.9%) and *Aspergillus* (36.4%) were the most common fungi isolated from the samples. *Rhodotorula, Syncephalastrum, Paecilomyces, Mucor,* and *Acremonium* were the least isolated fungi (1.5%).

Conclusion: This study showed the difference in the diversity and distribution of fungi in different environments. *Cladosporium* and *Aspergillus* were the most common fungi isolated, which need to be considered due to their ability to cause various diseases.

^{*} Corresponding Author: Department of Medical Mycology and Parasitology, School of Medicine, Jiroft University of Medical Sciences, Jiroft, Iran. Email: mehditaheri.mt@gmail.com; Tel/ Fax: +983443317902

Introduction

Fungi are eukaryotic, non-chlorophyllous, and heterotrophic microorganisms that depend entirely on external nutrients for survival [1]. Fungal spores are everywhere in nature [2]. Fungal spores can be passively released from natural and anthropogenic sources [3]. Air is an important channel for the transporting and dispersal of biological particles such as fungi [4]. Most spores are released into the air and remain for some time before being transported over different distances (short or long). Fungal spores can be aerosolized from the surfaces of their colonies by rainwater drops and transported over short distances. Spores are carried by wind over long distances. Forests, green spaces, and decaying plant material are the primary sources of airborne fungal spores [1]. Fungal spores can cause various human, animal, and plant diseases [2]. For example, humans can cause infection, autoimmune reactions, and allergies such as allergic bronchopulmonary aspergillosis, severe asthma with fungal sensitization, and allergic fungal rhinosinusitis [2, 5]. The size of fungal spores determines the depth of their penetration and deposition in the human respiratory system, which seriously affects health [4]. Tiny spores can penetrate the alveolar region during inhalation, whereas large spores are deposited in the nasopharynx [6]. The abundance of spores in the air can be affected by geographical location, weather, use of fungicides, season, and time of day [4,7,8]. Interest in fungal spores has been increasing in recent decades [1]. Due to the potential risks

of airborne fungal spores to human health and other organisms, it is necessary to regularly monitor the presence and diversity of fungal spores in the atmosphere [4, 9]. Several studies have investigated airborne fungi in various parts of the world and reported different results [9-11]. So far, a comprehensive study has not been conducted on airborne fungi in Jiroft city. Therefore, this study was carried out to investigate the diversity and distribution patterns of airborne fungi in Jiroft city in the south of Iran.

Materials and Methods

The present study was conducted in the spring of 2023 at Jiroft University of Medical Sciences. Jiroft is located approximately 1375 kilometers from Tehran at 690 m above sea level. This study collected fourteen air samples from the hospital, forty-two indoor samples (14 kitchen air samples, 14 hall air samples, 14 bathroom air samples), and 10 outdoor samples. The open plate sedimentation method was used to collect the samples. This way, plates containing Sabouraud dextrose agar (SDA) were opened for at least two minutes. Then, the plates were sealed, transferred to the mycology laboratory, and incubated at 27 °C. Cultures were periodically examined for fungal growth. The number of grown colonies was recorded. Grown fungi were identified using standard mycological techniques, including determining macroscopic and microscopic characteristics.

Statistical analysis

Data were analyzed using descriptive statistics and a one-sample t-test. All statistical analyses were performed in SPSS (version 16) and at a significance of 0.05.

Results

In this study, a total of sixty-six air samples were collected. Fungi were isolated from 53 (80.3%) samples. The highest and lowest percent of positive samples were found in outdoor (100%) and hospital (64.3%) air samples, respectively (Fig. 1). A total of 230 colonies were isolated from all culture media. The highest and lowest percent of isolated colonies were from indoors (53%) and hospital (12.2%), respectively (Fig. 2). The number of colonies isolated from the hospital was significantly less than the colonies isolated from outdoors and indoors (p = 0.000). The number of colonies recovered from the hall air samples was significantly less than those recovered from the kitchen air samples (p =

0.004). The number of colonies recovered from the bathroom air samples was significantly less than those recovered from the kitchen air samples (p = 0.04). No significant difference was observed between the number of colonies isolated from the bathroom and hall (p > 0.05). The number of colonies recovered from the bathroom and hall air samples was significantly less than those isolated from outdoor (p = 0.000). No significant difference was observed between the number of colonies isolated from the kitchen and outdoor air samples (p = 0.2). A total of eleven fungal genera were identified. *Cladosporium* (43.9%) and *Aspergillus* (36.4%) were the most common fungi isolated from the samples. Rhodotorula, Syncephalastrum, Paecilomyces, Mucor, and Acremonium were the least isolated fungi (1.5%). Penicillium, Bipolaris, Alternaria, and Rhizopus were other isolated fungi genera. In this study, some yeasts and some unrecognizable fungi were isolated (Table 1).

Fungus	Outdoor	Hospital	Hall	Bathroom	Kitchen	Total
	N(%)	N(%)	N(%)	N(%)	N(%)	N(%)
Cladosporium	5(17.23%)	5(17.23%)	6(20.7%)	6(20.7%)	7(24.1%)	29(100%)
Aspergillus	7(29.2%)	3(12.5%)	5(20.8%)	6(25%)	3(12.5%)	24(100%)
Alternaria	3(21.4%)	2(14.3%)	2(14.3%)	4(28.6%)	3(21.4%)	14(100%)
Unrecognizable	1(7.6%)	2(15.4%)	4(30.8%)	2(15.4%)	4(30.8%)	13(100%)
Penicillium	1(25%)	2(50%)	0(0%)	1(25%)	0(0%)	4(100%)
Yeasts	0(0%)	1(25%)	0(0%)	1(25%)	2(50%)	4(100%)
Bipolaris	1(33.3%)	1(33.3%)	0(0%)	0(0%)	1(33.3%)	4(100%)
Rhizopus	1(50%)	0(0%)	1(50%)	0(0%)	0(0%)	2(100%)
Rhodotorula	0(0%)	0(0%)	0(0%)	0(0%)	1(100%)	1(100%)
Syncephalastrum	0(0%)	1(100%)	0(0%)	0(0%)	0(0%)	1(100%)
Paecilomyces	0(0%)	0(0%)	0(0%)	0(0%)	1(100%)	1(100%)
Mucor	0(0%)	0(0%)	0(0%)	1(100%)	0(0%)	1(100%)
Acremonium	0(0%)	0(0%)	0(0%)	0(0%)	1(100%)	1(100%)

Table 1. Diversity, absolute and relative abundance of fungal genera recovered from the air of Jiroft city

International Journal of Medical Laboratory 2023;10(4): 340-346. © This work is licensed under CC BY-NC 4.0

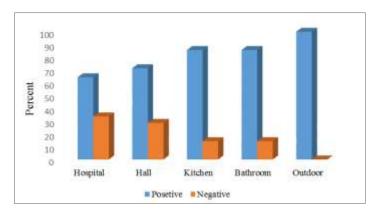


Fig. 1. Relative frequency of positive air samples in Jiroft city

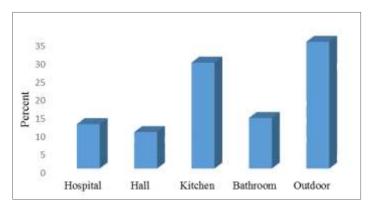


Fig. 2. Relative abundance of isolated colonies from air samples in Jiroft city

Discussion

It seems that this is the first comprehensive study that investigated culturable airborne fungi in Jiroft city. The present study isolated fungi from 80.3% of air samples. The difficulty of some fungi growing in artificial mediums and the need for some particular nutrients may be the reason for the negative effects of some media [12, 13]. The highest and lowest percent of positive samples were found in outdoor and hospital air samples, respectively. Many different nutrient sources outdoors, such as different types of plants, trees, and grasses, can cause various fungi to grow [14]. The type of building materials, ventilation, humidity control, special cleaning, the absence of flowers and plants in the hospital's interior, and the

restriction of different people entering the hospital may be among the reasons for the low percentage of positive samples of hospital air. In the current study, the highest and lowest percent of isolated colonies were from indoor and hospital, respectively. Some indoor fungal spores originate from persistent moisture in household structures; most have an outdoor source [14, 15]. Sometimes, it is impossible to accurately determine whether airborne spores have an internal origin or are imported from outdoors. The present study allowed us to identify 11 genera from airborne spores that constitute part of the environmental mycobiota of Jiroft City. Cladosporium and Aspergillus were the most common fungal genera isolated in all locations. These two fungal genera were also among the dominant fungi in other studies such as Kamali et al. [11], Shams-Ghahfarokhi et al. [15], Sepahvand et al. [16] and Garcia-Cruz et al. [17] studies. Although in some of the studies mentioned above [11, 16], Penicillium was also among the dominant fungi, this fungus was not significantly prevalent in our study. Alternaria was also significantly prevalent in the current study, consistent with the results of Sepahvand et al. [16] and Garcia-Cruz et al. [17]. However, it was inconsistent with the result of Shams-Ghahfarokhi et al. [15] study. The difference in the results of different studies can be related to the geographical region, the method of experimenting, the technician's skill in identifying fungi, temperature, humidity, vegetation, human and animal population, and the type of building materials. Air exposure to fungal material can significantly affect the health of individuals in a specific environment, especially in hospitals. Several fungi, like Aspergillus, Penicillium, Cladosporium, Alternaria, and Fusarium, are highlighted in the air quality of hospitals. The present study recovered Cladosporium, Aspergillus, Penicillium, and Alternaria from hospital air samples. Cladosporium involves fungi with wide distribution around the world, and the existence of this fungus has been confirmed in the literature as one of the most common fungi in hospital air and indoor air [18]. Cladosporium is aero-allergenic and can cause sinusitis, lung infections, severe allergic reactions in the respiratory tract, and intrabronchial lesions [19]. Alternaria is an outdoor fungus that spreads its

spores in warm and dry weather. However, it can also be found in damp and poorly ventilated houses [20]. In our study, the highest number of Alternaria was isolated from the bathroom. The presence of *Alternaria* in hospital air samples has been mentioned [21]. The genus Alternaria, like Cladosporium, is often associated with allergic respiratory diseases [20]. In the present study, Aspergillus was the second most common fungus isolated from air samples. The fungus can cause various respiratory allergic symptoms and infections in immunocompromised and nonimmunocompromised individuals [9, 22]. Penicillium was another fungus isolated in the present study. This fungus is important in producing various allergens, toxins, and volatile organic compounds [18, 23, 24]. In the present study, 13 colonies could not be identified due to the production of sterile mycelium and the lack of production of sporulation apparatus, as well as the indiscernibility of some conidia under a light microscope. Rhodotorula, Syncephalastrum, Paecilomyces, Mucor. Rhizopus, and Acremonium were other fungi isolated from air samples that do not seem to play an essential role in air quality due to their low prevalence.

Conclusion

This study elucidated the highly diverse community of airborne fungi in hospital, indoor, and outdoor air samples of Jiroft city in south Iran. In addition, this study showed the difference in the distribution of fungi in different environments and provided a baseline for future research on the air quality of this city. In addition, the results of this study can help evaluate potential health risks, especially in susceptible and immuno-compromised individuals. In this study, *Cladosporium* and *Aspergillus* were the most common fungi isolated, which need to be considered due to their ability to cause various diseases.

Ethical Consideration

Not applicable.

Funding

No financial interests are involved.

References

- [1]. Martinez-Bracero M, Markey E, Clancy JH, McGillicuddy EJ, Sewell G, O'Connor DJ. Airborne fungal spore review, new advances and automatisation. Atmosphere 2022; 13(2): 308.
- [2]. Singh N, Bhange K. Environmental fungal spore aerosolization: A review. J Bacteriol Mycol. 2023; 11(1): 20-22.
- [3]. Li X, Fu H. Fungal spore aerosolization at different positions of a growing colony blown by airflow. Aerosol Air Qual Res. 2020; 20(12): 2826-833.
- [4]. Odebode A, Adekunle A. Fungal spore diversity and abundance in five areas in Ibadan, South West, Nigeria. JPHE. 2023; 8(2): 168-80.
- [5]. Bongomin F, Gago S, Oladele RO, Denning DW. Global and multi-national prevalence of fungal diseases-estimate precision. J Fungi (Basel). 2017; 3(4): 57-66.
- [6]. Reponen T, Seo SC, Grimsley F, Lee T, Crawford C, Grinshpun SA. fungal fragments in moldy houses: A field study in homes in New Orleans and Southern Ohio. Atmos Environ. 2007; 41(37): 8140-149.
- [7]. Ma G, Gao X, Nan J, Zhang T, Xie X, Cai, Q. Fungicides alter the distribution and diversity of bacterial and fungal communities in ginseng fields. Bioengineered 2021; 12(1): 8043-8056.
- [8]. Sarda-Estève R, Baisnée D, Guinot B, Sodeau J, O'connor D, Belmonte J, et al. Variability and geographical origin of five years airborne fungal spore concentrations measured at Saclay, France from 2014 to 2018. Remote Sensing 2019; 11(14): 1671-676.
- [9]. Obi CM, Enweani-Nwokelo IB, Ogheneovo E, Oshim IO. Pathogenic potentials and DNA sequencing of airborne Aspergillus species from Indoor environments in Nigeria. WJBPHS. 2023; 13(2): 72-9.
- [10]. Reznik YV, Yermishev OV, Palamarchuk OV, Bobrovska OA, Rodinkova VV. Features

Conflict of Interest

The authors declare that there is no conflict of interest.

Acknowledgment

I would like to thank Ms. Mohammadi for his assistance in this study.

Authors' Contributions

Convincing and designing: M.K and M.T; collecting the data: M.K and M.T; analysis tools: M.K and M.T; performing the analysis: M.K and M.T; writing the article: M.K and M.T.

of the seasonal dynamics of airborne fungal spore concentrations in Ukraine. Biosystems Diversity 2023; 31(1): 71-83.

- [11]. Kamali M, Taheri Sarvtin M. A survey on airborne fungal spores in indoor air and outdoor air of Babol city. JJUMS. 2015; 2(1): 116-30.
- [12]. Chen D, Förster H, Adaskaveg JE. Baseline sensitivities of major citrus, pome, and stone fruits postharvest pathogens to natamycin and estimation of the resistance potential in Penicillium digitatum. Plant Dis. 2021; 105(8): 2114-121.
- [13]. Barzee TJ, Cao L, Pan Z, Zhang R. Fungi for future foods. Future Foods 2021; 1(1): 25-37.
- [14]. Tham R, Katelaris CH, Vicendese D, Dharmage SC, Lowe AJ, Bowatte G, et al. The role of outdoor fungi on asthma hospital admission in children and adolescents: A 5-year time stratified case-crossover analysis. Enviro Res. 2017; 154: 42-9.
- [15]. Shams-Ghahfarokhi M, Aghaei-Gharehbolagh S, Aslani N, Razzaghi-Abyaneh M. Investigation on distribution of airborne fungi in outdoor environment in Tehran, Iran. Environ Health Sci Eng. 2014; 12(1): 54.
- [16]. Sepahvand A, Shams-Ghahfarokhi M, Allameh A, Razzaghi-Abyaneh M. Diversity and distribution patterns of airborne microfungi in indoor and outdoor hospital environments in Khorramabad, Southwest Iran. Jundishapur J Microbiol. 2013; 6(2): 186-92.
- [17]. Garcia-Cruz C, Aguilar MN, Arroyo-Helguera O. Fungal and bacterial contamination on indoor surfaces of a hospital in Mexico. Jundishapur J Microbiol. 2012; 5(3): 460-64.
- [18]. Do Nascimento JPM, López AMQ, Andrade M. Airborne fungi in indoor hospital environments. Int J Curr Microbiol App Sci. 2019; 8(1): 2749-772.

- [19]. Ilies DC, Caciora T, Ilies A, Berdenov Z, Hossain MA, Grama V, et al. Microbial air quality in the built environment: Case study of Darvas-La Roche Heritage Museum House, Oradea, Romania. Buildings 2023; 13(3): 620.
- [20]. Abel-Fernández E, Martínez MJ, Galán T, Pineda F. Going over Fungal allergy: alternaria alternata and its allergens. J Fungi. 2023; 9(5): 582-89.
- [21]. Sakartepe E, Uzan AH, Gunyar OA. Airborne mycobiota determined in the neonatal unit of the ege university hospital. Fresenius Environ Bull. 2016; 25(8): 2833-841.
- [22]. De Linares C, Navarro D, Puigdemunt R, Belmonte J. Aspergillus conidia and allergens in outdoor environment: A health hazard? J Fungi. 2023; 9(6): 624.

- [23]. Salo MJ, Marik T, Mikkola R, Andersson MA, Kredics L, Salonen H, et al. Penicillium expansum strain isolated from indoor building material was able to grow on gypsum board and emitted guttation droplets containing chaetoglobosins and communesins A, B and D. Appl Microbiol. 2019; 127(4): 1135-147.
- [24]. Yin G, Zhao H, Pennerman KK, Jurick WM II, Fu M, Bu L, et al. Genomic analyses of penicillium species have revealed patulin and citrinin gene clusters and novel loci involved in oxylipin production. Journal of Fungi 2021; 7(9): 743.