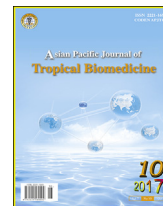




Contents lists available at [ScienceDirect](http://www.sciencedirect.com)

Asian Pacific Journal of Tropical Biomedicine

journal homepage: www.elsevier.com/locate/apjtb



Original article <http://dx.doi.org/10.1016/j.apjtb.2017.09.014>

Yeast-generated CO₂: A convenient source of carbon dioxide for mosquito trapping using the BG-Sentinel® traps



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ARTICLE INFO

Article history:

Received 9 Aug 2017

Received in revised form 30 Aug 2017

Accepted 8 Sep 2017

Available online 20 Sep 2017

Keywords:

Trinidad and Tobago

Carbon dioxide bait

BG sentinel trap

Attractant

ABSTRACT

Objectives: To evaluate carbon dioxide (CO₂) production from yeast/sugar mixtures and its efficiency as an attractant in BG-Sentinel traps.

Methods: The rate of CO₂ production was optimized for different yeast/sugar mixtures. The optimized mixture was then used as bait in BG-Sentinel traps. The efficiency of this bait was then compared to octenol baited traps.

Results: The yeast/sugar (5 g: 280 g) in 300 mL water generated the highest volume of CO₂. The CO₂ baited traps caught significantly more mosquitoes than octenol baited traps.

Conclusions: Yeast-produced CO₂ can effectively replace octenol baits in BG traps. This will significantly reduce costs and allow sustainable mass-application of the CO₂ baited traps in large scale surveillance programs.

1. Introduction

Chemical cues such as carbon dioxide (CO₂) are important for the host-finding behaviour of mosquitoes [1–4]. Surveillance programs which utilize mosquito trapping often include chemical baits such as CO₂, octenol, nonanol or lactic acid to increase catch rates [5–11]. In Trinidad and Tobago, surveillance and/or sampling exercises utilise the BG-Sentinel® trap [12] baited with either octenol or dry ice which generates CO₂. However, one of the limiting factors is the cost and availability of octenol and dry ice [13,14]. The BG-Sentinel® trap is a well-established monitoring tool for capturing mosquitoes [15], however, the effectiveness of yeast/sugar generated CO₂ bait has not been evaluated for use in Trinidad and Tobago.

Various studies have previously reported on the efficacy of octenol and carbon dioxide (CO₂) as attractants in mosquito traps such as the Fay-Prince trap, CDC-type and the Encephalitis Virus Surveillance traps [15–24]. These studies suggest that octenol may have species-specific effects. Kleine *et al.* [16] also reported that octenol differs in its effectiveness for

attracting different mosquito species. However, Octenol does not appear to be a strong attractant for *Stegomyia* mosquitoes which includes *Aedes aegypti* (*Ae. aegypti*), an important vector for the spread of tropical diseases such as dengue, zika and chikungunya, which has a high prevalence rate in the Caribbean region. Canyon and Hii [6] reported that octenol significantly decrease collection of *Ae. aegypti* when compared to carbon dioxide using Fay-Prince traps. Shone *et al.* [25] reported that species such as *Aedes albopictus* was attracted more to CO₂ and CO₂+ octanol baited CDC and Fay-Prince traps than unbaited or octenol-baited traps. These studies all suggest that the choice of bait can effectively increase the catch efficiency of a mosquito trap.

In Trinidad and Tobago, the preferred bait is mosquito traps is octanol, though dry ice is sometimes used. However this is neither cost effective nor sustainable when traps are deployed in remote areas or when large scale sampling is needed, as is the case when there is an upsurge in the number of cases of dengue, zika and chikungunya. In tropical environments, dry ice sublimates faster than in temperate areas and has to be replaced frequently. Moreover, dry ice has the disadvantage that the release rate of CO₂ is highly variable and diminishes over time [26,27]. When large scale trapping is required, such as during a national surveillance program, the use of both dry ice and octenol can become prohibitively expensive. To overcome these limitations, CO₂ produced by fermentation of sugar can

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Peer review under responsibility of Hainan Medical University. The journal implements double-blind peer review practiced by specially invited international editorial board members.

be a reliable alternate that is cheap, easy to manage and durable. This paper seeks to optimize CO₂ production from fermentation of sugar and compare its efficiency to octenol for capturing mosquito in baited BG-sentinel traps.

2. Materials and methods

2.1. Carbon dioxide production from yeast/sugar mixture

Three hundred (300) mL of 130, 190, 250 and 280 g/L sugar solutions each containing 3 g of baker's yeast was prepared in 500 mL Buchner flasks and maintained at 35 °C in a water bath. The rate of gas production was determined using a displacement method. The buchner flask was connected to a sealed conical flask filled with water and fitted with a displacement tube which emptied into a graduated cylinder. The volume of fluid displaced per unit time was used to calculate CO₂ production rates over 24 h.

2.2. Optimization of yeast for carbon dioxide production

Four reaction flasks containing 280 g/L sugar solution were prepared to assess the effects of yeast on the CO₂ production. Aliquot 1.5 g and 5 g of yeast was added to duplicate flask, and the rate of gas production was monitored over 24 h. Estimation of the carbon dioxide production was determined as described previously.

2.3. Field evaluation of yeast-generated carbon dioxide in mosquito collections

The optimized reaction mixture was then tested in field trials using the BG trap. Carbon dioxide baited traps were run simultaneously with octenol baited traps to compare the catch efficiency of the two baits. A total of 45 sampling efforts were conducted from January to May 2017 at two sample locations: (1) Open green house and (2) in dwellings. The total number of mosquitoes was counted and the number of species compared.

3. Results

The volume of carbon dioxide generated from the sugar solution with 3 g of yeast generally increased over the first (4–6) h then gradually decreased (Figure 1). Production rate after 1 h ranged between 3.1 mL/min (130 g/L solution) and 6.2 mL/min (280 g/L solution) (Figure 1). The 280 g/L solution generated significantly ($P < 0.05$) higher levels of CO₂ when compared to the 130 g/L solution. However CO₂ production levels in 280 g/L solution were not significantly higher than production from 190 g/L and 250 g/L solutions. The total volume of CO₂ generated varied between 3 L (130 g/L solution) and 5 L (280 g/L solution) over the first 7 h. After 24 h, CO₂ production rates significantly decreased, ranging between 0.73 mL/min (130 g/L solution) and 4.4 mL/min (280 g/L solution). However, over the 24 h period all the mixtures were still generation CO₂ at levels that were higher than the estimate CO₂ release rate of (1–1.8) mL/h from human skin [28]. This would suggest that the system would continue to attract mosquitoes over an extended period of time.

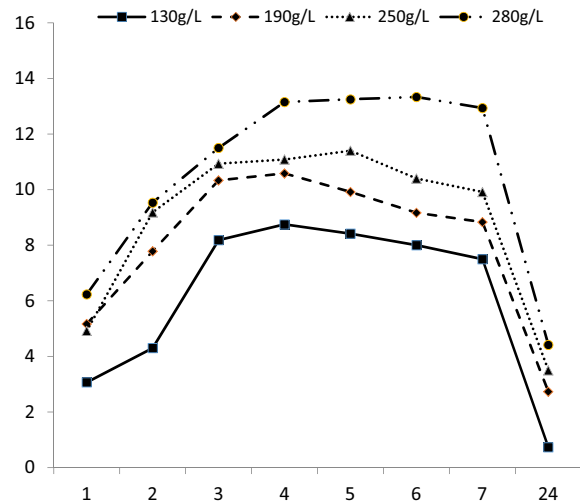


Figure 1. CO₂ production rate using different masses of sugar.

3.1. Optimization of yeast for carbon dioxide production

The amount of carbon dioxide generated can be influenced by the rate of fermentation and the amount of yeast added. From the first experiment, the solution containing 280 g of sugar produced the largest volume of CO₂ for the longest time period. Varying yeast concentration (1.5 g, 3.0 g and 5.0 g) significantly increased CO₂ production from the sugar mixture (Figure 2). Production rate after 1 h ranged between 3 mL/min (1.5 g yeast and 280 g/L sugar solution) and 10 mL/min (5 g yeast and 280 g/L sugar solution) (Figure 2). The solution containing 5 g of yeast produced significantly ($P < 0.05$) more CO₂ than the one containing 1.5 g of yeast (Figure 2). After 7 h CO₂ production rate ranged between 6.8 mL/min (1.5 g yeast and 280 g/L sugar solution) and 14.1 mL/min (5 g yeast and 280 g/L sugar solution) (Figure 2). After 24 h CO₂ production rates were still high, ranging between 5.9 mL/min (1.5 g yeast and 280 g/L sugar solution) and 5.5 mL/min (5 g yeast and 280 g/L sugar solution) (Figure 2). The highest level of CO₂ was produced after 3 h averaging 8.8 mL/min (1.5 g yeast) and 21.7 mL/min for the mixture with 5 g of yeast.

The total volume of CO₂ generated varied between 3 L (1.5 g yeast in 280 g/L sugar solution) and 7 L (5 g yeast in 280 g/L sugar solution) within the first 7 h. After 24 h production rates were significantly reduced ranging between 6 mL/min (1.5 g yeast in 280 g/L sugar solution) and 4.4 mL/min (3 g in 280 g/L sugar solution). However over the 24 h period all the reaction mixtures were still generating sufficiently high levels of CO₂.

3.2. Field evaluation of yeast-generated carbon dioxide, in mosquito collections

A total of 45 field trials (30 open green house and 15 in dwellings) were conducted using both CO₂ baited and octenol baited BG traps. *Culex quinquefasciatus* (*Cx. quinquefasciatus*) was the dominant species caught at both sites. A total of 842 mosquitoes were collected which consisted primarily of *Cx. quinquefasciatus* (84.1%) and *Ae. aegypti* (15.9%). The total number of mosquitoes collected with the CO₂ baited BG traps (620 mosquitoes) was three times higher than the number collected with the octenol baited (222 mosquitoes) traps. This

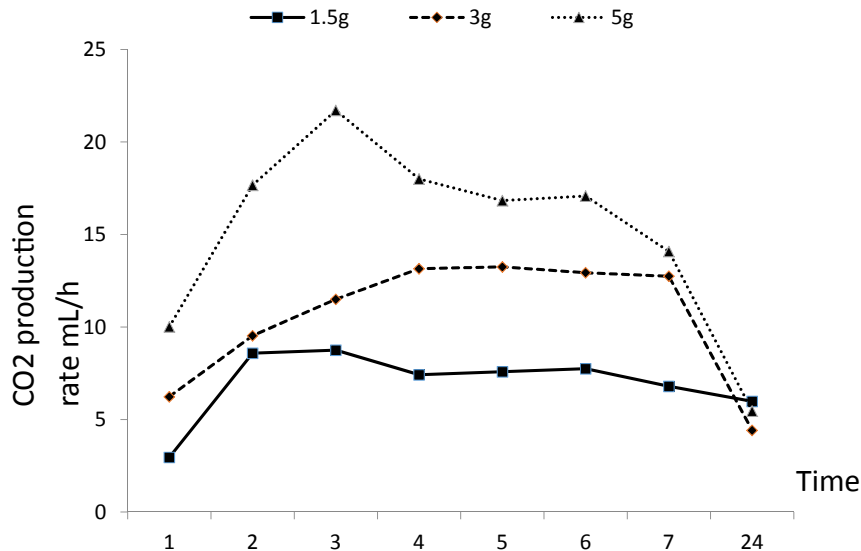


Figure 2. CO₂ production rated from varying masses of yeast.

would suggest that the catch efficiency of CO₂ baited traps was greater than the octenol bait.

The CO₂ baited traps attracted about 4 times more *Cx. quinquefasciatus* (555) than octenol (153) baited traps. However both baits attracted similar numbers of *Ae. aegypti*. In the open green house, the CO₂ baited traps collected twice as much *Cx. quinquefasciatus* (216) than the octenol (112) baited traps. The CO₂ bait traps collected 20% less *Ae. aegypti* (40) when compared to the octenol baited (60). At the site within the dwellings, the CO₂ baited collected eleven times more *Cx. quinquefasciatus* (339) when compared to the octenol baited traps (41). The CO₂ baited traps also collected three times more *Ae. aegypti* (25) than the octenol baited traps (9). The octenol baited traps had 42% males and 58% females while the CO₂ baited trap had 52% males and 48% females. However, all of *Cx. quinquefasciatus* collected were females.

4. Discussion

Mosquitoes respond to a complex set of cues such as carbon dioxide, lactic acid or temperature to locate a host. In Trinidad and Tobago, surveillance programs utilize the BG sentinel traps, which have been shown to be more effective in capturing *Aedes* sp. than other traps such as the CDC-LT [8,29]. The BG traps are normally baited with octenol, however, the cost can be prohibitive for large scale monitoring. This study showed that carbon dioxide generated from yeast/sugar mixtures can be a more efficient attractant than octenol in BG traps. The optimum mixture used in this study, which produced the highest amount CO₂ was 5 g yeast and 280 g/L sugar solution.

Various studies previously reported on the catch efficiency of different attractants. Several studies have also reported that few species were attracted to octenol alone, and catch rates increases when it is used in combination with other attractants [22,23,30]. Kline *et al.* [31–34] also reported that different mosquito species sometimes respond differently to different attractants. The basic response pattern was that very few species were attracted to octenol alone, but in combination with CO₂ a synergistic affects apparently occurred and catch efficiency increases 2-fold or greater. This present study showed that CO₂ was about 3 times more efficient at capturing mosquitoes than octenol alone

using the BG Traps. However, previous studies on African and Brazilian malaria vectors, such as *Anopheles arabiensis*, *Anopheles funestus*, *Anopheles darlingi* and *Anopheles aquasalis* have shown that CO₂ was insufficiently attractive as standalone bait. Better catch rates were obtained using CO₂ in mixed odour baits or together with body odours [35–37].

The BG traps though especially developed for capturing *Ae. aegypti*, have also been shown to capture *Culex* mosquitoes [12,38]. This present study showed that the BG traps baited with CO₂ had a catch rate about 8 times higher for *Cx. quinquefasciatus* than *Ae. aegypti*. Other studies have also reported that BG traps baited with CO₂ do have high catch efficiencies [39–41]. Ferreira de Ázara *et al.* [42] also reported that BG traps operated with CO₂ trapped 6 times more female *Culex* spp than *Ae. aegypti*. The high catch rates of up to 272 *Culex* females with CO₂ and up to 57 *Culex* females without CO₂ (mainly *Cx. quinquefasciatus*) per 24 h shows that the BG traps might be a useful tool for the monitoring of diseases that are transmitted by the species in urban areas in Brazil, like Oropouche fever or Bancroftian Filariasis. This study further emphasises that CO₂ attracted only female *Culex*, while it attracted both male and female aedes. This would suggest that CO₂ is also an attractant for *Culex* species which may be due to the BG's trap imitation of human odour plumes. Russell [43] also reported improved collection of *Cx. quinquefasciatus* in CO₂ baited traps for in French Polynesia while Muturi *et al.* [44] reported high levels if *Cx. quinquefasciatus* and *Culex annulirois* in Kenya. Zhang *et al.* [45] also showed that CDC-LT with dry ice was most effective for trapping of *Cx. quinquefasciatus* when compared with UV light traps and gravid traps in China. Smallegange *et al.* [46] also showed that traps baited with yeast-produced CO₂ caught significantly more mosquitoes than unbaited traps and traps baited with industrial CO₂. They suggested that yeast-produced CO₂ can effectively replace industrial CO₂ for sampling of mosquitoes such as *Anopheles gambiae*. The use of the yeast/sugar generated CO₂ would significantly reduce costs and allow sustainable mass-application of traps for mosquito sampling in remote areas.

Given the recent upsurge of vector borne diseases such as dengue, chikungunya, and zika, greater efforts are being made to

control mosquito populations with an increased in surveillance efforts. The success of the yeast/sugar mixture as bait in the BG traps can greatly reduce the cost of surveillance and increase the efficiency of mosquito capture.

Conflict of interest statement

The authors declare that there is no conflict of interest.

References

- [1] Gillies MT. The role of carbon dioxide in host-finding by mosquitoes (Diptera: Culicidae): a review. *Bull Entomol Res* 1980; **70**(4): 525-32.
- [2] Costantini C, Gibson G, Sagnon N, Della Torre A, Brady J, Coluzzi M. Mosquito responses to carbon dioxide in a west African Sudan Savanna Village. *Med Vet Entomol* 1996; **10**(3): 220-7.
- [3] Takken W, Knols BGJ. Odor-mediated behavior of Afrotropical malaria mosquitoes. *Ann Rev Entomol* 1999; **44**: 131-57.
- [4] Eiras AE, Jepson PC. Host location by *Aedes aegypti* (Diptera: Culicidae): a wind tunnel study of chemical cues. *Bull Entomol Res* 1991; **81**(2): 151-60.
- [5] Service MW. Review: importance of ecology in *Aedes aegypti* control. *Southeast Asian J Trop Med Public Health* 1992; **23**(4): 681-90.
- [6] Canyon DV, Hii JLK. Efficacy of carbon dioxide, 1-octen-3-ol, and lactic acid in modified Fay-Prince traps as compared to man-landing catch of *Aedes aegypti*. *J Am Mosq Control Assoc* 1997; **13**(1): 66-70.
- [7] Owino EA, Sang R, Sole CL, Pirk C, Mbogo C, Torto B. Field evaluation of natural human odours and the biogent-synthetic lure in trapping *Aedes aegypti*, vector of dengue and chikungunya viruses in Kenya. *Parasit Vec* 2014; **7**: 451.
- [8] Owino EA, Sang R, Sole CL, Pirk C, Mbogo C, Torto B. An improved odor bait for monitoring populations of *Aedes aegypti*-vectors of dengue and chikungunya viruses in Kenya. *Parasit Vec* 2015; **8**: 253.
- [9] Pombi M, Jacobs F, Verhulst NO, Caputo B, della Torre A, Takken W. Field evaluation of a novel synthetic odour blend and of the synergistic role of carbon dioxide for sampling host-seeking *Aedes albopictus* adults in Rome, Italy. *Parasit Vec* 2014; **7**: 580.
- [10] Sukuman D. A review on use of attractants and traps for host seeking *Aedes aegypti* mosquitoes Indian. *J Nat Prod Resour* 2016; **7**(3): 207-14.
- [11] Szali M, Samino S, Leksono AS. Attractiveness test of attractants toward dengue virus vector (*Aedes aegypti*) into lethal mosquito trap modifications (LMM). *Int J Mosq Res* 2014; **1**(4): 47-9.
- [12] Kröckel U, Rose A, Eiras AE, Geier M. New tools for surveillance of adult yellow fever mosquitoes: comparison of trap catches with human landing rates in an urban environment. *J Am Mosq Control Assoc* 2006; **22**(2): 229-38.
- [13] Gibson G, Torr SJ. Visual and olfactory responses of haematophagous Diptera to host stimuli. *Med Vet Entomol* 1999; **13**(1): 2-23.
- [14] Mboera LEG, Knols BGJ, Braks MAH, Takken W. Comparison of carbon dioxide-baited trapping systems for sampling outdoor mosquito populations in Tanzania. *Med Vet Entomol* 2000; **14**(3): 257-63.
- [15] Iyaloo DP, Facknath S, Bheecarry A. Field evaluation of BG Sentinel™ traps of four different black-and-white color combinations in Mauritius for enhanced *Ae. albopictus* mosquito collection. *Int J Mosq Res* 2017; **4**(1): 43-9.
- [16] Kline DL. Semiochemicals, traps/targets and mass trapping technology for mosquito management. *J Am Mosq Control Assoc* 2007; **23**(2): 241-51.
- [17] Kline DL. Olfactory attractants for mosquito surveillance and control: 1-octen-3-ol. *J Am Mosq Control Assoc* 1994; **10**(2): 280-7.
- [18] Kline DL. Evaluation of various models of propane-powered mosquito traps. *J Vector Ecol* 2002; **27**(1): 1-7.
- [19] Vaidyanathan R, Edman JD. Sampling methods for potential epidemic vectors of eastern equine encephalomyelitis virus in Massachusetts. *J Am Mosq Control Assoc* 1997a; **13**(4): 342-7.
- [20] Vaidyanathan R, Edman JD. Sampling with light traps and human bait in epidemic foci for eastern equine encephalomyelitis virus in southeastern Massachusetts. *J Am Mosq Control Assoc* 1997b; **13**(4): 348-55.
- [21] Kline DL, Lemire GF. Evaluation of attractant-baited traps/targets for mosquito management on Key Island, Florida, USA. *J Vector Ecol* 1998; **23**(2): 171-85.
- [22] Kline DL, Mann MO. Evaluation of butanone, carbon dioxide, and 1-octen-3-ol as attractants for mosquitoes associated with north central Florida bay and cypress swamps. *J Am Mosq Control Assoc* 1998; **14**(3): 289-97.
- [23] Rueda LM, Harrison BA, Brown JS, Whitt PB, Harrison RL, Gardner RC. Evaluation of 1-octen-3-ol, carbon dioxide, and light as attractants for mosquitoes associated with two distinct habitats in North Carolina. *J Am Mosq Control Assoc* 2001; **17**(1): 61-6.
- [24] van den Hurk AF, Beebe NW, Ritchie SA. Responses of mosquitoes of the *Anopheles farauti* complex to 1-octen-3-ol and light in combination with carbon dioxide in northern Queensland, Australia. *Med Vet Entomol* 1997; **11**(2): 177-80.
- [25] Shone SM, Ferrao PN, Lesser CR, Glass GE, Norris DE. Evaluation of carbon dioxide- and 1-octen-3-ol-baited centers for disease control fay-prince traps to collect *Aedes albopictus*. *J Am Mosq Contr Assoc* 2003; **19**(4): 445-7.
- [26] Mboera LEG, Takken W. Carbon dioxide chemotropism in mosquitoes (Diptera: Culicidae) and its potential in vector surveillance and management programmes. *Rev Med Vet Entomol* 1997; **85**: 355-68.
- [27] Saitoh Y, Hattori J, Chinone S, Nihei N, Tsuda Y, Kurahashi H, et al. Yeast-generated CO₂ as a convenient source of carbon dioxide for adult mosquito sampling. *J Am Mosq Control Assoc* 2004; **20**(3): 261-4.
- [28] Carlson DA, Schreck CE, Brenner RJ. Carbon dioxide released from human skin: effect of temperature and insect repellents. *J Med Entomol* 1992; **29**(2): 165-70.
- [29] Sinka ME, Bangs MJ, Manguin S, Chareonviriyaphap T, Patil AP, Temperley WH, et al. The dominant *Anopheles* vectors of human malaria in the Asia-Pacific region: occurrence data, distribution maps and bionomic precis. *Parasit Vectors* 2011; **4**: 89.
- [30] Kemme JA, Van Essen PH, Ritchie SA, Kay BH. Response of mosquitoes to carbon dioxide and 1-octen-3-ol in southeast Queensland, Australia. *J Am Mosq Control Assoc* 1993; **9**: 431-5.
- [31] Kline DL, Dame DA, Meisch MV. Evaluation of 1-octen-3-ol and carbon dioxide as attractants for mosquitoes associated with irrigated ricefields in Arkansas. *J Am Mosq Control Assoc* 1991a; **7**: 165-9.
- [32] Kline DL, Wood JR, Cornell JA. Interactive effects of 1-octen-3-ol and carbon dioxide on mosquito (Diptera: Culicidae) surveillance and control. *J Med Entomol* 1991b; **28**: 254-8.
- [33] Kline DL, Wood JR, Morris CD. Evaluation of 1-octen-3-ol as an attractant for *Coquillettidia perturbans*, *Mansonia* spp. and *Culex* spp. associated with phosphate mining operations. *J Am Mosq Control Assoc* 1990a; **6**: 605-11.
- [34] Kline DL, Takken W, Wood JR, Carlson DA. Field studies on the potential of butanone, carbon dioxide, honey extract, 1-octen-3-ol, lactic acid, and phenols as attractants for mosquitoes. *Med Vet Entomol* 1990b; **4**: 383-91.
- [35] Hiwat H, Andriessen R, Rijk M, Koenraadt CJ, Takken W. Carbon dioxide baited trap catches do not correlate with human landing collections of *Anopheles aquasalis* in Suriname. *Mem Inst Oswaldo Cruz* 2011; **106**(3): 360-4.
- [36] Service MW. *Field sampling methods*. London and New York: Elsevier; 1993.
- [37] Rubio-Palis Y, Curtis CF. Evaluation of different methods of catching anopheline mosquitoes in western Venezuela. *J Am Mosq Control Assoc* 1992; **8**(3): 261-7.
- [38] Williams CR, Long SA, Russel RC, Ritchie SA. Field efficacy of the BG-Sentinel compared with CDC backpack aspirators and

- CO₂-baited EVS traps for collection of adult *Aedes aegypti* in Cairns, Queensland. *Aust J Am Mosq Control Assoc* 2006; **22**(2): 296-300.
- [39] Obenauer PJ, Abdel-Dayem MS, Stoops CA, Villinski JT, Tageldin R, Fahmy NT, et al. Field responses of *Anopheles gambiae* complex (Diptera: Culicidae) in Liberia using yeast-generated carbon dioxide and synthetic lure-baited light traps. *J Med Entomol* 2013; **50**(4): 862-70.
- [40] Hoel DF, Marika JA, Dunford JC, Irish SR, Geier M, Obermayr U, et al. Optimizing collection of *Anopheles gambiae* s.s. (Diptera: Culicidae) in biogents sentinel traps. *J Med Entomol* 2014; **51**(6): 1268-2175.
- [41] Roiz D, Duperier S, Roussel M, Bousse's P, Fontenille D, Simard F, et al. Trapping the tiger: efficacy of the novel BG-sentinel 2 with several attractants and carbon dioxide for collecting *Aedes albopictus* (Diptera: Culicidae) in Southern France. *J Med Entomol* 2015; **53**(2): 460-5.
- [42] de Ázara TM, Degener CM, Roque RA, Ohly JJ, Geier M, Eiras ÁE. The impact of CO₂ on collection of *Aedes aegypti* (Linnaeus) and *Culex quinquefasciatus* Say by BG-Sentinel[®] traps in Manaus, Brazil. *Mem Inst Oswaldo Cruz* 2013; **108**(2): 229-32.
- [43] Russell RC. The relative attractiveness of carbon dioxide and octenol in CDC- and EVS-type light traps for sampling the mosquitoes *Aedes aegypti* (L.), *Aedes polynesiensis* marks, and *Culex quinquefasciatus* say in Moorea, French Polynesia. *J Vector Ecol* 2004; **29**(2): 309-14.
- [44] Muturi EJ, Mwangangi J, Shililu J, Muriu S, Jacob B, Mbogo CM, et al. Evaluation of four sampling techniques for surveillance of *Culex quinquefasciatus* (Diptera: Culicidae) and other mosquitoes in African rice agroecosystems. *J Med Entomol* 2007; **44**(3): 503-8.
- [45] Zhang HL, Zhang YZ, Yang WH, Feng Y, Nasci RS, Yang J, et al. Mosquitoes of Western Yunnan Province, China: seasonal abundance, diversity, and arbovirus associations. *PLoS One* 2013; **8**(10). e77017.
- [46] Smallegange RC, Schmied WH, van Roey KJ, Verhulst NO, Spitzen J, Mukabana WR, et al. Sugar-fermenting yeast as an organic source of carbon dioxide to attract the malaria mosquito. *Anopheles gambiae*. *Malar J* 2010; **9**: 292.