



## Research Article

# Evaluation of bioethanol production from rice field weed biomass

Phuong Thi Vu, Yuwalee Unpaprom, Rameshprabu Ramaraj

### Abstract

Bioethanol has attracted more attention as a clean-burning fuel that can benefit both environment and energy sector. Gooseweed and small-flowered nutsedge are abundant in rice fields in form of weeds and considered as a major agricultural problem. Thus, this paper aims to evaluate the possibility of ethanol production from these two weeds by calculating the theoretical ethanol yield from its reducing sugars and cellulose content. Experiment was conducted in rice fields in Chiang Mai province, Thailand and 207 kg/ha and 201 kg/ha biomass yield was obtained from gooseweed and small-flowered nutsedge plants. The theoretical ethanol yield of gooseweed and small-flowered nutsedge were 160 L/Mg and 223 L/Mg, respectively that suggest utilizing these materials as promising feedstocks for bioethanol production.

**Keywords** gooseweed, small-flowered nutsedge, theoretical ethanol yield

### Introduction

With the rapid development of population, additional energy has been needed in order to meet the growing demand of the world. Fossil fuels are the main source of energy all over the world. However, the use of fossil fuels has been associated with a lot of environmental issues which affects the whole biosphere and its inhabitants [1-2]. Another downside of using non-renewable energy is its limited supply. Especially nowadays, due to its high consumption, it is approaching their natural limits and it takes a considerable long time to be created. Thus, in order to meet the demand of energy as well as to control the quality of environment, biofuels should be considered as a feasible option. Biofuels has already been investigated around the world and continuously being utilized for the enhancement of global energy security. It can be used as an alternative source of energy for various purposes such as engine fuels, cooking, heating, electricity generating, etc. [4].

Most biofuels such as bioethanol, biogas, biodiesel, and biohydrogen are made from biomass and waste that helps to reduce the pressure on the environment [4]. Among different kinds of biofuel, bioethanol has drawn much widespread attention due to its promise of providing a clean transport fuel [8]. Even though its energy content is approximately same as gasoline, bioethanol has higher octane number (106-110) than gasoline which makes it an antiknock fuel [5-9]. Hence, it is often blended with gasoline or diesel with appropriate ratios in order to create new mixtures to reduce the harmful gas emission and increase the engine performance [10]. USA and Brazil are two top leading countries in bioethanol production from edible sources (corn and sugarcane) with 56.1 and 28.2 billion liters bioethanol production in 2015, respectively [11].

**Received:** 9 November 2017  
**Accepted:** 17 December 2017  
**Online:** 18 December 2017

#### Authors:

Phuong Thi Vu, Rameshprabu Ramaraj ✉  
School of Renewable Energy, Maejo University,  
Chiang Mai 50290, Thailand

Yuwalee Unpaprom  
Program in Biotechnology, Faculty of Science;  
Maejo University, Chiang Mai 50290, Thailand

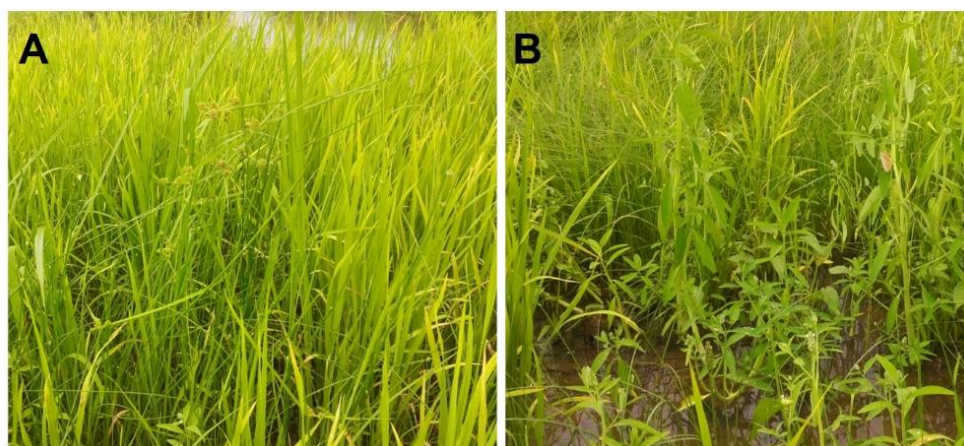
Rameshprabu Ramaraj  
Energy Research Center, Maejo University,  
Chiang Mai 50290, Thailand

✉ rameshprabu@mju.ac.th,  
rameshprabu@gmail.com

**Emer Life Sci Res (2017) 3(2): 42-49**

**E-ISSN: 2395-6658**  
**P-ISSN: 2395-664X**

**DOI:** <http://dx.doi.org/10.7324/ELSR.2017.324249>



**Figure 1. Small-flowered nutsedge (A); Gooseweed (B) in the rice field at Maejo University, Chiang Mai, Thailand**

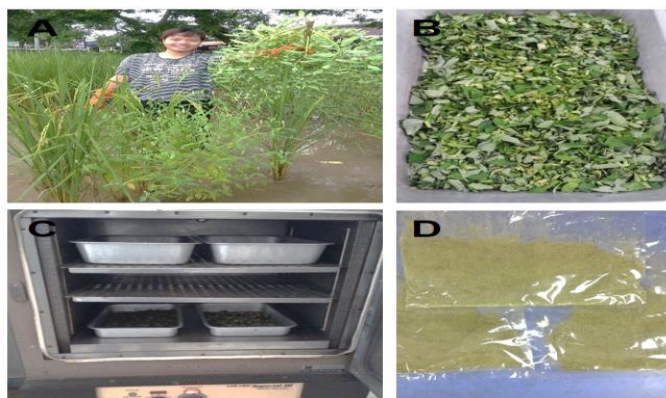
However, using edible biomass for bioethanol production has led to an argument of “food versus fuel” [12]. More lands and other sources such as water, fertilizers, and labors are needed to grow crops for energy [13-14]. Thus, lignocellulosic biomass, so-called second generation of bioethanol, has been preferred due to its abundance, low price, and worldwide distribution [15].

Gooseweed and small-flowered nutsedge both are short life-cycle plants and dominant in wet land areas. In general, they are considered as a problem in the rice field, as they compete with nutrients, water source, sunlight, etc. (Figure 1). Thus, in order to reduce the loss of rice yield, these materials are often taken out manually by farmers or chemical method which causes harmful effect on human health and increases the cost of labor. Hence, although being an invaluable waste, the feasibility of bioethanol production from these two materials should be investigated by calculating the theoretical ethanol yield.

## Methodology

### *Material collection and preparation*

Both gooseweed and small-flowered nutsedge were collected from the rice field at Maejo University, Nong Han, Sansai, Chiang Mai, Thailand (18° 53' 37.4"N; 99° 01' 13.4"E). The two materials were firstly washed with tap water to remove dirt and mud. They were then chopped into 1-2 cm long pieces and dried in hot air oven at 50°C for 3 days. Size reduction was carried out by high-speed blender (Otto BE-127, Thailand) (Figure 2, 3). Dried powder after blending was passed through a 1mm mesh sieve and stored in a desiccator for further experiment.



**Figure 2. Gooseweed: (A) Sample collection; (B) Chopping; (C) Drying in hot air oven; (D) Powdered samples**



**Figure 3. Small-flowered nutsedge: (A) Sample collection; (B) Chopping; (C) Drying in hot air oven; (D) Powdered samples**

### ***Biomass yield***

Biomass yield was calculated by the total mass of plants within a given unit of environment area. Since both gooseweed and small-flowered nutsedge grew in the stagnant area, especially in the rice fields located in Maejo University, Chiang Mai, Thailand (18°53'36.3"N 99°01'14.4"E). A 1 x 1m quadrat was placed in rice field randomly (Figure 4). The two plants were counted, collected and weighted as fresh samples followed by drying in hot air oven until it reached constant weight. The recorded data was used to calculate density (plant/m<sup>2</sup>) and biomass yield (kg/ha).



**Figure 4. 1 X 1 m quadrat in the rice field**

### ***Biochemical analysis***

Reducing sugar was determined by HPLC with following description. Sugars of liquid phase by pre-treatment were analyzed by high performance liquid chromatography (HPLC) (condition: mobile phase-5 mM H<sub>2</sub>SO<sub>4</sub>; flow rate-0.7 mL/min; temperature of column: 60°C; Hi-Plex H column). The amount of cellulose, hemicellulose, and lignin was calculated using the method of fiber analysis reported by Van Soest [18].





### Ethanol estimation procedure

For lignocellulosic biomass, cellulose, a main part of plant cell wall which is formed of many  $\beta$  (1 $\rightarrow$ 4) linked D-glucose units, is an important source of sugar for bioethanol production [15]. Besides, soluble reducing sugars or simple sugars such as monosaccharides (glucose, arabinose, fructose, etc.) that are found outside the cell wall are another source of fermentation substrate. Hence, it can be assumed that sugars from cellulose chains and soluble reducing sugars could be totally converted into bioethanol. As a result, a theoretical ethanol yield could be estimated from amount of cellulose and soluble reducing sugars present in the samples [19, 20]. The conversion of cellulose and reducing sugar into bioethanol were performed according to the below mentioned chemical equations (Eq1, Eq2, and Eq3). By using a balanced chemical equation where total mass of reactants and total mass of products are equal, so-called stoichiometry, theoretical bioethanol yield can be calculated as the below equations (Eq4, Eq5, and Eq6) [14, 19, 21, 22].

Ethanol density: 0.789 g/mL



Ethanol from cellulose (TEC) in 1 g of dry biomass

$$\text{TEC (g)} = \text{cellulose (g)} * 0.57 \quad (\text{Eq4})$$

Ethanol from reducing sugar (TER) in 1 g of dry biomass

$$\text{TER (g)} = \text{Reducing sugar (g)} * 0.51 \quad (\text{Eq5})$$

Ethanol yield from biomass (TEB)

$$\text{TEB (L/Mg)} = (\text{TEC} + \text{TER}) * 1267 \quad (\text{Eq6})$$

## Results and Discussion

### Characteristics of gooseweed

Gooseweed is a kind of tropical weed that grows invasively in damp land, especially in lowland rice field. Table 1 shows the basic classification of gooseweed. Its life cycle is coincident with rice plants and it is often dominant in rice field [24]. The appearance of this plant may cause many unexpected consequences for rice production due to the competition of essential nutrients with rice plants.

Table 1. Taxonomy of gooseweed

Classification	Gooseweed
Kingdom	Plantae
Phylum	Tracheophyta
Class	Magnoliopsida
Order	Campanulales
Family	Campanulaceae
Scientific Name	<i>Sphenoclea zeylanica</i> Gartn.

For these reasons, this plant had been recognized as one of the worst weeds in the world by Holm et al. [25]. A full description about dispersal, ecology, and morphology of gooseweed was reported by Carter et al. [26], since gooseweed had been considered as contaminant of rice feed in North America. In addition, reducing sugars including fructose, xylose, arabinose, and glucose were 19.02 mg/g dry biomass, 3.23 mg/g dry biomass, 2.72 mg/g dry biomass, and 3.63 mg/g dry biomass, respectively (Figure 5). Abundance in

quantity of this material can be a big advantage comparing to sugar/ starch-feedstock for bioethanol production [23].

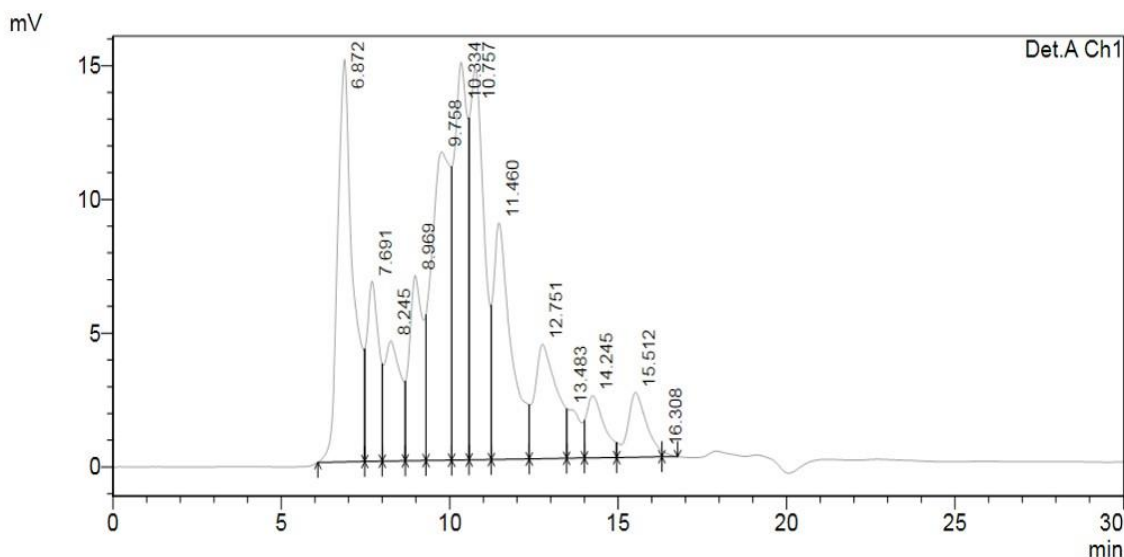


Figure 5. The peaks of sugars from gooseweed released after pretreatment (mobile phase-5 mM H<sub>2</sub>SO<sub>4</sub>; flow rate-0.7 mL/min; temperature of column: 60°C; Hi-Plex H column) [30]

### Characteristic of small-flowered nutsedge

Small-flowered nutsedge, named *Cyperus difformis* L (Table 2), is listed in the Holm's list of the world's worst weeds [25]. It is worldwide distributed and grows in several parts of Thailand [5]. It is an invasive plant which grows on wetland and highly considered as a problematic weed in rice fields that is found anywhere at the bank of water bodies, in the field with crops plant, and its resourceful nature makes it easy to cultivate [28-30]. Though this material can be a good substrate for bioethanol fermentation, very few studies have been done on this comparatively new material [30].

Table 2. Taxonomy of small-flowered nutsedge

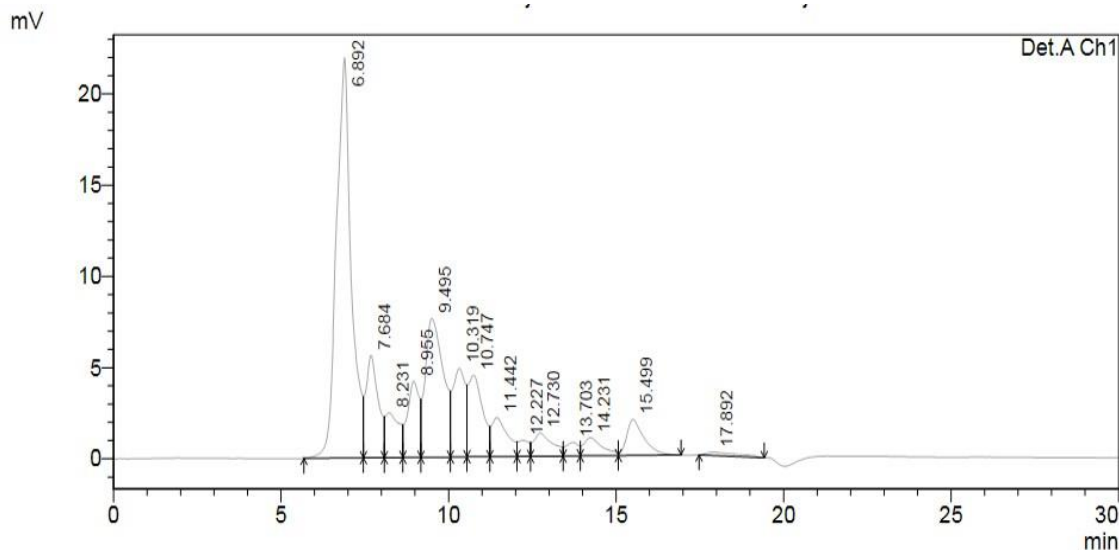
Classification	Gooseweed
Kingdom	Plantae
Phylum	Tracheophyta
Class	Liliopsida
Order	Cyperales
Family	Cyperaceae
Scientific Name	<i>Cyperus difformis</i> L..

The quality and quantity of sugars were analyzed by HPLC after pre-treatment with 1% NaOH and 1% H<sub>2</sub>O<sub>2</sub> (Figure 6). The reducing sugar present in small-flowered nutsedge included 12.1 mg/ g dry biomass, 4.7 mg/g dry biomass, 2.02 mg/g dry biomass, and 1.2 mg/g dry biomass of fructose, glucose, xylose, and arabinose were respectively (Figure 6).

### Biomass yield

The research was conducted in rice fields in which these two weed plants were dominant. The average density of gooseweed and small-flowered nutsedge were 59 plants/m<sup>2</sup> and 38 plant/m<sup>2</sup>, respectively. High density of these plants causes the loss of rice yield due to the competition of nutrients and other essential

elements between weeds and rice plants [31-35]. Region with gooseweed showed 207 kg/ha rice yield, while small-flowered nutsedge produced 201 kg/ha rice yield. Yields varied with season, types of rice plant, and the method of growing rice.



**Figure 6. The peaks of sugars from small-flowered nutsedge released after pretreatment (mobile phase-5 mM H<sub>2</sub>SO<sub>4</sub>; flow rate-0.7 mL/min; temperature of column: 60°C; Hi-Plex H column).**

**Ethanol yield estimation**

Table 3 shows cellulose, reducing sugar contents and theoretical ethanol yield of gooseweed and small-flowered nutsedge.

**Table 3. Cellulose, reducing sugar content and theoretical ethanol yield of gooseweed and small-flowered nutsedge**

Plant	Cellulose (g) *	Reducing sugar (g) *	TEC (g) *	TER (g) *	TEB (L/ Mg)**
<b>Small-flowered nutsedge</b>	0.22 ± 0.001	0.100 ± 0.001	0.125	0.051	223.5
<b>Gooseweed</b>	0.137 ± 0.003	0.096 ± 0.0	0.078	0.049	160.9

\*Performed as g per 1 g dry biomass.

\*Reducing sugar: glucose, fructose, xylose, and arabinose.

\*\* Theoretical ethanol yield (L) per Mg (Ton) of dry biomass.

The components of plant such as cellulose, hemicellulose, lignin, and soluble carbohydrate could be different due to season, environment condition, and age of plant [19]. The average theoretical ethanol yield from gooseweed and small-flowered nutsedge were 160 L/Mg and 223.5 L/Mg, respectively.

**Conclusion**

The yield of gooseweed and small-flowered nutsedge in the rice field were 207 kg/ha and 201 kg/ha, respectively. Several types of sugars were founded such as glucose, fructose, xylose, and arabinose in both materials. Gooseweed and small-flowered nutsedge contained 14% and 22% cellulose, respectively. Gooseweed and small-flowered nutsedge are almost untapped biomass feedstock for bioethanol production



via fermentation. The theoretical ethanol yield of gooseweed and small-flowered nutsedge were 160 L/ Mg and 223 L/Mg respectively. The feasibility of bioethanol production from these two materials should be investigated in future by performing other required laboratory experiments.

### Acknowledgments

The authors would like to acknowledge the support of School of Renewable Energy, Energy Research Center and Plant Physiology & Technology laboratory, Program in Biotechnology, Maejo University for providing facilities.

### References

- [1] Y. J. Kaufman, R. S. Fraser and R. L. Mahoney (1991). Fossil fuel and biomass burning effect on climate-heating or cooling. *J. Climate*, **4**: 578-588.
- [2] R. Wilson (1980). Health effects of fossil fuel burning: assessment and mitigation. Ballinger Publishing Company. ISBN: 9780884107149, <https://books.google.com.tr/books?id=B6iEAAAIAAJ>
- [3] M. F. Demirbas (2011). Biofuels from algae for sustainable development. *Appl. Energy*, **88**: 3473-3480.
- [4] R. Strzalka, D. Schneider and U. Eicker (2017). Current status of bioenergy technologies in Germany. *Renew. Sust. Energ. Rev.*, **72**: 801-820.
- [5] H. Zabed, J. N. Sahu, A. Suelya, A. N. Boycea and G. Faruq (2017). Bioethanol production from renewable sources: Current perspectives and technological progress. *Renew. Sust. Energ. Rev.*, **71**: 475-501.
- [6] K. Smith (2013). Biofuels, air pollution, and health: a global review. Springer Science & Business Media.
- [7] RFA (2017). Ethanol industry outlook, Building partnerships and growing markets, in Washington, DC, February 2017.
- [8] Y. N. Guragain, J. De Coninck, F. Husson, A. Durand and S. K. Rakshita (2011). Comparison of some new pretreatment methods for second generation bioethanol production from wheat straw and water hyacinth. *Bioresour. Technol.*, **102**: 4416-4424.
- [9] P. S. Nigam and A. Singh (2011). Production of liquid biofuels from renewable resources. *Prog. Energy Combust. Sci.*, **37**: 52-68.
- [10] J. Baeyens, Q. Kang, L. Appels, R. Dewil, Y. Lv and T. Tan (2015). Challenges and opportunities in improving the production of bio-ethanol. *Prog. Energy Combust. Sci.*, **47**: 60-88.
- [11] H. Chen and X. Fu (2016). Industrial technologies for bioethanol production from lignocellulosic biomass. *Renew. Sust. Energy Rev.*, **57**: 468-478.
- [12] L. R. Brown (1980). Food or fuel. *Worldwatch paper*, **35**: p.43.
- [13] G. Koçar and N. Civaş (2013). An overview of biofuels from energy crops: Current status and future prospects. *Renew. Sust. Energy Rev.*, **28**: 900-916.
- [14] M. Chen, L. Xia and P. Xue (2007). Enzymatic hydrolysis of corncob and ethanol production from cellulosic hydrolysate. *Int. Biodeterior. Biodegradation*, **59**: 85-89.
- [15] H. B. Aditiya, T. M. I. Mahlia, W. T. Chong, H. Nur and A. H. Sebayang (2016). Second generation bioethanol production: A critical review. *Renew. Sust. Energy Rev.*, **66**: 631-653.
- [16] P. Alvira, E. Tomás-Pejó, M. Ballesteros and M. J. Negro (2010). Pretreatment technologies for an efficient bioethanol production process based on enzymatic hydrolysis: A review. *Bioresour. Technol.*, **101**: 4851-4861.
- [17] S. H. Mood, A. H. Golfeshan, M. Tabatabaei, G. S. Jouzani, G. H. Najafi and M. Gholami et al. (2013). Lignocellulosic biomass to bioethanol, a comprehensive review with a focus on pretreatment. *Renew. Sust. Energy Rev.*, **27**: 77-93.
- [18] J. Yan, Z. Wei, Q. Wang, M. He, S. Li and C. Irbis (2015). Bioethanol production from sodium hydroxide/hydrogen peroxide-pretreated water hyacinth via simultaneous saccharification and fermentation with a newly isolated thermotolerant *Kluyveromyces marxianu* strain. *Bioresour. Technol.*, **193**: 103-109.



- [19] C. Toquero and S. Bolado (2014). Effect of four pretreatments on enzymatic hydrolysis and ethanol fermentation of wheat straw. Influence of inhibitors and washing. *Bioresour. Technol.*, **157**: 68-76.
- [20] Y. Zheng, Z. Pan and R. Zhang (2009). Overview of biomass pretreatment for cellulosic ethanol production. *Int. J. Agri. & Biol. Eng.*, **2**: 51-68.
- [21] P. J. Van Soest, J. B. Robertson and B. A. Lewis (1991). Methods for dietary fiber, neutral detergent fiber, and non-starch polysaccharides in relation to animal nutrition. *J. Dairy Sci.*, **74**: 3583-3597.
- [22] K. P. Vogel, B. S. Dien, H. G. Jung, M. D. Casler, S. D. Masterson and R. B. Mitchell (2011). Quantifying actual and theoretical ethanol yields for switchgrass strains using NIRS analyses. *BioEnergy Res.*, **4**: 96-110.
- [23] K. J. Ptasiński (2016). Efficiency of biomass energy: An exergy approach to biofuels, power, and biorefineries. John Wiley & Sons, ISBN: 9781119118169.
- [24] S. K. Thangavelu, A. S. Ahmed and F. N. Ani (2014). Bioethanol production from sago pith waste using microwave hydrothermal hydrolysis accelerated by carbon dioxide. *Appl. Energy*, **128**: 277-283.
- [25] B. C. Sah, N. N. Nichols, N. Qureshi, G. J. Kennedy, L. B. Iten and M. A. Cotta (2015). Pilot scale conversion of wheat straw to ethanol via simultaneous saccharification and fermentation. *Bioresour. Technol.*, **175**: 17-22.
- [26] P. T. Vu, Y. Unpaprom and R. Ramaraj (2018). Impact and significance of alkaline-oxidant pretreatment on the enzymatic digestibility of *Sphenoclea zeylanica* for bioethanol production. *Bioresour. Technol.*, **247**: 125-130.
- [27] K. Saito, K. Azoma and J. Rodenburg (2010). Plant characteristics associated with weed competitiveness of rice under upland and lowland conditions in West Africa. *Field Crops Res.*, **116**: 308-317.
- [28] L. G. Holm, D. L. Plucknett, J. V. Pancho and J. P. Herberger (1977). The world's worst weeds. University Press.
- [29] R. Carter, J. C. Jones and R. H. Goddard (2014). *Sphenoclea zeylanica* (Sphenocleaceae) in North America-Dispersal, ecology, and morphology. *Castanea*, **79**: 33-50.
- [30] C. Bryant (2010). Third Berkeley Conference on Bioeconomy.
- [31] O. Ueno and T. Takeda (1992). Photosynthesis pathways, ecological characteristics, and the geographical distribution of the Cyperaceae in Japan. *Oecologia*, **89**: 195-203.
- [32] B. Sanders (1994). The life cycle and ecology of *Cyperus difformis* (rice weed) in temperate Australia: a review. *Aust. J. Exp. Agr.*, **34**: 1031-1038.
- [33] R. Vishwakarma and R. Banerjee (2016). Enhancement of sugar content of *Cyperus* sp. through cellulolytic enzymes for bioethanol generation. *Lignocellulose*, **5**: 94-105.
- [34] B. Chauhan and D. Johnson (2009). Ecological studies on *Cyperus difformis*, *Cyperus iria* and *Fimbristylis miliacea*: three troublesome annual sedge weeds of rice. *Ann. Appl. Biol.*, **155**: 103-112.
- [35] M. O. Mabbayad and A. K. Watson (1995). Biological control of gooseweed (*Sphenoclea zeylanica* Gaertn.) with an *Alternaria* sp. *Crop Prot.*, **14**: 429-433.