# REVIEW ARTICLE

# The Aging Epigenome and The Rejuvenation Strategies

Anna Meiliana<sup>1,2,3,\*</sup>, Nurrani Mustika Dewi<sup>2,3</sup>, Andi Wijaya<sup>2,3</sup>

<sup>1</sup>Department of Pharmacology and Clinical Pharmacy, Faculty of Pharmacy, Universitas Padjadjaran, Jl. Raya Bandung-Sumedang Km 21,

Jatinangor 45363, Indonesia

<sup>2</sup>Prodia Clinical Laboratory, Jl. Supratman No 43, Bandung 40114, Indonesia <sup>3</sup>Prodia Education and Research Institute, Jl. Kramat Raya No. 150, Jakarta, 10430, Indonesia

\*Corresponding author. E-mail: anna.meiliana@prodia.co.id

Received date: Dec 16, 2021; Revised date: Feb 22, 2022; Accepted date: Feb 23, 2022

### Abstract

**ACKGROUND:** Aging is an unavoidable part of life, defined by a gradual loss in tissue and organ function and an increasing chance of death. Current studies of aging connected the genetic and epigenetic changes to cause this process.

**CONTENT:** When the aging-related epigenetic alterations is accumulated, it may result in irregulated gene expression, metabolic instability, stem cell senescence and exhaustion, and imbalance of tissue homeostasis, which all accelerate the aging process. Altered epigenetic gene regulatory mechanisms such as DNA methylation, histone

modification and chromatin remodeling, and non-coding RNAs can induce aging process, thus manipulating these processes give a chance for the success of age-delaying interventions.

**SUMMARY:** Given updated tools and technologies to investigate the epigenetic regulation affecting aging processes, new therapeutic strategies to delay this process can be developed to increase longevity and improve quality of life.

**KEYWORDS:** aging, epigenetic, senescence, autophagy, mitochondria, metabolism, rejuvenation

Indones Biomed J. 2022; 14(1): 11-28

### Introduction

Aging has been linked with a loss of function at the cellular, tissue, and organismal levels, as well as with a broad variety of illnesses, including cardiovascular and neurological diseases, metabolic disorders, and several types of cancer. Health span is a term that refers to the period of diseasefree physiological health (for example, normal cognition and mobility) and significantly indicates human aging. Understanding the factors regulating aging process, lifespan, and health span can open opportunities to reverse the hallmarks of aging and promote a longer youthful period.(1)

The aging hallmarks can be influenced by genetic and also epigenetic.(2) By controlling the transcriptional

machinery's access to DNA, gene expression dynamically or chronically by chromatin and epigenetic factors can impact, even through cell division. These factors include enzymes that alter DNA or histones (H2A, H2B, H3, and H4), and variant histones (H3.3, macroH2A, and H2A.Z) directly. Numerous epigenetic studies in all aging models showed that in local and global chromatin remodeling, histones become loss and altered. There is also global and local changes in DNA methylation, histone post-translational modification, and chromatin structure and remodeling, and substantial nuclear rearrangement, particularly in mammalian systems. (3) Non-coding RNAs, microRNAs, and circular RNAs, also provide layers of epigenetic control and affect the aging process.(4)

Recent investigations have discovered that significant chromatin alterations occur throughout aging, generally



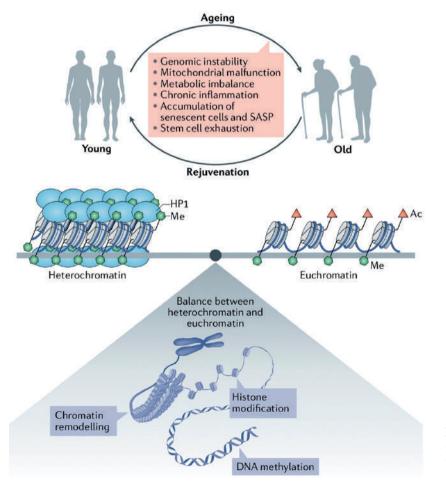
accompanied by gradual loss of constitutive heterochromatin (5,6), and that processes influence transgenerational inheritance, most crucially, the environmental and epigenetic variables (such as sirtuins) which may directly regulate aging kinetics. In yeast normal aging, declining histone was found, while restoration of histone levels promotes lifetime extension.(7) These findings reveal that aging may be slowed without changing genomic sequences, but due to the epigenetic forms. Metabolic modification, partial reprogramming, heterochronic parabiosis, pharmacological delivery, and senescent cell ablation (8), stem cell function restoration, mitochondrial activity rescue, suppression of active transposons, and repression of age-related chronic inflammation, 'resetting' the epigenome might lead to rejuvenation of cells, tissues, and species.(2)

### Epigenetic Regulation of Aging

Epigenetic pathways have been widely recognized to play essential roles in both health and illness, including aging process (9), and now even recognized as indicators of aging. Epigenetic drift due to long-term environmental stress exposure result in disrupted phenotype, gene expression, and even immune system (10). This links how the environmental can alter the epigenome and affect our healthspan.

The exact causation between these epigenetic modifications and the aging process, on the other hand, is still on investigation based on two evidence. First, epigenetic drift or mutations that accumulate with age can lead to genomic instability and changes in gene expression patterns defined by an increase in gene expression noise, both of which have been linked to aging. Second, epigenetic changes that accumulate in one generation may be transmitted down to the next generation, influencing the offspring aging acceleration.(2) Environmental perturbations can cause heritable epigenetic (11), and the children of elderly paternal mice have shorter lives and show aging characteristics earlier than the offspring of young paternal mice (12). The epigenetic bases of aging and rejuvenation can be found in Figure 1.

The epigenome is the cross-point of the genome and environment. Epigenome and transcriptome can be affected by lifestyle and behavioral factors such as prenatal exposures, childhood adversity, diet, physical activity, exercise, stress, exposure to pollutants and toxins, smoking,



**Figure 1. The epigenetic bases of aging and rejuvenation.**(2) (Adapted with permission from Springer Nature).

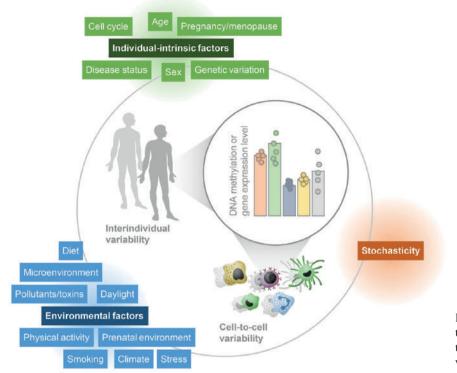


Figure 2. Overview of the different types and sources of epigenetic and transcriptional variability.(19) (Adapted with permission from Wiley Periodicals).

climate, season, daylight, culture, education, socioeconomic variables, result in phenotype (Figure 2). Some changes even can have a long-term influence, and be handed on to future generations.(13-19)

Post-synthesis chemical alteration of three kinds of essential macromolecules: DNA, RNA, and proteins, is required for the regulation and adaptation of practically all biological activities. Methylation, one of the most common modifications, comprises an alkylation process in which a methyl group replaces a hydrogen atom. Methyltransferases act as the "writers" in methylation process assisted by methyl "readers", while demethylases is the "erasers", utilizing S-adenosylmethionine (SAM) as the methyl donor. The fact that their dysregulation is connected to a variety of diseases emphasizes the relevance of the multiple methylation mechanisms.(20-24) DNA methyltransferase 1 (DNMT1), DNMT3A, and DNMT3B are three enzymes that methylate DNA and keep genomic methylation patterns under check. The methylation by adding a methyl group to the 5' position of the cytosine pyrimidine ring mostly in palindromic CpG dinucleotides (5mC). While the first step of active DNA demethylation started by the oxidation of 5-methyl group of 5mC by the ten-eleven translocation (TET) dioxygenases create 5-hydroxymethylcytosine (5hmC), 5-formylcytosine, or 5-carboxylcytosine.(25)

SAM is transformed to S-adenosylhomocysteine (SAH) during the methylation step, which limits methyltransferase

function.(26) As a result, alterations in the cellular SAM-to-SAH ratio can affect methyltransferases. CpG methylation of proto-oncogene promoters can increase cellular levels of SAM and inhibit proto-oncogenes.(27) Different enzymes can methylate different classes of macromolecule.(28) According to the 'heterochromatin loss hypothesis of aging' theory, aging causes cellular malfunction due to reduced heterochromatin and/or incorrect redistribution of heterochromatin-silencing proteins. While the processes causing heterochromatin remodeling are unknown, the interactions between the chromatin machinery and nuclear peripheral proteins such nuclear lamins may be involved (Figure 3).(1) Loss of heterochromatin with age is frequently accompanied with modification of histone methylations associated with 'active' chromatin.(29-31).

The methylation landscapes of DNA and histones alter as a cell matures. Early research claimed that global DNA hypomethylation is a sign of aging; however, more recent research revealed that DNA methylation loss occurs especially in constitutive-heterochromatin repeat regions, whereas promoter CpGs are hypermethylated. (32,33) The genomic transposition of retrotransposable elements is associated with the formation of DNA double-strand breaks.(34-36) This can induce genome instability and cause disease.(37) A stable epigenome, on the other hand, has been linked to lifespan and cancer resistance.(38)

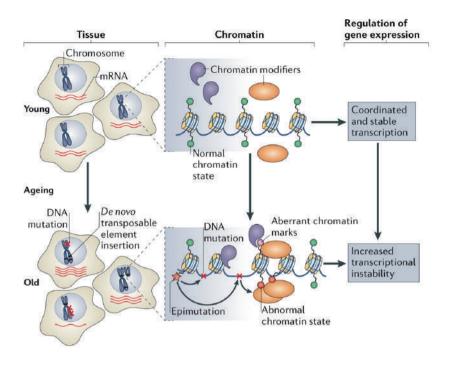


Figure 3. A model for the possible crosstalk between chromatin changes and transcription and genomic instability during aging.(1) (Adapted with permission from Springer Nature).

In aging blood and tissues, there has been an increase in locus-specific DNA hypermethylation (39,40), produces a transcriptional pattern that is age-related (40). In model organisms, a variety of DNA methylation 'clocks' have been identified (41), may be used to estimate age (42), predict mortality (43) and lifespan (44,45).

Two common chromatin alteration themes in aging influenced by environmental cues, nutritional signaling, and metabolic status include global upregulation of activating marks and downregulation of repressive marks, and gene-specific changes in chromatin which influence essential longevity genes expression. Significant epigenetic alterations that may be linked to aging identification bring up big challenges and opportunity to improve our agingrelated metabolism, such as which chromatin modification or enzymes can induce which pathway, especially related to the mitochondrial aging.(46)

### The Metabolic Regulation of Aging

A variety of metabolic changes seen in aging, including regulation of mitochondrial activity, a decrease in insulin sensitivity, and changes in substrate use.(47) Dissecting how metabolic pathways are influenced by environmental or genetic alterations to regulate longevity may provide greater information. In mice and rats, lifelong caloric restriction (CR) can lengthen lifespan by up to 50% compared to ad libitum-fed control animals in some circumstances (48), led more studies to figure out how reducing food intake leads to longer life, and how the genetic and physiological processes mediate this effect (49). The mechanistic target of rapamycin (mTOR) is one clear connection. mTOR is a serine-threonine kinase that acts as an intracellular energy sensor and is found in two different protein complexes in mammalian cells: mTORC1 and mTORC2. mTOR is activated in response to growth factor stimulation or an increase in intracellular amino acid levels, and it then governs a slew of downstream events that control development and metabolism.(50)

Another set of possible mediators relating nutrition availability to lifespan is the sirtuin family of proteins. Both histone and nonhistone protein deacetylation of nicotinamide adenine dinucleotide (NAD), and the acyl moieties removal from longer carbon lengths such as malonyl and succinyl groups, from several enzymes, all performed by sirtuin family members.(51) The relationship suggests that this protein family may be at the intersection of metabolism and lifespan, as answering how CR can increase the lifespan in yeast (52) and flies (53), although some studies questioned it (55).

Another critical linkage between nutrition and lifespan are the metabolic sensor adenosine monophosphate (AMP)activated protein kinase (AMPK), and autophagy process, which is the important lysosomal-dependent recycling process. When intracellular energy supplies (ATP) are exhausted, AMPK is activated and lower-energy adenine nucleotides (AMP and ADP) rise to compensate and regulates a number of metabolic pathways that, in general, enhance energy supply while decreasing demand.(54) The AMPK capacity to regulate autophagy later recognized as a key modulator of organismal aging.(55) Intriguingly, these effects can act in a non-cell-autonomous manner in the same way as mTOR signaling does.

A mutation in a receptor of *C. Elegans* receptor that is similar to the mammalian insulin/Insulin-like Growth Factor 1 (IGF-1) receptor double its lifespan compare to the wildtype, and this suggested a strong link between insulin/IGF-1 signaling and lifespan.(56,57) Surprisingly, genetic studies of long-lived humans have frequently linked mutations in the IGF-1 receptor or FOXO transcription factors to a longer lifespan and better health.(58,59) Finally, the importance of the somatotrophic axis in coordinating growth, metabolism, and lifespan is highlighted by secreted substances such as insulin/IGF-1, or, as noted in the case of dwarf mice, secretion of factors such as growth hormone. (60) Thus, neuroendocrine modulation is vital for energy homeostasis and aging.(61,62)

Bulk histone acetylation altered patterns are frequently seen in aging tissues and have been linked to agerelated diseases including cancer and dementia (1,20,63), suggesting a large-scale chromatin modifications (64). Class III HDACs (sirtuins) have previously been demonstrated to mediate the connection of histone acetylation to general metabolism (65), and that CR-induced improvements in healthy longevity via overexpression of the Sir2 deacetylase (66). Resveratrol from wine knows to boost the activity of AMPK, SIRT1, and peroxisome proliferator-activated receptor gamma coactivator  $1-\alpha$  (PGC- $1\alpha$ ), demonstrated to have a favorable metabolic influence on obese adults.(67) Somehow, different results were observed on C. elegans and D. melanogaster about how sirtuins involved in aging process due to strict control of the genetic background of the animals.(68,69)

Acetyl-CoA, amajor molecule produced predominantly in mitochondria, is used by KATs.(70) Acetyl-CoA that has not been metabolized by the tricarboxylic acid (TCA) cycle can be transferred to the cytosol by being converted to citrate, which is then cleaved back into oxaloacetate and acetyl-CoA by ATP citrate lyase (ACLY/ATPCL) in the cytosol.(71) Because acetyl-CoA may readily transport into the nucleus, reduction of ACLY/ATPCL causes reduced histone acetylation levels in animals (71), tying metabolism and histone modification together (72-74).

Flies with decreased ATPCL activity had longer lifespan, indicating that increased histone acetylation is linked to the aging process.(75) Another study found that increasing pyruvate carboxylase activity is connected to age-related memory impairment in Drosophila, links acetyl-CoA levels to organismal aging.(64) The discovery of major metabolic pathways connected to longevity has initiated interest to pharmacologically targeting these parameters with hopes of slowing aging and alleviating age-related disorders. The natural plant polyphenol resveratrol was the first chemical screen found as one such sirtuin activator. (76) The question of whether resveratrol works as a direct or indirect sirtuin activator however is still controversial. (77) Despite the lack of understanding of the mechanism, resveratrol has been shown to have positive cardiovascular and metabolic benefits in animal models (78,79), nonhuman primates (80,81) and human subjects (67).

Low-carbohydrate diets (LCDs) shift the metabolic away from carbohydrate oxidation toward fatty acid oxidation. The ketogenic diet (KD), the most extreme LCD, has been proven to create an anti-inflammatory metabolic state and boost ketone body levels in rats, similar to key aspects of CR.(82) KD significantly increased median lifespan and survival compared to controls. Only those mice with KD showed signs of physiological function preservation as they grew older. In a tissue-dependent way, the KD raised protein acetylation levels and controlled mTORC1 signaling. In this study, mice with a KD have a longer lifespan and are healthier.(83)

Another developing strategy is to increase intracellular NAD<sup>+</sup> metabolite levels to combat aging and age-related disease. In both model species and humans, biochemical levels of tissue NAD<sup>+</sup> appear to decrease with age.(84) Although the exact cause of this age-related loss is unknown, a decrease in NAD<sup>+</sup> metabolite levels appear to diminish the sirtuin function, as these proteins are mostly NAD<sup>+</sup>-dependent enzymes. NAD supplementation stimulates the UPR system, improves metabolic indices, and, most importantly, increases longevity.(84-87) Other research shows that restoring 'youthful' NAD<sup>+</sup> levels may help to delay the onset of age-related illnesses.(88)

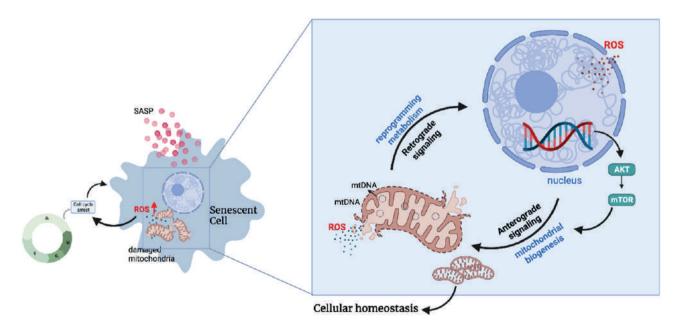
### Mitochondria: Masters of Epigenetics

Aging has been linked to a general decline of mitochondrial function.(46,89-91) A genetic mutation in the mitochondrial DNA polymerase induce accelerated aging in mice.(92) The altered serum metabolome of mice with an accelerated aging phenotype, including lower citrate levels, a sign of mitochondrial activity, supports the functional relevance of metabolic decline throughout aging.(93) Muscle fibers taken from aged adult individuals had lower oxygen consumption rates, implying that this tissue has lower mitochondrial activity.(94) Somehow, lowering metabolic activity by calorie restriction or inhibiting the activities of important metabolic regulators like IGF-1, mTOR, and AMPK has been linked to a longer lifespan.(95) Contradictively, enhanced mitochondrial activity at earlier stages of aging (midlife) associated with a shorter lifespan.(75,96) This might explain why, in midlife, lowering calorie intake and hence decreasing metabolic activity can delay age-related metabolic gain.(64)

As mentioned before, nuclear-encoded mitochondrial proteins have been implicated in the organisms' control of longevity.(97,98) Most of these genetic manipulations affected the expression of electron transport chain components, impairing mitochondrial function while also extending longevity. Human aging is often associated with mitochondrial function loss (99). Studies have recently discovered that mitochondrial stress exposure to *C. elegans* larvae induce a particular and long-lasting epigenetic response which protects the worm as they adult and increases longevity.(100-102)

Mitochondrial function depends on the coordination of nuclear and mitochondrial genomes, the transcription and translation of both genomes are coregulated. For example, the oxidative phosphorylation (OXPHOS) complexes were made up of protein components encoded in both genomes and hence must be built in a stoichiometric way.(103,104) The nucleus regulates mitochondrial function through a process known as "anterograde regulation" (signaling from the nucleus to the mitochondria), which stimulates biogenesis and regulates mitochondrial activity to fulfill cellular demands (Figure 4). Mitochondria, on the other hand, can affect nuclear gene expression via a "retrograde reaction" (signaling from mitochondria to the nucleus), changing cellular function via reprogramming metabolism. Mitonuclear communication, or bidirectional regulation, is a strong network that maintains cellular homeostasis while also controlling response to a range of stimuli.(105) In addition, environmental changes and cellular metabolism can alter the epigenetic regulation of mitochondrial function and the mechanisms that affect both nuclear and mitochondrial epigenome.(106)

Environmental variation including nutritional and stress, are translated into gene expression modifications through epigenome alteration. Epigenetic modifications control the expression of certain genes without affecting the DNA sequence, but only modification in the chromatin structure either in open or condensed state.(107) Posttranslational histone modifications and DNA methylation are the key epigenetic processes implicated in this shift. Histone acetyltransferases (HATs) and histone deacetylases (HDACs) are enzymes to modulate chromatin packing to control gene expression. HATs transfer an acetyl group from acetyl-CoA to the e-amino group of lysine residues, relaxing the chromatin and boosting accessibility to transcriptional activators by hiding the positive charge of the lysine.



**Figure 4. Throughout life, cells were exposed to stress, such as senescence.** Senescent cells release a lot of reactive oxygen species (ROS) and pro-inflammatory senescence associated secretory phenotype (SASP). ROS can damage mitochondria and nuclear DNA which enforce cell cycle arrest, and further induce mitochondrial biogenesis. (Created by author in biorender.com).

The function of HATs is dependent on acetyl-CoA levels. Glycolysis and pyruvate, fatty acid oxidation, acetate, ketone bodies, and amino acids are all examples of metabolic processes and intermediates that can produce acetyl-CoA. (108,109) Citrate synthase in mitochondria converts acetyl-CoA and oxaloacetate to citrate in the TCA cycle. Citrate is then exported to the cytosol, where it is transformed to acetyl-CoA and oxaloacetate in an inverse process by ATPcitrate lyase (ACLY).(71) Acetyl-CoA levels in cells are highly correlated with energy generation: The acetyl-CoA levels rise as the energy production is high, to increase histone acetylation and gene expression. When cellular metabolism is sluggish, decreasing acetyl-CoA levels reduce histone acetylation and inhibiting gene expression through chromatin condensation.(108) Mitochondrial activity, namely the TCA cycle, is a key regulator of histone acetylation and, as a result, global gene expression.

The most common methylations are found on histone 3 (H3) and 4 (H4) lysine residues. Histone methylation, unlike acetylation, works by changing a single lysine (K) or arginine (R) amino acid residue on a histone protein to activate or inhibit transcription. Lysine can be mono-, di-, or trimethylated during the methylation process, and arginine usually be mono- or dimethylated.(110,111)

Histone methyltransferases (HMTs) are enzymes that methylate histones, while histone demethylases (HDMs) removemethylmarks.SET domain lysine methyltransferases, non-SET domain lysine methyltransferases, and arginine methyltransferases are the three types of HMTs. SAM is used as a precursor in methyl group transfer in all of the aforementioned classes.(118,120) As a result, mitochondrial activity can affect SAM production and hence modulate histone methylation. Although SAM is produced in the cytosol through the methionine-homocysteine cycle, this cycle is dependent on the folate cycle and ATP, both of which are dependent on mitochondrial metabolism.(118,121)

Stress in mitochondria can alters the epigenome by causing substantial chromatin remodeling, as evidenced by alterations in H3K9me2 methylation patterns mediated by HMT MET-2 and nuclear cofactor LIN-65. H3K9me2 methylation increases chromatin compaction, so the transcriptional apparatus is unable to initiate transcription. During mitochondrial stress, ATFS-1 and DVE-1, the transcriptional regulators open and bind loci carrying UPRmt genes and downregulation of global transcription. These variables cause the UPRmt to be activated in order to maintain mitochondrial proteostasis and improve mitochondrial lifespan. Importantly, LIN-65 and DVE-1 nuclear translocation are interdependent, indicating that mitochondrial stress-induced chromatin reconfiguration and UPRmt transcriptional activation are highly controlled and interrelated processes.(101) Overall, mitochondrial function affects longevity by regulating the epigenome, linking two important characteristics of aging, mitochondrial failure and epigenetic changes (112), to lifetime determination.

## Regulation of Chromatin and Gene Expression by Metabolic Enzymes and Metabolites

Environmental changes can be sensed by cells and modulate particular epigenome conversion through a range of signaling components such as histone- and DNAmodifying enzymatic activity.(113) Cell homeostasis, growth, proliferation, migration, differentiation, and death, need cell metabolism regulation to support all critical cellular functions. Cell metabolism started at the gene expression level to adjust the cells' extracellular and intracellular signals due to environmental change. In turn, cell metabolism can alter chromatin changes and hence chromatin functions, either directly or indirectly, through the activity of metabolic enzymes and metabolites. Hence, DNA-based process and cell metabolism are interlinked as a respond to the environment, and any disruption to this process can induce disease development.(114)

Enzymes that alter DNA or histones via cofactors are involved in chromatin control. These enzymes either add small chemical units (PTMs) or modify the location or content of the nucleosome (*i.e.*, of histone variants). This regulation is thought to be partially dependent on the changing amounts of cellular metabolites that function as cofactors for enzymes like acetyl-CoA, or S-adenosyl methionine utilized by methyltransferases, flavin was utilized by demethylases, kinases utilize ATP as acetyl, methyl, or phospho groups donors, while deacetylases utilize nicotinamide adenine dinucleotide (NAD).(113)

Although it is plausible that a number of metabolites are involved in various elements of epigenetic regulation on a conceptual level.(74,115) The hypothesis is that metabolite levels may fluctuate in response to diverse physiological stressors, therefore affecting enzymes involved in chromatin remodeling. The subcellular concentration of certain metabolites may influence the activation or inhibition of enzyme activity at the local level. Thus, the presence of "niches" of chromatin-associated metabolites has been suggested, which might explain why otherwise indistinguishable histones and DNA are modified locus-specifically.(74) The metabolites may be related to enzyme activity involved in chromatin remodeling and DNA methylation on a functional level.

A metabolite is an intermediate or end product of metabolism. It serves as cofactors and regulators for enzymes, including those in chromatin remodeling, thus metabolites have important roles in methylation, hydroxylation, acetylation, crotonylation, 2-hydroxyisobutyrylation, O-GlcNAcylation, and poly (ADP) ribosylation (PARylation). Suggested that a diverse array of metabolites can regulate methyltransferase and demethylase activities to dynamically control histone and DNA methylation. Cell signaling regulates the production of these metabolites, and the enzymes responsible for their production can be mutated, thus determining biological outcomes.(114)

It was recently discovered that several metabolic enzymes, which are normally located in the cytoplasm or mitochondria, can also be present in the nucleus. These enzymes include glycolytic enzymes pyruvate kinase M2 isoform (PKM2), glyceraldehyde-3-phosphate dehydrogenase (GAPDH); TCA (also known as MAT2A), 6-phosphofructo-2-kinase/fructose-2,6-bisphosphatase 4 (PFKFB4), and fructose-1,6-bisphosphatase 1 (FBP1). These enzymes can provide metabolites to chromatin-modifying enzymes or transcription regulators in the nucleus, therefore regulating chromatin shape and gene expression. They may also play other roles in controlling chromatin and gene expression that are unrelated to metabolite synthesis, such as direct alteration of histones and transcription regulators (Figure 5).(114)

Numerous types of enzymes can have their activity regulated by variations in metabolite availability or by the usage of particular metabolites as coenzymes.(113) The class III HDACs are a group of mammalian proteins that were initially discovered due to their resemblance to the yeast gene Sir2. There are seven members of sirtuin family in mammals in mitochondria (Sirt3, Sirt4, and Sirt5), nucleus (Sirt1, Sirt6, and Sirt7), and in cellular compartments (Sirt1, Sirt6, and Sirt7) (116). Sir2 was first discovered as a NAD<sup>+</sup>-dependent deacetylase and associated with longevity regulation (117–119), although the effects in various organisms is still debated (68).

A key concept rooting chromatin's involvement in regulating cell physiology and homeostasis is that nuclear activity needs coordination and reaction to the overall metabolic state of the cell. However, nuclear activities involving DNA-associated chromatin transactions distinct by location and regulation from those involving cytoplasmbased intermediate metabolic enzymes. A novel hypothesis to explain this dichotomy is that changes in metabolism may

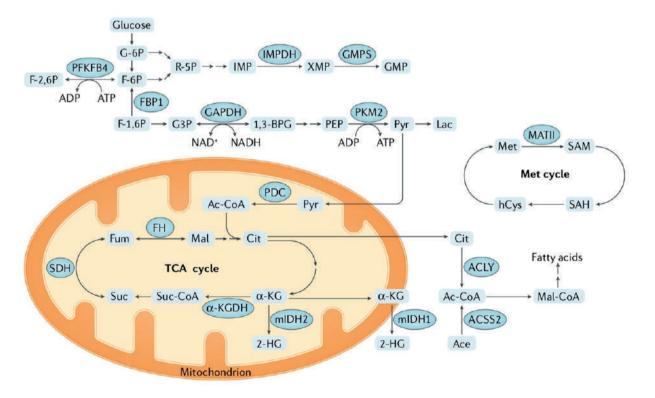


Figure 5. Core metabolic functions of metabolic enzymes that also function in epigenetic modifications.(114) (Adapted with permission from Springer Nature).

directly affect the amounts of acetyl-CoA, SAM, NAD, and FAD in the nucleus, which may subsequently affect DNA and histone modifications, and also transcription.(74)

Cell metabolism and gene expression should be tightly coupled and mutually regulated to perform essential biological function. Many factors can change the enzymatic activity and location at once such as nutrients, growth factors, and cytokines, and also stressors, both physiological or pathological. These alterations not only respond directly to cellular metabolic demands, but also govern cellular homeostasis, growth, proliferation, survival, and a variety of other activities via gene reprogramming. Enzymes' metabolites including acyl-CoA, SAM, fumarate, succinate, 2-HG, NAD<sup>+</sup>, NAM, and ketone bodies can function as direct substrates for DNA, histone nodification, or regulate the DNA activity. The enzymes can function for non-metabolic activities such as protein kinases for direct phosphorylate histones or being the transcription factors and co-activators for gene transcription machinery. When metabolic enzymes located in the nucleus, they can merge with transcription factors, histone writers, readers, and also erasers.(114) It is critical whether epigenetic modifications contribute to cellular aging, given the longestablished links between life duration, chromatin enzymes, and metabolic changes. The finding that yeast Sir2, a critical protein involved in heterochromatic silencing and lifespan promotion, is a NAD<sup>+</sup>-dependent HDAC has raised the possibility that chromatin plays a role in aging. (52, 117-120)

### Autophagy as Promoter of Longevity

Autophagy is the body's way of cleaning out damaged cells by catabolic process, in order to regenerate newer, healthier cells, and maintain cellular homeostasis. Autophagy facilitates lysosomes to do the break down and recycling of intracellular macromolecules and organelles. It was initially identified as a survival strategy in yeasts subjected to nutrient deprivation, a circumstance that accelerates the activity much above basal levels.(121) Numerous lines of evidence suggest that aging has an effect on the autophagy process, where autophagy-reporter analyses and gene expression studies in various species showed that autophagy has declined over time, and genetic experiments in shortlived animals showed that autophagy modulation can promote lifespan.(121)

Autophagy has four primary implications on agerelated processes. First, by increasing the cytoplasmic turnover, autophagy can neutralize the damaged organelles and macromolecular complexes. Second, when the homeostasis enhanced, genomic and epigenomic were stabilized, and the sensitivity of nutrition sensing increased. Third, autophagy significantly improve anti-inflammatory and immunological properties, therefore promoting system homeostasis. And fourth, and maybe most importantly, its actions may be connected to circadian clocks. The evidence for a bidirectional relationship between circadian clocks and autophagy-flux is growing.(122)

While the genetic data indicates that bulk autophagy is involved in aging processes, there is also evidence that impairments in the turnover of particular cargoes via selective autophagy may play a significant role in age-related diseases and aging. The accumulation of defective mitochondria is a common feature of aging and a variety of age-related illnesses.(89,91,112,123) Although the fundamental mechanisms causing age-related loss of mitochondrial function remain unknown and may include a variety of processes, a decrease in mitophagy has been hypothesized to play a critical role.(91,123,124) Mitophagy's molecular processes have been explored in depth in mammalian cell culture and genetic studies in model species (125-127), and found to be linked to the pathogenesis of age-related illnesses such as heart disease (128), retinopathy (129), fatty liver disease (130), pulmonary hypertension (131), kidney disease (132), and neurodegenerative disorders.

Numerous studies in varied species indicate that various pro-longevity treatments are related with certain alteration in lipid metabolism.(133) Autophagy's role in intracellular lipid droplet destruction, dubbed lipophagy, has been found in recent years.(134) The first conclusive evidence that lipid droplets can be recycled by autophagy came from experiments in cultured hepatocytes with decreased ATG5 levels.(135,136) The ability of autophagy to control lipid metabolism increases its physiological importance in the cellular energy balance modulation. Alterations in lipophagy may contribute to the aging metabolic syndrome (137), which is characterized by obesity, dysregulated lipoprotein metabolism, inappropriate glucose management, and hypertension. Additionally, lipophagy has been associated with cancer (138) and atherosclerosis (139).

Aggrephagy is the selective type of autophagy process which degrade protein aggregates, or oligomeric versions of proteins. Some protein aggregates in degenerated neurons such as tau,  $\alpha$ -synuclein, and mutant HTT can induce toxicity and neurodegenerative disorders such as Alzheimer disease, Parkinson disease, and Huntington disease.(140-142), so upregulating autophagy for these aggregates by chemical (143), genetic (144) or by hormetic heat shock (145) can improve neurodegeneration symptoms in a variety of animal models, including *C. elegans*, *D. melanogaster*, zebrafish, and mice (146). Indeed, aging is a significant risk factor for the majority of neurodegenerative disorders.

The molecular sensors that initiate autophagy in response to CR also regulate circadian clocks.(122) As a result, autophagy is regularly increased in a circadian clockdependent way, and autophagy induction has a circadian clock-regulating effect.(147) Autophagy, in a variety of forms, has lately emerged as a critical process of stress adaption throughout the eukaryotic kingdom, posing a potential (but complicated) target for the treatment of a variety of human diseases.(148,149) Macroautophagy, in particular (hereinafter referred to as autophagy), destroys potentially hazardous or disposable cytoplasmic components to maintain cellular homeostasis under physiological and stress circumstances.(150) Over the last decade, many studies has suggesting that autophagy and aging are inextricably connected. Numerous ATG genes have been demonstrated to be needed for the extended lifespan generated by conserved longevity paradigms in model organisms ranging from yeast to mice.(121)

### **Epigenetic Regulation of Senescence**

Aging-related cellular damage activates mechanisms that regulate cell proliferation and differentiation, as well as stem cell-mediated regeneration, in order to prevent the irreversible breakdown of cellular and tissue homeostasis. Cellular senescence and regeneration mechanisms are particularly relevant to aging and AADs.(151) Human diploid cells have a finite lifetime after they terminally exit the cell cycle.(152) This process is referred as cellular senescence. Although cellular senescence is considered to contribute to aging, this theory is still disputed. Numerous studies have demonstrated the accumulation of senescent cells in aged tissues *in vivo* (153-155), such as in progeria patients' fibroblasts which had shorter replicative lifespans (156).

Senescence is a cell cycle halt that occurs in reaction to stress and is connected with age-related tissue deterioration *in vivo*.(157) More than 30% chromatin rearrangement were altered in senescent cells and develop senescence-associated heterochromatin foci (SAHF), a highly condensed chromatin regions.(29) It was associated with heterochromatic histone modifications, heterochromatic proteins, histone variant macroH2A, high-mobility group A (HMGA) proteins, and late replicating regions in the genome.(158,159) These alterations are associated with the downregulation of lamin B1 transcription in senescence.(29,160,161) Autophagic degradation of lamin B1 was linked with destabilization of heterochromatic lamin-associated domains (LADs), and respressive histones.(162)

Senescent cells also known to have a significant abundance of H4K16ac (acetylation of lysine 16 on H4) near active gene promoter elements where it overlaps with HIRA and newly synthesized H3.3 peaks.(163) HIRA, a non-replicating histone chaperone that deposits variant histone H3.3 and H4, is essential for H4K16ac steadystate stability. These changes imply that the senescent state is associated with a dynamic and unbalanced chromatin environment that is qualitatively distinct from the proliferative state.(3)

Numerous processes promoting senescence have been found with the majority of which involve the buildup of damage to DNA and other cell macromolecules. (164-170) The chromatin shape is important for cell and organism life span control.(171-173) Aging is associated with heterochromatin reduction, as shown in senescent cells that expression of histone-encoding genes and the number of repressive heterochromatin marks, such as DNA methylation and histone methylation, including H3K9me3, H3K27me3, and H4K20me3 marks were reduced.(9)

Senescence is irreversible, caused by prolonged activation of the tumor suppressor RB–p16 and p53–p21 pathways.(174,175) This pathway activation can initiate the senescence-associated secretory phenotype (SASP) mediated by transforming growth factor  $\beta$  (TGF $\beta$ ) and nuclear factor- $\kappa\beta$  (NF- $\kappa$ B). Long term SASP activation can be harmful since it impairs intercellular communication between immune-responsive to senescent cells and, therefore preventing immune system-mediated to eliminate those senescent cells.

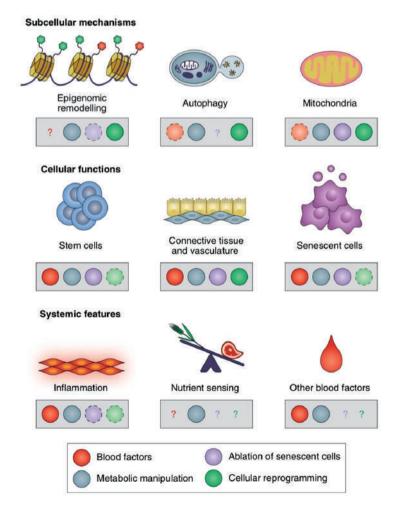
Failure to eliminate senescent cells during aging might exacerbate physiological deterioration by promoting cellular senescence in adjacent cells via gap junction, mediated cell-cell interactions.(112) Artificially induced senescence clearance in aged mice with mitotic checkpoint protein BubR1 deficiency increase its lifespan, as well as inhibits atherogenesis in low-density lipoprotein receptor-deficient mice, and preserves renal and cardiac function in wild-type mice.(179-181) The SASP may contribute to local and systemic malfunction and illness, and because senescent cells increase with age, targeting senescent cells serve as potential drug targets for promoting a prolonged, healthy lifespan.(176)

### **Emerging Rejuvenation Strategies**

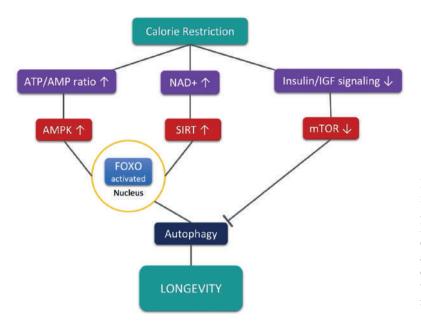
Aging was first considered as irreversible, but recent studies showed that it was not. Indeed, suppression of high-nutrient-sensing pathways (e.g., the IGF and mTOR pathways) and activation of low-nutrient-sensing proteins (e.g., AMPK and sirtuins) both increase longevity in many model animals.(177,178) Even when administered late in life, nutritional treatments such as dietary restriction and pharmaceutical interventions such as the mTOR inhibitor rapamycin can improve aging.(83,179-183) The question is whether the aging of cells, organs, and organisms can be reversed or 'rejuvenated' rather than just delayed. Numerous rejuvenation methods targeting these characteristics have recently developed, classified as systemic (blood) factors, metabolic alterations, senescent cell ablation, and cellular reprogramming. While various treatments appear to target disparate aspects of aging (95,184-187), then more questions raised, whether they share common mechanisms of action? Is there any cost associated with the rejuvenating effect? Can rejuvenation techniques ultimately be utilized to increase human health and lifespan, as well as to combat age-related diseases?

Figure 6 shows the mechanisms and target cells of the rejuvenation strategies. Restoring a young epigenome by maintaining the stem cell pool may be critical for tissue function and lifespan extension in aged animals. (8,188) Some studies investigate how epigenetic process involved in recovering the youthful function in old stem cells subjected to rejuvenating treatments. The epigenetic rejuvenation extends longevity by correcting other agingrelated characteristics, including as mitochondrial failure, genomic instability induced by retrotransposon activation, and chronic inflammation.

Dysfunctional mitochondria accumulated in aging cells.(91) After CR, partial reprogramming, heterochronic parabiosis, and pharmacological treatment, these mitochondria were restored.(88,189-191) In muscle and white fat tissues, CR promotes mitochondrial biogenesis. (192) Resveratrol supplementation increases the amount of mitochondria in muscle, increases physical activity, and increases the average longevity of mice.(78) Reprogramming mitochondria restores them to a condition similar in



**Figure 6.** Potential common mechanisms and target cells of the rejuvenation strategies.(8) (Adapted with permission from Springer Nature).



**Figure 7. The regulation of autophagy by calorie restriction.** In low energy intake condition, ATP/ AMP ratio and NAD+ level are higher while Insulin/IGF signaling was lower. This activates the energy sensors AMPK and SIRT, but inhibit mTOR activity. AMPK and SIRT upregulate the expression of forkhead box transcription factors (FOXOs) in the nucleus. SIRT1 deacetylates the core autophagy machinery and thereby increases autophagy.

embryonic stem cells.(193) Additionally, heterochronic parabiosis decreases the swelling and vacuolization of mitochondria in skeletal muscle of elderly mice.(194)

Attenuation of oxidative phosphorylation (91), and regulation of mitochondrial unfolded protein response are two mitochondria-dependent strategies for lifespan extension. (189) By providing substrates for epigenetic changes, mitochondria govern lifespan mechanistically, acting as essential platforms for metabolism, epigenetic regulation, and aging.(114) TCA cycle's metabolic intermediates and byproducts function as cofactors and substrates for a variety of epigenetic enzymes.(64,88,91,192,195)

NAD<sup>+</sup>, the cofactor for sirtuins, is another critical molecule that connects epigenetic control to mitochondria. (189) Increased NAD<sup>+</sup> levels increase mitochondrial function, replenish stem cell pools, and prolong the lives of mice.(196,197) Nicotinamide mononucleotide (NMN) plasma levels decline with aging. NAD<sup>+</sup> precursors include nicotinamide riboside and NMN.(198) Supplementing aged mice with NAD<sup>+</sup> precursors slows mitochondrial function decreases, ameliorates muscular, neuronal, and melanocyte stem cell senescence, ameliorates age-related physiological losses, and prolongs longevity.(88,198-204)

The rejuvenation techniques we discussed here have the potential to promote epigenetic reprogramming, implying that the epigenetic program can be experimentally reprogrammed to a younger state throughout later stages of life. Not only does a cell's epigenomic landscape represent its identity, but also its health and biological age.(1,40,42) Senescent cells have a distinct chromatin state (158,205), and their secreted cytokines (for example, IL-6) can cause epigenomic alterations.(206,207) It has been hypothesized that epigenomic modification in cell results in the rejuvenating effect.(208) Dietary treatments such as calorie restriction and dietary restriction-mimicking drugs such as resveratrol combining with NAD<sup>+</sup> (Figure 7) or Acetyl-CoA supplementation have been proved in many studies to increase the autophagy activities and rejuvenate our cells, hence increase the longevity.(1,181,209)

Nutrigenomic and nutrigenetic assessments, together with gut microbiomes and metabolites profiling are what we currently have to construct a personalize lifestyle and determine what kind of diet and supplementation as well as exercise we need (210) since genetic and environmental treatments have been demonstrated to extend life expectancy, by acting primarily through epigenome modification.(211) Aging process is flexible and that old cells, tissues, and organs may be rejuvenated. Additionally, they offer the intriguing prospect of translation for the treatment of human aging and age-related illnesses. Undoubtedly, the future years will bring exciting advances in our continuous efforts to better understand, postpone, and maybe reverse aging.(8)

### Conclusion

Aging process is characterized by tissues and organs functional decline, and the increased risk of aging-associated disorders. Our increasing knowledge about how epigenetic changes along our aging process guide future research on treatments to rejuvenate the epigenome and delay the aging processes. Many treatments were proposed to delay the onset of aging, prevent the aging-associated diseases, and extend the healthspan as well as lifespan. These treatments including metabolic manipulation, partial reprogramming, heterochronic parabiosis, pharmaceutical administration and senescent cell ablation.

## Authors Contribution

AM drafted and wrote the manuscript, NMD edited the manuscript, AW proposed the manuscript topic, supervised, and edited the manuscript.

#### References

- Benayoun BA, Pollina EA, Brunet A. Epigenetic regulation of ageing: linking environmental inputs to genomic stability. Nat Rev Mol Cell Biol. 2015; 16(10): 593–610.
- Zhang W, Qu J, Liu GH, Belmonte JCI. The ageing epigenome and its rejuvenation. Nat Rev Mol Cell Biol. 2020; 21(3): 137–50.
- Sen P, Shah PP, Nativio R, Berger SL. Epigenetic mechanisms of longevity and aging. Cell. 2016; 166(4): 822–39.
- Zhang W, Song M, Qu J, Liu GH. Epigenetic modifications in cardiovascular aging and diseases. Circ Res. 2018; 123(7): 773–86.
- Trojer P, Reinberg D. Facultative heterochromatin: is there a distinctive molecular signature? Mol Cell. 2007; 28(1): 1–13. doi: 10.1016/j.molcel.2007.09.011.
- Allshire RC, Madhani HD. Ten principles of heterochromatin formation and function. Nat Rev Mol Cell Biol. 2018; 19(4): 229– 44.
- Feser J, Truong D, Das C, Carson JJ, Kieft J, Harkness T, *et al.* Elevated histone expression promotes life span extension. Mol Cell. 2010; 39(5): 724–35.
- Mahmoudi S, Xu L, Brunet A. Turning back time with emerging rejuvenation strategies. Nat Cell Biol. 2019; 21(1): 32–43.
- Sidler C, Kovalchuk O, Kovalchuk I. Epigenetic regulation of cellular senescence and aging. Front Genet. 2017; 8: 138. doi: 10.3389/ fgene.2017.00138.
- Fraga MF, Esteller M. Epigenetics and aging: the targets and the marks. Trends Genet. 2007; 23(8): 413–8.
- Pal S, Tyler JK. Epigenetics and aging. Sci Adv. 2016; 2(7): e1600584. doi: 10.1126/sciadv.1600584.
- Xie K, Ryan DP, Pearson BL, Henzel KS, Neff F, Vidal RO, *et al.* Epigenetic alterations in longevity regulators, reduced life span, and exacerbated aging-related pathology in old father offspring mice. PNAS. 2018; 115(10): E2348–57.
- Feil R, Fraga MF. Epigenetics and the environment: emerging patterns and implications. Nat Rev Genet. 2012; 13(2): 97–109.
- Li CCY, Cropley JE, Cowley MJ, Preiss T, Martin DIK, Suter CM. A sustained dietary change increases epigenetic variation in isogenic mice. PLOS Genetics. 2011; 7(4): e1001380. doi: 10.1371/journal. pgen.1001380.
- Brodin P, Davis MM. Human immune system variation. Nat Rev Immunol. 2017; 17(1): 21–9.
- Jones MJ, Goodman SJ, Kobor MS. DNA methylation and healthy human aging. Aging Cell. 2015; 14(6): 924–32.

- Voisin S, Eynon N, Yan X, Bishop DJ. Exercise training and DNA methylation in humans. Acta Physiol (Oxf). 2015; 213(1): 39–59.
- Azzi A, Dallmann R, Casserly A, Rehrauer H, Patrignani A, Maier B, *et al.* Circadian behavior is light-reprogrammed by plastic DNA methylation. Nat Neurosci. 2014; 17(3): 377–82.
- Ecker S, Pancaldi V, Valencia A, Beck S, Paul DS. Epigenetic and transcriptional variability shape phenotypic plasticity. BioEssays. 2018; 40(2): 1700148. doi: 10.1002/bies.201700148.
- Dawson MA, Kouzarides T. Cancer epigenetics: from mechanism to therapy. Cell. 2012; 150(1): 12–27.
- 21. Jin B, Li Y, Robertson KD. DNA methylation: superior or subordinate in the epigenetic hierarchy? Genes Cancer. 2011; 2(6): 607–17.
- Luo C, Hajkova P, Ecker JR. Dynamic DNA methylation: In the right place at the right time. Science. 2018; 361(6409): 1336–40.
- Stojković V, Fujimori DG. Mutations in RNA methylating enzymes in disease. Curr Opin Chem Biol. 2017; 41: 20–7.
- Xie P, Zang LQ, Li XK, Shu Q. An epigenetic view of developmental diseases: new targets, new therapies. World J Pediatr. 2016; 12(3): 291–7.
- Kohli RM, Zhang Y. TET enzymes, TDG and the dynamics of DNA demethylation. Nature. 2013; 502(7472): 472–9.
- Deguchi T, Barchas J. Inhibition of transmethylations of biogenic amines by S-adenosylhomocysteine. Enhancement of transmethylation by adenosylhomocysteinase. J Biol Chem. 1971; 246(10): 3175–81.
- Wang Y, Sun Z, Szyf M. S-adenosyl-methionine (SAM) alters the transcriptome and methylome and specifically blocks growth and invasiveness of liver cancer cells. Oncotarget. 2017; 8(67): 111866– 81.
- Michalak EM, Burr ML, Bannister AJ, Dawson MA. The roles of DNA, RNA and histone methylation in ageing and cancer. Nat Rev Mol Cell Biol. 2019; 20(10): 573–89.
- Shah PP, Donahue G, Otte GL, Capell BC, Nelson DM, Cao K, *et al.* Lamin B1 depletion in senescent cells triggers large-scale changes in gene expression and the chromatin landscape. Genes Dev. 2013; 27(16): 1787–99.
- Ni Z, Ebata A, Alipanahiramandi E, Lee SS. Two SET domain containing genes link epigenetic changes and aging in Caenorhabditis elegans. Aging Cell. 2012; 11(2): 315–25.
- Pu M, Ni Z, Wang M, Wang X, Wood JG, Helfand SL, et al. Trimethylation of Lys36 on H3 restricts gene expression change during aging and impacts life span. Genes Dev. 2015; 29(7): 718– 31.
- Wilson ID. Drugs, bugs, and personalized medicine: pharmacometabonomics enters the ring. Proc Natl Acad Sci USA. 2009; 106(34): 14187–8.
- Unnikrishnan A, Hadad N, Masser DR, Jackson J, Freeman WM, Richardson A. Revisiting the genomic hypomethylation hypothesis of aging. Ann NY Acad Sci. 2018; 1418(1): 69–79.
- De Cecco M, Criscione SW, Peterson AL, Neretti N, Sedivy JM, Kreiling JA. Transposable elements become active and mobile in the genomes of aging mammalian somatic tissues. Aging (Albany NY). 2013; 5(12): 867–83.
- Belgnaoui SM, Gosden RG, Semmes OJ, Haoudi A. Human LINE-1 retrotransposon induces DNA damage and apoptosis in cancer cells. Cancer Cell Int. 2006; 6(1): 13. doi: 10.1186/1475-2867-6-13.
- Gasior SL, Wakeman TP, Xu B, Deininger PL. The human LINE-1 retrotransposon creates DNA double-strand breaks. J Mol Biol. 2006; 357(5): 1383–93.
- Belancio VP, Deininger PL, Roy-Engel AM. LINE dancing in the human genome: transposable elements and disease. Genome Med. 2009; 1(10): 97. doi: 10.1186/gm97.

- Tan L, Ke Z, Tombline G, Macoretta N, Hayes K, Tian X, *et al.* Naked mole rat cells have a stable epigenome that resists iPSC reprogramming. Stem Cell Rep. 2017; 9(5): 1721–34.
- Beerman I, Bock C, Garrison BS, Smith ZD, Gu H, Meissner A, *et al.* Proliferation-dependent alterations of the DNA methylation landscape underlie hematopoietic stem cell aging. Cell Stem Cell. 2013; 12(4): 413–25.
- Hannum G, Guinney J, Zhao L, Zhang L, Hughes G, Sadda S, *et al.* Genome-wide methylation profiles reveal quantitative views of human aging rates. Mol Cell. 2013; 49(2): 359–67.
- Field AE, Robertson NA, Wang T, Havas A, Ideker T, Adams PD. DNA methylation clocks in aging: categories, causes, and consequences. Mol Cell. 2018; 71(6): 882–95.
- Horvath S, Raj K. DNA methylation-based biomarkers and the epigenetic clock theory of ageing. Nat Rev Genet. 2018; 19(6): 371–84.
- Marioni RE, Shah S, McRae AF, Chen BH, Colicino E, Harris SE, et al. DNA methylation age of blood predicts all-cause mortality in later life. Genome Biol. 2015; 16(1): 25. doi: 10.1186/s13059-015-0584-6.
- Steensma DP, Bejar R, Jaiswal S, Lindsley RC, Sekeres MA, Hasserjian RP, *et al.* Clonal hematopoiesis of indeterminate potential and its distinction from myelodysplastic syndromes. Blood. 2015; 126(1): 9–16.
- Chen BH, Marioni RE, Colicino E, Peters MJ, Ward-Caviness CK, Tsai PC, et al. DNA methylation-based measures of biological age: meta-analysis predicting time to death. Aging (Albany NY). 2016; 8(9): 1844–65.
- Bratic A, Larsson NG. The role of mitochondria in aging. J Clin Invest. 2013; 123(3): 951–7.
- Riera CE, Dillin A. Tipping the metabolic scales towards increased longevity in mammals. Nat Cell Biol. 2015; 17(3): 196–203.
- Speakman JR, Mitchell SE. Caloric restriction. Mol Aspects Med. 2011; 32(3): 159–221.
- Finkel T. The metabolic regulation of aging. Nat Med. 2015; 21(12): 1416–23.
- Albert V, Hall MN. mTOR signaling in cellular and organismal energetics. Curr Opin Cell Biol. 2015; 33: 55–66.
- Hirschey MD, Zhao Y. Metabolic regulation by lysine malonylation, succinylation, and glutarylation. Mol Cell Proteom. 2015; 14(9): 2308–15.
- Lin SJ, Defossez PA, Guarente L. Requirement of NAD and SIR2 for life-span extension by calorie restriction in Saccharomyces cerevisiae. Science. 2000; 289(5487): 2126–8.
- Rogina B, Helfand SL. Sir2 mediates longevity in the fly through a pathway related to calorie restriction. PNAS. 2004; 101(45): 15998–6003.
- Hardie DG. AMPK: positive and negative regulation, and its role in whole-body energy homeostasis. Curr Opin Cell Biol. 2015; 33: 1–7. doi: 10.1016/j.ceb.2014.09.004.
- Ulgherait M, Rana A, Rera M, Graniel J, Walker DW. AMPK modulates tissue and organismal aging in a non-cell-autonomous manner. Cell Rep. 2014; 8(6): 1767–80.
- Kenyon C, Chang J, Gensch E, Rudner A, Tabtiang R. A C. elegans mutant that lives twice as long as wild type. Nature. 1993; 366(6454): 461–4.
- Kimura KD, Tissenbaum HA, Liu Y, Ruvkun G. daf-2, an insulin receptor-like gene that regulates longevity and diapause in Caenorhabditis elegans. Science. 1997; 277(5328): 942–6.
- Suh Y, Atzmon G, Cho M-O, Hwang D, Liu B, Leahy DJ, et al. Functionally significant insulin-like growth factor I receptor mutations in centenarians. Proc Natl Acad Sci USA. 2008; 105(9):

3438-42.

- Wheeler HE, Kim SK. Genetics and genomics of human ageing. Philos Trans R Soc Lond B Biol Sci. 2011; 366(1561): 43–50.
- Brown-Borg HM. The somatotropic axis and longevity in mice. Am J Physiol Endocrinol Metab. 2015; 309(6): E503–10.
- Satoh A, Brace CS, Rensing N, Clifton P, Wozniak DF, Herzog ED, et al. Sirt1 extends life span and delays aging in mice through the regulation of Nk2 homeobox 1 in the DMH and LH. Cell Metab. 2013; 18(3): 416–30.
- Zhang G, Li J, Purkayastha S, Tang Y, Zhang H, Yin Y, *et al.* Hypothalamic programming of systemic ageing involving IKK-β, NF-κB and GnRH. Nature. 2013; 497(7448): 211–6.
- Gräff J, Tsai LH. Histone acetylation: molecular mnemonics on the chromatin. Nat Rev Neurosci. 2013; 14(2): 97–111.
- Peleg S, Feller C, Ladurner AG, Imhof A. The Metabolic impact on histone acetylation and transcription in ageing. Trends Biochem Sci. 2016; 41(8): 700–11.
- Verdin E. NAD+ in aging, metabolism, and neurodegeneration. Science. 2015; 350(6265): 1208–13.
- Guarente L. Sir2 links chromatin silencing, metabolism, and aging. Genes Dev. 2000; 14(9): 1021–6.
- 67. Timmers S, Konings E, Bilet L, Houtkooper RH, van de Weijer T, Goossens GH, *et al.* Calorie restriction-like effects of 30 days of resveratrol supplementation on energy metabolism and metabolic profile in obese humans. Cell Metab. 2011; 14(5): 612–22.
- Burnett C, Valentini S, Cabreiro F, Goss M, Somogyvári M, Piper MD, *et al.* Absence of effects of Sir2 overexpression on lifespan in C. elegans and Drosophila. Nature. 2011; 477(7365): 482–5.
- Partridge L, Gems D. Benchmarks for ageing studies. Nature. 2007; 450(7167): 165–7.
- Fan J, Krautkramer KA, Feldman JL, Denu JM. Metabolic regulation of histone post-translational modifications. ACS Chem Biol. 2015; 10(1): 95–108.
- Wellen KE, Hatzivassiliou G, Sachdeva UM, Bui TV, Cross JR, Thompson CB. ATP-citrate lyase links cellular metabolism to histone acetylation. Science. 2009; 324(5930): 1076–80.
- Everett JR. Pharmacometabonomics in humans: a new tool for personalized medicine. Pharmacogenomics. 2015; 16(7): 737–54.
- Sutendra G, Kinnaird A, Dromparis P, Paulin R, Stenson TH, Haromy A, *et al.* A nuclear pyruvate dehydrogenase complex is important for the generation of acetyl-CoA and histone acetylation. Cell. 2014; 158(1): 84–97.
- Katada S, Imhof A, Sassone-Corsi P. Connecting threads: epigenetics and metabolism. Cell. 2012; 148(1): 24–8.
- Peleg S, Feller C, Forne I, Schiller E, Sévin DC, Schauer T, *et al.* Life span extension by targeting a link between metabolism and histone acetylation in Drosophila. EMBO Rep. 2016; 17(3): 455–69.
- Howitz KT, Bitterman KJ, Cohen HY, Lamming DW, Lavu S, Wood JG, *et al.* Small molecule activators of sirtuins extend Saccharomyces cerevisiae lifespan. Nature. 2003; 425(6954): 191– 6.
- Dang W. The controversial world of sirtuins. Drug Discov Today Technol. 2014; 12: e9–17.
- Lagouge M, Argmann C, Gerhart-Hines Z, Meziane H, Lerin C, Daussin F, *et al.* Resveratrol improves mitochondrial function and protects against metabolic disease by activating SIRT1 and PGC-1α. Cell. 2006; 127(6): 1109–22.
- Pearson KJ, Baur JA, Lewis KN, Peshkin L, Price NL, Labinskyy N, *et al.* Resveratrol delays age-related deterioration and mimics transcriptional aspects of dietary restriction without extending lifespan. Cell Metab. 2008; 8(2): 157–68.
- 80. Jimenez-Gomez Y, Mattison JA, Pearson KJ, Martin-Montalvo A,

Palacios HH, Sossong AM, *et al.* Resveratrol improves adipose insulin signaling and reduces the inflammatory response in adipose tissue of rhesus monkeys on high-fat, high-sugar diet. Cell Metab. 2013; 18(4): 533–45.

- Mattison JA, Wang M, Bernier M, Zhang J, Park SS, Maudsley S, et al. Resveratrol prevents high fat/sucrose diet-induced central arterial wall inflammation and stiffening in nonhuman primates. Cell Metab. 2014; 20(1): 183–90.
- Meidenbauer JJ, Ta N, Seyfried TN. Influence of a ketogenic diet, fish-oil, and calorie restriction on plasma metabolites and lipids in C57BL/6J mice. Nutr Metab. 2014; 11(1): 23. doi: 10.1186/1743-7075-11-23.
- Roberts MN, Wallace MA, Tomilov AA, Zhou Z, Marcotte GR, Tran D, *et al.* A ketogenic diet extends longevity and healthspan in adult mice. Cell Metab. 2017; 26(3): 539-46.e5.
- 84. Cantó C, Houtkooper RH, Pirinen E, Youn DY, Oosterveer MH, Cen Y, et al. The NAD+ precursor nicotinamide riboside enhances oxidative metabolism and protects against high-fat diet induced obesity. Cell Metab. 2012; 15(6): 838–47.
- Gomes AP, Price NL, Ling AJY, Moslehi JJ, Montgomery MK, Rajman L, *et al.* Declining NAD+ induces a pseudohypoxic state disrupting nuclear-mitochondrial communication during aging. Cell. 2013; 155(7): 1624–38.
- Mouchiroud L, Houtkooper RH, Moullan N, Katsyuba E, Ryu D, Cantó C, *et al.* The NAD+/sirtuin pathway modulates longevity through activation of mitochondrial UPR and FOXO signaling. Cell. 2013; 154(2): 430–41.
- Yoshino J, Mills KF, Yoon MJ, Imai S. Nicotinamide mononucleotide, a key NAD+ intermediate, treats the pathophysiology of diet- and age-induced diabetes in mice. Cell Metab. 2011; 14(4): 528–36.
- Imai S, Guarente L. NAD+ and sirtuins in aging and disease. Trends Cell Biol. 2014; 24(8): 464–71.
- Wallace DC, Fan W, Procaccio V. Mitochondrial energetics and therapeutics. Annu Rev Pathol Mech Dis. 2010; 5(1): 297–348.
- Koopman WJH, Willems PHGM, Smeitink JAM. Monogenic mitochondrial disorders. N Engl J Med. 2012; 366(12): 1132–41.
- Sun N, Youle RJ, Finkel T. The mitochondrial basis of aging. Mol Cell. 2016; 61(5): 654–66.
- Trifunovic A, Wredenberg A, Falkenberg M, Spelbrink JN, Rovio AT, Bruder CE, *et al.* Premature ageing in mice expressing defective mitochondrial DNA polymerase. Nature. 2004; 429(6990): 417–23.
- Jiang N, Yan X, Zhou W, Zhang Q, Chen H, Zhang Y, et al. NMRbased metabonomic investigations into the metabolic profile of the senescence-accelerated mouse. J Proteome Res. 2008; 7(9): 3678– 86.
- 94. Joseph AM, Adhihetty PJ, Buford TW, Wohlgemuth SE, Lees HA, Nguyen LMD, *et al.* The impact of aging on mitochondrial function and biogenesis pathways in skeletal muscle of sedentary high- and low-functioning elderly individuals. Aging Cell. 2012; 11(5): 801–9.
- Longo VD, Antebi A, Bartke A, Barzilai N, Brown-Borg HM, Caruso C, *et al.* Interventions to slow aging in humans: are we ready? Aging Cell. 2015; 14(4): 497–510.
- Correia-Melo C, Marques FDM, Anderson R, Hewitt G, Hewitt R, Cole J, *et al.* Mitochondria are required for pro-ageing features of the senescent phenotype. EMBO J. 2016; 35(7): 724–42.
- Dillin A, Hsu AL, Arantes-Oliveira N, Lehrer-Graiwer J, Hsin H, Fraser AG, *et al.* Rates of behavior and aging specified by mitochondrial function during development. Science. 2002; 298(5602): 2398–401.
- Lee SS, Lee RYN, Fraser AG, Kamath RS, Ahringer J, Ruvkun G. A systematic RNAi screen identifies a critical role for mitochondria in

C. elegans longevity. Nat Genet. 2003; 33(1): 40-8.

- Short KR, Bigelow ML, Kahl J, Singh R, Coenen-Schimke J, Raghavakaimal S, *et al.* Decline in skeletal muscle mitochondrial function with aging in humans. PNAS. 2005; 102(15): 5618–23.
- 100. Merkwirth C, Jovaisaite V, Durieux J, Matilainen O, Jordan SD, Quiros PM, *et al.* Two conserved histone demethylases regulate mitochondrial stress-induced longevity. Cell. 2016; 165(5): 1209– 23.
- 101. Tian Y, Garcia G, Bian Q, Steffen KK, Joe L, Wolff S, *et al.* Mitochondrial stress induces chromatin reorganization to promote longevity and UPRmt. Cell. 2016; 165(5): 1197–208.
- Tatar M, Sedivy JM. Mitochondria: masters of epigenetics. Cell. 2016; 165(5): 1052–4.
- Couvillion MT, Soto IC, Shipkovenska G, Churchman LS. Synchronized mitochondrial and cytosolic translation programs. Nature. 2016; 533(7604): 499–503.
- Richter-Dennerlein R, Oeljeklaus S, Lorenzi I, Ronsör C, Bareth B, Schendzielorz AB, *et al.* Mitochondrial protein synthesis adapts to influx of nuclear-encoded protein. Cell. 2016; 167(2): 471–83.e10.
- Quirós PM, Mottis A, Auwerx J. Mitonuclear communication in homeostasis and stress. Nat Rev Mol Cell Biol. 2016; 17(4): 213– 26.
- Matilainen O, Quirós PM, Auwerx J. Mitochondria and epigenetics crosstalk in homeostasis and dtress. Trends Cell Biol. 2017; 27(6): 453–63.
- Margueron R, Reinberg D. Chromatin structure and the inheritance of epigenetic information. Nat Rev Genet. 2010; 11(4): 285–96.
- 108. Menzies KJ, Zhang H, Katsyuba E, Auwerx J. Protein acetylation in metabolism — metabolites and cofactors. Nat Rev Endocrinol. 2016; 12(1): 43–60.
- 109. Pietrocola F, Galluzzi L, Bravo-San Pedro JM, Madeo F, Kroemer G. Acetyl coenzyme A: a central metabolite and second messenger. Cell Metab. 2015; 21(6): 805–21.
- Teperino R, Schoonjans K, Auwerx J. Histone methyl transferases and demethylases; can they link metabolism and transcription? Cell Metab. 2010; 12(4): 321–7.
- Greer EL, Shi Y. Histone methylation: a dynamic mark in health, disease and inheritance. Nat Rev Genet. 2012; 13(5): 343–57.
- López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. Cell. 2013; 153(6): 1194–217.
- 113. Berger SL, Sassone-Corsi P. Metabolic dignaling to vhromatin. Cold Spring Harb Perspect Biol. 2016; 8(11): a019463. doi: 10.1101/ cshperspect.a019463.
- 114. Li X, Egervari G, Wang Y, Berger SL, Lu Z. Regulation of chromatin and gene expression by metabolic enzymes and metabolites. Nat Rev Mol Cell Biol. 2018; 19(9): 563–78.
- Lu C, Thompson CB. Metabolic regulation of epigenetics. Cell Metab. 2012; 16(1): 9–17.
- Finkel T, Deng CX, Mostoslavsky R. Recent progress in the biology and physiology of sirtuins. Nature. 2009; 460(7255): 587–91.
- Chalkiadaki A, Guarente L. The multifaceted functions of sirtuins in cancer. Nat Rev Cancer. 2015; 15(10): 608–24.
- Imai S, Armstrong CM, Kaeberlein M, Guarente L. Transcriptional silencing and longevity protein Sir2 is an NAD-dependent histone deacetylase. Nature. 2000; 403(6771): 795–800.
- Schwer B, Verdin E. Conserved metabolic regulatory functions of sirtuins. Cell Metab. 2008; 7(2): 104–12.
- 120. Landry J, Sutton A, Tafrov ST, Heller RC, Stebbins J, Pillus L, et al. The silencing protein SIR2 and its homologs are NAD-dependent protein deacetylases. PNAS. 2000; 97(11): 5807–11.
- 121. Hansen M, Rubinsztein DC, Walker DW. Autophagy as a promoter of longevity: insights from model organisms. Nat Rev Mol Cell Biol.

2018; 19(9): 579–93.

- 122. Kalfalah F, Janke L, Schiavi A, Tigges J, IX A, Ventura N, *et al.* Crosstalk of clock gene expression and autophagy in aging. Aging (Albany NY). 2016; 8(9): 1876–95.
- Green DR, Galluzzi L, Kroemer G. Mitochondria and the autophagyinflammation-cell death axis in organismal aging. Science. 2011; 333(6046): 1109–12.
- Palikaras K, Lionaki E, Tavernarakis N. Coordination of mitophagy and mitochondrial biogenesis during ageing in C. elegans. Nature. 2015; 521(7553): 525–8.
- Pickrell JA, Oehme FW. Invited response to definition of hormesis (EJ Calabrese and LA Baldwin). Hum Exp Toxicol. 2002; 21(2): 107–9.
- 126. Ashrafi G, Schwarz TL. The pathways of mitophagy for quality control and clearance of mitochondria. Cell Death Differ. 2013; 20(1): 31–42.
- Youle RJ, Narendra DP. Mechanisms of mitophagy. Nat Rev Mol Cell Biol. 2011; 12(1): 9–14.
- 128. Dutta D, Calvani R, Bernabei R, Leeuwenburgh C, Marzetti E. Contribution of impaired mitochondrial autophagy to cardiac aging: mechanisms and therapeutic opportunities. Circ Res. 2012; 110(8): 1125–38.
- 129. Singh LP, Devi TS, Yumnamcha T. The role of Txnip in mitophagy dysregulation and inflammasome activation in diabetic retinopathy: a new perspective. JOJ Ophthalmol. 2017; 4(4): 1–11. doi: 10.19080/jojo.2017.04.555643.
- 130. Pang L, Liu K, Liu D, Lv F, Zang Y, Xie F, *et al.* Differential effects of reticulophagy and mitophagy on nonalcoholic fatty liver disease. Cell Death Dis. 2018; 9(2): 90. doi: 10.1038/s41419-017-0136-y.
- Marshall E. Metabolic research. Canadian group claims "unique" database. Science. 2007; 315(5812): 583–4.
- 132. Chen K, Dai H, Yuan J, Chen J, Lin L, Zhang W, et al. Optineurinmediated mitophagy protects renal tubular epithelial cells against accelerated senescence in diabetic nephropathy. Cell Death Dis. 2018; 9(2): 105. doi: 10.1038/s41419-017-0127-z.
- Hansen M, Flatt T, Aguilaniu H. Reproduction, fat metabolism, and life span: what is the connection? Cell Metab. 2013; 17(1): 10–9.
- 134. Schulze RJ, Sathyanarayan A, Mashek DG. Breaking fat: The regulation and mechanisms of lipophagy. Biochim Biophys Acta Mol Cell Biol Lipids. 2017; 1862(10 Pt B): 1178–87.
- 135. Singh R, Kaushik S, Wang Y, Xiang Y, Novak I, Komatsu M, et al. Autophagy regulates lipid metabolism. Nature. 2009; 458(7242): 1131–5.
- 136. Kaushik S, Cuervo AM. The coming of age of chaperone-mediated autophagy. Nat Rev Mol Cell Biol. 2018; 19(6): 365–81.
- 137. Singh R, Cuervo AM. Lipophagy: connecting autophagy and lipid metabolism. Int J Cell Biol. 2012; 2012: 282041. doi: 10.1155/2012/282041.
- Maan M, Peters JM, Dutta M, Patterson AD. Lipid metabolism and lipophagy in cancer. Biochem Biophys Res Commun. 2018; 504(3): 582–9.
- 139. Chen K, Yuan R, Zhang Y, Geng S, Li L. Tollip deficiency alters atherosclerosis and steatosis by disrupting lipophagy. J Am Heart Assoc. 2017; 6(4): e004078. doi: 10.1161/JAHA.116.004078.
- 140. Ravikumar B, Duden R, Rubinsztein DC. Aggregate-prone proteins with polyglutamine and polyalanine expansions are degraded by autophagy. Hum Mol Genet. 2002; 11(9): 1107–17.
- 141. Webb JL, Ravikumar B, Atkins J, Skepper JN, Rubinsztein DC. α-synuclein is degraded by both autophagy and the proteasome. J Biol Chem. 2003; 278(27): 25009–13.
- 142. Berger Z, Ravikumar B, Menzies FM, Oroz LG, Underwood BR, Pangalos MN, *et al.* Rapamycin alleviates toxicity of different

aggregate-prone proteins. Hum Mol Genet. 2006; 15(3): 433-42.

- 143. Ravikumar B, Vacher C, Berger Z, Davies JE, Luo S, Oroz LG, et al. Inhibition of mTOR induces autophagy and reduces toxicity of polyglutamine expansions in fly and mouse models of Huntington disease. Nat Genet. 2004; 36(6): 585–95.
- 144. Lopez A, Lee SE, Wojta K, Ramos EM, Klein E, Chen J, et al. A152T tau allele causes neurodegeneration that can be ameliorated in a zebrafish model by autophagy induction. Brain. 2017; 140(4): 1128–46.
- 145. Kumsta C, Chang JT, Schmalz J, Hansen M. Hormetic heat stress and HSF-1 induce autophagy to improve survival and proteostasis in C. elegans. Nat Commun. 2017; 8: 14337. doi: 10.1038/ncomms14337.
- 146. Menzies FM, Fleming A, Caricasole A, Bento CF, Andrews SP, Ashkenazi A, et al. Autophagy and neurodegeneration: pathogenic mechanisms and therapeutic opportunities. Neuron. 2017; 93(5): 1015–34.
- Lopez-Otín C, Kroemer G. Decelerating ageing and biological clocks by autophagy. Nat Rev Mol Cell Biol. 2019; 20(7): 385–6.
- 148. Levy JMM, Towers CG, Thorburn A. Targeting autophagy in cancer. Nat Rev Cancer. 2017; 17(9): 528–42.
- 149. Galluzzi L, Bravo-San Pedro JM, Levine B, Green DR, Kroemer G. Pharmacological modulation of autophagy: therapeutic potential and persisting obstacles. Nat Rev Drug Discov. 2017; 16(7): 487– 511.
- Galluzzi L, Pietrocola F, Levine B, Kroemer G. Metabolic control of autophagy. Cell. 2014; 159(6): 1263–76.
- 151. Kubben N, Misteli T. Shared molecular and cellular mechanisms of premature ageing and ageing-associated diseases. Nat Rev Mol Cell Biol. 2017; 18(10): 595–609.
- Hayflick L. The limited in vitro lifetime of human diploid cell strains. Exp Cell Res. 1965; 37(3): 614–36.
- Herbig U. Cellular senescence in aging primates. Science. 2006; 311(5765): 1257. doi: 10.1126/science.1122446.
- 154. Farr JN, Fraser DG, Wang H, Jaehn K, Ogrodnik MB, Weivoda MM, et al. Identification of senescent cells in the bone microenvironment: senescent in the bone microenvironment. J Bone Miner Res. 2016; 31(11): 1920–9.
- 155. Biran A, Zada L, Abou Karam P, Vadai E, Roitman L, Ovadya Y, et al. Quantitative identification of senescent cells in aging and disease. Aging Cell. 2017; 16(4): 661–71.
- 156. Huang S, Risques RA, Martin GM, Rabinovitch PS, Oshima J. Accelerated telomere shortening and replicative senescence in human fibroblasts overexpressing mutant and wild-type lamin A. Exp Cell Res. 2008; 314(1): 82–91.
- van Deursen JM. The role of senescent cells in ageing. Nature. 2014; 509(7501): 439–46.
- 158. Chandra T, Kirschner K, Thuret J-Y, Pope BD, Ryba T, Newman S, *et al.* Independence of repressive histone marks and chromatin compaction during senescent heterochromatic layer formation. Mol Cell. 2012; 47(2): 203–14.
- Zhang R, Chen W, Adams PD. Molecular dissection of formation of senescence-associated heterochromatin foci. Mol Cell Biol. 2007; 27(6): 2343–58.
- 160. Freund A, Laberge RM, Demaria M, Campisi J. Lamin B1 loss is a senescence-associated biomarker. Mol Biol Cell. 2012; 23(11): 2066–75.
- 161. Shimi T, Butin-Israeli V, Adam SA, Hamanaka RB, Goldman AE, Lucas CA, *et al.* The role of nuclear lamin B1 in cell proliferation and senescence. Genes Dev. 2011; 25(24): 2579–93.
- 162. Dou Z, Xu C, Donahue G, Shimi T, Pan JA, Zhu J, *et al.* Autophagy mediates degradation of nuclear lamina. Nature. 2015; 527(7576): 105–9.

- 163. Rai TS, Cole JJ, Nelson DM, Dikovskaya D, Faller WJ, Vizioli MG, *et al.* HIRA orchestrates a dynamic chromatin landscape in senescence and is required for suppression of neoplasia. Genes Dev. 2014; 28(24): 2712–25.
- 164. Lundblad V, Szostak JW. A mutant with a defect in telomere elongation leads to senescence in yeast. Cell. 1989; 57(4): 633–43.
- Harley CB, Futcher AB, Greider CW. Telomeres shorten during ageing of human fibroblasts. Nature. 1990; 345(6274): 458–60.
- 166. Sohal RS, Weindruch R. Oxidative stress, caloric restriction, and aging. Science. 1996; 273(5271): 59–63.
- 167. Vaziri H, Benchimol S. From telomere loss to p53 induction and activation of a DNA-damage pathway at senescence: the telomere loss/DNA damage model of cell aging. Exp Gerontol. 1996; 31(1– 2): 295–301.
- 168. Sedelnikova OA, Horikawa I, Zimonjic DB, Popescu NC, Bonner WM, Barrett JC. Senescing human cells and ageing mice accumulate DNA lesions with unrepairable double-strand breaks. Nat Cell Biol. 2004; 6(2): 168–70.
- 169. Seluanov A, Mittelman D, Pereira-Smith OM, Wilson JH, Gorbunova V. DNA end joining becomes less efficient and more error-prone during cellular senescence. Proc Natl Acad Sci USA. 2004; 101(20): 7624–9.
- Sfeir A, de Lange T. Removal of shelterin reveals the telomere endprotection problem. Science. 2012; 336(6081): 593–7.
- 171. Sedivy JM, Banumathy G, Adams PD. Aging by epigenetics—A consequence of chromatin damage? Exp Cell Res. 2008; 314(9): 1909–17.
- 172. Feser J, Tyler J. Chromatin structure as a mediator of aging. FEBS Letters. 2011; 585(13): 2041–8.
- O'Sullivan RJ, Karlseder J. The great unravelling: chromatin as a modulator of the aging process. Trends Biochem Sci. 2012; 37(11): 466–76.
- 174. Tchkonia T, Zhu Y, van Deursen J, Campisi J, Kirkland JL. Cellular senescence and the senescent secretory phenotype: therapeutic opportunities. J Clin Invest. 2013; 123(3): 966–72.
- 175. Krishnamurthy J, Ramsey MR, Ligon KL, Torrice C, Koh A, Bonner-Weir S, *et al.* p16INK4a induces an age-dependent decline in islet regenerative potential. Nature. 2006; 443(7110): 453–7.
- Baumann K. Rejuvenating senolytics. Nat Rev Mol Cell Biol. 2018; 19(9): 543. doi: 10.1038/s41580-018-0047-5.
- 177. Kenyon CJ. The genetics of ageing. Nature. 2010; 464(7288): 504– 12.
- 178. Gems D, Partridge L. Genetics of longevity in model organisms: debates and paradigm shifts. Annu Rev Physiol. 2013; 75: 621–44.
- 179. Brandhorst S, Choi IY, Wei M, Cheng CW, Sedrakyan S, Navarrete G, et al. A periodic diet that mimics fasting promotes multi-system regeneration, enhanced cognitive performance, and healthspan. Cell Metab. 2015; 22(1): 86–99.
- 180. Cheng CW, Villani V, Buono R, Wei M, Kumar S, Yilmaz OH, et al. Fasting-mimicking diet promotes Ngn3-driven β-cell regeneration to reverse diabetes. Cell. 2017; 168(5): 775-788.e12.
- 181. Cerletti M, Jang YC, Finley LWS, Haigis MC, Wagers AJ. Short-term calorie restriction enhances skeletal muscle stem cell function. Cell Stem Cell. 2012; 10(5): 515–9.
- 182. Newman JC, Covarrubias AJ, Zhao M, Yu X, Gut P, Ng CP, et al. Ketogenic diet reduces midlife mortality and improves memory in aging mice. Cell Metab. 2017; 26(3): 547–57.e8.
- 183. Harrison DE, Strong R, Sharp ZD, Nelson JF, Astle CM, Flurkey K, *et al.* Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. Nature. 2009; 460(7253): 392–5.
- 184. de Cabo R, Carmona-Gutierrez D, Bernier M, Hall MN, Madeo F. The search for antiaging interventions: from elixirs to fasting regimens.

Cell. 2014; 157(7): 1515-26.

- 185. Conboy MJ, Conboy IM, Rando TA. Heterochronic parabiosis: historical perspective and methodological considerations for studies of aging and longevity. Aging Cell. 2013; 12(3): 525–30.
- 186. de Keizer PLJ. The fountain of youth by targeting Senescent cells? Trends Mol Med. 2017; 23(1): 6–17.
- 187. Mahmoudi S, Brunet A. Aging and reprogramming: a two-way street. Curr Opin Cell Biol. 2012; 24(6): 744–56.
- Oh J, Lee YD, Wagers AJ. Stem cell aging: mechanisms, regulators and therapeutic opportunities. Nat Med. 2014; 20(8): 870–80.
- Galluzzi L, Yamazaki T, Kroemer G. Linking cellular stress responses to systemic homeostasis. Nat Rev Mol Cell Biol. 2018; 19(11): 731–45.
- 190. Weir HJ, Yao P, Huynh FK, Escoubas CC, Goncalves RL, Burkewitz K, et al. Dietary restriction and AMPK increase lifespan via mitochondrial network and peroxisome remodeling. Cell Metab. 2017; 26(6): 884–96.e5.
- 191. Labbadia J, Brielmann RM, Neto MF, Lin YF, Haynes CM, Morimoto RI. Mitochondrial stress restores the heat shock response and prevents proteostasis collapse during aging. Cell Rep. 2017; 21(6): 1481–94.
- Guarente L. Mitochondria—a nexus for aging, calorie restriction, and sirtuins? Cell. 2008; 132(2): 171–6.
- 193. Masotti A, Celluzzi A, Petrini S, Bertini E, Zanni G, Compagnucci C. Aged iPSCs display an uncommon mitochondrial appearance and fail to undergo in vitro neurogenesis. Aging (Albany NY). 2014; 6(12): 1094–108.
- 194. Sinha I, Sinha-Hikim AP, Wagers AJ, Sinha-Hikim I. Testosterone is essential for skeletal muscle growth in aged mice in a heterochronic parabiosis model. Cell Tissue Res. 2014; 357(3): 815–21.
- 195. Ren R, Ocampo A, Liu GH, Izpisua Belmonte JC. Regulation of stem cell aging by metabolism and epigenetics. Cell Metab. 2017; 26(3): 460–74.
- 196. Zhang H, Ryu D, Wu Y, Gariani K, Wang X, Luan P, et al. NAD+ repletion improves mitochondrial and stem cell function and enhances life span in mice. Science. 2016; 352(6292): 1436–43.
- 197. Igarashi M, Miura M, Williams E, Jaksch F, Kadowaki T, Yamauchi T, *et al.* NAD+ supplementation rejuvenates aged gut adult stem cells. Aging Cell. 2019; 18(3): e12935. doi: 10.1111/acel.12935.
- Rajman L, Chwalek K, Sinclair DA. Therapeutic potential of NADboosting molecules: the in vivo evidence. Cell Metab. 2018; 27(3): 529–47.
- 199. Yoshino J, Baur JA, Imai SI. NAD+ intermediates: the biology and therapeutic potential of NMN and NR. Cell Metab. 2018; 27(3): 513–28.
- 200. Mitchell SJ, Bernier M, Aon MA, Cortassa S, Kim EY, Fang EF, et al. Nicotinamide improves aspects of healthspan, but not lifespan, in mice. Cell Metab. 2018; 27(3): 667–76.e4.
- 201. Saini JS, Corneo B, Miller JD, Kiehl TR, Wang Q, Boles NC, et al. Nicotinamide ameliorates disease phenotypes in a human iPSC model of age-related macular degeneration. Cell Stem Cell. 2017; 20(5): 635–47.e7.
- Saint-Geniez M, Rosales MAB. Eyeing the fountain of youth. Cell Stem Cell. 2017; 20(5): 583–4.
- 203. Katsyuba E, Mottis A, Zietak M, De Franco F, van der Velpen V, Gariani K, *et al.* De novo NAD+ synthesis enhances mitochondrial function and improves health. Nature. 2018; 563(7731): 354–9.
- 204. Martens CR, Denman BA, Mazzo MR, Armstrong ML, Reisdorph N, McQueen MB, *et al.* Chronic nicotinamide riboside supplementation is well-tolerated and elevates NAD+ in healthy middle-aged and older adults. Nat Commun. 2018; 9(1): 1286. doi: 10.1038/s41467-018-03421-7.

- 205. Narita M, Nuñez S, Heard E, Narita M, Lin AW, Hearn SA, et al. Rb-mediated heterochromatin formation and silencing of E2F target genes during cellular senescence. Cell. 2003; 113(6): 703–16.
- 206. Foran E, Garrity-Park MM, Mureau C, Newell J, Smyrk TC, Limburg PJ, et al. Upregulation of DNA methyltransferase-mediated gene silencing, anchorage-independent growth, and migration of colon cancer cells by interleukin-6. Mol Cancer Res. 2010; 8(4): 471–81.
- 207. Hodge DR, Xiao W, Clausen PA, Heidecker G, Szyf M, Farrar WL. Interleukin-6 regulation of the human DNA methyltransferase (HDNMT) gene in human erythroleukemia cells. J Biol Chem. 2001; 276(43): 39508–11.
- 208. Ocampo A, Reddy P, Martinez-Redondo P, Platero-Luengo A, Hatanaka F, Hishida T, *et al.* In vivo amelioration of age-

associated hallmarks by partial reprogramming. Cell. 2016; 167(7): 1719–33.e12.

- 209. Hahn O, Grönke S, Stubbs TM, Ficz G, Hendrich O, Krueger F, *et al.* Dietary restriction protects from age-associated DNA methylation and induces epigenetic reprogramming of lipid metabolism. Genome Biol. 2017; 18(1): 56. doi: 10.1186/s13059-017-1187-1.
- 210. Rando TA, Chang HY. Aging, rejuvenation, and epigenetic reprogramming: resetting the aging clock. Cell. 2012; 148(1): 46– 57.
- 211. Greer EL, Maures TJ, Hauswirth AG, Green EM, Leeman DS, Maro GS, *et al.* Members of the H3K4 trimethylation complex regulate lifespan in a germline-dependent manner in C. elegans. Nature. 2010; 466(7304): 383–7.