

Phosphodiesterase 8B Polymorphism rs4704397 Is Associated with Infertility in Subclinical Hypothyroid Females: A Case-Control Study

Tabassum Mansuri, M.Sc., Shahnawaz D. Jadeja, M.Sc., Mala Singh, Ph.D., Rasheedunnisa Begum, Ph.D., Pushpa Robin, Ph.D.*

Department of Biochemistry, Faculty of Science, The Maharaja Sayajirao University of Baroda, Gujarat, India

Abstract

Background: Subclinical hypothyroidism (SCH) remains largely unnoticed as a major cause of infertility due to asymptomatic. Polymorphisms of phosphodiesterase 8B gene (*PDE8B*) have been linked with various diseases, including female infertility. Hence, we aimed to study prevalence of SCH, in infertile females, explore association of *PDE8B* rs4704397 A/G and rs6885099 G/A polymorphisms with infertility in females suffering from SCH and genotype-phenotype correlation of the polymorphisms with thyroid stimulating hormone (TSH) levels in Gujarat population.

Materials and Methods: In this retrospective study, TSH level was estimated from plasma of 230 infertile and 100 control females by enzyme-linked fluorescence immunoassay (ELFA) to find out the prevalence of SCH. Further, based on TSH levels, thyroid function test (TFT) was performed in controls and infertile females with subclinical hypothyroidism (IF-SCH). *PDE8B* rs4704397 and rs6885099 polymorphisms were genotyped by PCR-RFLP and ARMS-PCR, respectively in 74 controls and 60 IF-SCH females.

Results: We observed i. significantly high prevalence of SCH (32%) in the infertile females, ii. significantly lower frequency of 'G' allele ($P=0.006$), while the frequency of 'A' allele ($P<0.0001$) was higher in IF-SCH females, compared to the controls, for rs4704397 A/G SNP, iii. no significant difference in the genotype ($P=0.214$; OR=2.51; CI=0.74–8.42) and the allele frequency ($P=0.129$; OR=1.51; CI=0.92–2.47) of rs6885099 G/A SNP, iv) low linkage disequilibrium for the polymorphisms, v. significantly higher frequency of 'AA' haplotype ($P=0.0001$; OR=3.84; CI=1.86–8.01), while the 'GG' haplotype ($P=0.0023$; OR=0.33; CI=0.16–0.69) was significantly lower in IF-SCH females and vi. no significant difference in the TSH level of IF-SCH females with respect to the genotypes.

Conclusion: The present study reports an association of *PDE8B* rs4704397 polymorphism with infertility in SCH females. The study categorically shows a higher prevalence of SCH in infertile females of Gujarat and advocates the importance of screening for SCH in infertility management.

Keywords: Genetic Polymorphisms, Infertility, Thyroid

Citation: Mansuri T, Jadeja Sh.D, Singh M, Begum R, Robin P. Phosphodiesterase 8B polymorphism rs4704397 is associated with infertility in subclinical hypo-thyroid females: a case-control study. *Int J Fertil Steril.* 2020; 14(2): 122-129. doi: 10.22074/ijfs.2020.6015. This open-access article has been published under the terms of the Creative Commons Attribution Non-Commercial 3.0 (CC BY-NC 3.0).

Introduction

Apart from its multiple functions, thyroid hormones play crucial role in reproduction. Hence, altered thyroid hormone levels can greatly affect reproductive function (1). Thyroid diseases in women with reproductive age are very common due to the complex interplay of various hormones (2). Abnormal thyroid functions of hyper or hypothyroidisms are symptomatic and they may have an adverse effect on the reproductive health contributing to infertility (3-4). However, subclinical hypothyroidism (SCH) is silent and hence it is often undiagnosed. It is a common thyroid disorder

often found to coexist with various other morbidities. It is an asymptomatic condition where the patient has a normal serum free T_4 (fT_4 /thyroxin) levels, but high thyroid stimulating hormone/thyrotropin (TSH) levels (5). TSH is considered as a sensitive indicator of the thyroid status and SCH. Normal TSH levels in serum are finely regulated in humans. Nevertheless, serum thyroid parameters show substantial inter-individual variability (6), in which genetic variations are proved as the major factors in several populations. It has been shown that altered TSH levels are related to genetic factors in up to 65% of the cases (7-9).

Received: 14/June/2019, Accepted: 22/December/2019

*Corresponding Address: Department of Biochemistry, Faculty of Science, The Maharaja Sayajirao University of Baroda, Vadodara-390002, Gujarat, India

Email: pushparobin@gmail.com



Royan Institute
International Journal of Fertility and Sterility
Vol 14, No 2, July-September 2020, Pages: 122-128

Different cohort studies reported phosphodiesterase 8B (*PDE8B*) as a genetic modulator of TSH levels. *PDE8B* gene encodes a cyclic adenosine monophosphate (cAMP) specific phosphodiesterase (PDE) enzyme (10). *PDE8B* affects cAMP levels in the thyroid gland resulting in changes in the levels of thyroid hormones, which in turn affects the release of TSH from the pituitary gland. *PDE8B* is mainly expressed in thyroid and brain (11, 12). Several single nucleotide polymorphisms (SNPs) for *PDE8B* have been demonstrated to associate with increased levels of serum TSH. More than 360,000 SNPs were tested for their associations with serum TSH levels with an additive model. The obtained results revealed three SNPs (i.e. rs4704397, rs6885099 and rs2046045) with genome-wide significance ($P < 10^{-10}$). These three SNPs were reported to be in strong linkage disequilibrium. Of the three SNPs, rs4704397 showed strongest association and it could explain 2.3% of the variations in TSH levels (13). *PDE8B* rs4704397 polymorphism has been found to associate with myocardial infarction, height (14), pregnancy (15, 16), recurrent miscarriage (17) and obesity in children (18), apart from thyroid function. Another *PDE8B* polymorphism, rs6885099 has also been shown to increase TSH levels, but to a lesser extent, in different populations (13). The relevance of human reproduction to PDE has been well-documented (19-22). While the underlying mechanism regulating oocyte maturation is not clearly known yet, the second messenger cyclic adenosine monophosphate (cAMP) role in oocyte maturation is well known (23) and thus research investigating the role of rs4704397 in the oocyte maturation might give an insight to primary infertility caused by hypothyroidism.

Numerous studies have reported the importance of screening for SCH, and the worldwide prevalence of SCH in infertile-females has been reported to be as high as 26.7% in various populations (24-27). In India, prevalence of SCH is high and reported to be 25% (28-33). However, there is no study on the status of SCH per se or its prevalence amongst infertile females in western part of India. Furthermore, there is no report on the role of *PDE8B* polymorphisms in female infertility. We therefore, aimed to estimate the prevalence of SCH in infertile females and explore association of *PDE8B* rs4704397 and rs6885099 polymorphisms in infertile females of Gujarat population.

Materials and Methods

Study subjects

The present retrospective study is a matched, case-control study. Two hundred and thirty infertile females were recruited from Dr. Mahesh Pandya's Ghanshyam Clinic (a fertility management center; Vadodara, India) along with 100 control females recruited from various health check-up camps. Random sampling method was

followed for selection of the groups. The study protocol was explained and informed consent was obtained from all participants of the study. Seventy four out of 230 infertile females were found to have (IF) for the TSH level with the inclusion criteria of primary infertility diagnosis and duration of more than one year of unprotected intercourse without pregnancy, while 76 out of 100 controls were found to be euthyroid (with normal thyroid hormone levels). Exclusion criteria were male factor infertility, any tubal anomaly congenital or urogenital tract anomaly and history of thyroid disease/medication/surgery.

For this study, IF-SCH females/case group are defined as the infertile females who have subclinical hypothyroidism with no other clinical difficulty. In addition, they should not be under any type of medication, including thyroid disorder. Whereas, the control group includes fertile, perous, healthy euthyroid females with no medical history for thyroid or any other disorder. Control group does not include any subclinical hypothyroid female.

Sample size for the present study was calculated using G-Power software with Alpha 0.05 and effect size of 0.9. The effect size was calculated based on the observed genotype frequencies (34).

Thyroid function test

Five ml blood samples was collected by venous puncture from fasting individuals and serum was separated for thyroid function test (TFT). Estimation of serum TSH, free T_3 (fT_3) and fT_4 were carried out by enzyme-linked fluorescence immunoassay (ELFA) on mini VIDAS® immuno-analyzer (BioMérieux India Pvt. Ltd., India). Females having TSH values between 3.5 and 10 μ IU/ml with normal fT_4 , along with an opinion from gynecologist and endocrinologist were considered as IF-SCH females. Fertile females having TSH values within the normal/euthyroid range (i.e. 0.35-3.5 μ IU/ml) and fT_4 levels within the normal range were included as controls in the present study. The reference range for serum thyroid hormones (fT_3 and fT_4) and TSH levels for different conditions are shown in Table S 1 (See Supplementary Online Information at www.celljournal.org). The confounding variables such as age, body mass index (BMI), smoking and hemoglobin (Hb) levels showed no significant difference between control and IF-SCH females (Table S2, See Supplementary Online Information at www.celljournal.org).

Genotyping *PDE8B* rs4704397 and rs6885099 polymorphisms

DNA was extracted from peripheral blood mononuclear cells (PBMCs) using 'IAamp DNA Blood Kit (QIAGEN Inc., USA) as per manufacturer's instructions. *PDE8B* rs4704397 A/G genotyping was done by polymerase chain reaction-restriction fragment length polymorphism

(PCR-RFLP) while *PDE8B* rs6885099 (G/A) genotyping was done by amplification refractory mutation system (ARMS)-PCR. Amplification was performed using Mastercycler Gradient PCR (Eppendorf, Germany) according to the following protocol: initial denaturation at 94°C for 10 minutes, followed by 30 cycles of denaturation at 94°C for 45 seconds, annealing at 60°C for 45 seconds and 72°C for 1 minute. The amplified products were analyzed by electrophoresis in a 2.0% agarose gel stained with ethidium bromide. The respective primers and restriction enzyme (RE) used for genotyping are shown in Table S3. 15 µl of the amplified products was digested for 16 hours at 37°C, using 1 U restriction enzyme. For PCR-RFLP based genotyping, the digested products (300 bp and 219 bp) with 100 bp DNA ladder (Bioron, Germany) were loaded in 3.5% agarose gels stained with ethidium bromide and visualized under UV transilluminator. Furthermore, genotyping of *PDE8B* rs6885099 G/A was done by Amplification refractory mutation system (ARMS-PCR) in 60 IF-SCH females and 76 control females. Human growth hormone (HGH) was used, as a reaction control in the ARMS-PCR (35). Amplification was performed using Mastercycler Gradient PCR according to the following protocol: initial denaturation at 94°C for 10 minutes, followed by 35 cycles of 94°C for 30 seconds, primer dependent annealing for 30 seconds and 60°C for 1 minute. The amplified products were analyzed by electrophoresis in a 3.5% agarose gel stained with ethidium bromide using 100 bp DNA ladder.

Statistical analysis

Hardy-Weinberg equilibrium (HWE) test was evaluated for the polymorphisms using chi-square test equating the observed and expected genotype frequencies. The genotype and allele risk associations were calculated by chi-square test using Prism 5 software (GraphPad Software Inc, USA; 2007). For genetic analysis, Bonferroni's correction was applied and statistical significance was considered at P-value less than 0.025. The linkage disequilibrium (LD) and haplotype analysis were carried out using <http://analysis.bio-x.cn/myAnalysis.php> (36). Levels of TSH and thyroid hormones were analyzed by non-parametric unpaired t-test and one-way ANOVA using Prism 5 software (GraphPad Software Inc.; 2007).

In-silico analysis

Web-based in-silico prediction tool HaploReg v4.1 (<https://www.pubs.broadinstitute.org/mammals/haploreg/haploreg.php>) was employed to predict the effect of non-coding rs4704397 polymorphism. Tissue specific effect of rs4704397 was assessed by an eQTL database-GTeX portal (<https://www.gtexportal.org>).

Ethical consideration

It was ensured that the study design complies with the ethical standards of the Institutional Ethical Committee for Human Research (IECHR), Faculty of Science, The Maharaja Sayajirao University of Baroda, Vadodara, Gu-

jarat, India (FS/IECHR/BC/PR/1) and with the 1964 Helsinki declaration.

Results

Estimation of thyroid stimulating hormone, free T3 and free T4 levels

Analysis of TSH, fT3 and fT4 levels in the studied subjects revealed that among 230 females with primary infertility, 58% (n=133) were euthyroid, 32% (n=74) were SCH, 6% (n=14) were overt hypothyroid and the rest 4% (n=9) females were hyperthyroidism (Fig.1 A, Table S3) (See Supplementary Online Information at www.cell-journal.org). IF-SCH females had significantly higher (P<0.0001; Fig.1B) TSH levels (mean ± SEM: 5.34 ± 0.21 µIU/ml) compared to the control females (mean ± SEM: 1.91 ± 0.08 µIU/ml) and they had no significant difference in fT3 levels (P=0.1159, mean ± SEM: 3.036 ± 0.0462pg/ml; Fig.1C) compared to the controls (mean ± SEM: 2.935 ± 0.0436). There was no significant difference between fT4 levels (P=0.0741, mean ± SEM: 1.22 ± 0.0249) in IF-SCH females compared to controls (mean ± SEM: 1.195±0.0318 ng/dl).

PDE8B rs4704397 SNP in infertile females with sub-clinical hypothyroidism females

Genotyping *PDE8B* rs4704397 polymorphism was carried out in 60 IF-SCH females and 76 healthy fertile females (Fig.2A). Other variables such as age (P=0.419), BMI (P=0.309), smokers (0%) and Hb (P=0.117) levels were not significantly different between the subjects of each genotypes (Table S4). The observed genotype frequencies of *PDE8B* rs4704397 SNP in IF-SCH females were slightly deviated from HWE (P=0.049; Table 1), whereas the control population was under HWE (P=0.062; Table 1). Ancestral allele 'A' and genotype 'AA' were considered as the reference allele and genotype respectively. The frequency of AG and GG genotypes were significantly lower in IF-SCH females, compared to controls (P=<0.0001 and P=0.006 respectively; Table 1). The frequency of 'G' allele was also significantly lower in IF-SCH females, compared to the control females (23% vs. 47%, P<0.0001, OR=0.34). Hence, "G" allele was identified to have a protective effect and 'A' allele was identified as the risk allele for SCH and infertility in females.

PDE8B rs6885099 SNP in infertile females with subclinical hypothyroidism

Genotyping of *PDE8B* rs6885099 polymorphism was carried out in 60 IF-SCH and 76 control females (Fig.2B). The observed genotype frequencies of *PDE8B* rs6885099 polymorphism among the control and IF-SCH females were in accordance with HWE (P=0.248 and P=0.134 respectively; Table 2). Distribution of genotype as well as allele frequencies revealed no significant difference among the IF-SCH and control females (Table 2).

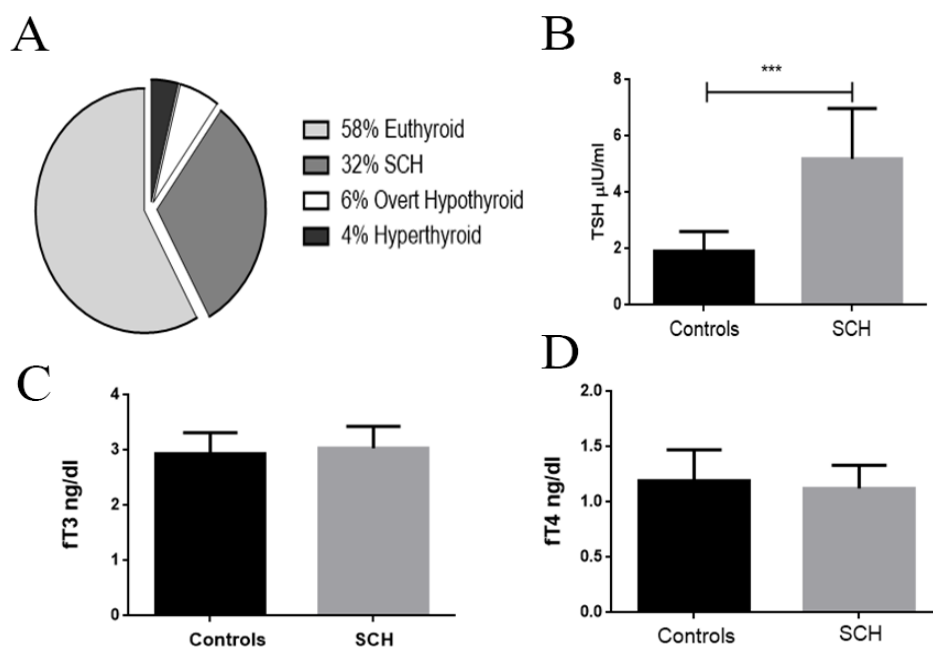


Fig.1: Estimation of TSH and thyroid hormone levels. **A.** Prevalence of thyroid dysfunction among the infertile females. **B.** TSH level in controls and IF-SCH females. **C.** ft_3 levels in the controls and IF-SCH females. **D.** ft_4 levels in controls and IF-SCH females. TSH; Thyroid stimulating hormone, IF-SCH; Infertile females with subclinical hypothyroidism, ft_3 ; Free T_3 , ft_4 ; and Free T_4 .

Table 1: Distribution of genotype and allele frequencies for PDE8B rs4704397 A/G polymorphism

Genotype or allele	IF-SCH females (Freq. %)	Control females (Freq. %)	P value	Odds Ratio	95% CI	P value HWE
Genotype	n= 60	n=76				
AA	38 (63%)	17 (22%)	R	1	0.07-0.35	0.062 (C)
AG	16 (27%)	46 (61%)	<0.0001 ^a	0.16	0.07-0.63	
GG	06 (10%)	13 (17%)	0.006 ^a	0.21		0.049 (P)
Allele						
A	92 (77%)	80 (53%)	R	1	-	
G	28 (23%)	72 (47%)	<0.0001 ^b	0.34	0.19-0.57	

n; number of IF-SCH females/control females, R; reference group, Freq.; Frequency, CI; Confidence interval, P; IF-SCH females, C; Control females, ^a IF-SCH female vs. control females (genotype) using chi-squared test with 2x2 contingency table, and ^b IF-SCH females vs. control females (allele) using chi-squared test with 2x2 contingency table, and IF-SCH; Infertile females with subclinical hypothyroidism.

Table 2: Distribution of genotypes and alleles for PDE8B rs6885099 G/A polymorphism

Genotype or allele	IF-SCH females (Freq. %)	Control females (Freq. %)	P value	Odds Ratio	95% CI	P value HWE
Genotype	n= 60	n=76				
GG	17 (28%)	32 (42%)	R	1	-	-
GA	35(58%)	38 (50%)	0.1914 ^a	1.73	0.82-3.65	0.248 (C)
AA	08 (13%)	06 (8%)	0.2145 ^a	2.51	0.74-8.42	
Allele						0.134 (P)
A	69 (58%)	102 (67%)	R	1	-	
G	51 (42%)	50 (33%)	0.1292 ^b	1.51	0.92-2.47	

F-SCH; Infertile females with subclinical hypothyroidism; n; number of IF-SCH females/Control females, R; reference group, Freq.; Frequency, CI; Confidence interval, P; IF-SCH females and C; Control females, ^a IF-SCH female vs. control females (genotype) using chi-squared test with 2x2 contingency table, and ^b IF-SCH females vs. control females (allele) using chisquared test with 2x2 contingency table.

Table 3: Distribution of haplotype frequencies for PDE8B rs4704397 and rs6885099 polymorphisms

Haplotype [rs4704397(A/G); rs6885099 (G/A)]	IF-SCH Female Freq. (%)	Control females Freq. (%)	P value for association	P value (Global)	Odds Ratio [95% CI]
AG	48 (46%)	49 (21%)	0.4434	7.5 × 10 ⁻⁵	1.230 [0.72-2.09]
AA	31 (30%)	12 (10%)	0.0001		3.84 [1.86-8.01]
GG	12 (12%)	34 (28%)	0.0023		0.33 [0.160-0.69]
GA	13 (12%)	25 (21)	0.0876		0.53 [0.25-11.10]

Freq.; Frequency, CI; Confidence interval (Frequency <0.03 in both control and case has been dropped and it was ignored in the analysis), and IF-SCH; Infertile females with subclinical hypothyroidism

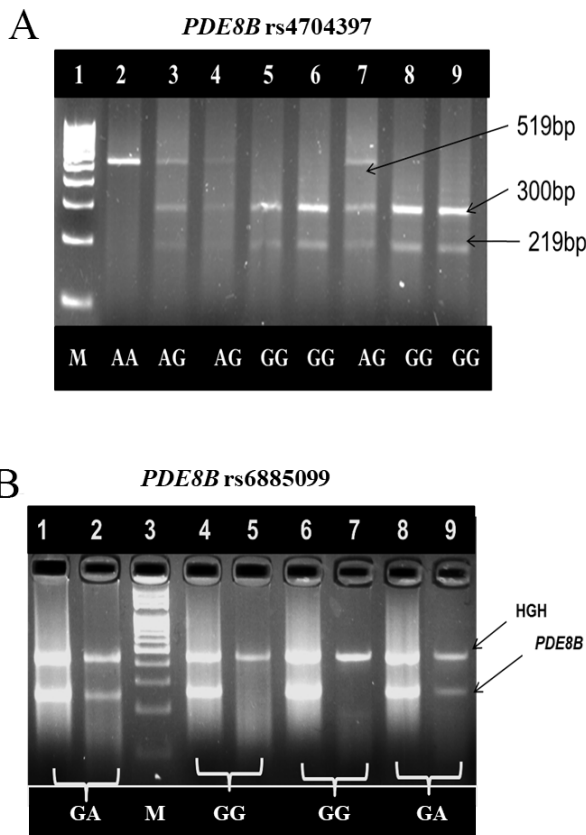


Fig. 2: Representative gel images for PDE8B rs4704397 and rs6885099 genotyping. **A.** PCR-RFLP analysis of PDE8B rs4704397 SNP on 3.5% agarose gel. Lane 1 shows 100 bp ladder, lane 2 shows homozygous (AA) genotype, lanes 3, 4 and 7 show heterozygous (AG) genotypes, lanes 5, 6, 8 and 9 show homozygous (GG) genotypes. **B.** ARMS-PCR analysis of PDE8B rs6885099 SNP on 3.5% agarose gel. Lanes 1 and 2 show homozygous (GA); lane 4, 5, 6 and 7 show homozygous (GG) genotypes and lane 3 shows 100 bp ladder, lanes 8 and 9 show heterozygous (GA) genotypes. PCR-RFLP; Polymerase chain reaction-restriction fragment length polymorphism.

Linkage disequilibrium and haplotype analysis

Linkage disequilibrium (LD) analysis revealed that two investigated PDE8B polymorphisms (i.e. rs4704397 and rs6885099) were in low LD association ($D' = 0.060$, $r^2 = 0.003$). Haplotype analysis revealed that the frequency of ‘AA’ haplotype was significantly higher in the patients and risk of IF-SCH females was increased by 3.84 fold ($P = 0.0001$, $OR = 3.84$; $CI = 1.86-8.01$; Table 3). The

frequency of ‘GG’ haplotype was significantly lower in IF-SCH females, compared to the controls suggesting its protective effect ($P = 0.0023$, $OR = 0.33$; $CI = 0.16-0.69$; Table 3).

Genotype-phenotype correlation analysis

TSH levels in IF-SCH females were analyzed with respect to the genotypes of PDE8B rs4704397 A/G and rs6885099 G/A. No significant difference in TSH levels was observed with respect to genotypes of the both SNPs (Fig.3).

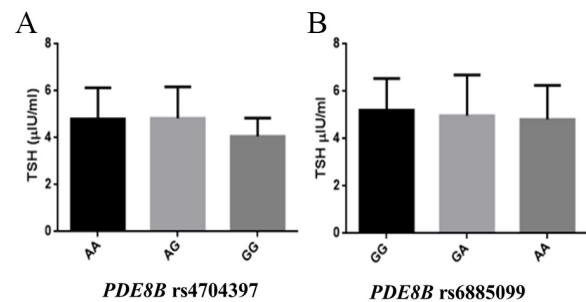


Fig. 3: Correlation of PDE8B rs4704397 and rs6885099 with TSH levels in IF-SCH females. No significant difference of TSH levels was observed with respect to PDE8B polymorphisms **A.** rs4704397 and **B.** rs6885099. TSH; Thyroid stimulating hormone, IF-SCH; Infertile females with subclinical hypothyroidism

In-silico analysis

Analysis of functional consequences of PDE8B rs4704397 by HaploReg v4.1 predicted that PDE8B rs4704397 could alter heat shock factor-type (HSF) motif and enhancer state by H3K27 acetylation (H3K27ac) in inferior temporal lobe of brain (https://www.pubs.broadinstitute.org/mammals/haploreg/detail_v4.1.php?query=&id=rs4704397). eQTL database GTEx portal showed significantly elevated PDE8B transcripts in thyroid tissue of individuals carrying ‘A’ allele, compared to ‘G’ allele (<https://www.gtexportal.org/home/snp/rs4704397>).

Discussion

The present study shows a high prevalence rate of SCH in infertile females (32%) in comparison with the healthy controls (Table S1) and the association of rs4704397 SNP with infertility in IF-SCH females of Gujarat region. In developing countries, one among four couples suffers from infertility and in these couples, hypothyroidism is one of the key perpetrators. In a study performed by Verma et al. (28), out of 394 infertile women, 23.9% were hypothyroid (TSH > 4.2 μ IU/ml). An intervention to rectify the hypothyroidism resulted in 76.6% of the conceived infertile women. Primary health caregivers most often pick up overt hypothyroidism easily; however, SCH with its subtle symptoms most often goes unnoticed. The prevalence of SCH amongst infertile females is common, but there is a scarcity on available data. However, there are a few studies reporting the prevalence of hypothyroidism, ranging from 15-25% in Indian population (28-33). As SCH is largely asymptomatic, it goes undiagnosed, resulting in infertility. It is essential to include evaluation of thyroid related hormones as a standard practice along with other tests to ascertain the causes of infertility.

SCH occurs due to multiple factors. Some of them include congenital agenesis, defect in synthesis due to iodine deficiency or anti-thyroid drugs, autoimmune diseases, post-surgery, hypopituitarism, TSH deficiency, environmental pollutants, mutations and SNPs (37). Of these factors, the present study focuses on the SNPs. To evaluate possible correlation between the polymorphisms associated with increased TSH levels and infertility, two SNPs (rs4704397 and rs6885099) of the *PDE8B* were studied in healthy controls and IF-SCH females. Higher frequency of the "A" allele for *PDE8B* rs4704397 polymorphism in SCH related infertile patients which revealed "A" as a risk allele for infertility in IF-SCH females. However, *PDE8B* rs6885099 was not associated with infertility. Earlier, *PDE8B* rs4704397 was also found to associate with recurrent miscarriage (17). *PDE8B* is found in the thyroid but not pituitary. In addition, given the importance of cAMP activity in TSH signaling, it is suggested that the *PDE8B* rs4704397 polymorphism could reduce cAMP levels in the thyroid resulting in a decreased response of thyroid gland to TSH stimulation, which leads to an increase of TSH set point for the same free T3 and T4 levels (18). Polymorphism in *PDE8B*, rs4704397 results in an increase in *PDE8B* enzyme expression. We propose that this could result in a faster degradation of cAMP, which decreases the synthesis and release of T3 and T4. In such a scenario, the negative inhibition of Thyrotropin-releasing hormone (TRH) will not take place and this will result in increased levels of TRH and hence TSH. As a consequence, T3 and T4 levels become normal. The increased level of TSH results in development of SCH. *PDE8B* rs4704397 polymorphism might induce phos-

phodiesterase activity in *PDE8B*, thereby reducing the ability of thyroid gland to generate free T4 when stimulated by TSH. This results in SCH, which can be the cause of infertility in IF-SCH patients. Arnaud et al. in a GWAS study reported that *PDE8B* rs4704397 could affect plasma TSH levels. Each copy of the minor allele "A" may lead to a mean increase of 0.13 mIU/l TSH levels (13). However, we did not observe significant correlation of the *PDE8B* rs4704397 SNP with circulating TSH levels. This might be due to the limited sample size in the present study. *PDE8B* rs4704397 SNP was also found to be associated with various conditions like cardiovascular, body height, pregnancy, recurrent miscarriage, obesity in children, etc. (14-18). Though the exact underlying mechanism of *PDE8B* rs4704397 SNP affecting TSH levels is not clear, in-silico tools predicted that this variation might lead to enhancement of *PDE8B* expression by influencing epigenetic level. The role of *PDE8B* in human placenta and ovaries is still to be understood, while human reproduction relevance to *PDE* has been proposed (19-22). The underlying mechanism of regulating oocyte maturation is not clearly documented yet, but the second messenger cAMP role in oocyte maturation is well known (23). Thus, investigating the role of rs4704397 in the oocyte maturation could be an interesting area of research as far as female infertility is concerned.

On the other hand, medications given to alter the levels of reproductive hormones have serious repercussions on the health of females with long-term implications (38). Treatment of infertility is usually done by direct targeting the reproductive system, instead of looking for the involvement of other factors, such as genetic polymorphisms, as a cause of infertility. This genetic approach could be used to identify IF-SCH patients and treat infertility with greater success and fewer side-effects without disturbing the reproductive system. Since, small sample size was a limiting factor for the present study, we suggest investigating larger number of infertile females in different populations. This might provide a significant insight into understanding the role of *PDE8B* in infertility.

Conclusion

The present study establishes an association of *PDE8B* rs4704397 with infertility in IF-SCH females and reiterates the importance of screening SCH, as a diagnostic tool in infertility management.

Acknowledgements

We thank all the subjects for their participation in the study. We thank Dr. Mahesh Pandya, Dr. Jigisha Pandya and Dr. Jignesh Pandya of Ghanshyam clinic (A Fertility management center), Vadodara, India for their whole-hearted support and permission to carry out counseling

and blood collection for recruitment of the subjects. This work financially was supported by a grant to Tabassum Mansuri from the Department of Science and Technology, Government of India, New Delhi under Women Scientist Scheme-A (SR /LS-117 /2012). The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

Authors' Contributions

T.M., P.R. R.B.; Contributed to conception and design. T.M.; Contributed to all experimental works and drafted the manuscript. T.M., S.D.J. and M.S.; Contributed to data collection, statistical analysis and interpretation of data. R.B. P.R.; Were responsible for overall supervision. All authors read and approved the final manuscript.

References

1. Jefferys A, Vanderpump M, Yasmin E. Thyroid dysfunction and reproductive health. *Obstet Gynaecol.* 2015; 17(1): 39-45.
2. Silva JF, Ocarino NM, Serakides R. Thyroid hormones and female reproduction. *Biol Reprod.* 2018; 99(5): 907-921.
3. Weiss RV, Clapauch R. Female infertility of endocrine origin. *Arq Bras Endocrinol Metab.* 2014; 58(2): 144-152.
4. Saran S, Gupta BS, Philip R, Singh KS, Bende SA, Agroiya P, et al. Effect of hypothyroidism on female reproductive hormones. *Indian J Endocrinol Metab.* 2016; 20(1): 108-113.
5. Stavreus Evers A. Paracrine interactions of thyroid hormones and thyroid stimulation hormone in the female reproductive tract have an impact on female fertility. *Front Endocrinol (Lausanne).* 2012; 3: 50.
6. Practice Committee of the American Society for Reproductive Medicine. Subclinical hypothyroidism in the infertile female population: a guideline. *Fertil Steril.* 2015; 104(3): 545-553.
7. Bernadette Biondi. The Normal TSH reference range: what has changed in the last decade? *J Clin Endocrinol Metab.* 2013; 98(9): 3584-3587.
8. Panicker V. Genetics of thyroid function and disease. *Clinical Biochem Rev.* 2011; 32(4): 165-175.
9. Malinowski JR, Denny JC, Bielinski SJ, Basford MA, Bradford Y, Peissig PL, et al. Genetic variants associated with serum thyroid stimulating hormone (TSH) levels in European Americans and African Americans from the eMERGE Network. *PLoS One.* 2014; 9(12): e111301.
10. Medici M, Visser WE, Visser TJ, Peeters RP. Genetic determination of the hypothalamic-pituitary-thyroid axis: where do we stand? *Endocr Rev.* 2015; 36(2): 214-244.
11. Vezzosi D, Bertherat J. Phosphodiesterases in endocrine physiology and disease. *European journal of endocrinology.* *Eur J Endocrinol.* 2011; 165(2): 177-188.
12. Lakics V, Karran EH, Boess FG. Quantitative comparison of phosphodiesterase mRNA distribution in human brain and peripheral tissues. *Neuropharmacology.* 2010; 59(6): 367-374.
13. Arnaud-Lopez L, Usala G, Ceresini G, Mitchell BD, Pilia MG, Piras MG, et al. Phosphodiesterase 8B gene variants are associated with serum TSH levels and thyroid function. *Am J Hum Genet.* 2008; 82(6): 1270-1280.
14. Jorde R, Schirmer H, Wilsgaard T, Joakimsen RM, Mathiesen EB, Njølstad I, et al. The phosphodiesterase 8B gene rs4704397 is associated with thyroid function, risk of myocardial infarction, and body height: the Tromsø study. *Thyroid.* 2014; 24(2): 215-222.
15. Shields BM, Freathy RM, Knight BA, Hill A, Weedon MN, Frayling TM, et al. Phosphodiesterase 8B gene polymorphism is associated with subclinical hypothyroidism in pregnancy. *J Clin Endocrinol Metab.* 2009; 94(11): 4608-4612
16. Yang S, Tao J, Zhang J, Fan J, Qian W, Shu K. Genetic association study of phosphodiesterase 8B gene with subclinical hypothyroidism in pregnant women. *Endocrine Res.* 2015; 40(4): 199-203.
17. Granfors M, Karypidis H, Hosseini F, Skjöldebrand-Sparre L, Stavreus-Evers A, Bremme K, et al. Phosphodiesterase 8B gene polymorphism in women with recurrent miscarriage: a retrospective case control study. *BMC Med Genet.* 2012; 13: 121.
18. Grandone A, Perrone L, Cirillo G, Di Sessa A, Corona AM, Amato A, et al. Impact of phosphodiesterase 8B gene rs4704397 variation on thyroid homeostasis in childhood obesity. *European J Endocrinol.* 2012; 166(2): 255-260.
19. Hayashi M, Shimada Y, Nishimura Y, Hama T, Tanaka T. Genomic organization, chromosomal localization, and alternative splicing of the human phosphodiesterase 8B gene. *Biochem Biophys Res Commun.* 2002; 297(5): 1253-1258.
20. Soderling SH, Bayuga SJ, Beavo JA. Cloning and characterization of a cAMP-specific cyclic nucleotide phosphodiesterase. *Proc Natl Acad Sci USA.* 1998; 95(15): 8991-8996.
21. Gamanuma M, Yuasa K, Sasaki T, Sakurai N, Kotera J, Omori K. Comparison of enzymatic characterization and gene organization of cyclic nucleotide phosphodiesterase 8 family in humans. *Cell Signal.* 2003; 15(6): 565-574.
22. Horvath A, Giatzakis C, Tsang K, Greene E, Osorio P, Boikos S. A cAMP-specific phosphodiesterase (PDE8B) that is mutated in adrenal hyperplasia is expressed widely in human and mouse tissues: a novel PDE8B isoform in human adrenal cortex. *Eur J Hum Genet.* 2008; 16(10): 1245-1253.
23. Shu YM, Zeng HT, Ren Z, Zhuang GL, Liang XY, Shen HW, et al. Effects of cilostamide and forskolin on the meiotic resumption and embryonic development of immature human oocytes. *Hum Reprod.* 2008; 23(3): 504-513.
24. Papi G, degli Uberti E, Betterle C, Carani C, Pearce EN, Braverman LE, et al. Subclinical hypothyroidism. *Curr Opin Endocrinol Diabetes Obes.* 2007; 14(3): 197-208.
25. Orouji Jokar T, Fourman LT, Lee H, Mentzinger K, Fazeli PK. Higher TSH levels within the normal range are associated with unexplained infertility. *J Clin Endocrinol Metab.* 2017; 103(2): 632-639.
26. Deeba F, Fatima P, Banu J, Ishrat S, Begum N, Anwary SA. Thyroid status and treatment response of hypothyroid infertile women in tertiary care center of bangladesh. *Bangladesh J Obstet Gynaecol.* 2016; 31(2): 86-89.
27. Feldthusen AD, Pedersen PL, Larsen J, Toft Kristensen T, Ellervik C, Kvetny J. Impaired fertility associated with subclinical hypothyroidism and thyroid autoimmunity: the Danish general suburban population study. *J Pregnancy.* 2015; 2015: 132718.
28. Verma I, Sood R, Juneja S, Kaur S. Prevalence of hypothyroidism in infertile women and evaluation of response of treatment for hypothyroidism on infertility. *Int J Appl Basic Med Res.* 2012; 2(1): 17-19.
29. Priya DM, Akhtar N, Ahmad J. Prevalence of hypothyroidism in infertile women and evaluation of response of treatment for hypothyroidism on infertility. *Indian J Endocrinol Metab.* 2015; 19(4): 504-506.
30. Pushpagiri N, Gracelyn LJ, Nagalingam S. Prevalence of subclinical and overt hypothyroidism in infertile women. *Int J Reprod Contraception Obstet Gynecol.* 2015; 4(6): 1733-1738.
31. Bharti G, Singh K, Kumari R, Kumar U. Prevalence of hypothyroidism in subfertile women in a tertiary care centre in North India. *Int J Res Med Sci.* 2017; 5(5): 1777-1780.
32. Malaiarasi N, Santhanalakshmi L. The association of thyroid dysfunctions with infertility in females. *Int J Adv Res.* 2016; 4(7): 1017-1024.
33. Abdul R, Seema M. Effect of clinical/sub-clinical hypothyroidism on fertility in infertility case and the response of treatment for hypothyroidism on fertility in cases of infertility. *IOSR Journal of Dental and Medical Sciences (IOSR-JDMS).* 2015; 14(2): 5-8.
34. Faul F, Erdfelder E, Lang AG, Buchner A. G*power 3: a flexible statistical power analysis program for the social, behavioral, and biomedical sciences. *Behav Res Methods.* 2007; 39(2): 175-191.
35. Jadeja SD, Mansuri MS, Singh M, Dwivedi M, Laddha NC,

- Begum R. A case-control study on association of proteasome subunit beta 8 (PSMB8) and transporter associated with antigen processing 1 (TAP1) polymorphisms and their transcript levels in vitiligo from Gujarat. *PLoS One*. 2017; 12(7): e0180958.
36. Barrett JC, Fry B, Maller JD, Daly MJ. Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics*. 2004; 21(2): 263-265.
37. Biondi B, Cooper DS. The clinical significance of subclinical thyroid dysfunction. *Endocr Rev*. 2008; 29(1): 76-131.
38. Reigstad MM, Storeng R, Myklebust TÅ, Oldereid NB, Omland AK, Rødsahl TE, et al. Cancer risk in women treated with fertility drugs according to parity status-a registry-based cohort study. *Cancer Epidemiol Biomarkers Prev*. 2017; 26 (6): 953-962.
-