

RESEARCH ARTICLE

Ser81 Survivin Induced Protein Kinase A (PKA)-dependent Phosphatidylinositol 3-kinase (PI3K) ActivityFerry Sandra^{1,2,3,4,*}, Roya Khosravi-Far⁴¹Department of Biochemistry and Molecular Biology, Faculty of Dentistry, Trisakti University, Jl. Kyai Tapa No.260, Jakarta, Indonesia²BioCORE Laboratory, Faculty of Dentistry, Trisakti University, Jl. Kyai Tapa No.260, Jakarta, Indonesia³Prodia Clinical Laboratory, Prodia Tower, Jl. Kramat Raya No.150, Jakarta, Indonesia⁴Department of Pathology, Harvard Medical School, BIDMC, Research North, 99 Brookline Ave., Boston, MA 02215, USA

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Abstract

BACKGROUND: Our previous report showed that phosphorylated-survivin at Ser81 induces survivin back loop to activate protein kinase A (PKA) in the cytoprotection mechanism. Activated PKA could possibly induce the cytoprotection *via* Phosphatidylinositol 3-kinase (PI3K). Therefore our current study was conducted to investigate the possibility of survivin-PKA-PI3K signaling pathway.

METHODS: Viral productions by BOSC23 cells of Survivin, Antisense Survivin (Survivin-AS) and Ser81Ala mutant (Survivin-S81A) in pMSCV-IRES-GFP vector with cytomegalovirus (CMV) promoter were conducted. L929 cells were pretreated with/without PKI 6-22 amide and infected with viral particle of Survivin, Survivin-AS, Survivin-S81A or vector only. Cells were harvested, lysed and immunoprecipitated with anti-PI3K (p85) antibody and immunoblotted to detect PI3K (p85) and phospho-(Tyr) p85 PI3K. To confirm PI3K activation, PI3K Activity Assay was conducted by using phosphoinositide fraction containing PtdIns(4,5)P₂ and [³²P]ATP.

RESULTS: Immunoblot and PI3K activity results showed similar results. Upon infection of virus with survivin, a markedly increased level of tyrosine phosphorylation of p85 PI3K or PI3K activity in L929 cells was seen. Low levels of tyrosine phosphorylation of p85 PI3K or PI3K activity were observed for Survivin-AS and Survivin-S81A-viral-infected L929 cells. With higher concentrations of Survivin-viral-infection, levels of tyrosine phosphorylation of p85 PI3K or PI3K activity in L929 cells were gradually

Abstrak

LATAR BELAKANG: Penelitian kami sebelumnya memperlihatkan bahwa survivin yang terfosforilasi pada *Ser81*, dapat menginduksi *survivin back loop* sehingga mengaktifkan *protein kinase A* (PKA) dalam mekanisme pertahanan sel. PKA yang teraktivasi dapat menginduksi pertahanan sel lewat *Phosphatidylinositol 3-kinase* (PI3K). Oleh karena itu, kami melakukan penelitian ini untuk mengetahui kemungkinan adanya jalur pensinyalan *survivin*-PKA-PI3K.

METODE: Dilakukan produksi virus *Survivin*, *Antisense Survivin* (Survivin-AS) dan *Ser81Ala mutant* (Survivin-S81A) pada plasmid *pMSCV-IRES-GFP* dengan promoter *cytomegalovirus* (CMV) oleh sel BOSC23. Sel L929 diberikan perlakuan dengan/tanpa *PKI 6-22 amide* sebelum diinfeksi dengan virus *Survivin*, Survivin-AS, Survivin-S81A atau vektor saja. Sel kemudian dikumpulkan, dilisis, dan di-immunopresipitasi dengan antibodi anti-PI3K (p85) dan di-immunoblot untuk mendeteksi PI3K (p85) and *phospho-(Tyr) p85 PI3K*. Untuk mengkonfirmasi adanya aktivasi PI3K, tes aktivitas PI3K dilakukan dengan menggunakan fraksi fosfoinositida dan *PtdIns(4,5)P₂* dan [³²P]ATP.

HASIL: Hasil imunoblot dan aktivitas PI3K memperlihatkan hasil yang serupa. Setelah infeksi dengan virus *Survivin*, terlihat peningkatan level p85 PI3K yang terfosforilasi pada tirosin. Peningkatan aktivitas PI3K juga terlihat. Didapatkan level yang rendah pada p85 PI3K yang terfosforilasi pada tirosin dan aktivitas PI3K pada infeksi virus Survivin-AS dan Survivin-S81A. Dengan konsentrasi virus *Survivin* yang semakin tinggi, maka level p85 PI3K

increased. However, when L929 cells were pretreated with PKI 6-22 amide, prior to Survivin-viral-infection, level of tyrosine phosphorylation level of p85 PI3K or PI3K activity was detected much lower.

CONCLUSION: Our result suggest that Ser81 Survivin play role in inducing PI3K activation and the Survivin-PI3K signaling pathway was PKA-dependent.

KEYWORDS: Ser81, Survivin, PKA, PI3K, L929

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Introduction

Among Inhibitor of mammalian Inhibitor of Apoptosis Proteins (IAPs) family, survivin is the smallest member, containing a single Baculovirus IAP Repeat (BIR), which is the hallmark of these molecules.(1,2) Transcription of the survivin gene is strictly cell cycle-regulated, and peaks at mitosis.(1,3) Survivin plays several roles in homeostatic networks, including the control of mitosis, the regulation of apoptosis and the cellular stress response.(1) Therefore, the presence of survivin in cancer has been linked to resistance to apoptosis, metastasis, cell cycle checkpoints bypass and resistance to therapy. In addition, in some cancers, survivin have identified survivin as a 'risk factor' for poor prognosis and disease recurrence.(1,4)

In response to cell death stimulation, survivin physically complexes with X-linked IAP (XIAP), then in turn promotes XIAP stability and synergistic inhibition of caspase-9 activation.(1,5) Besides that, Phosphatidylinositol 3-kinase (PI3K), Akt, β -catenin, T-cell factor (Tcf), Lymphoid enhancer-binding factor (Lef) and many others implicated in the expression of survivin.(6-8) Our previous report showed that Survivin dominant negative enhanced Tumor necrosis factor (TNF)-related apoptosis-inducing ligand (TRAIL)'s activity in inducing apoptosis.(7) In accordance, a recent report showed a positive feedback loop connecting survivin expression in tumor cells to PI3K/Akt, followed by secretion of Vascular Endothelial Growth Factor (VEGF) and angiogenesis.(6)

Amino acid residue Thr34 that lies within survivin BIR domain, was shown as an important amino acid in regulation of apoptosis.(9) Phosphorylated-survivin at Thr34 bound to caspase-9 in HeLa cells.(9) In addition, initiated from Scansite result, Ser81 was shown as another

yang terfosforilasi pada tirosin dan aktivitas PI3K semakin meningkat pula. Akan tetapi, ketika sel L929 diberikan perlakuan dengan *PKI 6-22 amide*, sebelum infeksi dengan *Survivin*, level p85 PI3K yang terfosforilasi pada tirosin dan aktivitas PI3K akan rendah sekali.

KESIMPULAN: *Ser81 Survivin* sangat penting dalam menginduksi aktivasi PI3K dan jalur pensinyalan Survivin-PI3K merupakan jalur yang bergantung pada PKA.

KATA KUNCI: *Ser81, Survivin, PKA, PI3K, L929*

potential target to inhibit survivin-modulated cytoprotection as well as to sensitize efficacy of TRAIL or other related apoptotic inducers.(7) Phosphorylated-survivin at Ser81 was later known mediating survivin back loop to activate protein kinase A (PKA) in the cytoprotection mechanism.(10)

As being reported, phosphodiesterase 4 (PDE4) control intracellular cyclic adenosine monophosphate (cAMP) levels by catalyzing their hydrolysis and inactivating second messengers.(11) PDE4 inhibition promotes PKA-dependent resolution of established neutrophilic inflammation by inhibition of the PI3K/Akt/myeloid cell leukemia 1 (Mcl-1) pathway.(11) Due to our previous study in PKA-activation upon phosphorylation of survivin at Ser81 (10), we assume that the activated PKA could possibly induce cytoprotection via PI3K. Therefore our current study was conducted to investigate the possibility of survivin-PKA-PI3K signaling pathway.

Methods

Preparation of Survivin Constructs, Viral Production and L929 Cells Infection

All preparations of Antisense survivin (Survivin-AS) and Ser81Ala mutants (Survivin-S81A) were described in our previous report (7,10). Similar protocols were conducted. Briefly, Survivin, Survivin-AS, Ser81Ala mutants (Survivin-S81A) cDNAs were inserted in pMSCV-IRES-GFP vector with cytomegalovirus (CMV) promoter. Each vector was transformed in DH5a *Escherichia coli*, purified and confirmed. Each cDNA was transfected in BOSC23 cells by calcium phosphate method with addition of pCIS-EcotR for 10 hours at 37°C. Viruses were harvested at 72 hours after transfection and tittered using NIH3T3 cells.

L929 cells were cultured in α -DMEM containing 10% horse serum. Infection was carried out using viral product of BOSC23 cells (Survivin, Survivin-AS, Survivin-S81A or vector only) for 48 hours.

Cell Treatment, Lysate Preparation and PI3K (p85) Immunoprecipitation

L929 cells were pretreated with/without 4 μ M PKI 6-22 amide, a PKA inhibitor (Santa Cruz Biotechnology, Dallas, TX) and infected with viral particle of Survivin, Survivin-AS, Survivin-S81A or vector only. Cells were then harvested and lysed in a cold lysis buffer (20 mM Tris-HCl buffer pH 8.0, 500 mM NaCl, 1 mM EDTA, 1 mM EGTA, 10 mM β -glycerophosphate, 10 mM NaF, 10 mM pNPP, 0.4 mM sodium orthovanadate, 1 mM benzamide, 2 mM phenylmethylsulfonyl fluoride (PMSF), aprotinin, 1 mg/ml leupeptin, 1 mM dithiothreitol (DTT) and 10% Nonidet P-40). The cell lysate supernatant was collected by centrifugation and incubated with monoclonal anti-PI3K (p85) antibody (Cell Signaling, Beverly, MA) immobilized with protein G Sepharose (Sigma, St. Louis, MO) in immunoprecipitation buffer (20 mM Tris-HCl buffer pH 8.0, 500 mM NaCl, 1 mM EDTA, 1 mM EGTA, 10 mM β -glycerophosphate, 10 mM NaF, 10 mM pNPP, 0.4 mM sodium orthovanadate, 1 mM benzamide, 2 mM PMSF, 10 mg/ml aprotinin, 1 mg/ml leupeptin, 1 mM DTT and 0.1% Nonidet P-40).

Phospho-(Tyr) p85 PI3K Immunoblot

PI3K (p85)-immunoprecipitated samples were separated by SDS-polyacrylamide gel electrophoresis (PAGE) and transferred to a polyvinylidene difluoride (PVDF) sheet. After blocking with 5 % skim milk in Tris-buffered saline (TBS, 150 mM NaCl and 50 mM Tris-HCl, pH 7.4), the sheet was incubated with monoclonal anti-PI3K (p85) (Cell Signaling) or rabbit polyclonal anti-phospho-(Tyr) p85 PI3K binding motif antibodies. The secondary antibody was horseradish peroxidase-conjugated sheep anti-mouse (Amersham, Piscataway, NJ) or donkey anti-rabbit IgG antibody (Amersham). The bound antibodies were visualized using the ECL system (Amersham), captured using Alliance 4.7 (UVItech, Ltd., Cambridge, UK) and quantified using UViband software (UVItech, Ltd.).

PI3K Activity Assay

PI3K (p85)-immunoprecipitated samples were washed with serial solutions as follows, solution A: PBS containing 1% Nonidet P-40 and 1 mM DTT, solution B: consisting of 0.1 M Tris-HCl buffer (pH 7.6), 0.5 M LiCl and 1 mM

DTT, and solution C: containing 10 mM Tris-HCl buffer (pH 7.6), 0.1 M NaCl and 1 mM DTT. This was followed by incubation with 0.5 mg/ml phosphoinositide fraction containing PtdIns(4,5)P₂. Reaction buffer (50 mM MgCl₂, 100 mM HEPES buffer pH 7.6, 250 μ M ATP containing 5 μ Ci of [³²P]ATP) was added and incubated at 30°C for 10 min. Labelled-samples were then spotted on a thin layer chromatography (TLC) plate. The assay was continued as described by Chen *et al.* (12).

Results

Survivin induced tyrosine phosphorylation of p85 PI3K

Similar levels of p85 PI3K were observed for all lanes (Figure 1), showing that the same volume of proteins were electrophoresed and detected. Meanwhile, basal tyrosine phosphorylation of p85 PI3K (lane 1 from left) was seen in L929 cells. Upon infection of virus with vector only (lane 2 from left), the level of tyrosine phosphorylation of p85 PI3K in L929 cells was remained similar to the basal. However, upon infection of virus with survivin, a markedly increased level of tyrosine phosphorylation of p85 PI3K in L929 cells was seen (lane 3 from left). Low levels of tyrosine phosphorylation of p85 PI3K, even lower than the basal, were observed for Survivin-AS (lane 4 from left) and Survivin-S81A-viral-infected L929 cells (lane 5 from left).

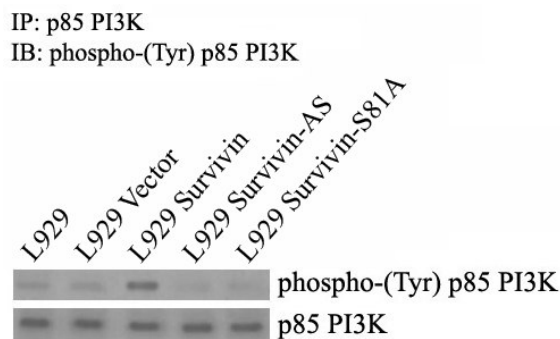


Figure 1. Survivin induced PI3K phosphorylation. L929 cells were infected with 200x10⁷ viral particle/ml retrovirus of Survivin, Survivin-AS, Survivin-S81A or vector merely for 48 hours and cell sorted. Sorted cells were then lysed, immunoprecipitated using anti-p85 PI3K antibody and immunoblotted using anti-phospho-(Tyr) p85 PI3K antibody. Detailed procedures were described in "Methods". This experiment was repeated 3 times.

Survivin induced PI3K activity

As shown in Figure 2, basal PtdIns(3,4,5)P₃ (PIP₃) smeary dot was seen (lane 1 from left). A clear PIP₃ dot was seen for Survivin-viral-infected L929 cells (lane 3 from left). Meanwhile, smeary PIP₃ dots were observed for Vector

(lane 2 from left), Survivin-AS (lane 4 from left) and Survivin-S81A-viral-infected L929 cells (lane 5 from left).

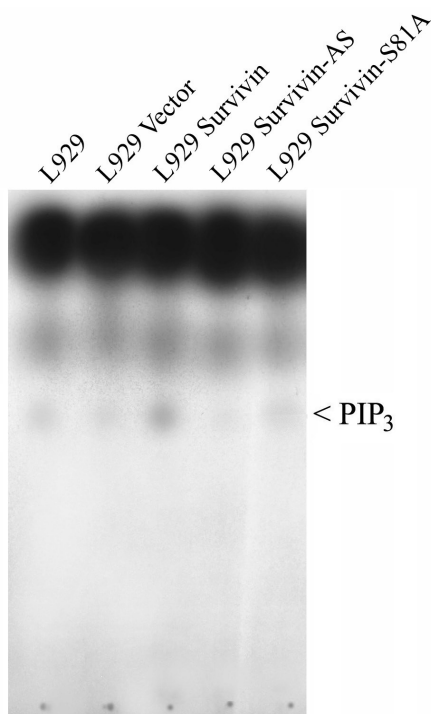


Figure 2. Survivin induced PI3K activity. L929 cells were infected with 200x10⁷ viral particle/ml retrovirus of Survivin, Survivin-AS, Survivin-S81A or vector merely for 48 hours and cell sorted. Sorted cells were then lysed and subjected to PI3K activity assay. Detailed procedures were described in "Methods". This experiment was repeated 3 times.

IP: p85 PI3K

IB: phospho-(Tyr) p85 PI3K

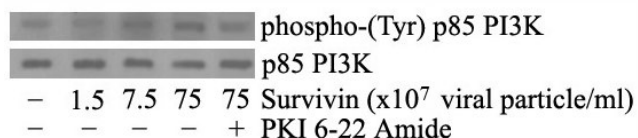


Figure 3. Survivin induced PI3K phosphorylation via PKA
L929 cells were pretreated with/without 4 μ M PKI 6-22 amide for 2 hours prior to infection with different concentrations of Survivin retrovirus for 48 hours as indicated in the panel. Cell were sorted, lysed, immunoprecipitated using anti-p85 PI3K antibody and immunoblotted using anti-phospho-(Tyr) p85 PI3K antibody. Detailed procedures were described in "Methods". This experiment was repeated 3 times.

PKI 6-22 amide inhibited Survivin-induced tyrosine phosphorylation of p85 PI3K

To confirm same proteins were loaded, levels of p85 PI3K were observed. Yet, we found similar levels of p85 PI3K (Figure 3). Basal tyrosine phosphorylation of p85 PI3K (lane 1 from left) was seen in L929 cells. Upon infection of higher concentrations of virus with survivin, levels of

tyrosine phosphorylation of p85 PI3K in L929 cells were gradually increased (lane 2-4 from left). A clear band of tyrosine phosphorylation of p85 PI3K was detected for 75x10⁷ viral particle/ml Survivin-viral-infected L929 cells (lane 4 from left). However, when L929 cells were pretreated with 4 μ M PKI 6-22 amide, a synthetic peptide that acts as PKA inhibitor, prior to 75x10⁷ viral particle/ml Survivin-viral-infection, tyrosine phosphorylation level of p85 PI3K was detected much lower (lane 5 from left) compared to the one without PKI 6-22 amide pretreatment (lane 4 from left).

PKI 6-22 amide inhibited Survivin-induced PI3K activity

Basal PIP₃ smeary dot was seen (Figure 4, lane 1 from left). Upon infection of higher concentrations of virus with survivin, levels of PIP₃ dots were gradually clearly seen (lane 3-5 from left). A clear PIP₃ dot was detected for 75x10⁷ viral particle/ml Survivin-viral-infected L929 cells (lane 5 from left). However, when L929 cells were pretreated with 4 μ M PKI 6-22 amide, prior to 75x10⁷ viral particle/ml Survivin-viral-infection, smeary PIP₃ dot was resulted (lane 2 from left).

Discussion

Evidence for the existence of a positive feedback loop connecting survivin expression in tumor cells to PI3K/Akt was reported.(6) This signal enhanced β -catenin-Tcf/Lef-dependent transcription followed by secretion of VEGF and angiogenesis.(6) In accordance, our current results had shown that Survivin could induce tyrosine phosphorylation of p85 PI3K and PI3K activation specifically. While Survivin induced tyrosine phosphorylation of p85 PI3K and PI3K activity, Vector merely and Survivin-AS did not induce neither tyrosine phosphorylation of p85 PI3K nor PI3K activity. Based on our previous report, phosphorylation of Ser81 was crucial for activation of PKA.(10) In our current results, we found phosphorylation of Ser81 was also crucial in inducing tyrosine phosphorylation of p85 PI3K and activating PI3K. Survivin-PKA-PI3K signal pathway possibly play important role in anti-apoptosis/cytoprotection. In the opposite, Survivin-S81A-viral-infection was reported to enhance TRAIL's activity in inducing apoptosis.(7)

PKA was reported to have potential in regulating insulin receptor substrate-1 (IRS-1) and growth factor receptor bound protein 2-associated binding protein 2 (GAB2), adaptors that normally integrate receptor/nonreceptor tyrosine kinase signaling into PI3K/Akt.(13)

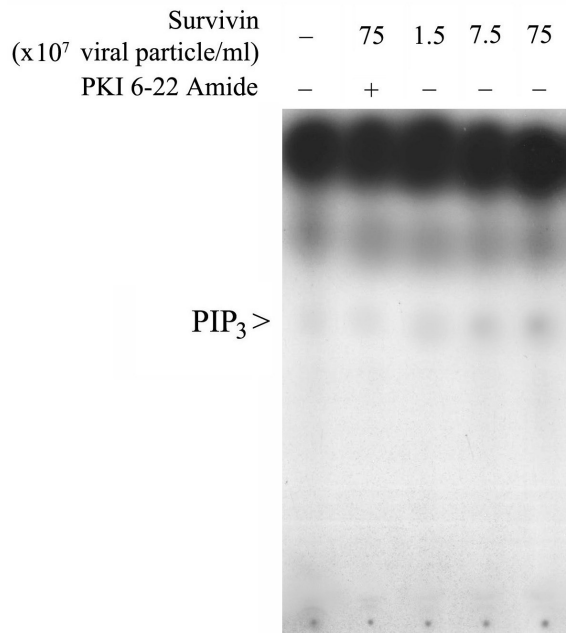


Figure 4. Survivin induced PI3K activity via PKA. L929 cells were pretreated with/without 4 μ M PKI 6-22 amide for 2 hours prior to infection with different concentrations of Survivin retrovirus for 48 hours as indicated in the panel. TNF- α was Cell were sorted, lysed and subjected to PI3K activity assay. Detailed procedures were described in "Methods". This experiment was repeated 3 times.

This reveals a new route for PKA to activate a pathway that promotes proliferation, inhibits apoptosis and enhances translation.(13) This report is in accordance with our previous report, showing that phosphorylated-Survivin at Ser81 induced PKA back loop.(10) In addition, our current results emphasized that Survivin-induced PI3K activation was mediated by PKA, since PKI 6-22 amide inhibited Survivin-induced pathway, as shown by lower tyrosine phosphorylation of p85 PI3K and PI3K activity upon pretreatment of PKI 6-22 amide prior to Survivin-viral infection.

Phosphatase and tensin homolog deleted on chromosome 10 (PTEN)/PI3K/Akt constitute an important pathway regulating the signaling of multiple biological processes such as apoptosis, metabolism, cell proliferation and cell growth.(14-17) PTEN is a non-redundant phosphatase, counteracting one of the most critical cancer-promoting pathways: the PI3K/Akt signaling pathway and its main substrate PIP3 is the product of PI3K.(14-19) Increase in PIP3 recruits Akt to the membrane where is activated by other kinases also dependent on PIP3.(14) Another positive feedback mechanism that decrease of E-cadherin, lowers PTEN and thereby increases PIP3, further activating Akt.(20) Many components of this pathway have been described

as causal factors in cancer. (14,15,17,18, 20) Activation of PI3K and AKT are reported to occur in breast, ovarian, blood, pancreatic, esophageal, gallbladder and other cancers. (14,17) Taken together, our current results suggest that Ser81 Survivin play an important role in inducing PI3K activation and the Survivin-PI3K signaling pathway was PKA-dependent. This provides important information to understand the mechanism induced by Survivin in mediating cytoprotection. However, further investigation to disclose downstream of Survivin-PKA-PI3K signaling pathway is necessary.

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