



Document heading doi: 10.1016/S2305-0500(13)60126-8

Induction of ovarian activity in Bulgarian Murrah buffaloes by hormonal treatment in the early postpartum period

Stanimir A Yotov^{1*}, Anatoli S Atanasov¹, Yordanka Y Ilieva²

¹Department of Obstetrics, Reproduction and Reproductive Disorders, Faculty of Veterinary Medicine, Trakia University, Student Campus, 6000 Stara Zagora, Bulgaria

²Agricultural Institute, Tzar Simeon Veliki 3, 9700 Shumen, Bulgaria

ARTICLE INFO

Article history:

Received 5 February 2013
 Received in revised form 7 February 2013
 Accepted 10 February 2013
 Available online 20 June 2013

Keywords:

Buffaloes
 Postpartum
 Hormonal treatment
 Ovarian activity

ABSTRACT

Objective: To determine the possibilities for induction of ovarian activity in Bulgarian Murrah buffaloes by hormonal treatment in the early postpartum period. **Methods:** Twenty six clinically healthy animals, weighing 480–520 kg, 3–5 years of age, with normal parturition, without clinical signs of endometritis during the entire experimental period were divided into 2 groups: control ($n=10$) and experimental ($n=16$). Primiparous and multiparous buffaloes were proportionally allotted into both groups. By 5, 21, and 28 postpartum days, control animals were intramuscularly treated with 2 mL physiological saline, whereas experimental buffaloes received 500 mg PGF₂ α by the 5th postpartum day, 100 μ g GnRH by the 21st postpartum day and a second prostaglandin dose 7 days later. All buffaloes were submitted to transrectal ultrasound examination on days 5, 21 and 28 after calving. Evaluation of ovarian activity was made on the basis of follicle type and detection of a newly formed corpus luteum. The percentage of animals with small, medium or dominant follicle and corpus luteum after either spontaneous or induced ovulation was determined. Insemination management included introduction of two proven fertile bulls into the herd at the start of the treatment. Pregnancy ultrasound checks were performed on postpartum days 58, 68, 120 and 150. The proportion of buffaloes with clinical signs of spontaneous or induced oestrus until the 35th postpartum day, pregnancy rates by the 35th and 90th postpartum days and service periods <60 days, 60–90 days and >90 days were registered. **Results:** On the 5th day after calving, small follicles were predominating in the ovaries of buffaloes from both groups. Similar findings were established on the 21st day in the control group, whereas the share of experimental buffaloes with small follicles was significantly ($P<0.01$) lower. By the 21st day, there were no medium follicles in untreated animals, while in hormonally treated ones they were found out in 25% of cases. The proportion of experimental buffaloes with dominant follicle (56.3%) by the 21st day was statistically significantly higher ($P<0.01$) compared to that of controls (10%). Cumulative percentage of animals with detected corpus luteum (66.8%) until the 28th postpartum day differed considerably ($P<0.01$) from that in controls (10.0%). There was a substantial between-group difference ($P<0.05$) between the total percentage of buffaloes with clinical signs of oestrus and pregnancy rates until the 35th day after calving. The share of buffaloes with service period <60 days was higher ($P<0.05$) compared to the untreated group. The analysis of results demonstrated that the PGF₂ α –GnRH–PGF₂ α treatment on 5, 21 and 28 postpartum days, was able to induce ovarian activity and oestrus in Bulgarian Murrah buffaloes. The total proportion of animals with signs of oestrus (both spontaneous and induced) and pregnancy rates by the 35th postpartum day were significantly higher ($P<0.05$) in the treated group compared to controls. **Conclusions:** The tested hormonal treatment protocol, combined with the presence of fertile bulls in the herd, could be successfully used for reduction of service period and calving interval in clinically healthy buffaloes.

*Corresponding author: Stanimir A. Yotov, Department of Obstetrics, Reproduction and Reproductive Disorders, Faculty of Veterinary Medicine, Trakia University, Student Campus, 6000 Stara Zagora, Bulgaria.

Tel.: +359 42 699 513
 Fax: +359 42 670 624
 E-mail: stanrad@abv.bg

1. Introduction

The extended calving interval is the main reason for the low reproductive efficiency in buffaloes^[1,2]. Numerous authors affirm that the first 90 days after calving are particularly important for optimization of reproductive process^[3–5].

The high concentrations of placental and ovarian steroid during late gestational phase have a negative impact on hypothalamic–pituitary system and subsequently, on the recovery of ovarian activity^[2,6,7]. The studies of Agrawal et al.^[8] on ovaries obtained from slaughtered buffaloes on the day of calving showed the presence of a corpus luteum graviditatis 3.5 mm of size and insignificant follicular development. Similar findings are reported by Jainudeen et al.^[9] which, using rectal examination, palpated a small rough structure protruding over the ovarian surface, <3 mm in diameter, by the 10th day. Usmani et al.^[10] established a complete corpus luteum graviditatis regression within (21.8±1.1) days, whereas Momongan et al.^[11] indicated an interval of (7±2) days, and Khasatiya et al.^[4] (9.39±0.27) days. According to Georgiev et al.^[12] the regression of corpus luteum graviditatis begins as early as the end of the pregnancy. Lohan et al.^[13] reported that by the 6th day after calving, the average number of small and medium follicles was (8.10±5.67) and (1.00±0.00), respectively. All cycling animals between the 14th and the 20th postpartum day had a follicle (>8.5 mm) in diameter, that is interpreted as a parameter of ovarian activity resumption. In half of studied animals, spontaneous ovulation occurred after the 65th postpartum day. Having investigated the recovery of ovarian activity, Presicce et al.^[3] demonstrated that the number of antral follicles (≥2 mm) gradually decreased until the 60th day after calving. Follicles ≥3 mm of size increased during the first two postpartum weeks and were considerably more numerous in the ovary, contralateral to the corpus luteum graviditatis, and the balance was reinstated by the end of the first month.

The onset of the first ovulation and the first estrus in buffaloes after calving was extensively researched^[14–18], but the subject is still not completely clarified. The pattern of restoration of cycling activity in buffaloes showed that during the formation of the corpus luteum after the first ovulation, 26%–86% of animals exhibited shortened luteal phase (6–13 days), occasionally accompanied by low progesterone levels^[19,20]. According to Presicce et al.^[3] 30% of primiparous and 10% of multiparous buffaloes remained anovulatory until the 60th day after calving.

The postpartum period in large ruminants is of primary importance for shortening of calving period^[2,21]. Most investigations^[4,16,18,19,22] on hormonally–induced ovarian activity in buffaloes have been conducted after the 30th postpartum day, and obtained results are often contradictory. The data regarding the hormonal induction of ovarian activity in buffaloes during the early postpartum period is limited.

The aim of the current study was to determine the possibilities for induction of ovarian activity in Bulgarian Murrah buffaloes by hormonal treatment during the first 30 days after calving.

2. Materials and methods

The trial was performed with 26 Bulgarian Murrah

buffaloes, weighing 480–520 kg, at the age of 3–5 years. The average daily milk production was (5.6±0.6) kg with milking morning and evening. They were clinically healthy, with normal parturition, normally expelled placenta, without clinical signs of endometritis during the entire experimental period. Females were housed together with bulls over the entire period of the study and fed uniformly. The farm management included separation of calves from the dams immediately after the birth, feeding colostrum during the first 3 days, and a milk replacer thereafter. The daily ration comprised concentrate, rough feeds (straw, hay and 8–hour grazing on pasture); water was offered *ad libitum*. Buffaloes were reared in a farm located at latitude of 42.183 N, longitude 25.567 E. and experiment was conducted between April and August.

The buffaloes were divided into 2 groups: control ($n=10$) and experimental ($n=16$). Primiparous and multiparous buffaloes were proportionally allotted into both groups. By postpartum days 5, 21, and 28 control animals were intramuscularly treated with 2 mL physiological saline. Experimental buffaloes were hormonally treated with 500 mg PGF₂α (2 mL PGF Veyx forte, Veyx–Pharma GmbH, Schwarzenborn, Germany) by the 5th postpartum day, 100 μg GnRH(Depherelin, Veyx–Pharma GmbH, Schwarzenborn, Germany) by the 21st postpartum day and a second prostaglandin dose 7 days after GnRH administration.

All buffaloes were submitted to transrectal ultrasound (SonoScape A5 Vet, SonoScape Co. LTD, Shenzhen, China) with multifrequency (7–12 MHz) linear transducer. Initial examinations were performed on days 5, 21 and 28 after calving.

The evaluation of ovarian status was based on the visualization of small (<6 mm diameter), medium (6–9 mm diameter) or dominant follicle (>10 mm diameter) with or without a corpus luteum into one of the both ovaries. The percentage of animals with the respective follicular structures during the different time intervals and with a corpus luteum after spontaneous, induced ovulation and cumulative values was determined.

The detection of estrous activity until the 35th postpartum day was performed by daily observation during the grazing period. By the beginning of the treatment, two proven fertile bulls were introduced into the groups to mount the cycling females. The percentage of animals exhibiting the standing estrus and mated by a bull was recorded. The percentage of animals with spontaneous oestrus, induced oestrus and the cumulative values were registered.

Pregnancy ultrasound checks were performed on postpartum days 58, 68, 120 and 150. Positive diagnosis was based on visualisation of enlarged uterine lumen filled with amniotic fluid and an embryo (foetus).

The pregnancy rates by the 35th and 90th postpartum days and the cumulative pregnancy rates were calculated after the last ultrasound examination. The percentage of buffaloes with service periods <60 days, 60–90 days and >90 days were registered in both groups.

Statistical analysis was performed with Stat–Soft 1984–2000 Inc. statistical software (Copyright©1990–1995 Microsoft. Corp.) by means of non–parametric analysis for comparison of two proportions, using Student's *t*–criterion. Differences were considered significant in *P*–values < 0.05.

3. Results

The data for reproductive traits showed that by the 5th day after calving, small follicles were predominant ovarian structures in both groups (Table 1). Almost the same finding (90% buffaloes with small follicles) was established by the 21st postpartum day in control groups, whereas in the treated group, their number was substantially reduced ($P<0.01$) to 18.8%. The difference ($P<0.05$) was also present by the 28th postpartum day. Three buffaloes from the experimental group did not exhibit follicular activity. By the 21st day, there were no medium follicles in control animals, but they were present in 25% of hormonally stimulated animals.

A similar trend was determined by the 28th day, but the differences between groups was not significant ($P>0.06$).

A dominant follicle was observed in two untreated buffaloes (20%): one by the 21st postpartum day and another one – by the 28th day. In the experimental group, the number of animals with dominant follicle by the 21st day (56.3%) was significantly higher ($P<0.01$) than that in controls (10%) and decreased to 31.3% by the 28th day. Only one of untreated buffaloes had a corpus luteum after spontaneous ovulation by the 28th day. In the experimental group, it was detected in 1 animal by the 21st day, while the other 9 corpora lutea detected by the 28th day were after GnRH-induced ovulation. Eight of ovulated animals had a follicle >10 mm on the day of gonadotropin administration, and in one buffaloes, the follicle diameter was 9 mm. The cumulative percentage of treated buffaloes with corpus luteum until the 28th day (62.5%), was considerably higher ($P<0.05$) vs that in controls (10%).

Clinical signs of spontaneous oestrus within 28–35 postpartum days occurred in 20% of untreated animals. At the same time, 12.5% of experimental animals with medium-size follicles by the 21st day exhibited estrous activity on the day of prostaglandin administration and were determined as buffaloes with spontaneous oestrus. Seven animals (43.8%) with corpus luteum and follicle (>9 mm) by the 28th day were cycling 3 to 5 days after the prostaglandin treatment and were hence determined as subjects with hormonally induced oestrus. There was a significant difference ($P<0.05$) between the cumulative percentage of control and experimental animals with estrous activity until the 35th day (20% and 56.3%, respectively).

The data from the first two ultrasound checks showed that until the 35th day, 10% of control and 37.5% of buffaloes with PGF₂ α induced oestrus have conceived ($P<0.05$). The ultrasound performed on postpartum days 120 and 150 did not exhibit significant between-group differences in pregnancy rates (20.0% and 18.8%) until the 90th postpartum day. Nevertheless, the total relative share of pregnant buffaloes after hormonal treatment (43.8%), was higher than pregnancy rate of controls (30%).

The proportion of buffaloes with service period <60 days was higher in the experimental than in control group ($P<0.05$), whereas the number of animals from both groups with service periods of 60–90 days was similar. There was however a trend to reduction of the number of hormonally treated buffaloes with service period over 90 days.

Table 1

Ovarian structures and reproductive parameters in the control and experimental buffaloes during the postpartum period.

Parameters	Control group postpartum days (n=10)			Experimental group postpartum days (n=16)		
	5	21	28	5	21	28
Ovarian structures (%)						
Small follicles(< 6mm)	100.0 (10/10)	90.0 (9/10) ^a	70.0 (7/10) ^c	93.7 (15/16)	18.8 (3/16) ^b	18.8 (3/16) ^d
Medium follicles(6–9 mm)	0.0 (0/10)	0.0 (0/10)	20.0 (2/10)	6.3 (5/16)	25.0 (4/16)	50.0 (8/16)
Dominant follicle (> 10 mm)	0.0 (0/10)	10.0 (1/10) ^a	10.0 (1/10)	0.0 (0/16)	53.6 (9/16) ^b	31.3 (5/16)
Cumulative percentage of animals with corpus luteum until the 28 th day(%)		10.0 (1/10) ^c			62.5 (10/16) ^b	
Estrous activity until the 35 th day (%)						
Spontaneous		20.0 (2/10)			12.5 (2/16)	
Induced		–			43.8 (7/16)	
Cumulative		20.0 (2/10) ^c			56.3 (9/16) ^d	
Pregnancy rate (%)						
Until the 35th day		10.0 (1/10) ^c			37.5 (6/16) ^d	
Between 35th and 90th days		20.0 (2/10)			18.8 (3/16)	
Cumulative		30.0 (3/10)			56.3 (9/16)	
Service period (%)						
<60 days		10.0 (1/10) ^c			37.5 (6/16) ^d	
60–90 days		20.0 (2/10)			18.8 (3/16)	
>90 days		70.0 (7/10)			43.8 (7/16)	

Different superscripts indicate significant difference between control and experimental values from the same period, a,b $P<0.01$; c,d $P<0.05$.

4. Discussion

The presence of small follicles into the ovaries in most of buffaloes from both groups by the 5th day after calving, indicate lack of ovarian activity. These findings are comparable to the results of other researchers^[8,9,13] and support the thesis for postpartum suppression of ovarian activity by high progesterone levels during the pregnancy.

The significantly higher number of experimental buffaloes with dominant follicle by the 21st postpartum day affirms enhance ovarian activity in treated animals. It could be attributed to lysis of the corpus luteum of pregnancy after the prostaglandin administration and lower progesterone concentrations. According to Zerbe *et al.*^[23], the rapid drop of progesterone concentrations results a GnRH release, increase of LH and FSH pulsations and resumption of ovarian activity. Iqbal *et al.*^[24] demonstrated that the application of PGF2 α two hours after calving led to increase uterine involution, earlier resumption of follicular activity, expression of estrous activity and shorter calving interval. The data from the ultrasound examination of control buffaloes by the 28th day support this statement.

The presence of a dominant follicle in one animal only in control group, and especially the considerably higher percentage of buffaloes with small follicles ($P < 0.05$) indicates a low level of ovarian activity. The inadequate LH pulses during that period could be a cause^[25,26], for the decreased release of estrogens by smaller follicles^[27], respectively inadequate hypothalamic stimulation. We assume that the inadequate ovarian activity is resulting from the maintenance of sufficiently high blood progesterone concentrations in untreated buffaloes over a prolonged time interval, but a categorical conclusion could not be made without blood progesterone assay. The absence of substantially increased number of animals with medium and dominant follicles in this group by the 28th day is also in support of the hypothesis. Palta and Madan^[28] also report a strongly reduced pituitary sensitivity to gonadotropin-releasing hormone during the pregnancy and about its gradual restoration from the 2nd to the 35th day after calving. The lack of ovarian response in three of treated buffaloes could be explained by an inadequate response of the pituitary gland to gonadotropin stimulus in the early postpartum period^[29], seasonal effect^[30] or individual sensitivity of some animals to hormonal therapy^[31].

The presence of a newly formed corpus luteum is indicating a spontaneous ovulation ^[13,32]. Its presence by the 21st postpartum day in the experimental and by the 28th day in control group is a sign of spontaneous ovulation of follicles occurring at an earlier stage. A similar phenomenon has been established by Presicce *et al.*^[3] between the 15th and the 25th day, but in general, the newly formed luteal structure has a very short lifespan ^[19,20]. The presence of corpora lutea by the 28th days in the treated group

showed a GnRH-induced ovulation of follicles with diameter > 9 mm. The results support the findings of Gimenes *et al.*^[33] for ovulation of Murrah buffalo heifers with follicles from 8.5 to 10 mm induced by exogenous LH administration. Campanile *et al.*^[34] have utilized GnRH agonists or human chorionic gonadotropin (hCG) in multiparous Italian Mediterranean buffalo cows and ovulation of follicles between 4.2 mm and 13.0 mm of size. On the other hand, the lack of corpus luteum in two of animals with follicles > 8 mm and a third one with dominant follicle after the GnRH treatment indicated that successful ovulation is dependent not only on follicle size. One of possible reasons could be the inadequate production of gonadotropin hormone receptors, as shown by Sartori *et al.*^[35] in investigations with dairy cows. Campanile *et al.*^[36] supposed that the inadequate production of estradiol by follicle could result in the lack of preovulatory LH peak. In our opinion, these follicles have not ovulated or were not luteinized after the GnRH application^[37,38], but continued their development until the preovulatory follicular stage. The hypothesis for the lack of sufficient number of receptors, LH surge and subsequent ovulation, was supported by the spontaneous estrus expression in 12.5% of buffaloes on the day of prostaglandin treatment. Probably, these cycles were anovulatory because the animals were not pregnant during the subsequent ultrasound examinations. The considerably higher ($P < 0.05$) cumulative percentage of treated buffaloes with corpus luteum until the 28th day (62.5%), that in controls (10%) also state to benefits of PG-GnRH treatment on days postpartum 5 and 21, respectively.

The clinical estrus expression in 43.8% of buffaloes with corpus luteum until the 35th day shows a successful oestrus induction after PGF2 α application. These results differ substantially from reported data about 100% expression of oestrus after treatment with 3 mL GnRH analogue by the 30th postpartum day, 2.5 mL PGF2 α seven days latter^[5]. Applying a GnRH-PG protocol in the same postpartum period, Usmani *et al.*^[19] detected estrus only in 27.8% of treated buffaloes. The causes for discrepancies could be due to the time of treatment, the type and dose of used hormone or the methods for estrus detection and insemination. The small proportion of animals with estrous activity from the 35th to the 90th postpartum days in both groups despite the presence of the bull indicated a strong seasonal influence on the resumption of ovarian activity. Seasonal effects on ovarian functions are extensively studied and reviewed by El-Wishy^[2] and Perera^[39]. The lack of oestrus behaviour in experimental buffaloes with detected corpus luteum by the 28th day during the next 7 days could be attributed both to seasonal influence and insufficient functional activity of the corpus luteum, responsible for its inadequate sensitivity to prostaglandin injection. According to Cohan *et al.*^[30] the efficacy of PGF2 α treatment was considerably reduced out of the breeding season, and buffaloes with smaller corpus

luteum and low blood progesterone concentrations exhibit a less reproductive response^[40]. Regardless of that, the used hormonal treatment protocol resulted in cumulative estrous activity in significantly more buffaloes (56.3%) as compared to untreated animals (20%).

The results from the first two ultrasound pregnancy checks in the control group showed that the animals could conceive after the first spontaneous ovulation in normal uterine involution. This is in agreement with our previous findings in Bulgarian Murrah buffaloes, demonstrating complete uterine involution by the 34th day after calving^[41] and spontaneous ovulation between days 19 and 34 (Yotov *et al.*, unpublished data). Statistically differences ($P < 0.05$) between pregnancy rates of control and treated buffaloes until the 35th day, showed that the hormonal treatment during the early postpartum period could induce ovulatory oestrus with subsequent fertilization. The cumulative percentage of pregnant animals until the 90th day corresponded to rate of 53.7% reported by Usmani *et al.*^[19], but is in disagreement with the data of other researchers^[5]. Unlike us, the last authors has studied swamp buffaloes reared under different climatic conditions, using higher doses of GnRH and PGF2 α analogues and beginning of the treatment by the 30th postpartum day. According to Zicarelli *et al.*^[42], a large part (30%) of non-pregnant animals after prostaglandin-induced cyclic activity during the transition season, remain acyclic and are difficult to be fertilized. Irrespective of that, the results suggested a positive trend to more pregnancies after hormonal stimulation of ovarian function during the first 30 postpartum days. An additional argument in support of the benefits of such a treatment is the significantly higher proportion of buffaloes with service period <60 days in the experimental group and the relatively higher parts of animals with service period >90 days in the control group. The positive effect of hormonal treatment during in the latter period (between day 30th and 60th postpartum) for reduction of service period and calving interval is demonstrated in numerous investigations^[4,18,24].

The analysis of results demonstrated that the PGF2 α –GnRH–PGF2 α treatment on postpartum days 5, 21 and 28, respectively, was able to induce ovarian activity and oestrus in Bulgarian Murrah buffaloes. The cumulative percentage of animals with estrous activity and pregnancy rates by the 35th postpartum day were significantly higher in the treated group compared to controls. The used hormonal treatment protocol, combined with the presence of fertile bulls in the herd, could be successfully used for reduction of service period and calving interval in clinically healthy buffaloes.

Conflict of interest statement

We declare that we have no conflict of interest

Acknowledgments

Research conducted by the authors was supported by Research Grant # 2012–10, Trakia University, Stara Zagora, Bulgaria. The authors gratefully acknowledge to people from Dimitriev farm to their help during the experiment.

References

- [1] Borghese A. (ed). In: *Technical series 67*. Rome: Food and Agriculture Organization; 2005, p. 77–94.
- [2] El-Wishy AB. The postpartum buffalo: II. Acyclicity and anestrus. *Anim Reprod Sci* 2007; **97**: 216–236.
- [3] Presicce GA, Bella A, Terzano GM, De Santis G, Senatore EM. Postpartum ovarian follicular dynamics in primiparous and pluriparous Mediterranean Italian buffaloes (*Bubalus bubalis*). *Theriogenology* 2005; **63**: 1430–1439.
- [4] Khasatiya CT, Kavani FS, Dhani AJ, Derashri HJ, Panchal, MT, Desai PM. Studies on puerperal events and reproductive efficiency following hormone therapy at day 42 postpartum in surti buffaloes. *Int J Agri Biol* 2006; **8**: 132–137.
- [5] Yendraliza, Zesfin BP, Udin Z, Jaswandi, Arman C. Effect of combination of GnRH and PGF2 α for estrus synchronization on onset of estrus and pregnancy rate in different postpartum swam buffalo in Kampar regency. *J Indonesian Trop Anim Agric* 2011; **36**: 8–13.
- [6] Karsch FJ, Foster DL, Bittman EL, Goodman RR. A role for estradiol in enhancing hormone pulse frequency during the follicular phase of the estrous cycle of the sheep. *Endocrinology* 1983; **113**: 1333–1339.
- [7] Nett TM. Function of the hypothalamic–hypophysial axis during the postpartum period in ewes and cows. *J Reprod Fert* 1987; **34**: 201–213.
- [8] Agrawal KP, Riazada BC, Pandey MD. Postparturient changes in the ovary and related endocrine glands in the buffalo. *Indian J Anim Sci* 1979; **49**: 25–36.
- [9] Jainudeen MR, Bongso TA, Tan HS. Postpartum ovarian activity and uterine involution in the suckled swamp buffalo (*Bubalus bubalis*). *Theriogenology* 1983; **5**: 181–190.
- [10] Usmani RH, Ahmed M, Inskoop EK, Dailey RA, Lewis PE, Lewis GS. Uterine involution and postpartum ovarian activity in Nili Ravi buffaloes. *Theriogenology* 1985; **24**: 435–445.
- [11] Momongan VG, Sarabia AS, Roxas NP, Palad OA, Obsioma AR, Nava ZM, et al. Increasing the productive efficiency of Caraboas under small holder farming systems. In: *Domestic buffalo production in Asia*. Vienna: IAEA; 1990, p. 167–178.
- [12] Georgiev B, Marinov M, Kacheva D, Maslev Tz, Danev A. A study of ovarian activity of buffalo by ultrasonography investigation. In: *Proceeding of the 10th Jubilee International Symposium of Immunology of Reproduction*. Varna, Bulgaria, vol. 1, 1993, p. 118–113.
- [13] Lohan IS, Malik RK, Kaker ML. Uterine involution and ovarian follicular growth during early postpartum period of Murrah

- buffaloes. *Asian–Aust J Anim Sci* 2004; **17**: 313–316.
- [14]Ashturkar RW, Aher VD, Bhokre AP. Studies on infertility problems in non-descript buffaloes and cows. *Indian Vet J* 1995; **72**: 1050–1052.
- [15]Qureshi MS, Safi GM, Dhanani J, Kaka I. Reproductive performance of dairy buffaloes in the Northern Hilly areas of Pakistan. *Buffalo J* 1999; **15**: 391–396.
- [16]Anwar A, Ullah N, Mehmood A, Andrabi SMH. Postpartum anoestrus in Nili–Ravi buffaloes maintained under rural and peri-urban management. *Pakistan Vet J* 2003; **3**: 114–117.
- [17]Honparkhe M, Singh J, Dadarwal D, Dhaliwal GS, Kumar A. Estrus induction and fertility rates in response to exogenous hormonal administration in postpartum anestrous and subestrus bovines and buffaloes. *J Vet Med Sci* 2008; **70**: 1327–1331.
- [18]Shah SK, Nakao T. A clinical study of anoestrus buffaloes in southern Nepal. *J Reprod Dev* 2010; **56**: 208–211.
- [19]Usmani RH, Ahmed N, Shafiq P, Mirza MA. Effect of subclinical uterine infection on cervical and uterine involution, estrous activity and fertility in postpartum buffaloes. *Theriogenology* 2001; **55**: 563–71.
- [20]Uçar M, Kucukkebacı M, Gundogan M, Saban E. Using milk progesterone assay at the time of oestrus and post-mating for diagnosing early pregnancy in Anatolian water buffaloes. *Turk J Vet Anim Sci* 2004; **28**: 513–518.
- [21]Martin AD, Lystad ML, Reksen O, Ropstad E, Waldmann A, Nafstad O, et al. Assessment of progesterone profiles and postpartum onset of luteal activity in spring calving Hereford beef suckler cattle. *Acta Vet Scand* 2010; **52**: 42–42.
- [22]Zaabel SM, Hegab AO, Montasser AE, El-Sheikh H. Reproductive performance of anestrous buffaloes treated with CIDR. *Anim Reprod* 2009; **6**: 460–464.
- [23]Zerbe H, Gregory C, Grunert E. Zur behandlung ovarieller bedingter zyklusstörungen beim milchrind mit progesteron-abgebenden vorrichtungen. *Tierarztl Umsch* 1999; **54**: 189–192. (In German).
- [24]Iqbal S, Allem M, Saaed MA. Role of single injection of prostaglandin F₂ alfa on breeding efficiency of postpartum buffaloes. *Pakistan Vet J* 2003; **23**: 197–201.
- [25]Batra SK, Pandey RS. Luteinizing hormone in blood plasma of postpartum buffaloes (*Bubalus bubalis*). *Theriogenology* 1983; **19**: 193–200.
- [26]Palta P, Bansal N, Prakash BS, Manik RS, Madan ML. Interrelationship between follicular size and follicular fluid estradiol-17 β , progesterone and testosterone concentrations in individual buffalo ovarian follicles. *Asian–Aust J Anim Sci* 1998; **11**: 293–299.
- [27]Dharani S, Kathiresan D, Devanathan TG, Balachandaran C, Satheeshkumar S. Ovulatory response of the first follicular wave growing and regressing phase follicle for GnRH administration in cyclic buffaloes. *Buffalo Bull* 2010; **29**: 199–205.
- [28]Palta P, Madan ML. Effect of gestation on GnRH induced LH and FSH release of buffalo (*Bubalis bubalis*). *Theriogenology* 1996; **46**: 993–998.
- [29]Shah NH, Willemse AH, Van De, Weil DFM. A review of the factors influencing fertility in the postpartum buffalo. *Buffalo J* 1990; **2**: 103–115.
- [30]Chohan KR, Iqbal J, Choudhary RA, Khan AH. Oestrous response and fertility in true anestrous buffaloes following hormonal treatment during summer. *Pak Vet J* 1995; **15**: 68–72.
- [31]Khan MI–R, Rana MA, Ahmad N. Ultrasonic monitoring of follicles and corpora lutea during synchronization in summer anoestrus Nili–Ravi buffaloes and their subsequent superovulatory response. *Pakistan Vet J* 2005; **25**: 82–86.
- [32]Rahman MS, Shohag AS, Kamal MM, Parveen N, Shamsuddin M. Application of ultrasonography to investigate postpartum anoestrus in water buffaloes. *Reprod Dev Biol* 2012; **36**: 103–108.
- [33]Gimenes LU, Carvalho NAT, Sà Filho MF, Torres JJRS, Ayres H, Vannucci FS, et al. Follicle selection by ultrasonography and plasmatic characteristics and ovulatory capacity in buffaloes. *Ital J Anim Sci* 2007; **6**: 629–631.
- [34]Campanile G, Vecchio D, Di Palo R, Neglia G, Gasparini B, Prandi A, et al. Delayed treatment with GnRH agonist, hCG and progesterone and reduced embryonic mortality in buffaloes. *Theriogenology* 2008; **70**: 1544–1549.
- [35]Sartori R, Fricke PM, Ferreira JCP, Ginther OJ, Wiltbank MC. Follicular deviation and acquisition of ovulatory capacity in bovine follicles. *Biol Reprod* 2001; **65**: 1403–1409.
- [36]Campanile G, Baruselli PS, Neglia G, Vecchio D, Gasparini B, Gimenes LU, et al. Ovarian function in the buffalo and implications for embryo development and assisted reproduction. *Anim Reprod Sci* 2010; **121**: 1–11.
- [37]Tiwari H, Guilbault LA, Proulx J, Ramkumar R, Dufour JJ. Histological populations and atresia of ovarian follicles in postpartum cattle treated with an agonist of gonadotropin-releasing hormone. *J Anim Sci* 1994; **72**: 192.
- [38]Sharma RK, Singh JK, Khanna S, Singh I. Ovarian response of prepubertal Murrah heifers to exogenous GnRH. *Anim Reprod Sci* 2012; **133**: 153–158.
- [39]Perera BMAO. Reproductive cycles of buffalo. *Anim Reprod Sci* 2011; **124**: 194–199.
- [40]Brito LFC, Satrapa R, Marson EP, Kastelic JP. Efficacy of PGF₂ α to synchronize estrus in water buffalo cows (*Bubalus bubalis*) is dependent upon plasma progesterone concentration, corpus luteum size and ovarian follicular status before treatment. *Anim Reprod Sci* 2002; **73**: 23–35.
- [41]Atanasov A, Dineva J, Yotov S. Ultrasonic evaluation of uterine involution in Bulgarian Murrah buffalo after administration of oxytocin. *Anim Reprod Sci* 2012; **133**: 71–76.
- [42]Zicarelli L, Campanile G, Infascelli F, Esposito L. Influenza del periodo, dell'età e della distanza dal parto sull'anaestro primaverile della bufala. *Riv Zoot Vet* 1988; **16**: 21–31.