

Original Article

Asian Pacific Journal of Reproduction

Journal homepage: www.apjr.net



doi: 10.4103/2305–0500.281078

Effects of ciprofloxacin on testicular tissue and sperm quality in rabbits

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ABSTRACT

Objective: To investigate the hormonal, histopathological toxicity and sperm quality of ciprofloxacin in male genital system in the rabbit model.

Methods: Twenty adult New Zealand male rabbits were randomly divided into the treatment and control groups, and respectively received 30 mg/kg/day ciprofloxacin and normal saline intraperitoneally for 14 days. Blood samples and testicular tissues were taken for testosterone by enzyme linked immunosorbent assay method, and measurement of histopathologic and histomorphometric investigations was made on the 14th day of the experiment and 56 days after the last dose of ciprofloxacin. Additionally, epididymis sperm sample was collected for quality analysis.

Results: On day 14 of the experiment, histopathologic examination revealed severe degenerative changes in the epithelium of seminiferous tubules and loss of all germ cells in most tubes in the treatment group. In the histomorphometric study, significant reductions in the seminiferous tubules diameter, epithelium height, and Johnsen' score were observed. On day 56 after the last dose of ciprofloxacin, the seminiferous tubules revealed regeneration in the treatment group and no significant difference was observed in the spermatogenesis parameters except epithelial height parameter between the control group and the treatment group. All sperm quality parameters were significantly decreased on day 56 after the last dose of ciprofloxacin. Testosterone levels did not significantly change during the follow-ups period.

Conclusions: There is potential reversible testicular toxicity for ciprofloxacin in the rabbit model in according to relative regeneration of spermatogenic epithelium after 56 days of last dose of ciprofloxacin. Sperm quality will be improved with a delay after complete regeneration of seminiferous tubules.

KEYWORDS: Ciprofloxacin; Testis; Rabbits; Histopathology; Testosterone; Histomorphometry

1. Introduction

Infertility is a significant problem with a 20% incidence among couples and reported more in men[1,2]. Various agents lead to infertility in males including defects in the spermatogenesis, sperm transportation, number and morphology of sperm[1], infections of the urinary system[3,4] and anatomical factors such as varicoceles and ductal obstructions[5]. Also, male fertility may be affected by some environmental risk factors such as cigarette smoking, radiation, nutritional condition, estrogens, heavy metals, scrotal temperature and anti-bacterial drugs[6,7]. Antibiotics may induce injury to the spermatogenesis in men and animal species. One of the antibiotics is ciprofloxacin that belongs to the second-generation fluoroquinolones[8]. This broad-spectrum antibiotic enters the seminal fluid and decreases concentration, motility and viability of sperm[9]. Long use of some quinolones causes toxicity in the spermatogenesis and testicular tissue[10].

Rabbits are excellent models for many aspects of research in reproductive toxicology. In the present study, the side effects of ciprofloxacin were investigated on male rabbit system by histopathological and histomorphometric evaluation.

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How to cite this article: Kheirandish R, Emadi L, Akhtardanesh B, Azizi S, Imani M, Mahmoodabadi F, et al. Effects of ciprofloxacin on testicular tissue and sperm quality in rabbits. *Asian Pac J Reprod* 2020; 9(2): 83-88.

Article history: Received: 22 June 2019; Revision: 1 December 2019; Accepted: 9 December 2019; Available online: 30 March 2020

2. Materials and methods

2.1. Experimental animals

Twenty white adult New Zealand male rabbits (*Oryctolagus caniculus*) (12 months old, 2-4 kg) were kept in the cages for one week until be adapted to the environment under the standard condition [12 h light-12 h dark/ (22 ± 2) °C] and free access to the balanced diet and water *ad libitum*. The animals were randomly divided into the control and ciprofloxacin (treatment) groups ($n=10$ in each group). The ciprofloxacin group received intraperitoneal ciprofloxacin at dose of 30 mg/kg/day for 14 days. The control group received the same volume of normal saline. The 30 mg/kg was treatment dose with a course of 14 days for usually infections in human[11].

2.2. Blood collection

Blood samples (5 mL) were taken from the jugular vein of each rabbit in the control and treatment groups for hormonal analysis on the 14th day after the beginning of treatment (the 2nd week) and 56th day following the last ciprofloxacin dose. 56 days was chosen according to a cycle of spermatogenesis in rabbit that is was 48 days. Serum was separated by the centrifuge at 3 000 rpm for 10 min and testosterone level was measured by DRG testosterone enzyme linked immunosorbent assay kit (EIA-1559).

2.3. Histopathological and histomorphometric investigations

Five animals were selected from each ciprofloxacin and control groups on the 14th day of the experiment and 56 days after the last ciprofloxacin dose to conduct histopathological and histomorphometric investigation. After euthanasia with sodium thiopental overdose, testes from rabbits of both groups were fixed in 10% neutral buffered formalin. After fixation, the tissue samples were processed *via* the standard procedure and embedded in the paraffin wax. Tissue sections in 5 µm thickness were stained with hematoxylin-eosin (H & E) and studied by light microscopy (Olympus, Model: CX31) with 40, 100, 400 magnifications in according to investigated parameters. For the histomorphometric study, diameter and height epithelium in 10 smallest and roundest seminiferous tubules were investigated. In addition, the diameter of 10 spermatogonia[12] and Johnsen's score were determined in the control and ciprofloxacin testes[13]. The level of sperm maturation was graded between 1 and 10, according to the most advanced germ cell in the tubule.

2.4. Sperm sample collection and preparation

Sperm samples were obtained from 10 cases of 20 animals of each group during the experimental period in the 14th day of the experiment and 56 days after the last ciprofloxacin treatment. In order to limit blood contamination, the cauda epididymis was

dissected from the connective tissues, rinsed in the 37 °C normal saline and dried carefully. Samples of mature spermatozoa were collected from the caudal region of epididymis by finely mincing it in phosphate-buffered saline at 37 °C. Sperm evaluation was carried out by Computer Assisted Semen Analysis (CASA) system (HFT CASA V6.50, Hooshmand Fanavar Tehran Co., Iran) in each experimental group.

2.5. Sperm analysis

Sperm concentration, motile sperm ratio and sperm motion characteristics were measured with the CASA system. Briefly, 10 µL of sperm sample was pipetted into a sperm counting chamber with 10 µm depth that had been prewarmed to 37 °C on a heated microscopic stage (HT 50, Minitube Inc., Germany) and evaluated by a light microscopy (DN-PW117M, Pro Way Optics and Electronics Co., China) with magnification of ×40. At least 1 000 spermatozoa were observed in eight sequentially selected microscopic fields. The spermatozoa motility patterns were monitored by a digital video camera system (SDS-313B, Samsung Techwin Co., South Korea) connected to the light microscope. Five motion parameters obtained from CASA sperm tracker were as follows: (1) curvilinear velocity (VCL, micrometers per second): the average velocity over the total distance moved, including all deviations of sperm head movement; (2) straight-line velocity (VSL, micrometers per second): the average velocity calculated using the straight line distance between the beginning and end of the sperm track; (3) average path velocity (VAP, micrometers per second): the average velocity of the sperm head along its average path; (4) amplitude of lateral head displacement (ALH, micrometers): the mean value of the extreme side to side movement of the sperm head in each beat and (5) straightness of the path velocity (VSL/VAP) (STR). Additionally, sperms based on motion properties were divided into four categories: Class A: Motile sperm with high-velocity progressive motion; Class B: Motile sperm with low-velocity progressive motion; Class C: Sperm moving in place; Class D: Non-motile sperm.

2.6. Statistical analysis

The results were analyzed by SPSS 17.0 (SPSS Inc., Chicago, IL, USA). Independent Student's *t*-test were used for investigated parameters for comparing the control and ciprofloxacin groups. Data are expressed as mean standard deviation (mean±SD). *P*-value less than 0.05 was considered as statistically significant different level.

2.7. Ethics statement

This study was approved by the Ethical Committee of Shahid Bahonar Veterinary School (ethical approval No. IR.UK.REC.1395.001). All animals received human care in compliance with the Guide for Care and Use of Laboratory Animals published by the National Institutes of Health.

3. Results

3.1. Histopathological findings

In the control group, the microscopic study revealed normal structure of the seminiferous tubules, spermatogenesis and Sertoli cells during the experiment (Figure 1A). Fourteen days after initiation of ciprofloxacin injection, the ciprofloxacin group showed severe degenerative changes in the epithelium of the seminiferous tubules. The most tubules were wrinkled and collapsed. All the germ cells had decreased and only the spermatogonia were observed. Sertoli cells had a clear nucleus and marked nucleolus. Giant-like cells were also visible in the seminal lumen (Figure 1B). Edema and hemorrhage had occurred in the interstitial tissues between the tubules (Figure 1B). 56 days after the last ciprofloxacin dose, most of the tubules were regenerated in the ciprofloxacin group. All stages of germ cells from spermatogonia to spermatozoa were present. The epithelium of the seminiferous tubules showed the structure near to the control group (Figure 1C).

3.2. Histomorphometric evaluation

Fourteen days after initiation of ciprofloxacin injection, Johnsen's score (indicator for spermatogenesis), the diameter and epithelium height of the seminal tubules significantly decreased in comparison to the control group ($P < 0.05$). However, the diameter of spermatogonium nucleus in the ciprofloxacin group was significantly higher than that in the control group ($P < 0.05$) (Table 1).

And 56 days following the last ciprofloxacin dose, evidence of regeneration was observed in the tubular epithelium and these tubules were reconstructed. The most spermatogenesis parameters were the same as the normal control group. There were no significant differences in the diameter of the seminal tubes, spermatogonia nucleus and the Johnsen score between the ciprofloxacin group and the normal control group ($P > 0.05$) (Table 1). In the ciprofloxacin group, the epithelial height had increased in comparison to the second week but it was significantly lower than the control group at this time ($P < 0.05$) (Table 1).

3.3. Sperm quality

Table 2 showed the effect of ciprofloxacin 14 days after initiation of drug injection and 56 days after the end of drug administration in comparison with the control group in sperm concentration, motile sperm ratio and sperm motion characteristics. Evaluation of epididymal sperm quality parameters 14 days after initiation of drug injection revealed that sperm concentration, motile sperm ratio and sperm motion parameters were slightly decreased. When the testis be was damaged, it takes took a long time about a spermatogenic cycle until sperm quality will would be affected. Therefore, during 2 weeks, sperm quality remains remained near to normal range.

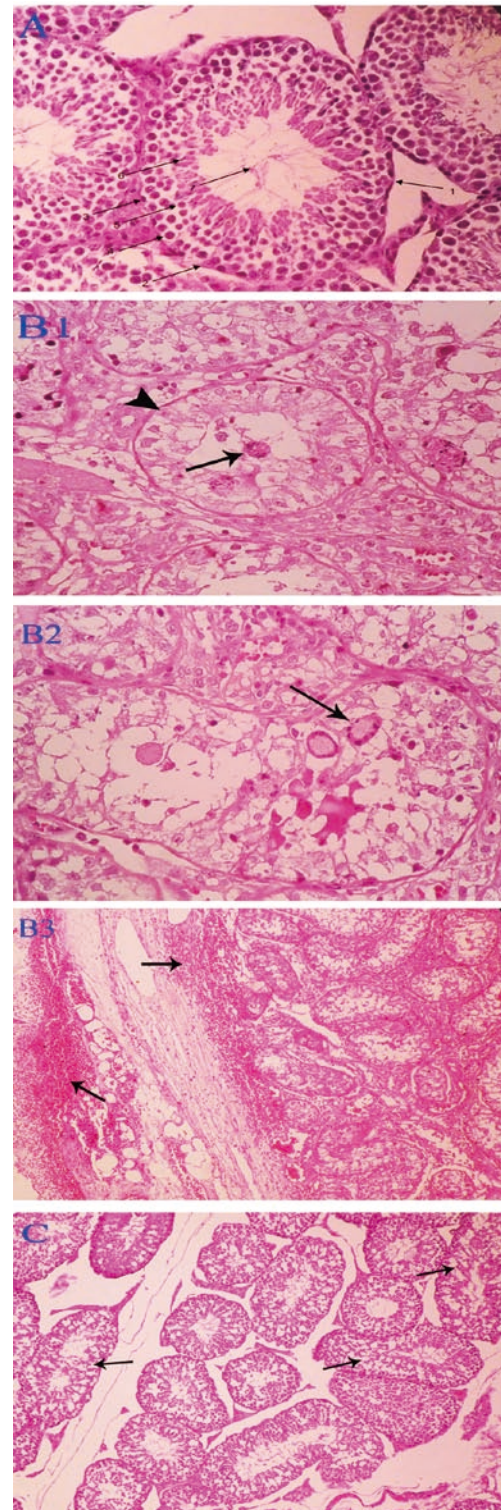


Figure 1. Histopathological findings of ciprofloxacin on the testis. A: The control group: 1) Normal seminiferous tubules in rabbit, 2) Sertoli cell, 3) Spermatogonium cell, 4) Primary spermatocyte, 5) Round spermatid, 6) Rod spermatid, 7) Spermatozoa (H & E, $\times 400$). B: Ciprofloxacin-treated group: B1) Severe degeneration of germinal epithelium, sloughing of epithelial cells into lumen (arrow) and remained Sertoli cell with a marked nucleolus (arrowhead) (H & E, $\times 400$); B2): Degeneration of the germ cells and presence of multinucleated cells in seminal lumen (arrow) (H & E, $\times 400$); B3): Wrinkled and collapsed tubules due to severe degeneration of the germ cell lines as well as hemorrhage (arrows) between tubules (H & E, $\times 100$). C: Ciprofloxacin-treated group for 56 days following last ciprofloxacin. Mild degenerative changes as vacuolation and increasing space (arrows) in germinal epithelium of seminiferous tubules (H & E, $\times 400$).

Table 1. Seminiferous tubules diameter, germinal epithelium height, diameter of spermatogonia nuclei and Johnsen's score in the control and ciprofloxacin (30 mg/kg) groups.

| Parameters | 14 days of experiment | | 56 days after last ciprofloxacin dose | |
|--|-----------------------|---------------------|---------------------------------------|---------------------|
| | Control | Ciprofloxacin | Control | Ciprofloxacin |
| Seminiferous tubules diameter (μm) | 235.90 \pm 55.81 | 197.76 \pm 66.00* | 223.34 \pm 32.37 | 222.11 \pm 27.96# |
| Germinal epithelium height (μm) | 71.12 \pm 26.49 | 62.10 \pm 20.18* | 74.16 \pm 20.96 | 68.09 \pm 22.41** |
| Diameter of spermatogonia nuclei (μm) | 5.85 \pm 1.36 | 6.38 \pm 3.79* | 6.70 \pm 1.73 | 6.64 \pm 1.31# |
| Johnsen's score | 6.96 \pm 1.56 | 5.07 \pm 1.07* | 7.10 \pm 1.71 | 6.94 \pm 1.33# |

Data are expressed as mean \pm SD. *: indicates a significant difference ($P<0.05$) in comparison with the control group and #: indicates a significant difference ($P<0.05$) with ciprofloxacin after 14 days. Non-parametric *U*-Mann-Whitney test for Johnsen's score and independent Student's *t*-test for other parameters were used for comparing the control and ciprofloxacin groups.

Table 2. Effect of ciprofloxacin on sperm motion patterns at each evaluated time.

| Parameters | Control group | Treatment 1* | Treatment 2* |
|---------------------------------------|--------------------------------|--------------------------------|--------------------------------|
| Sperm concentration ($\times 10^6$) | 155.57 \pm 5.86 ^a | 150.07 \pm 6.90 ^a | 112.14 \pm 2.57 ^b |
| Motile sperm ratio (%) | 71.16 \pm 2.39 ^a | 69.41 \pm 1.96 ^a | 34.91 \pm 3.61 ^b |
| Class A (%) | 32.46 \pm 2.12 ^a | 27.62 \pm 1.28 ^a | 9.84 \pm 0.92 ^b |
| Class B (%) | 27.08 \pm 1.73 ^a | 26.27 \pm 2.39 ^a | 15.21 \pm 1.04 ^b |
| Class C (%) | 11.63 \pm 2.39 ^a | 15.51 \pm 2.32 ^a | 9.86 \pm 2.58 ^b |
| Class D (%) | 28.83 \pm 2.39 ^a | 30.58 \pm 1.96 ^a | 65.08 \pm 3.61 ^b |
| VCL ($\mu\text{m/s}$) | 29.31 \pm 2.20 ^a | 29.46 \pm 1.31 ^a | 14.11 \pm 1.16 ^b |
| VSL ($\mu\text{m/s}$) | 13.17 \pm 1.28 ^a | 12.68 \pm 0.93 ^a | 5.07 \pm 0.45 ^b |
| VAP ($\mu\text{m/s}$) | 19.28 \pm 1.56 ^a | 19.24 \pm 1.34 ^a | 11.60 \pm 0.95 ^b |
| ALH (μm) | 1.34 \pm 0.09 ^a | 1.42 \pm 0.08 ^a | 0.70 \pm 0.03 ^b |
| STR (%) | 68.11 \pm 1.50 ^a | 65.97 \pm 2.40 ^a | 44.21 \pm 3.80 ^b |

Data are expressed as mean \pm SD; values in a row followed by different superscripts (a, b) differ significantly ($P<0.05$). *Treatment 1: 2 weeks after ciprofloxacin medication. *Treatment 2: 10 weeks after ciprofloxacin medication. Class A: Motile sperm with high velocity progressive motion. Class B: Motile sperm with low velocity progressive motion. Class C: Sperm moving in place. Class D: Non motile sperm. VCL: The average velocity over the total distance moved. VSL: The average velocity calculated using the straight line distance between the beginning and end of the sperm track. VAP: The average velocity of the sperm head along its average path. ALH: The mean value of the extreme side to side movement of the sperm head in each beat. STR: Straightness of the path velocity.

However, a mild elevation was seen in non-motile sperm count. While 56 days after the end of drug administration, sperm concentration significantly decreased in comparison with the control group. Motile sperm ratio (class A and B) was significantly decreased in the treatment group, whereas the proportion of non-motile sperms (class D) was significantly increased compared to the control group. Sperm velocity parameters (VCL, VSL and VAP) significantly decreased in comparison with the control group. Motion characteristics such as ALH that showed the ability of sperm cells in penetration through the oocyte significantly decreased in the treatment group. The significant reduction of straightness of the path velocity (STR) was observed in comparison with the control group (Table 2).

3.4. Serum testosterone level

Level of the serum testosterone on the 14th day of the experiment and 56 days after the last ciprofloxacin dose showed no significant difference statistically between the ciprofloxacin and control groups ($P>0.05$) (Table 3).

Table 3. Serum testosterone level (ng/mL) of ciprofloxacin groups following administration in 14th day of experiment and 56 days after last ciprofloxacin dose (30 mg/kg) (10th week).

| Groups | 14th day of experiment | 56 days after last ciprofloxacin dose |
|---------------|------------------------|---------------------------------------|
| Control | 1.73 \pm 0.46 | 1.26 \pm 0.91 |
| Ciprofloxacin | 1.43 \pm 0.19 | 1.31 \pm 0.44 |

Data are expressed as mean \pm SD.

4. Discussion

Ciprofloxacin is a wide spectrum antibiotic used for genitourinary tract infections. This broad-spectrum antibiotic can enter into the seminal fluid and cause toxicological effects on male reproductive organs. This antibiotic inhibits DNA gyrase and topoisomerase IV that are need for replication of prokaryotic DNA[14,15]. The common side effects of all quinolones are nausea, vomiting, dizziness and convulsions[16]. Some other ciprofloxacin adverse effects were described in the male reproductive system[8,9]. In the present study, a potential reversible testicular toxicity of ciprofloxacin was demonstrated in the rabbit model 56 days after last dose of ciprofloxacin administration. The cycle of the spermatogenesis in rabbit is 48 days. Therefore, sperm quality remains in low quality until the regeneration of damaged seminiferous tubules be completed.

Previously, some researchers reported that ciprofloxacin is a safe antibiotic[17]. In the study of Waite et al, the effects of ciprofloxacin (500 mg orally every 12 h for 4 days) on the cortisol and testosterone concentrations were investigated in the eight healthy men. Similar to our study, it was concluded that a low dose of ciprofloxacin does not have any anti-androgenic side effects and testosterone concentration was not significantly different from the control group after the first and last doses of ciprofloxacin[17]. Furthermore, few *in vivo* studies described that ciprofloxacin is a proper drug without breakage or disruption in DNA[18].

In some studies, ciprofloxacin administration did not change the blood testosterone level. It may show this antibiotic inhibits mixed-function oxidase and affects activities of necessary enzymes for metabolism of some substrates such as theophylline and antipyrine, but the activities of responsible enzymes in synthesis of testosterone are not affected[17]. Conversely, some studies revealed that ciprofloxacin causes disorders in testicular function and structure in rats[19] that changes the level of serum testosterone[20]. Zobeiri *et al* described that ciprofloxacin decreases the steroidal activity of the Leydig cells and leads to an imbalance of gonadotropins and testosterone in blood[21]. Leydig cells synthesize testosterone and control physiological functions of Sertoli cells. Elias and Nelson showed ciprofloxacin decreases testis weight, sperm count and significantly reduces testosterone level according to dose and time in guinea pig[8]. Khaki *et al* showed oral usage of 12.5 mg/kg ciprofloxacin for 60 days significantly decreases spermatogenic cells, motility, viability, sperm concentration, and increases apoptosis of germ cells[9]. In addition, other parts of the genital system including epididymis, seminal vesicles and prostate can be damaged[22].

Production of peroxide radicals may be responsible for the disadvantage effects of ciprofloxacin in the testis[20]. *In vitro* genotoxicity investigations show this antibiotic breaks DNA single-strand and makes deviations in chromosome[23,24]. Ciprofloxacin increases apoptosis by activation of caspase-3[25,26]. Zobeiri *et al* reported that ciprofloxacin (206 mg/kg, *p.o.* for 45 consecutive days) causes severe damages to DNA of immature sperm cells and results in karyorrhexis, vacuolation of primary spermatocyte and degenerative alterations in germ cells. These injuries decrease fertility and embryonic development of testis[27].

The World Health Organization recognizes that CASA offers improved precision over manual methods and can now be applied to routine analyses such as sperm counts, provided both adequate quality control procedures and high measurement standards are followed[28]. Accordingly, in the present study, CASA was used for the evaluation of ciprofloxacin treatment effects on sperm quality parameters. In the present study, all sperm quality parameters evaluated by CASA significantly reduced 56 days after discontinuation of ciprofloxacin medication. This decline in quality was a result of disruption of the spermatogenesis process, which is clearly identified in the pathology assessment. There are several studies in the literature that confirm the results of our work[9,12,29,30]. Khaki *et al* observed that administration of ciprofloxacin at dose of 12.5 mg/kg for two months had a severe disruptive effect on sperm concentration, motility and viability in male rats[9]. In addition, ciprofloxacin can lead to the damage of DNA content of sperm head and subsequent reduction of embryonic quality[27]. It is also clearly shown that sperm DNA damage is correlated with poor sperm quality parameters[31].

In conclusion, we describe the pathological findings of ciprofloxacin in the present study, which are associated with testicular damages. Our results suggest potential testicular toxicity for ciprofloxacin.

Conflict of interest statement

All the authors declare that there is no conflict of interest.

Funding

This work was supported by Shahid Bahonar University of Kerman and funded by grant number (AZ-91-11).

Authors' contributions

Reza Kheirandish and Shahrzad Azizi conducted pathological evaluations; Ladan Emadi and Baharak Akhtardanesh made the study design and sperm quality assessment; Masoud Imani made revisions to the paper; Fatemeh Mahmoodabadi, Fereshteh Irani and Homa Shokrizadeh made practical procedure.

References

- [1] Vaziri MH, Sadighi Gilani MA, Kavousi A, Firoozeh M, Khani Jazani R, Vosough Taqi Dizaj A, et al. The relationship between occupation and semen quality. *Int J Fertil Steril* 2011; **5**: 66-71.
- [2] Najafi G, Nejati V, Shalizar Jalali A, Zahmatkesh E. Protective role of royal jelly in oxymetholone induced oxidative injury in mouse testis. *Iran J Toxicol* 2014; **8**: 1073-1080.
- [3] Choi GY, Cho JH, Jang JB, Lee KS. Effects of *Panax ginseng* on the sperm motility and spermatogenesis in the SD rat. *Korean J Orient Med* 2004; **25**: 90-94.
- [4] Lang T, Dechant M, Sanchez V, Wistuba J, Boiani M, Pilatz A, et al. Structural and functional integrity of spermatozoa is compromised as a consequence of acute uropathogenic *E. coli*-associated epididymitis. *Biol Reprod* 2013; **89**: 59.
- [5] Olayemi FO. A review on some causes of male infertility. *Afr J Biotechnol* 2010; **9**: 2834-3842.
- [6] Wang C, Yang L, Wang S, Zhang Z, Yu Y, Wang M, et al. The classic EDCs, phthalate esters and organochlorines, in relation to abnormal sperm quality: A systematic review with meta-analysis. *Sci Rep* 2016; **6**: 19982.
- [7] Shine R, Peek J, Birdsall M. Declining sperm quality in New Zealand over 20 years. *N Z Med J* 2008; **121**: 50-56.
- [8] Elias A, Nelson B. Toxicological effect of ciprofloxacin on testicular function of male guinea pigs. *Asian J Exp Biol Sci* 2012; **3**: 384-390.
- [9] Khaki A, Heidari M, Ghaffari Novin M, Khaki AA. Adverse effects of ciprofloxacin on testis apoptosis and sperm parameters in rats. *Iran J Reprod Med* 2008; **6**: 71-76.
- [10] El-Harouny MA, Zalata AA, Naser ME, El-Atta HMA, El-Shawaf IM, Mostafa T. Long-term ofloxacin testicular toxicity: An experimental study. *Andrologia* 2010; **42**: 92-96.
- [11] Maria PG, Marchini G. Clinical pharmacology of ciprofloxacin in

- neonates: Effects and pharmacokinetics. *Int J Pediatr* 2017; **5**: 5023-5041.
- [12] Abd-Allah AR, Aly HA, Moustafa AM, Abdel-Aziz AA, Hamada FM. Adverse testicular effects of some quinolone members in rats. *Pharm Res* 2000; **41**: 211-219.
- [13] Dohle GR, Elzanaty S, Van Casteren NJ. Testicular biopsy: Clinical practice and interpretation. *Asian J Androl* 2012; **14**: 88-93.
- [14] Sharma PC, Jain A, Jain S, Pahwa R, Yar MS. Ciprofloxacin: Review on developments in synthetic, analytical, and medicinal aspects. *J Enzym Inhib Med Chem* 2010; **25**: 577-589.
- [15] Shrinivas S, Revanasiddappa M. Analytical stability indicative method development and validation by high-pressure liquid chromatography for assay in ciprofloxacin hydrochloride drug substances. *Am J Analyt Chem* 2015; **6**: 719-730.
- [16] Childs S. Safety of the fluoroquinolone antibiotics: Focus on the molecular structure. *Infect Urol* 2000; **13**: 3-10.
- [17] Waite NM, Edwards DJ, Arnott WS, Warbasse LH. Effects of ciprofloxacin on testosterone and cortisol concentrations in healthy males. *Antimicrob Agents Chemother* 1989; **33**: 1875-1877.
- [18] Herbold BA, Brendler-Schwaab SY, Ahr HJ. Ciprofloxacin: *In vivo* genotoxicity studies. *Mutat Res* 2001; **498**: 193-205.
- [19] Demir A, Turker P, Sirvanci S, Onol FF, Demir A, Turker P, et al. The effects of acute epididymorchitis and ciprofloxacin treatment on testicular histomorphology and sperm parameters in rats. *Eur Urol* 2006; **5**: 214-241.
- [20] Weyer AI, Ugnia LI, Garcia Ovando H, Gorla NB. Ciprofloxacin increases hepatic and renal lipid hydroperoxides levels in mice. *Biocell* 2002; **26**: 225-228.
- [21] Zobeiri F, Sadrkhanlou RA, Salami S, Mardani K. Long-term effect of ciprofloxacin on testicular tissue: Evidence for biochemical and histochemical changes. *Int J Fertil Steril* 2013; **6**: 294-303.
- [22] Abu-Aita NA, Ahmed KA, Mouneir SM. The protective effect of ginger and *N*-acetyl cysteine on ciprofloxacin-induced reproductive toxicity in male rats. *J Am Sci* 2011; **7**: 741-752.
- [23] Sanchez G, Hidalgo ME, Vivanco JM, Escobar J. Induced and photoinduced DNA damage by quinolones: Ciprofloxacin, ofloxacin and nalidixic acid determined by comet assay. *Photoch Photobiol* 2005; **81**: 819-822.
- [24] Itoh T, Mitsumori K, Kawaguchi S, Sasaki YF. Genotoxic potential of quinolone antimicrobials in the *in vitro* comet assay and micronucleus test. *Mutat Res* 2006; **603**: 135-144.
- [25] Olivia A, Liping Z, Samir A, David PWJ, Tuan HK, Fazlul HS. Role of mitochondria in ciprofloxacin induced apoptosis in bladder cancer cell. *J Urol* 2002; **167**: 1288-1294.
- [26] Zhang JH, Zhang Y, Herman B. Caspases apoptosis and aging. *Ageing Res Rev* 2003; **2**: 357-366.
- [27] Zobeiri F, Sadrkhanlou RA, Salami S, Mardani K, Ahmadi A. The effect of ciprofloxacin on sperm DNA damage, fertility, potential and early embryonic development in NMRI mice. *Vet Res Forum* 2012; **3**: 131-135.
- [28] World Health Organization. Optional procedures. In: Cooper TG. (ed.) *WHO laboratory manual for the examination and processing of human semen*. Cambridge: World Health Organization; 2010, p. 136-137.
- [29] Vartan SV, Ali AH, Jawad AM. Effect of ciprofloxacin on semen analysis in human healthy volunteers. *Bas J Surg* 2003; **9**: 175-178.
- [30] Eskandari M, Ghalyanchi Langeroudi A, Zeighami H, Rostami A, Kazemi M, Eyni H, et al. Co-administration of ginseng and ciprofloxacin ameliorates epididymo-orchitis induced alterations in sperm quality and spermatogenic cells apoptosis following infection in rats. *Andrologia* 2016; **49**: 1-11.
- [31] Aydos OS, Yükselten Y, Kaplan F, Sunguroğlu A, Aydos K. Analysis of the correlation between sperm DNA integrity and conventional semen parameters in infertile men. *Turk J Urol* 2015; **41**(4): 191-197.