Tumor Markers- A Review

Nagalaxmi Velpula¹, Chandra Neela Yathavakilla^{2,*}, Aditi Ramesh³

¹Professor & HOD, ²PG Student, ³Assitant Professor, Dept. of Oral Medicine & Radiology, Sri Sai College of Dental Surgery, Vikarabad, Telangana

*Corresponding Author:

Email: chandraneelay@gmail.com

Abstract

The role of tumor markers in clinical oncology has increased tremendously over last decade, a trend that continues to grow as technology progresses and our understanding about human body and the disease processes increase. Tumour markers have wide applications in cancer care, starting from screening, choosing modality of management, assessment of prognosis to follow-up after treatment. Their judicious use in clinical practice needs a thorough understanding of the basics of pathophysiology and techniques of identification in any given malignancy. Of the numerous tumor markers identified, described and extensively researched upon, only a handful of them are used in routine clinical practice; and even of these, only a few have support of established consensus guidelines for use in day- to-day care of patients. This article discusses the uses of tumor markers, classification, some commonly used tumour markers and general principles for their optimal use.

Keywords: Tumor marker, Serum markers, Salivary markers, Early diagnosis, Malignancy

Introduction

Tumor markers are molecules that may be present in higher than usual concentrations in the tissue, serum, urine, or other body fluids of patients with cancer. (1-4) Apart from being useful as screening tests in diagnosing malignancy, they can also be used in assessing prognosis, guiding choice of treatment, and to monitor progress during and after treatment. In a relevant clinical setting, they undoubtedly help treating physician in management of malignancy, but also understanding the limitations of tumour markers is of utmost importance for their judicious use.

Ideal Tumour Marker

Although the characteristics of an ideal tumor marker depend to some degree on the classification and application of the marker, the general properties of such an ideal tumor marker include:⁽⁵⁾

- 1. Specific production by premalignant or malignant tissue early in the progression of disease;
- 2. Produced at detectable levels in all patients with a specific malignancy;
- 3. Expression in an organ site-specific manner;
- 4. Evidence of presence in bodily fluids obtained noninvasively or in easily accessible tissue;
- 5. Levels related quantitatively to tumor volume, biological behavior, or disease progression;
- Relatively short half-life, reflecting temporal changes in tumor burden and response to therapy; and
- 7. Existence of a standardized, reproducible, and validated objective and quantitative assay.

Uses of Tumour Markers

- 1. Screening for early malignancy
- 2. Acting as a diagnostic aid for malignancy

- 3. Predicting therapeutic efficacy-e.g., tissue markers such as estrogen receptor and HER-2 (human epidermal growth factor receptor-2) predicting response to endocrine therapy and trastuzumab respectively.⁽⁶⁾
- 4. Monitoring therapy in advanced malignancy. (3)
- 5. Determining prognosis in malignancy
- 6. Maintaining surveillance following surgery of the primary tumor and for detecting disease recurrence.

Limitations of Tumour Markers

- 1. Tumor markers are not in themselves specific enough to permit a diagnosis of malignancy to be made and not an alternative for histopathological diagnosis (biopsy).
- 2. False positives lead to unnecessary anxiety and further investigations
- 3. False negatives lead to false assurance, delayed diagnosis, and advancement of malignancy
- 4. They may be elevated in benign conditions
- 5. Economic implications

Classification of Tumour Markers

Tumor markers can be broadly classified based on the type of tissue as follows:⁽⁸⁾

- a. Epithelial markers
 - Cell surface markers Histocompatibility
 - Intracellular markers Cytokeratins
 - Basement membrane markers Type 4 collagen
 - Matrix markers Tenascin
 - Membrane antigen Blood group antigens.
- b. Connective tissue markers
 - Intermediate filament proteins Desmin
 - Other filament proteins Laminin
 - Cellular enzymes Amylase, lysozyme

- Cytoplasmic non-filamentous non-enzymatic proteins Myoglobin, S100 protein
- Membrane antigen Leukocyte specific antigen.
- c. Salivary gland markers
 - Epithelial markers Cytokeratins
 - Myoepithelial cell markers Actin, myosin
 - Serum acinar cell markers Salivary amylase
 - Myoepithelial cells + acinar cells S100 protein.

Classification depending on Functional Utility

Tumour markers can also be classified depending on their functional utility into following types according to NCCN (National Comprehensive Cancer Network). (9)

Diagnostic Markers: These markers are mainly useful in establishing the disease from analysis of patient's sample (serum, body fluid, tissue etc.). Demonstration of Philadelphia chromosome in chronic myelogenous leukemia through fluorescence in situ hybridization (FISH) technique is an example of diagnostic marker. (10)

Prognostic Markers: Prognostic markers aid in assessment of disease outcomes, such as overall survival independent of the management options. (11) An example of a prognostic marker is *p53* mutations, which correlates with aggressive disease course and poor outcomes. (12)

Predictive Markers: Predictive markers help in assessing response of a tumour to a particular modality of management, thus aid in treatment planning. (11) An example is favourable response of *HER2* (Human epidermal growth factor receptor 2) positive breast cancer patients to trastuzumab. (13-15)

Companion Diagnostic Markers: Companion diagnostic markers may be diagnostic, prognostic, or predictive, but are used to identify a subgroup of patients for whom a therapy has shown benefit (*BRAF* V600E mutation in melanoma). (16, 17)

There are numerous tumour markers which may be diagnostic, prognostic or predictive that are used in day to day clinical practice for various neoplastic conditions. Most commonly used tumour markers are given in **Table 1**.⁽¹⁸⁾

Molecular basis of Tumour Markers: Genetic alteration in a tumor cell affects directly or indirectly the gene expression pattern of the tumor cell or the surrounding tissue. (19-20) These genetic alterations can be reflected at various levels [Table 2], from genetic defects like mutation, deletion of gene to viral genomic incorporation, forming the molecular basis of tumour markers. (21)

Methods of Detection of Tumour Markers: There are several methods used in the detection of tumour markers, of which serological enzyme assay is the most commonly used method. Tumour markers can also be detected through immunohistochemistry (*IHC*), radioimmunoassay (*RIA*), or enzyme-linked

immunosorbent assay (*ELISA*). (22-24,26,27) Immunological detection is based on Monoclonal antibodies specifically binding to epitopes on tumor markers, which can be identified with dyes in immunohistochemistry (*IHC*), radioactive tags in radioimmunoassay (*RIA*), or enzymes in enzyme-linked immunosorbent assay (*ELISA*). Flow cytometry is an alternative method to analyze the presence and percentage of antibody-tagged cells in a suspension. (26–28)

The above mentioned methods are highly sensitive and are useful in semi-quantitative or quantitative estimation of tumour markers. IHC is the most commonly used method in the detection of tumour markers nowadays. Immunohistochemistry (IHC) in oncology is used to categorize undifferentiated malignant tumors, leukemias and lymphomas, to determine the site of origin of metastatic tumors and also to detect the molecules of prognostic or therapeutic significance (e.g., Estrogen/progesterone receptors (ER/PR) in breast cancer). (22-26)

Source of Tumour Markers: Tumor markers can be detected either in tissue (tissue tumor markers; for example, in solid tumors, lymph nodes, bone marrow or circulating tumor cells in the blood) or in body fluids like ascitic or pleural fluid or serum (serological tumor markers). (26,28) Tissue tumor markers are of prime importance to a diagnostic pathologist, while the serological tumor markers are more often used by a clinician.

Specific tumour markers in Oral neoplasms

While many tumour markers appear attractive theoretically, none of them is either specific or sensitive enough to be used as a mass screening test in low risk population. Some of the tumour markers and their significance in management of oral neoplasms is given in **Table 3**.

Salivary biomarkers

With new techniques of detecting small quantities of salivary components including proteins and messenger Ribonucleic acid (mRNA), almost anything that can be measured in blood can be measured in saliva. Hence, saliva is considered as the blood stream of oral cavity. Alterations in the levels of certain mRNA molecules and of certain proteins have been detected in several cancers, including oral cancers indicating their possible use in cancer detection and follow-up. Tumour markers CA 15-3, Her2/neu and CA 125 are found in saliva.

In breast cancer cases, Her2/neu is the first salivary biomarker reported. The levels of CA 15-3, Her2/neu are raised and p53 levels are low. Raised levels of CA 125 in the saliva is noticed in ovarian cancer cases. (39)

Saliva can also be used for the detection of oral cancer. p53 gene is mutated in the salivary DNA of oral cancer patients. IL-8, SAT, IL-1B, OAZ 1, H3F3A,

DUSP and S100P are some of the OSCC associated RNAs present in saliva. (40) Increased salivary levels of cell cycle regulatory proteins Cyclin D1 and Ki67, lactate dehydrogenase (LDH), matrix metalloproteinase (MMP)-9, reduction in DNA repair enzyme, 8-oxoquanine DNA glycosylase (OGG1) and tumor suppressor protein, Mapsin is noticed in oral cancer patients. (39)

Human Plasma Proteome

The human plasma proteome holds the promise of a revolution in disease diagnosis and therapeutic monitoring, provided that major challenges in proteomics and related disciplines can be addressed.

Plasma is not only the primary clinical specimen, but also represents largest and deepest version of the human proteome present in any sample. In addition to the classical "plasma proteins", it contains all tissue proteins (as leakage markers) plus very numerous distinct immunoglobulin sequences. Current progress in proteomics has been largely due to recent developments in mass spectrometry (MS) - based technologies. New techniques for the ionization of proteins and peptides, such as matrix assisted laser desorption-ionization (MALDI) and electrospray ionization (ESI) combined with time of flight (TOF), as well as new hybrid mass spectrometers, are now becoming the tools of choice for protein characterization.

Urine

Lida W Chan et al (2004)⁽²⁹⁾ determined the predictive value of urinary levels of two angiogenic factors, vascular endothelial growth factor (VEGF) and matrix metalloproteinase (MMPs), in a longitudinal study to determine their correlation with 1-year progression-free survival in patients with cancer. This small exploratory study suggests that the angiogenic urinary trends of VEGF and MMPs may be useful predictive markers for progression-free survival in cancer patients after the completion of radiotherapy.

Judicious use of tumor marker

One has to keep in mind following things while asking for tumour markers:

- 1. They can be raised in several benign or nonneoplastic conditions
- Population based screening of asymptomatic people with most serum tumour markers is not recommended owing to low diagnostic sensitivity and specificity
- 3. Pitfalls in assessing prognosis: results within normal limits do not exclude malignancy or progression
- 4. Tumour markers cannot replace biopsy or histopathology for establishing the primary diagnosis of cancer
- Tumour marker results are often method dependent
 patients should, ideally, be monitored using the

- same method and its name indicated on the report form
- Tumour marker results should always be interpreted in the context of all available information including clinical findings, imaging investigations, and other blood tests (such as renal and liver function and hematological tests).

Table 1: Table representing most commonly used tumour markers in clinical practice. (18)

Cancer	Non neoplastic			
	Conditions			
Gestational trophoblastic disease, Gonadal germ cell tumor	Pregnancy			
Medullary cancer Thyroid				
Pheochromocytoma				
Oncofetal Antigens				
Hepatocellular carcinoma, Gonadal germ cell tumor	Cirrhosis, Hepatitis			
Adenocarcinomas of the Colon, Pancreas, Lung, Breast, Ovary	Pancreatitis, Hepatitis, IBD, Smoking			
Prostate cancer	Prostatitis, Prostatic hypertrophy			
Small-cell cancer of the lung, Neuroblastoma				
Lymphoma, Ewing's sarcoma	Hepatitis, Hemolytic anemia,			
Tumor-Associated Proteins				
Prostate cancer	Prostatitis, Prostatic hypertrophy			
Myeloma	Infection, MGUS			
Ovarian cancer, some lymphomas	Menstruation, Peritonitis, Pregnancy			
Colon, Pancreatic, Breast cancer	Pancreatitis, Ulcerative colitis			
Hodgkin's disease, Anaplastic large-cell lymphoma				
Hairy cell leukemia, Adult T cell leukemia/lymphoma				
	trophoblastic disease, Gonadal germ cell tumor Medullary cancer Thyroid Pheochromocytoma Hepatocellular carcinoma, Gonadal germ cell tumor Adenocarcinomas of the Colon, Pancreas, Lung, Breast, Ovary Prostate cancer Small-cell cancer of the lung, Neuroblastoma Lymphoma, Ewing's sarcoma eins Prostate cancer Myeloma Ovarian cancer, some lymphomas Colon, Pancreatic, Breast cancer Hodgkin's disease, Anaplastic large-cell lymphoma Hairy cell leukemia, Adult T cell			

MGUS: Monoclonal gammopathy of uncertain significance.

Table 2: Molecular basis of Tumour Markers⁽²¹⁾

	Table 2: Molecular basis of Tumour Markers		
Levels of	Examples		
classification			
DNA	Promoter Hyper-methylation, e.g.,		
Epigenetic	GSP1, DAP in lung cancer; p15,		
	p16 in liver cancer		
Endogenous	Mutations, e.g., NADH		
Mitochondrial	dehydrogenase 4 (ND4) in urine in		
genetic	bladder cancer		
Oncogene	Mutation, e.g., K-ras in pancreatic		
	cancer;		
	micro-satellite alterations in head		
	and neck cancers		
Exogenous	EBV in NPC, Burkitt's lymphoma;		
viral RNA	HPV in cervical cancer		
Cell based	Tissue-specific markers, e.g., PSA		
endogenous	mRNA in prostate cancer,		
	cytokeratin 20 mRNA in breast		
	cancer		
Cell free	Circulating mRNA, e.g.,		
	Tyrosinase mRNA in melanoma		
Exogenous	Viral RNA, e.g., EBV-coded RNA		
viral	in NPC		
Translational			
protein			
Native protein	PSA in prostate cancer, CEA in		
	colonic cancer		
Glycan	Aberrant glycosylation, e.g.,		
110	monosialytactec AFP in HCC		

AFP: Alfa fetoprotein; CEA: Carcinoembryonic antigen;

EBV: Epstein-Barr Virus; HCC: Hepatocellular carcinoma;

HPV: Human papilloma virus; mRNA: Messenger RNA;

NPC: Nasopharyngeal carcinoma; PSA: Prostate-specific antigen

Table 3: Tumour markers significant in oral neoplasms

Tumour Markers	Significance	
VEGF, EGFR, TGF-α	Prognostic information in	
, Cyclin D1 ⁽³⁰⁾	SCCHN	
Mutation in the p53	Identifying individuals at	
tumour suppressor	high risk of SCCHN	
gene ⁽³¹⁾		
bcl-2 ⁽³²⁾	Prognostic indicator in	
	early SCCHN	
Proto-oncogene eIF4E	SCCHN, Premalignant	
$(4E)^{(33)}$	lesions of the larynx	
Beta 2-	Oral sub mucous fibrosis	
microglobulin ⁽³⁴⁾	and Oral cancer	
Cathepsin D ⁽³⁵⁾	Independent predictor of	
	cervical lymph node	
	metastasis in SCCHN	
Cytokeratins-CK19	Markers of sequential	
and CK8 ⁽³⁶⁻³⁷⁾	premalignant changes in	

	head and neck cancer
Non expression of	May be an early event
CK5 ⁽³⁸⁾	occurring in tobacco-
	associated pathological
	changes in the buccal
	mucosa
MMP1 (Matrix	Diagnostic markers of
Metallo Proteins)	SCCHN

SCCHN: Squamous Cell Carcinoma of Head and Neck

Conclusion

Hence with an evolution in the understanding of genetics and molecular basis of human malignancies, tumor markers have been better understood and are being progressively used not only in determination of the risk of tumors but also in treatment guidelines and decisions. A wide variety of tumor markers have been described in the literature but only a few have proved to be clinically useful, and therefore tumor markers cannot be construed as primary modalities but can be used as adjuncts in diagnosis and treatment planning of cancer.

References

- Diamandis EP, Hoffman BR, Sturgeon CM. National Academy of Clinical Biochemistry laboratory medicine practice guidelines for the use of tumor markers. Clinical chemistry. 2008 Nov 1;54(11):1935-9.
- Lai LC, Cheong SK, Goh KL etal. Clinical usefulness of tumour markers. The Malaysian journal of pathology. 2003 Dec;25(2):83-105.
- Duffy MJ. Role of tumor markers in patients with solid cancers: a critical review. European journal of internal medicine. 2007 May 31;18(3):175-84.
- 4. Sturgeon CM, Lai LC, Duffy MJ. Serum tumour markers: how to order and interpret them. Bmj. 2009 Sep 22;339(7725):852-8.
- Bigbee W, Herberman RB. Characteristics of the ideal tumor marker.
- Duffy MJ, Crown J. A personalized approach to cancer treatment: how biomarkers can help. Clinical chemistry. 2008 Nov 1;54(11):1770-9.
- Duffy MJ. Evidence for the clinical use of tumour markers. Annals of clinical biochemistry. 2004 Sep 1;41(5):370-7.
- Cuperlovic-Culf M, Belacel N, Ouellette RJ.
 Determination of tumour marker genes from gene expression data. Drug discovery today. 2005 Mar 15;10(6):429-37.
- Febbo PG, Ladanyi M, Aldape KD et al. NCCN Task Force report: Evaluating the clinical utility of tumor markers in oncology. Journal of the National Comprehensive Cancer Network. 2011 Nov 1;9(Suppl 5):S-1.
- Faderl S, Talpaz M, Estrov Z et al. The biology of chronic myeloid leukemia. New England Journal of Medicine. 1999 Jul 15;341(3):164-72.
- Altman DG, McShane LM, Sauerbrei W et al. Reporting recommendations for tumor marker prognostic studies (REMARK): explanation and elaboration. BMC medicine. 2012 May 29;10(1):1.
- Goldstein I, Marcel V, Olivier M et al. Understanding wild-type and mutant p53 activities in human cancer: new landmarks on the way to targeted therapies. Cancer gene therapy. 2011 Jan 1;18(1):2-11.

- Joensuu H, Kellokumpu-Lehtinen PL, Bono P et al. Adjuvant docetaxel or vinorelbine with or without trastuzumab for breast cancer. New England Journal of Medicine. 2006 Feb 23:354(8):809-20.
- Piccart-Gebhart MJ, Procter M, Leyland-Jones B et al. Trastuzumab after adjuvant chemotherapy in HER2positive breast cancer. New England Journal of Medicine. 2005 Oct 20:353(16):1659-72.
- 15. Romond EH, Perez EA, Bryant J et al. Trastuzumab plus adjuvant chemotherapy for operable HER2-positive breast cancer. New England Journal of Medicine. 2005 Oct 20;353(16):1673-84.
- Chapman PB, Hauschild A, Robert C et al. Improved survival with vemurafenib in melanoma with BRAF V600E mutation. New England Journal of Medicine. 2011 Jun 30;364(26):2507-16.
- Pollock PM, Meltzer PS. A genome-based strategy uncovers frequent BRAF mutations in melanoma. Cancer cell. 2002 Jul 31;2(1):5-7.
- Longo DL, Fauci AS, Kasper DL, Hauser SL, Jameson JL, Loscalzo J. Harrison's Principles of Internal Medicine 19E Vol 2 EB. McGraw Hill Education; 2015.
- Sharma S. Tumor markers in clinical practice: General principles and guidelines. Indian Journal of Medical and Paediatric Oncology. 2009 Jan 1;30(1):1.
- Evers BM, Townsend CM, Thompson JC. Small intestine. Sabiston textbook of surgery. 2004;2:1334-42.
- Johnson PJ. A framework for the molecular classification of circulating tumor markers. Annals of the New York Academy of Sciences. 2001 Sep 1;945(1):8-21.
- Waxman J. Tumor markers. Quart J Med. 1995;88:233–41.
- Sokoll LJ, Chan DW. Clinical chemistry: Tumor markers. Clinical Oncology. 3rd ed. Philadelphia: Elsevier Churchill Livingstone. 2004;329-39.
- Schrohl AS, Holten-Andersen M, Sweep F et al. Tumor markers from laboratory to clinical utility. Molecular & Cellular Proteomics. 2003 Jun 1;2(6):378-87.
- Evers BM, Townsend CM, Thompson JC. Small intestine. Sabiston textbook of surgery. 2004;2:1334-42.
- Cooper DL. Tumor markers. In: goldman, et al., editors. In: Cecil textbook of medicine. 22nd ed. Philadelphia: WB Saunders Company; 2004.
- Lee P, Pincus MR, McPherson RA. Diagnosis and management of cancer using serologic tumor markers. Henry's clinical diagnosis and management by laboratory methods. 21st ed. Philadelphia: Saunders-Elsevier. 2007:1362-3.
- Lindblom A, Liljegren A. Tumor markers in malignancies. Br Med J. 2000;320:424–7.
- 29. Chan LW, Moses MA, Goley E et al. Urinary VEGF and MMP levels as predictive markers of 1-year progressionfree survival in cancer patients treated with radiation therapy: a longitudinal study of protein kinetics throughout tumor progression and therapy. Journal of clinical oncology. 2004 Feb 1;22(3):499-506.
- Roger P, Montcourrier P, Maudelonde T et al. Cathepsin D immunostaining in paraffin-embedded breast cancer cells and macrophages: correlation with cytosolic assay. Human pathology. 1994 Sep 30;25(9):863-71.
- Weidner N, Folkman J, Pozza F et al. Tumor angiogenesis: a new significant and independent prognostic indicator in early-stage breast carcinoma. Journal of the National Cancer Institute. 1992 Dec 16;84(24):1875-87.
- Eisma RJ, Spiro JD, Kreutzer DL. Vascular endothelial growth factor expression in head and neck squamous cell

- carcinoma. The American journal of surgery. 1997 Nov 30;174(5):513-7.
- Gadbail AR, Chaudhary M, Patil S et al. Actual Proliferating Index and p53 protein expression as prognostic marker in odontogenic cysts. Oral Diseases. 2009 Oct 1;15(7):490-8.
- 34. Gandour-Edwards R, Trock B, Donald PJ. Predictive value of cathepsin-D for cervical lymph node metastasis in head and neck squamous cell carcinoma. Head & neck. 1999 Dec 1;21(8):718-22.
- Kartner N, Ling V. Multidrug resistance in cancer. Sci Am. 1989 Mar 1;260(3):44-51.
- 36. Hoffmann TK, Ballo H, Braunstein S et al. Serum level and tissue expression of c-erbB-1 and c-erbB-2 protooncogene products in patients with squamous cell carcinoma of the head and neck. Oral oncology. 2001 Jan 31;37(1):50-6.
- 37. Ito T, Kawabe R, Kurasono Y et al. Expression of heat shock proteins in squamous cell carcinoma of the tongue: an immunohistochemical study. Journal of oral pathology & medicine. 1998 Jan 1;27(1):18-22.
- 38. Jass JR. Broadsheet number 52: Molecular genetics of colorectal cancer. Pathology. 1999 Jan 1;31(4):354-64.
- 39. Devaraj SD. Salivary Biomarkers-A Review.
- Elashoff D, Zhou H, Reiss J et al. Prevalidation of salivary biomarkers for oral cancer detection. Cancer Epidemiology Biomarkers & Prevention. 2012 Apr 1;21(4):664-72.