



Ameliorative Role of Curcumin on Liver of Rats Treated with Gentamicin, Cefotaxime and Metronidazole

Hosny Abd El Fadil¹, Abdel Alim F A¹, Raslan Y A², Amany M Gad^{2*}, Ahmady Y A²

¹Department of Pharmacology, Faculty of Veterinary Medicine, Zagazig University, Zagazig, Egypt

²Department of Pharmacology, National Organization for Drug Control and Research (NODCAR), Giza, Egypt

Abstract The liver is the most vulnerable organ against the side effects of xenobiotics. Aminoglycosides especially gentamicin are the widely used antibiotics against Gram negative bacteria. Cefotaxime is a broad spectrum cephalosporin antibiotic used mainly against Gram positive bacteria. Metronidazole is a chemotherapy used in treatment of protozoal and anaerobic bacterial infections. Combination therapy has complementary and synergistic mechanisms of action. The Present investigation focused on evaluating the effect of curcumin on hepatic alterations of gentamicin, cefotaxime, metronidazole, and their combinations. For this study, eighty eight male rats were classified into eleven groups. (First): Served as control, (Second): Curcumin (200mg/kg.b.wt.,p.o.), (Third): Gentamicin (80 mg/kg.b.wt., i.m), (Fourth): Cefotaxime (540 mg/kg.b.wt., i.m), (Fifth): Metronidazole (135 mg/kg.b.wt., p.o.), (Sixth): Gentamicin with cefotaxime, (Seventh): Gentamicin with cefotaxime and metronidazole, (Eighth): Curcumin one hour prior gentamicin, (Ninth): Curcumin one hour prior cefotaxime, (Tenth): Curcumin one hour prior gentamicin and cefotaxime, (Eleventh): Curcumin one hour before gentamicin, cefotaxime and metronidazole for 14 successive days. Serum was prepared for measuring of Alkaline phosphate (ALP), Alanine aminotransferase (ALT) and Aspartate aminotransferase (AST), liver was removed for homogenate measurements, histopathological & immunohisto-chemical examination. Administration of curcumin indicated a significant ($p < 0.05$) decrease of ALP, ALT, AST, malondialdehyde (MDA) and tumor necrosis factor- α (TNF- α) and elevation in glutathione (GSH), catalase (Cat), superoxide dismutase (SOD), and glutathione peroxidase (GP^x) of liver homogenate. Our data showed that curcumin could protect the liver alterations by blocking oxidative damages.

Keyword: Gentamicin, Cefotaxime, Metronidazole, Curcumin, Liver

Introduction

Aminoglycoside antibiotics belong to one of the oldest classes of antibacterial agents used in curative therapies. Gentamicin is one of aminoglycosides was originated from *Micromonospora purpurea*. It is effective against most of the life threatening microbial pathogens, particularly Gram-negative bacterial infections and is also active against *Staphylococcus* and *Enterococcus*, especially in synergy with β -lactams [1,2,3]. Cefotaxime is a member of third generation cephalosporin broad spectrum β -lactam antibiotic. It is considered to be equivalent to cefotriaxone in terms of safety and efficacy [4]. Metronidazole is a 5-nitroimidazole chemotherapy widely used veterinarians and medicines as anti-protozoal and bactericidal chemotherapy [5].

Curcumin is the effective compound of *Curcuma longa*. It is used for curing many diseases including renal disorders. Curcumin has antioxidant, anti-inflammatory and cell membrane stabilizing equivalent to vitamins C, E, and beta-carotene [6,7].



The target of the present investigation was to evaluate the possible ameliorative role of curcumin against renal oxidative stress induced by combination of gentamicin, cefotaxime and metronidazole in male rats.

Materials and Methods

Drugs

Gentamicin (80 mg / kg, I.M.) [8], Cefotaxime (540 mg / kg, I.M) [9], were obtained from Egyptian Int. Pharmaceutical Industrial Co. (10th of Ramadan, Egypt), Metronidazole (135 mg / kg, orally; Alex. Co. for pharmaceutical industries (Egypt) [10]. Curcumin (200 mg/kg, orally; purchased from Sigma Chemicals Co. (USA) [11].

Experimental Animals

Eighty eight adult male albino rats (weighing 200±10 gm) were used in the present investigation. They were obtained from the Animal Breeding Unite, National Organization for drug control and research (Giza, Egypt), and used after one week of quarantine and acclimation, animal groups were divided in separating cages, in controlled temperature (23-25 °C), humidity (60%), light and dark cycles of 12 hours each. The animals were fed on standard pelleted diet and water *ad libitum*. The experiment was conducted in accordance with the ethical guidelines for investigations in laboratory animals and were approved by the Ethical Committee of Faculty of Veterinary Medicine, Zagazig University, Egypt and comply with the Guide for the Care and Use of Laboratory Animals [12].

Animal Grouping

At eighth day of acclimatization, eighty eight adult male albino rats were divided equally into eleven groups.

Group 1: (Control)

Group 2: received curcumin.

Group 3: received gentamicin.

Group 4: received cefotaxime.

Group 5: received metronidazole.

Group 6: received gentamicin with cefotaxime administration.

Group 7: received gentamicin with cefotaxime and metronidazole.

Group 8: received curcumin and gentamicin.

Group 9: received curcumin and cefotaxime.

Group 10: received curcumin gentamicin and cefotaxime.

Group 11: received curcumin, gentamicin, cefotaxime and metronidazole.

Experiment continued for fourteen days, at twenty-four hours after last dose, the animals were anaesthetized using Phenobarbital, blood samples were collected freshly from retro-orbital plexus in non-heparinized tubes and serum was separated by centrifugation for 20 min at 4000 r.p.m. for measuring albumin, total protein, ALP, ALT and AST. the two livers immediately removed and washed in ice saline, and one kept in formalin for histopathology and histochemistry, the second was homogenized in phosphate buffer [13], for determination of reduced glutathione (GSH) [14], catalase (Cat) [15], glutathione peroxidase (GPx) [16], superoxide dismutase (SOD) [17], Malondialdehyde (MDA) [18] and Tumor necrosis factor alpha (TNF- α) [19]. Liver was examined histopathologically and immunohistochemically [20].

Statistical Analysis

Results were expressed as mean \pm standard errors of the means (S.E.M.). Comparison between more than two different groups was carried out using the one-way analysis of variance (ANOVA) followed by Tukey-Kramer's Multiple Comparison Test, where $P < 0.05$ was considered significant [21].



Results

Effect of curcumin on liver function tests in male albino rats treated with gentamicin, cefotaxime and / or metronidazole

Table (1) displayed that, on comparison to the control group, curcumin, metronidazole, curcumin with cefotaxime, curcumin with cefotaxime and gentamicin and curcumin with cefotaxime, gentamicin, and metronidazole groups revealed non-significant differences in ALP, ALT and AST. Intramuscular administration of gentamicin and Cefotaxime groups evoked a significant increase in ALP, ALT and AST comparing with that of the normal control group, whereas oral administration of curcumin before gentamicin, cefotaxime, gentamicin with cefotaxime, gentamicin with cefotaxime and metronidazole groups induced a significant decrease in ALP, ALT and AST comparing with gentamicin treated group.

Table 1: Effect of curcumin on liver function tests in male albino rats treated with gentamicin, cefotaxime and / or metronidazole

Groups	ALP (U/L)	ALT (U/L)	AST (U/L)
(1) Control (Cont.)	78.60±1.301 ^d	27.89±0.788 ^d	78.97±1.178 ^c
(2) Curcumin (Cur.)	81.24±1.881 ^{cd}	30.89±0.647 ^{cd}	80.87±1.877 ^c
(3) Gentamicin (Gen.)	109.70±2.114 ^a	54.84±1.162 ^a	120.70±1.326 ^a
(4) Cefotaxime (Cef.)	86.91±2.099 ^c	36.86±0.989 ^{bc}	92.98±3.134 ^b
(5) Metronidazole (Met.)	84.46±0.352 ^{cd}	31.26±2.991 ^{cd}	85.39±1.941 ^{bc}
(6) Gen.+Cef.	101.00±0.715 ^b	44.32±1.695 ^b	103.00±1.921 ^b
(7) Gen.+Cef.+Met.	102.30±1.087 ^b	44.58±1.435 ^b	104.50±1.770 ^b
(8) Cur.+ Gen.	93.61±1.550 ^c	36.47±2.238 ^c	96.13±3.568 ^b
(9) Cur.+ Cef.	78.69±0.822 ^d	28.23±0.492 ^d	77.21±5.775 ^c
(10) Cur.+Gen.+Cef.	84.62±1.800 ^{cd}	34.89±0.606 ^{cd}	88.78±1.549 ^c
(11)Cur.+Gen.+Cef.+Met.	84.90±1.509 ^{cd}	34.21±1.423 ^{cd}	88.96±1.242 ^c

Means with different superscripts in the column are significant (p >0.05)

Effect of curcumin on albumin and total protein in male albino rats treated with gentamicin, cefotaxime and / or metronidazole

Table (2) showed evoked that, curcumin, cefotaxime, metronidazole, curcumin with cefotaxime, curcumin with cefotaxime and gentamicin and curcumin with cefotaxime, gentamicin, and metronidazole groups revealed non-significant differences in albumin but gentamicin induced a significant increase comparing to the control rats.

In curcumin and metronidazole groups displayed non- significant changes in total albumin but gentamicin and cefotaxime groups induced a significant decrease in total protein comparing to control rats. Administration of curcumin before gentamicin, cefotaxime, gentamicin with cefotaxime, gentamicin with cefotaxime and metronidazole groups showed a significant increase in total protein comparing to gentamicin group

Table 2: Effect of curcumin on albumin and total protein in male albino rats treated with gentamicin, cefotaxime and / or metronidazole

Groups	Albumin	Total protein
(1) Control (Cont.)	4.223±0.095 ^a	6.718±0.110 ^a
(2) Curcumin (Cur.)	4.286±0.088 ^a	6.366±0.135 ^{ab}
(3) Gentamicin (Gen.)	2.642±0.164 ^b	3.185±0.101 ^c
(4) Cefotaxime (Cef.)	3.845±0.040 ^a	5.971±0.059 ^b
(5) Metronidazole (Met.)	4.038±0.053 ^a	6.395±0.180 ^{ab}
(6) Gen.+ Cef.	3.865±0.145 ^a	4.051±0.067 ^d
(7) Gen.+Cef.+Met.	3.813±0.208 ^a	3.910±0.031 ^d
(8) Cur.+ Gen.	4.065±0.114 ^a	5.046±0.060 ^c
(9) Cur.+ Cef.	4.244±0.053 ^a	6.736±0.093 ^a
(10) Cur.+Gen.+Cef.	3.897±0.146 ^a	6.169±0.170 ^{ab}
(11) Cur.+Gen.+Cef.+Met.	3.879±0.121 ^a	6.145±0.174 ^{ab}

Means with different superscripts in the column are significant (p < 0.05)



Effect of curcumin on GSH, Cat, GPx, SOD, MDA, and TNF- α of liver in male albino rats treated with gentamicin, cefotaxime and/or metronidazole

Table (2) showed that, oral metronidazole treated group produced a non-significant difference in liver GSH, Cat, SOD, GPx and MDA, respectively, for the normal control group, but gentamicin or cefotaxime groups evoked a significant inhibition in GSH, Cat, GPx, SOD and increase in MDA and TNF- α versus control group except GPx for cefotaxime group, whereas oral administration of curcumin prior gentamicin, gentamicin with cefotaxime or gentamicin, cefotaxime with metronidazole groups induced a significant increase in GSH, Cat, GPx, SOD and decrease in MDA and TNF- α as compared to gentamicin groups.

Table 3: Effect of curcumin on GSH, Cat, SOD, GPx, MDA and TNF- α of liver in male albino rats treated with gentamicin, cefotaxime and/or metronidazole.

Groups	GSH (mg/g)	Cat. (U/g)	SOD (U / g)	GPx (U / g)	MDA (nmol/g)	TNF- α (Pg/g)
(1) Control(Cont.)	54.31±1.622 ^a	2.190±0.171 ^a	332.9±25.69 ^a	55.94±2.438 ^a	22.24±0.425 ^d	38±2.30 ^e
(5) Gentamicin(Gen)	25.46±1.227 ^d	0.570±0.056 ^d	89.14±4.59 ^c	7.36±0.785 ^c	78.68±5.612 ^a	206±4.04 ^a
(6) Cefotaxime (Cef.)	46.29±1.241 ^b	1.710±0.055 ^b	231.8±17.84 ^b	36.47±2.430 ^{ab}	31.16±0.344 ^c	71.3±1.85 ^d
(7)Metronidazol(Met.)	50.34±2.077 ^{ab}	1.895±0.062 ^{ab}	321.0±6.86 ^a	36.47±3.033 ^{ab}	25.96±0.926 ^d	50±1.52 ^d
(8) Gen. + Cef.	36.68±0.539 ^c	1.178±0.099 ^c	214.0±21.71 ^b	31.61±2.430 ^b	66.08±1.566 ^b	102.7±3.18 ^c
(9)Gen+Cef+ Met.	36.09±1.464 ^c	1.184±0.053 ^c	206.1±7.92 ^b	31.61±2.433 ^b	67.30±0.631 ^b	101.7±3.71 ^c
(10) Cur.+ Gen.	38.24±2.042 ^c	1.506±0.077 ^b	190.2±13.72 ^b	38.90±0.158 ^{ab}	37.212±2.007 ^c	144.3±3.48 ^b
(12) Cur.+Cef.	53.99±0.983 ^a	2.258±0.109 ^a	332.9±13.72 ^a	36.47±2.341 ^{ab}	22.23±0.542 ^d	47.6±1.85 ^e
(14)Cur+Gen+Cef	48.08±1.038 ^{ab}	1.746±0.102 ^{ab}	301.2±7.93 ^{ab}	55.12±3.243 ^a	28.48±2.224 ^{cd}	43±9.29 ^e
(16)Cur+Gen+Cef+ Met	48.53±1.925 ^{ab}	1.790±0.071 ^{ab}	299.3±19.67 ^{ab}	55.12±3.243 ^a	29.31±2.377 ^{cd}	44±5.56 ^e

Means with different superscripts in the column are significant (p < 0.05)

Effect of curcumin on histopathological picture of liver in male albino rats treated with gentamicin, cefotaxime and/or metronidazole

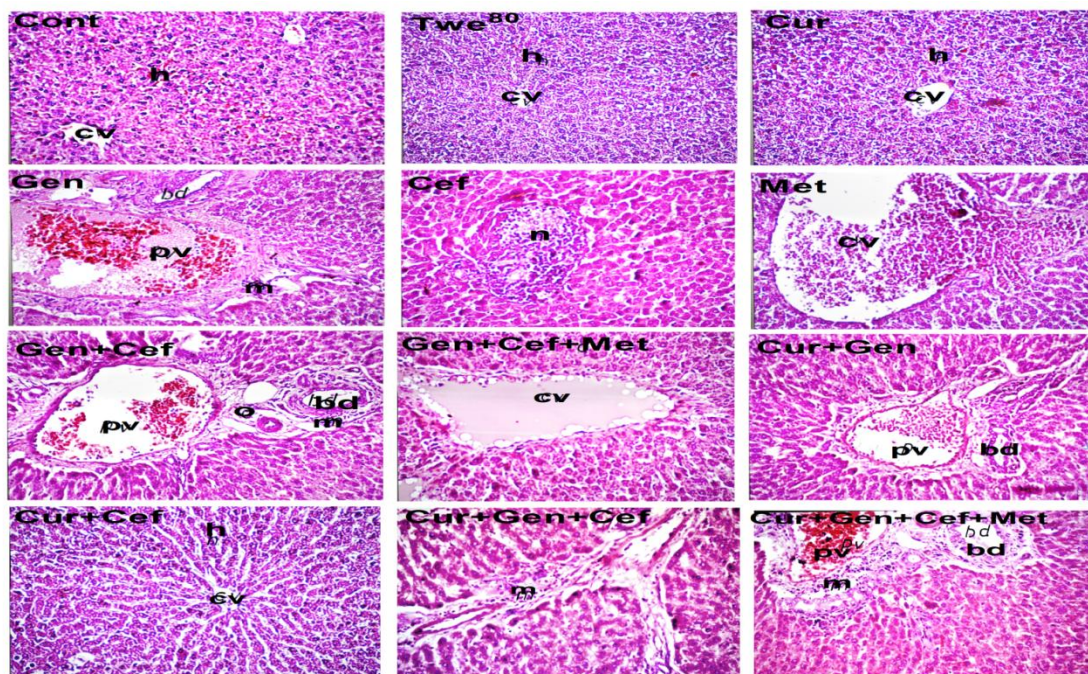


Figure 1: Liver sections of normal control, twe⁸⁰ and curcumin groups, showed no histopath-ological alterations and showed normal histological structure of the central vein (cv) and surrounding hepatocytes (h) but section of liver of



rat treated with gentamicin group, showed severe dilatation and congestion in portal vein (pv) with inflammatory cells infiltration (m) as well as dilatation in the bile ducts (bd) and cefotaxime treated group evoked focal necrosis (n) in hepatic parenchyma, On the other hand, liver section of rat treated with gentamicin and cefotaxime group, elicited severe dilatation and congestion in the portal vein as well as oedema with few inflammatory cells infiltration surrounding the dilated bile ducts. The liver of rats were received gentamicin, cefotaxime and metronidazole displayed severe dilatation in the central vein (cv) by haemolysed blood with granular degeneration (d) in the hepatocytes. Liver section of rats were administered curcumin and gentamicin group, showed congestion and dilatation in portal vein (pv) and bile duct (bd) with moderate inflammatory cells infiltration in portal area while curcumin and cefotaxime group, showed normal histological structure but curcumin, gentamicin and cefotaxime individuals, showed few inflammatory cells infiltration (m) in the portal area and Liver section of rat treated with curcumin, gentamicin, cefotaxime and metronidazole group, showed few inflammatory cells infiltration (m) in the portal area with congestion and dilatation in portal vein (pv) and normal hepatocytes (H&E x40).

Effect of curcumin on the severity of Immunohistopathological reaction using caspase-3 in liver of different experimental groups

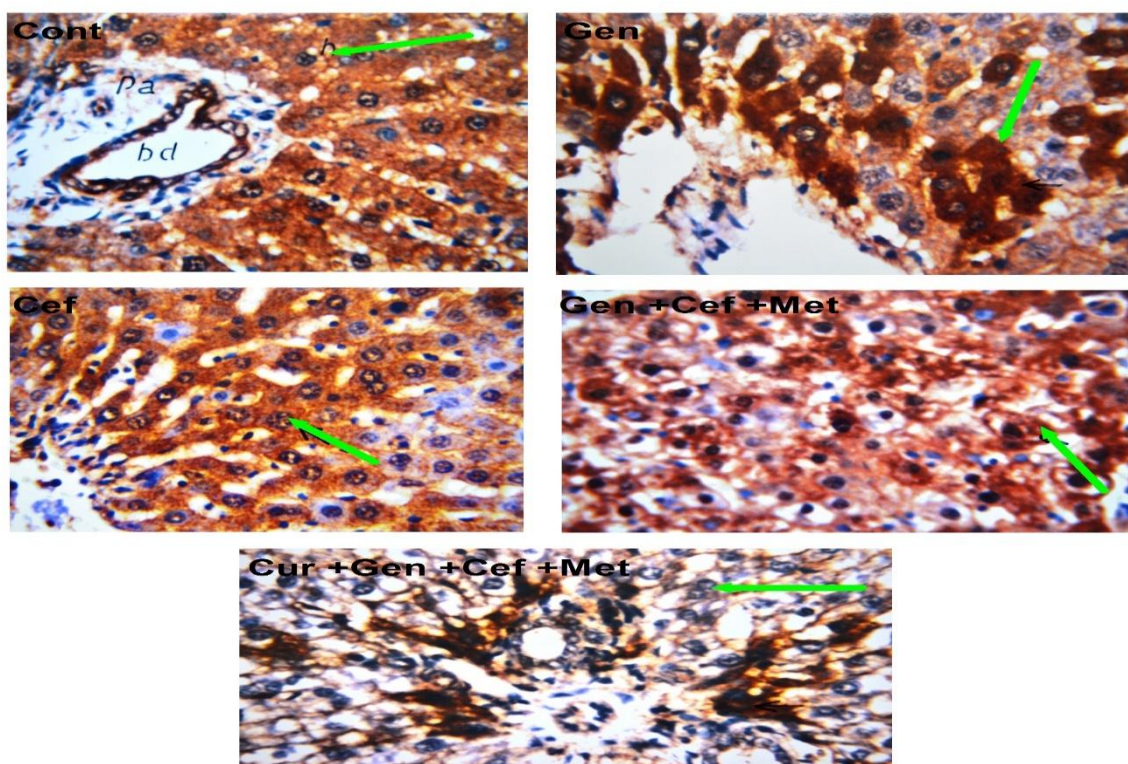


Figure 2: Liver section of control group, showed negative immunohistochemical reaction but Liver section of gentamicin group, displayed severe positive immuno-histochemical while section of rat treated with cefotaxime group, elicited mild positive immunohistochemical reaction in addition to liver section of rat treated with gentamicin, cefotaxime and metronidazole showed positive immunohistochemical reaction and liver section of rat treated with curcumin, gentamicin, cefotaxime and metronidazole evoked mild positive immunohistochemical reaction in the few hepatocytes (arrow) using caspase-3 antibody. (X 80).

Discussion

Aminoglycosides especially gentamicin have long been used in curing of gram negative bacteria resulting from its chemical stability and rapid onset of bactericidal action. Unlike its beneficial effect, gentamicin has hepatotoxic



action. Our biochemical results showed that gentamicin induced hepatic disturbances which were confirmed histopathologically and immunohistochemically.

Biochemically, serum hepatic biomarkers, ALP, ALT and AST were significantly increased while total protein and albumin were significantly decreased in the gentamicin accompanied rats versus curcumin accompanied rats. Our data interpret the increasing of liver biomarkers resulting from hepatocytes injuries and escaping of enzymes in the circulation in agreement with the previous reporters [22-24]. Also, our results indicated a significant increase in MDA and TNF- α and increase in GSH, Cat, SOD and GP^x of liver homogenate of rats treated with gentamicin but these investigations were attenuated by co-administration of curcumin.

In agreement with our data [25] reported that, gentamicin induced an increase of hepatic MDA level and decrease in GSH, Cat.; SOD, GPx, which described by hepatocytes damages and inflammatory cells infiltration. On the same ground [26] described gentamicin toxicity by stimulating synthesis of H₂O₂ and hydroxyl radicals in the mitochondria leading to mesangial cells contraction, altering the glomerular filtration rate. In this way, gentamicin induced a strong accumulation of oxidants (MDA and NO) in the liver while SOD activities and GSH contents were decreased [27].

Organism has a lot of anti-oxidative defense mechanisms for controlling reactive oxygen species preventing cellular damage, including the non-enzymatic (mainly reduced glutathione) and enzymatic defenses (including superoxide dismutase, glutathione reductase, glutathione S-transferase, catalase and glutathione peroxidase) [28]. Also, our results agreed with [29,30] who recorded that the gentamicin group was the most strongest caspase-3-stained tissues. These data showed that, co-administration of gentamicin and cefotaxime resulted in a significant improvement of parameters may be due to cefotaxime decreases the reabsorption of gentamicin in renal tubules in accordance with [31,32] who showed that, cefepime (cephalosporin) induced an improvement in the activities of superoxide dismutase and catalase along with decrease in MDA levels in amikacin (aminoglycoside) group. Our results coordinate with those obtained by [33] reported that, administration of curcumin to gentamicin-treated rats increased liver GSH level and GPx, Cat, and SOD activities. Furthermore, it has been suggested that lipid peroxidation might be a predisposing factor for induction of liver tissue damage. The role of curcumin in reducing the membrane damage is related to its ability to scavenge lipid peroxidation initiating agents. We observed a decrease in MDA and TNF- α levels in the liver tissue of rats treated with curcumin and gentamicin compared with gentamicin alone group. In the present study, the results of histopathological and histochemical examination showed a clear evidence of ameliorative effect of curcumin against gentamicin nephrotoxicity. The protective role of curcumin may be depend on its ability to eliminate the hydroxyl radical [34], superoxide radical [35], singlet oxygen [36], nitrogen dioxide [37], and nitric oxide [38]. Regarding to our results evoked the best effect of curcumin administration one hour before treatment due to its antioxidant, anti-inflammatory and anti-apoptotic effects.

Conclusion

The present study reveals that curcumin has potent protective effect against hepatotoxic alterations of gentamicin in rats.

References

1. Begg, E.J. & Barclay, M.L. (1995). Aminoglycosides-50 years on. *British Journal of Clinical Pharmacology*, 39(6):597-603.
2. Oliveira, F.P., Cipullo, J.P. & Burdmann, E.A. (2006). Aminoglycoside nephrotoxicity. *Brazil Journal of Cardiovascular Surgery*, 21(4):444-452.
3. Selby, N.M., Shaw, S., Woodier, N., Fluck, R.J. & Kolhe, N.V.(2009). Gentamicin-associated acute kidney injury. *Quarterly Journal of Medicine*, 102(12):873-880.
4. Williams, J.D. (1997). Beta- Lactamase inhibition and *in-vitro* activity of sulbactam. *Clinical infectious diseases Journal*, 24: 494-497.
5. Sarah. L. C., Kiera. L. D., Shannon, F. H., Dino, P. P., & Gary E. G. (2004). Treatment of Infections Caused by Metronidazole-Resistant *Trichomonas vaginalis*. *Clinical Microbiology Reviews*, 17(4):783-793.



6. Akram, M., Shahab, U., Afzal, A., Khan, U., Abdul Hannan, E. & Asif, M. (2010). Curcuma Longa and Curcumin: A review article. *Romanian Journal of Biology - Plant Biology*, 55(2): 65-70.
7. Ahmad, A., Husain, A., Mujeeb, M., Khan, S.A., Najmi, A.K., Siddique, N.A., Damanhour, Z.A. & Anwar, F. (2013). A review on therapeutic potential of *Nigella sativa*: A miracle herb. *Asian Pacific Journal of Tropical Biomedicine*, 3:337-352.
8. Hamdy, M.M., El-Sers, D.A. & Abdelhamid, M.M. (2013). Protective effect of curcumin and GinkgoBilObal. Extract against gentamicin-induced nephrotoxicity in Rats. *Assuit Medical Journal*, 37(1): 1-12.
9. Paget, G.E. & Barnes, J.M. (1964). Toxicity tests. In: Evaluation of drug activities and pharmacometrics. Eds.: Laurant, D.R. and Bacharach, A.L. *Academic Press, London and New York*, 135-166.
10. Oda, S.S. (2012). Histopathological and Biochemical Alterations of Metronidazole-Induced Toxicity in Male Rats. *Global Vetrinaria*, 9(3): 303- 310.
11. Venkatesan, N., Punithavathi, D. & Arumugam, V. (2000). Curcumin prevents adriamycin nephrotoxicity in rats. *British Journal of Pharmacology*, 129 (2): 231-234.
12. Institute of Laboratory Animal Resources (ILAR). (1996). Guide for the Care and Use of Laboratory Animal Resources, 8th edition. Ei-Washington, D.C.: *National Academy Press*.
13. Noeman, S.A., Hamooda, H.E. & Baalash, A.A. (2011). Biochemical study of Oxidative Stress Markers in the Liver, Kidney and Heart Fat Diet Induced Obesity in Rats. *Diabetology and Metabolic syndrom*, *Tanta University Journal, Egypt*, 3(17): 1-8.
14. Beutere, E., Duron, O. & Kelly, B.M. (1963). Improved method for the determination of blood glutathione. *The Journal of laboratory and clinical medicine*, 61:882-888.
15. Aebi, H. (1984). Catalase *in vitro*. *Methods in Enzymology*, 105: 121-126.
16. Paglia, D. E. & Valentine, W.N. (1967). Studies on the quantitative and qualitative characterization of erythrocyte glutathione peroxidase. *The Journal of laboratory and clinical medicine*, 70(1): 158-169.
17. Nishikimi, M., Roa, N.A. & Yogi, K. (1972). Measurement of superoxide dismutase. *Biochemical and biophysical research communications*, 46: 849-854.
18. Satoh, K. (1978). Plasma lipid peroxide in cerebrovascular disorder determined by a new colorimetric method. *International Journal of Clinical Chemistry*, 90: 37-43.
19. Taylor, P.C. (2001). Anti-TNF therapy for rheumatoid arthritis and other inflammatory diseases. *Molecular biotechnology*, 19:153-168.
20. Thompson, C.M., Dixon, L.R., Wasserfall, C., Monroe, M., McGuigan, J.M., Schatz, D., Crawford, J.M. & Atkinson, M.A. (2009). Pancreatic adenocarcinoma patients with localised chronic severe pancreatitis show an increased number of single beta cells, without alterations in fractional insulin area. *European Association for the Study of Diabetes*, 52: 262-270.
21. Snedecore, G.W. (1969). *Statistical Methods*. 4th Ed., Iowa State Collage Press, Ames, Iowa State College Press, Ames, Iowa.
22. Khan, M. R. I., Islam, M. A., Hossain, M. S., Asadujjaman, M., Wahed, M. I. I. & Rahman, B. M. (2010). Antidiabetic Effects of the Different Fractions of Ethanolic Extracts of *Ocimum sanctum* in Normal and Alloxan Induced Diabetic Rats. *Journal of Scientific Research*, 2(1):158-168.
23. Noorani, A.A., Gupta, K.A., Bhadada, K. & Kale, M.K. (2011). Protective effect of methanolic leaf extract of *Caesalpinia bonduc* (L.) on gentamicin-induced hepatotoxicity and nephrotoxicity in rats. *Iranian Journal of Pharmacology & Therapeutics*, 10(1):21-25.
24. Ademiluyi, A.O., Oboh, G., Owoloye, T.R. & Agbebi, O.J. (2013). Modulatory effects of dietary inclusion of garlic (*Allium sativum*) on gentamycin-induced hepatotoxicity and oxidative stress in rats. *Asian Pacific journal of tropical biomedicine*, 3(6):470-475.



25. Kandemir, F.M., Ozkaraca, M., Yildirim, B.A., Hanedan, B., Kirbas, A., Kilic, K., Aktas, E. & Benzer, F. (2015). Rutin attenuates gentamicin-induced renal damage by reducing oxidative stress, inflammation, apoptosis, and autophagy in rats. *Renal failure Journal*, 37(3), 518-525.
26. Walker, D. & Shah, V. (1988). Evidence suggesting a role for hydroxyl radical in gentamicin induced acute renal failure in rats. *European Society for Clinical Investigation*, 81: 334- 341.
27. Kalkan, Y., Kapakin, K.A., Kara, A., Atabay, T., Karadeniz, A., Simsek, N., Karakus, E., Can, I., Yildirim, S., Ozkanlar, S. & Sengul, E. (2012). Protective effect of Panax ginseng against serum biochemical changes and apoptosis in kidney of rats treated with gentamicin sulphate. *Journal of Molecular Histology*, 43(5): 603-613.
28. Ramakrishnan, G.; Raghavendran, H.R.; Vinodhkumar, R. & Devaki, T. (2006). Suppression of N-nitrosodiethylamine induced hepatocarcinogenesis by silymarin in rats. *Chemico-Biological Interactions*, 161: 104-114.
29. Masjedi, F., Gol, A. & Dabiri, S. (2013). Preventive Effect of Garlic (*Allium sativum L.*) on Serum Biochemical Factors and Histopathology of Pancreas and Liver in Streptozotocin- Induced Diabetic Rats. *Iranian Journal of Pharmaceutical Research*, 12(3):325-338.
30. Christina, L. K., Brittany, J. C., Jessica, L. G., Caitlin, E. T., Dominic, A. M. & Douglas, A. C. Comparison of activated caspase detection methods in the gentamicin-treated chick cochlea. *Hear Research*, 2008, 240(1-2): 1-11.
31. Chaudhary, M., Soni, A. & Dwivedi, V. Fixed Dose Combination of Cefepime Plus Amikacin Prevent Oxidative Stress in Liver of Mus musculus Mice. *Current Clinical Pharmacology*, 2008, 3(3):211-214.
32. Chaudhary, M., Soni, A., Dwivedi, V. & Sehgal, R. Anti-Oxidant Property of New Cephalosporin-Aminoglycoside Fixed Dose Combination. *Current Drug Therapy*, 2009, 4(1):2-6.
33. Cekmen, M., Ilbey, Y.O., Ozbek, E., Simsek, A., Somay, A. & Ersoz, C. (2009). Curcumin prevents oxidative renal damage induced by acetaminophen in rats. *Food and Chemical Toxicology*, 47(7):1480-1484.
34. Reddy, A.C. & Lokesh, B.R. (1994). Studies on anti-inflammatory activity of spice principles and dietary n-3 polyunsaturated fatty acids on carrageen an induced inflammation in rats. *Annals of Nutrition and Metabolism Journal*, 38: 349-358.
35. Sreejayan, N. & Rao, M. (1996). Free radical scavenging activity of curcuminoids. *Journal of the American Association of Pharmaceutical*, 46: 169-172.
36. Rao, M.V., Hale, B.A. & Ormrod, D.P. (1995). Amelioration of ozone – induced oxidative damage in wheat plants grown under high carbon dioxide: Role of antioxidant enzymes. *Journal of plant physiology*, 109: 421-432.
37. Unnikrishnan, M.K. & Rao, M. (1995). Curcumin inhibits nitrogen dioxide induced oxidation of hemoglobin. *Molecular and cellular biochemistry*, 146(1): 35-37.
38. Sreejayan, N. & Rao, M.N. (1997). Nitric oxide scavenging by curcuminoids, *Journal of Pharmacy and Pharmacology*, 49:105-107.

