

Review

Tannins as Natural Helpers in Reinforcement of Human Health

Neli Vilhelmova-Ilieva*, Angel S. Galabov

Department of Virology, The Stephan Angeloff Institute of Microbiology, Bulgarian Academy of Sciences, Sofia, Bulgaria

Abstract

Since ancient times, folk medicine has used the potency of a wide range of plants for treatment of various diseases. In recent years, the development of technology has made it possible to identify the active ingredients determining these properties of plants. One of the largest group of substances isolated from plants are polyphenols. Tannins are a type of polyphenols that have already proven to possess different biological activities important to human health, such as antioxidant, antitumoral, antimicrobial, antiviral, etc.. As an integral part of many food products, tannins are included in the daily diet of many people suffering from various diseases. Tannins can naturally enhance the body's ability to cope with various infections and diseases, improving the immune response and also having a positive effect on patients with heart disease and high blood pressure, which makes them promising therapeutic agents. All of the activities described above allow tannins to be considered as promising therapeutic agents.

Keywords: tannins, ellagitannins, plant substances, biological activities, antiviral effects, therapeutic agents

Резюме

От древни времена народната медицина използва ефективността на голям асортимент от растения за лечение на различни заболявания. През последните години развитието на технологиите направи възможно идентифицирането на активните съставки, които определят тези свойства на растенията. Една от най-големите групи вещества, изолирани от растения, са полифенолите. Танините са вид полифеноли, вече доказали различни свои биологични активности важни за човешкото здраве, като антиоксиданта, антитуморна, антимикробна, антивирусна и други. Като неделима част от много хранителни продукти танините са включени в ежедневната диета на много хора, страдащи от различни заболявания. Танините подобряват способността на организма да се справя с различни инфекции и заболявания, усилвайки имунния отговор. Също така те имат положителен ефект върху сърдечно-съдовите заболявания и високото кръвно налягане. Описаните активности позволяват танините да се смятат за обещаващи лечебни средства.

Introduction

Tannins are a group of polyphenols with molecular weight ranging between 500 and 3000. They are soluble in water and alcohols and are located in the root, bark, stem and outer layers of the plant tissue. They form complexes with proteins, carbohydrates, alkaloids and gelatin (Widsten *et al.*, 2014). Tannins are divided into two groups: hydrolysable, which can be hydrolyzed to glucose and gallic acid or other polyphenolic acids, and condensed tannins (proanthocyanidins), composed of flavonoid monomers, mostly (+) catechin or (-) epikatehin

connected by carbon-carbon bonds (Okuda *et al.*, 1991). Hydrolysable tannins can be subdivided into gallotannins or ellagitannins, depending on whether they contain gallic or ellagic acid in their structure. Ellagitannins are made up of two galloyl units connected together and forming the basic structural unit of ellagitannins – hexahydroxydifenic acid (HHDP). They are important secondary metabolites of plants because of their property to induce resistance against microorganisms due to their ability to bind with proteins and polysaccharides, there-

by blocking their growth.

Many tannins are isolated and structurally characterized from plant species used in traditional medicine. Grape, strawberry, cranberry, blueberry, pomegranate, chestnut, walnut, Brazil nut, pecan nut are with the highest content of tannins (Ascacio-Valdés *et al.*, 2011).

This group of plant products has attracted the attention of an increasing number of research teams due to their multifunctional properties that make them promising medicinals.

Antioxidant activity

Free radicals, which are present in the human body, are considered to be responsible for the appearance of more than 100 chronic disorders in humans, including atherosclerosis, arthritis, jaundice, liver damage, central nervous system injury, gastritis, neoplasms, etc. The reasons for their formation are different: environmental pollutants, radiation, chemicals, toxins, fried and spicy foods and physical stress can cause depletion of antioxidants in the immune system, changes in gene expression and production of abnormal proteins. The struggle with degenerative diseases requires increased levels of antioxidants in the body. Foods of plant origin usually contain natural antioxidants. The role of antioxidants is to inactivate reactive oxygen species, and thus slow down or prevent oxidative damage (Jain *et al.*, 2015).

Tannins have the ability to prevent the formation of free radicals, as well as to reduce the damage they cause in the cells (Yokozawa *et al.*, 2002; Sepúlveda *et al.*, 2011). This effect is stronger when tannins contain a greater number of phenolic hydroxyl groups (Hatano *et al.*, 1989). Products derived from the metabolism of tannins also possess antioxidant properties (Ito, 2011).

Antidiabetic activity

According to the World Health Organization (WHO), about 80% of the population uses herbal medicines of traditional medicine for treatment of various diseases (Sani, 2015).

Extracts from various plant species with proved high content of tannins show antidiabetic activity (Rao *et al.*, 2003; Raut and Gaikwad, 2006). C-glycosidic ellagitannins lagerstroemin, flosin B and reginin A from *Lagerstroemia speciosa* used for treatment of diabetes in the Philippines are activators of glucose transport (Yoshida *et al.*, 2010).

Antihypertensive activity

The treatment of hypertension includes many drugs of natural origin, as digitoxin from *Digitalis*

purpurea (foxglove), reserpine from *Rauwolfia serpentina* (snakeroot), aspirin from *Salix alba* (willow bark), tetramethylpyrazine, also known as Ligustrazine, from *Jatropha podagrica*, and tetrandrine from *Stephania tetradra* (Kwan, 1994; O’Kane *et al.*, 2003; Iwalokun *et al.*, 2011).

Along with other representatives of natural origin, certain types of tannins also show a positive effect against hypertension. Castalagin, chebulinic acid and corilagin isolated from the leaves of *Lumnitzera racemosa* are hydrolysable tannins with pronounced antihypertensive activity (Lin *et al.*, 1993).

Antitumor activity

One of the most remarkable abilities of tannins is their activity against various tumors as cervical cancer, prostate cancer, malignant cells in skin, breast, stomach, lung, esophagus, liver, etc. (Barrajon-Catalan *et al.*, 2010; Ascacio-Valdés *et al.*, 2011; Yildirim and Kutlu, 2015).

Ellagitannins possess the ability to bind to proteins located on the surface of the cell membrane thus preventing the proliferation of metastatic cells. They can induce apoptosis in tumor cells by inhibiting the factors responsible for the formation of metastases. Another mechanism suggests that during DNA replication ellagitannins bind to carcinogens thus preventing their mutation effect (Sepúlveda *et al.*, 2011). Corilagin induces cell cycle arrest at the G2/M phase and the apoptosis in cancer cells lines of ovarian cancer SKOv3ip, Hey and HO-8910PM (Jia *et al.*, 2013). In literature there are many data on ellagitannins with antitumor activity – geraniin, corilagin (Okabe *et al.*, 2001), oenothelin A and B, woodfordin C, D and F (Miyamoto *et al.*, 1997). Extracts containing tannins induce apoptotic activity in cells of breast and prostate cancer (Bawadi *et al.*, 2005).

There is evidence that ellagitannins reduce the negative effects of chemotherapy and mitigate the effects of radiation exposure in anticancer therapy (Varadkar and Dubey, 2001).

Cell proliferation and differentiation activity

Geraniin and furosin isolated from *Phyllanthus muellerianus* stimulated the proliferation and differentiation of human keratinocytes and dermal fibroblasts, as well as the biosynthesis of collagen (Agyare *et al.*, 2011).

Immunomodulatory activities

Ellagitannins show immunomodulatory activities using different mechanisms as a promoter of the formation of catechin-polysaccharide complex that is a potential immunostimulant (Monobe *et al.*,

2008); enhance the functionality of macrophages (Ushio et al., 1991); for release of nitric oxide (NO), tumour necrosis factor (TNF) and interferon (IFN) (Kolodziej and Kiderlen, 2005); stimulate the secretion of cytokines IL-1, IL- β 2 and α TNF- by human peripheral mononuclear cells (Wang et al., 2002).

Enzyme inhibitory activity

Representatives of this group of compounds are also effective inhibitors of certain enzymes. Woodfruticosin (woodfordin C) has anti-topoisomerase II activity; eugeniflorin D1 and D2 isolated from *Eugenia uniflora* L. and oenothien B effectively inhibit Epstein-Barr virus (EBV) DNA polymerase. Enzymes 5 α -reductase and aromatase have an important role in the development of benign prostatic hyperplasia, whose potential inhibitors are oenothien A and B isolated from *Epilobium* species.

It has been suggested that an important factor in the expression of genes, DNA replication and differentiation of cells is the enzyme poly(ADP-ribose) glycohydrolase that can be inhibited by oligomeric ellagitannins oenothien B and nobotanin B, E and K. The enzyme α -glucosidase (maltase), possibly important in the development of type-2 diabetes, is inhibited by chebulagic acid (isolated from *Terminalia chebula*), tellimagrandin I and eugeniin (casuarictin) (Yoshida et al., 2010).

Antimicrobial activity

Many tannins exhibit activity against various bacteria and fungi. Three galotannins: penta-, hexa-, and hepta-O-galloylglucose, isolated from mango (*Mangifera indica* L.), prevent the proliferation of Gram-positive food spoilage bacteria, and reduce the growth of Gram-negative *Escherichia coli* as the most likely inhibitory effects of these hydrolyzable tannins due to their iron-complexing properties (Engels et al., 2009). Tannins isolated from *Solanum trilobatum* Linn show antibacterial activity against *Staphylococcus aureus*, *Streptococcus pyogenes*, *Salmonella typhi*, *Pseudomonas aeruginosa*, *Proteus vulgaris* and *E. coli* (Doss et al., 2009).

The growth of *E. coli*, *Candida albicans*, *Cryptococcus neoformans* bacteria and *Aspergillus fumigatus* fungus is affected by the action of pomegranate, punicalagin, punicalin, gallagic and ellagic acids. Derivatives of ellagic acid demonstrated activity against *Klebsiella pneumoniae*, *Bacillus cereus*, *S. typhi* and *S. pyogenes* (Ascacio-Valdés et al., 2011).

Antiparasitic activity

Ellagitannins manifested antiparasitic activity against intracellular amastigotes of *Leishmania Donovan* (Kolodziej et al., 2001) or direct toxicity for extracellular promastigote *Leishmania donovani* or *L. major* strains (Kolodziej and Kiderlen, 2005). It has been shown that when administered via the feed of the plants rich in tannins they inhibit the development of the various stages of gastrointestinal nematodes of ruminants (Hoste et al., 2012).

Antiviral activity

In recent years, antiviral activity of various tannins has been increasingly often reported. It is a potential inhibitor of various enveloped viruses due to its capability of binding with different proteins and altering their structure, thus inactivating them.

Many studies have been conducted on tannins against the replication of human immunodeficiency virus (HIV) and the results of the various teams indicate that the targets of action of tannins in replicative cycle of HIV are several. There is evidence of ellagitannins that suppressed HIV replication by inhibiting reverse transcriptase (Asanaka et al., 1988; Notka et al., 2004). Other authors reported on ellagitannins (geranin and corilagin) that reduce HIV replication by inhibiting the HIV-1 protease and HIV-1 integrase enzyme (Notka et al., 2004), and ellagitannins isolated from *Tuberaria lignosa* that inhibit the entry of HIV in MT-2 cells (Bedoya et al., 2010).

The replication of human, porcine and duck influenza A virus *in vitro* is prevented by hydrolysable tannin strictinin (Saha et al., 2010).

In herpesvirus infections, chemotherapy was developed based on the application of nucleoside analogues that reduce the duration of symptoms and lead to faster healing of the lesions. The most commonly used nucleoside is acyclovir (ACV). The disadvantage of nucleoside analogues is the relatively rapid selection of resistant mutants (Abraham et al., 2007). Patients with compromised immune systems, and especially HIV- positive ones, are at the highest risk (Ziyaeyan et al., 2007).

Due to the frequent failure of treatment with nucleoside analogues, there is a need for the development of new inhibitors of herpes viruses (Knickelbein et al., 2009).

For hundreds of years, man has used natural sources for the treatment of various diseases. Fractions and crude extracts, and isolated and purified compounds from them were tested for antiviral activity against many viruses, including herpes

(Hassan *et al.*, 2015). A significant part of these extracts showed remarkable anti-herpesvirus activity. Grape, apple, strawberry juices and extract of *Azadirachta indica* possess anti-HSV activities. The aqueous extract of *Carissa edulis* root from Kenya shows *in vitro* and *in vivo* antiviral activity against acyclovir (ACV) sensitive and ACV resistant strain of HSV-1 and HSV-2. The extract of *Ceratostigma willmattianum* demonstrates suppression of viral adsorption, replication and transcription of HSV-1 and HSV-2 (Chattopadhyay *et al.*, 2010). Essential oils of ginger, thyme, hyssop and sandalwood effectively inhibit drug-resistant clinical HSV-1 strain (Schnitzler *et al.*, 2007). Extracts of *Cardamine angulata*, *Conocephalum conicum*, *Lysichiton americanum*, *Polypodium glycyrrhiza* and *Verbascum thapsus* exhibit antiviral activity against HSV (Jassim and Naji, 2003), extracts of the Taiwanese folk remedy *Boussingaultia gracilis* and *Serissa japonica* and the extract of *Senna petersiana* are used for sexually transmitted diseases (Chattopadhyay *et al.*, 2010). The methanolic crude extract of *Malolotus peltatus* possesses weak anti-HSV activity (Bag *et al.*, 2012). *Eucalyptus globulus* is a traditional herb that has been used in Iran for many years. The methanolic extract of this plant inhibits HSV-1 replication in cell cultures in various dilutions (Davood *et al.*, 2012). Acetone, ethanol and methanol extracts of *Phyllanthus urinaria* inhibit HSV-2 infection *in vitro* (Yang *et al.*, 2005). An extract of *Ribes nigrum* L., known as blackcurrant in Europe and Kurokarin® in Japan, inhibits HSV-1 attachment to the cell membrane and inhibits virus replication of HSV-1, HSV-2 and VZV by suppression of protein synthesis in infected cells in the early stage of infection (Suzutani *et al.*, 2003). The ethanolic extract of *Rheum officinale* and methanol extract of *Paeonia suffruticosa* inhibit the attachment and penetration of HSV-1, and other studies show that the aqueous extract of *P. suffruticosa* and ethanolic extract of *Melia toosendan* affect the attachment and replication of HSV -1 and HSV-2. A garlic extract and an extract of *Terminalia chebula* showed anti-HCMV activity (Chattopadhyay *et al.*, 2010), as well as extracts of *Euphorbia australis* and *Scaevola spinescens* (Jassim and Naji, 2003).

Various tannins were tested for anti-herpesviral activity. Seven ellagitannins isolated from *Phyllanthus myrtifolius* and *P. urinary*, and eugeniflorin D (1) and D (2) isolated from *Eugenia uniflora* L. are active against DNA polymerase of EBV (Lee *et al.*, 2000). Ellagitannin geraniin possesses a virucidal effect against herpesviruses (Notka *et al.*, 2004).

Chebulagic acid and punicalagin – two hydrolysable tannins isolated from *Terminalia chebula* Retz. inactivated HSV-1 entry and the cell-to-cell spread as their targets are HSV-1 glycoproteins (Lin *et al.*, 2011). Hydrolyzable tannin casuarinin from *Terminalia arjuna* Linn prevent the attachment of HSV-2 in the cell, and also violates the late stages of infection (Cheng *et al.*, 2002). Eugenin (from *Geum japonicum* and *Syzygium aromaticum*) showed a significant inhibitory effect on the activity of DNA polymerase of HSV-1 (Kurokawa *et al.*, 1998).

Our research in this area is mainly related to a substance that belongs to the group of nonahydroxyterphenol-bearing C-glucosidic ellagitannins. This substance is castalagin and is extracted from powdered pedunculate oak (*i.e.*, *Quercus robur*). It provides an activity against the replication of HSV-1 greater than that of acyclovir and well-defined, but relatively lower activity against HSV-2 replication (Vilhelmova *et al.*, 2011). Successfully combined with acyclovir, castalagin exerts a strong synergistic effect (Vilhelmova *et al.*, 2011). In addition, castalagin shows clear activity against strains of HSV-1 and HSV-2 resistant to acyclovir and its combination with acyclovir has a clear synergistic effect (Vilhelmova-Ilieva *et al.*, 2014).

We have also studied the two substances vesicalagin and grandinin from the same group, whose activity is not as strong as that of castalagin, but is significant and selective. They also demonstrate a synergistic effect when combined with ACV as in sensitive and in resistant to ACV strains of HSV -1 and HSV-2 (Vilhelmova *et al.*, 2011; Vilhelmova-Ilieva *et al.*, 2014).

Conclusion

Plant species from almost all plant families are at the heart of alternative medicine in different countries. Roots, stems, bark, leaves, flowers, fruits and grains possess medicinal properties, from which substances with different structure are isolated, an important part of which is occupied by tannins. They possess various biological activities, including anti-herpetic activity results in a damage at different stages of the virus replicative cycle. Further research remains to be conducted to discover new active substances, as well to establish the exact mechanism of action and the optimal concentrations in which active ingredients can be applied. With its diversity Nature provides a vast world in which future antiviral agents are hiding.

References

- Abraham, A. M., S. Kavitha, P. Joseph, R. George, D. Pillay, J. Malathi, M. V. Jesudason, G. Sridharan (2007). Aciclovir resistance among indian strains of herpes simplex virus as determined using a dye uptake assay. *Indian J. Med. Microbiol.* **25**: 260-262.
- Agyare, C., M. Lechtenberga, A. Detersa, F. Petereita, A. Hensel (2011). Ellagitannins from *Phyllanthus muellerianus* (Kuntze) Exell.: Geraniin and furosin stimulate cellular activity, differentiation and collagen synthesis of human skin keratinocytes and dermal fibroblasts. *Phytomedicine* **18**: 617-624.
- Asanaka, M., T. Kurimura, R. Kobayashi, T. Okuda, M. Mori, H. Yokoi (1988). Fourth International Conference on Immunopharmacol. Osaka, Japan, 1988. Abstr. p. 47.
- Ascacio-Valdés, J., J. J. Buenrostro-Figueroa, A. Aguilera-Carbo, A. Prado-Barragán, R. Rodríguez-Herrera, C. N. Aguilar (2011). Ellagitannins: biosynthesis, biodegradation and biological properties. *J. Med. Plant Res.* **5**: 4696-4703.
- Bag, P., D. Chattopadhyay, H. Mukherjee, D. Ojha, N. Mandal, M. C. Sarkar, T. Chatterjee, G. Das, S. Chakraborti (201). Anti-herpes virus activities of bioactive fraction and isolated pure constituent of *Mallotus peltatus*: an ethnomedicine from Andaman Islands. *Virology J.* **9**: 98.
- Barrajon-Catalan, E., S. Fernandez-Arroyo, D. Saura, E. Guillen, A. Fernandez-Gutierrez, A. Segura-Carretero, V. Micol (2010). Cistaceae aqueous extracts containing ellagitannins show antioxidant and antimicrobial capacity and cytotoxic activity against human cancer cells. *Food Chem. Toxicol.* **48**: 2273-2282.
- Bawadi, H. A., R. R. Bansode, A. Trappey II, R. E. Truax, J. N. Losso (2005). Inhibition of Caco-2 colon, MCF-7 and Hs578T breast and DU 145 prostatic cancer cell proliferation by water-soluble black bean condensed tannins. *Cancer Lett.* **218**: 153-162.
- Bedoyaa, L. M., M. J. Abadb, S. Sánchez-Palominoa, J. Alcamia, P. Bermejob (2010). Ellagitannins from *Tuberaria lignosa* as entry inhibitors of HIV. *Phytomedicine* **17**: 69-74.
- Cheng, H. Y., Lin, I. I., Lin, T. I. (2002). Antiherpes simplex virus type 2 activity of casuarinin from the bark of *Terminalia arjuna* Linn. *Antiviral Res.* **55**: 447-455.
- Chattopadhyay, D., S. Das, S. Chakraborty, S. K. Bhattacharya (2010). Ethnomedicines for the development of anti-herpesvirus agents, in: Chattopadhyay, D. (Ed.), *Ethnomedicine: A Source of Complementary Therapeutics*, Research Signpost, Kerala, India, pp, 117-147.
- Davood, A. A., Z. M. Javad, A. Alimohammad, A. A. Abbas, M. Hamidreza (2012). Evaluation effect of hydroalcoholic extract of *Eucalyptus globules* and *Artemisia dracunculul* compare with acyclovir against herpes simplex virus type 1. *Med. Plant Res.* **2**: 6-10.
- Doss, A., H. M. Mubarak, R. Dhanabalan (2009). Antibacterial activity of tannins from the leaves of *Solanum trilobatum* Linn. *Ind. J. Sci. Technol.* **2**: 41-43.
- Engels, C., M. Knödler, Y.-Y. Zhao, R. Carle, M. G. Gänzle, A. Schieber (2009). Antimicrobial activity of gallotannins isolated from mango (*Mangifera indica* L.) *Kernels. J. Agric. Food Chem.* **57**: 7712-7718.
- Fry, J. (1975). Deaths and complications from hypertension. *J. R. Coll. Gen. Pract.* **25**: 489-494.
- Hassan, S. T., R. Masarčíková, K. Berchová. (2015). Bioactive natural products with anti-herpes simplex virus properties. *J. Pharm. Pharmacol.* **67**: 1325-1336.
- Hatano, T., R. Edamatsu, M. Hiramatsu, A. Mori, Y. Fujita, T. Yasuhara, T. Yoshida, T. Okuda (1989). Effects of tannins and related polyphenols on superoxide anion radical, and on 1,1-diphenyl-2-picrylhydrazyl radical. *Chem. Pharm. Bul.* **37**: 2016-2021.
- Hoste, H., C. Martinez-Ortiz-De-Montellano, F. Manolaraki, S. Brunet, N. Ojeda-Robertos, I. Fourquaux, J. F. J. Torres-Acosta, C. A. Sandoval-Castro (2012). Direct and indirect effects of bioactive tannin-rich tropical and temperate legumes against nematode infections. *Vet. Parasitol.* **186**: 18-27.
- Ito, H. (2011). Metabolites of the ellagitannin geraniin and their antioxidant activities. *Planta Med.* **77**: 1110-1115.
- Iwalokun, B. A., S. A. Hodonu, S. Nwoke, O. Ojo, P. U. Ago-mo (2011). Evaluation of the possible mechanisms of anti-hypertensive activity of *Loranthus micranthus*: an African mistletoe. *Biochem. Research. Int.* Article ID 159439.
- Jain S, A. Jain, S. Jain, N. Malviya, V. Jain, D. Kumar (2015). Estimation of total phenolic, tannins, and flavonoid contents and antioxidant activity of *Cedrus deodara* heart wood extracts. *Egypt. Pharmaceut. J.* **14**: 10-14.
- Jassin, S. A. A., M. A. Naji (2003). Novel antiviral agents: a medicinal plant perspective. *J. Appl. Microbiol.* **95**: 412-427.
- Jia, L., H. Jin, J. Zhou, L. Chen, Y. Lu, Y. Ming, Y. Yu (2013). A potential anti-tumor herbal medicine, corilagin, inhibits ovarian cancer cell growth through blocking the TGF- β signaling pathways. *BMC Complement. Altern. Med.* **13**:33. doi: 10.1186/1472-6882-13-33.
- Knickelbein J. E., R. L. Hendricks, P. Charukamnoetkanok (2009). Management of herpes simplex virus stromal keratitis: an evidence-based review. *Surv. Ophthalmol.* **54**: 226-234.
- Kolodziej, H., O. Kayser, A. F. Kiderlen, H. Ito, T. Hatano, T. Yoshida, L. Y. Foo (2001). Antileishmanial activity of hydrolyzable tannins and their modulatory effect on nitric oxide and tumor necrosis factor- α released in macrophages *in vitro*. *Planta Med.* **67**: 825-832.
- Kolodziej, H., A. Kiderlen (2005). Antileishmanial activity and immune modulatory effects of tannins and related compounds on *Leishmania* parasitised RAW 264.7 cells. *Phytochem.* **66**: 2056-2071.
- Kurokawa, M., T. Hozumi, P. Basnet, M. Nakano, S. Kadota, T. Namba, T. Kawana, K. Shiraki (1998). Purification and characterization of eugenin as an anti-herpesvirus compound from *Geum japonicum* and *Syzygium aromaticum*. *J. Pharmacol. Exp. Ther.* **284**: 728-735.
- Kwan, C.-Y. (1994). Plant-derived drugs acting on cellular Ca²⁺ mobilization in vascular smooth muscle: tetramethylpyrazine and tetrandrine. *Stem Cells* **12**: 64-67.
- Lee, M. H., J. F. Chiou, K. Y. Yen, L. L. Yang (2000). EBV DNA polymerase inhibition of tannins from *Eugenia uniflora*. *Cancer Lett.* **154**: 131-136.
- Lin, L.-T., T.-Y. Chen, C.-Y. Chung, R. S. Noyce, T. B. Grindley, C. McCormick, T.-C. Lin, G.-H. Wang, C.-C. Lin, C. D. Richardson (2011). Hydrolyzable tannins (chebulagic acid and punicalagin) target viral glycoprotein-glycosaminoglycan interactions to inhibit herpes simplex virus

- 1 entry and cell-to-cell spread. *J. Virol.* **85**: 4386-4398.
- Lin, T. C., F. L. Hsu, J. T. Cheng (1993). Antihypertensive activity of corilagin and chebulinic acid, tannins from *Lumnitzera racemosa*. *J. Nat. Prod.* **56**: 629-632.
- Lyra R. M., D, Oliveira, N. Lins, N. Cavalcanti (2006). Prevention of type 2 diabetes mellitus. *Arq. Bras. Endocrinol. Metab.* **50**: 239-249.
- Miyamoto, K., T. Murayama, T. Yoshida, T. Hatano, T. Okuda Anticarcinogenic activities of polyphenols in food and herbs. In *Antinutrients and Phytochemicals in Food*, ed. By Shahidi, F., American Chemical Society: Washington, DC., 1997, pp. 245-259.
- Monobe, M., K. Ema, F. Kato, M. Maeda-Yamamoto (2008). Immunostimulating activity of a crude polysaccharide derived from green tea (*Camellia sinensis*) extract. *J. Agric. Food Chem.* **27**: 1423-1427.
- Notka, F., G. Meier, R. Wagner (2004). Concerted inhibitory activities of *Phyllanthus amarus* on HIV replication in vitro and ex vivo. *Antiviral Res.* **64**: 93-102.
- Okabe, S., M. Suganuma, Y. Imayoshi, S. Taniguchi, T. Yoshida, H. Fujiki (2001). New TNF- α releasing inhibitors, geraniin and corilagin, in leaves of *Acer nikoense* Megusuri-no-ki. *Biol. Pharm. Bull.* **24**: 1145-1148.
- O’Kane, P. D., L. R. Queen, Y. Ji, V. Reebye, P. Stratton, G. Jackson, A. Ferro (2003). Aspirin modifies nitric oxide synthase activity in platelets: effects of acute versus chronic aspirin treatment. *Cardiovas. Res.* **59**: 152-159.
- Okuda, T., T. Yoshida, T. Hatano (1991). Chemistry and biological activity of tannins in medicinal plants. *Econ. Medicinal Plant. Res.* **5**: 129-165.
- Rao, B. K., P. R. Sudarshan, M. D. Rajasekhar, N. Nagaraju, Ch. A. Rao (2003). Antidiabetic activity of *Terminaliapalida* fruit in alloxan induced diabetic rats. *J. Ethnopharmacol.* **85**: 169-172.
- Raut, N. A., N. J. Gaikwad (2006). Antidiabetic activity of hydro-ethanolic extract of *Cyperus rotundus* in alloxan induced diabetes in rats. *Fitoterapia* **77**: 585-588.
- Saha, R. K., T. Takahashi, Y. Kurebayashi, K. Fukushima, A. Minami, N. Kinbara, M. Ichitani, Y. M. Sagesaka, T. Suzuki (2010). Antiviral effect of strictinin on influenza virus replication. *Antiviral Res.* **88**: 10-18.
- Sani, U. M. (2015). Phytochemical screening and antidiabetic effects of extracts of the seeds of *Citrullus lanatus* in alloxan-induced diabetic albino mice. *J. App. Pharm. Sci.* **5**: 51-54.
- Schnitzler, P., C. Koch, J. Rsichling (2007). Susceptibility of drug-resistant clinical herpes simplex virus type 1 strain to essential oils of ginger, thyme, hyssop, and sandalwood. *Antimicrob. Agents Chemother.* **51**: 1859-1862.
- Sepúlveda, L., A. Ascacio, R. Rodriguez-Herrera, A. Aguilera-Carbó, C. N. Aguilar (2011). Ellagic acid: Biological properties and biotechnological development for production processes. *Afr. J. Biotechnol.* **10**: 4518-4523.
- Suzutani, T., M. Ogasawara, I. Yoshida, M. Azuma, Y. M. Knox (2003). Anti-herpesvirus activity of an extract of *Ribes nigrum* L. *Phytother. Res.* **17**: 609-613.
- Ushio, Y., T. Fang, T. Okuda, H. Abe (1991). Modificational changes in function and morphology of cultured macrophages by geraniin. *Jpn. J. Pharmacol.* **57**: 187-196.
- Varadkar, P., P. Dubey (2001). Modulation of radiation-induced protein kinase C activity by phenolics. *J. Radiol. Prot.* **21**: 361-370.
- Vilhelmova, N., R. Jacquet, S. Quideau, A. Stoyanova, A. S. Galabov (2011). Three-dimensional analysis of combination effect of ellagitannins and acyclovir on herpes simplex virus types 1 and 2. *Antiviral Res.* **89**: 174-181.
- Vilhelmova-Ilieva N., R. Jacquet, S. Quideau, A. S. Galabov (2014). Ellagitannins as synergists of ACV on the replication of ACV-resistant strains of HSV 1 and 2. *Antiviral Res.* **110**: 104-114.
- Wang, C. C., L. G. Chen, L. L. Yang (2002). *In vitro* immunomodulatory effects of cuphiin D 1 on human mononuclear cells. *Anticancer Res.* **22**: 4233-4236.
- Widsten P., C. D. Cruz, G. C. Fletcher, M. A. Pajak, T. K. McGhie (2014). Tannins and extracts of fruit byproducts: antibacterial activity against foodborne bacteria and antioxidant capacity. *J. Agric. Food Chem.* **62**: 11146-11156.
- Yang, C.-M., H.-Y. Cheng, T.-C. Lin, L.-C. Chiang, C.-C. Lin (2005). Acetone, ethanol and methanol extracts of *Phyllanthus urinaria* inhibit HSV-2 infection *in vitro*. *Antiviral Research.* **67**: 24-30.
- Yildirim, I., T. Kutlu (2015). Anticancer agents: saponin and tannin. *Int. J. Biol. Chem.* **9**: 332-340.
- Yokozawa, T., C. P. Chen, D. Y. Rhyu, T. Tanaka, Y. C. Park, K. Kitani (2002). Potential of sanguin H-6 against oxidative damage in renal mitochondria and apoptosis mediated by peroxynitrite *in vivo*. *Nephron* **92**: 133-141.
- Yoshida, T., Y. Amakura, M. Yoshimura (2010). Structural features and biological properties of ellagitannins in some plant families of the order *Myrtales*. *Int. J. Mol. Sci.* **11**: 79-106.
- Ziyaeyan, M., A. Alborzi, A. Japoni, M. Kadivar, M. A. Davarpanah, B. Pourabbas, A. Abassian (2007). Frequency of acyclovir-resistant herpes simplex viruses isolated from the general immunocompetent population and patients with acquired immunodeficiency syndrome. *Int. J. Dermatol.* **46**: 1263-1266.