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## Asian Pacific Journal of Tropical Medicine

journal homepage: [www.elsevier.com/locate/apjtm](http://www.elsevier.com/locate/apjtm)

Document heading doi:

Bioactivity guided isolation of mosquito larvicide from *Piper longum*Madhu SK<sup>1\*</sup>, Vijayan VA<sup>2</sup>, Shaukath AK<sup>3</sup><sup>1</sup>Vector Biology Research Lab, Department of Studies in Zoology, University of Mysore, Manasagangotri, Mysore 570 006, Karnataka, India<sup>2</sup>Department of Studies in Zoology, University of Mysore, Manasagangotri, Mysore 570 006, Karnataka, India<sup>3</sup>Department of Chemistry, Yuvaraja's College, University of Mysore, Mysore-05, India

## ARTICLE INFO

## Article history:

Received 1 November 2010

Received in revised form 16 December 2010

Accepted 15 January 2011

Available online 20 February 2011

## Keywords:

*Piper longum*

Bioassay

Pipyahyine

*Culex quinquefasciatus*

Filariasis

Larvicide

## ABSTRACT

**Objective:** To isolate the larvicidal component from the fruits of *Piper longum* (*P. longum*) against the filariasis vector, *Culex quinquefasciatus* (*C. quinquefasciatus*). **Methods:** Pulverized fruits of *P. longum* were subjected to soxhlet extraction using series of organic solvents of increasing polarity. All the solvent extracts were verified for their larvicidal efficacy against 4th instar larvae of *C. quinquefasciatus* employing standard WHO procedure. Bioassay-guided fractionation through column chromatography lead to the isolation of a bioactive amide, pipyahyine from the petroleum ether extract. **Results:** Petroleum ether extract was found to be the most active fraction among all the extracts tested with LC<sub>50</sub> and LC<sub>90</sub> being 1.03 and 2.04 ppm respectively. Whereas, pipyahyine, an isolated component of the same fraction was found to be even more effective than the parent extract in terms of LC<sub>50</sub> being 0.58 and 1.88 ppm respectively. **Conclusions:** From the results, it is evident that *P. longum* can be considered as a powerful arsenal for the control of mosquito population.

## 1. Introduction

Mosquitoes not only create a nuisance as biting insects but are etiologic agents for some of the devastating diseases of human history such as malaria, filariasis, chikunguniya, dengue etc. Out of 120 million actual cases of lymphatic filariasis in 83 countries, India alone contributes around 39%[1]. *Culex quinquefasciatus* (*C. quinquefasciatus*) is a domestic mosquito vector of lymphatic filariasis. It not only cause a high level of mortality and morbidity but also responsible for great socio-economic loss. Indeed, the present recrudescence of this disease include increasing number of vector breeding places and large number of parasite infected tourists/immigrants. On the other hand, management of filariasis through chemotherapy is usually effective at early infection. However, toxic side effects are often experienced during such treatments[2]. Consequently, the most reliable approach to diminish the disease incidence is to interrupt the vector lifecycle. The systematic application of insecticides is a common and

widely accepted approach to control mosquito population, as it will provide rapid solution. On the other hand, the chemical control measures although highly effective, vector control is still facing a threat, as selective pressure imposed by conventional insecticides is enhancing resistance in various mosquito species resulting in disease outbreak[3]. In certain cases, resistance to microbial agents by several mosquito species have also been elucidated[4]. The recent negative impact of chemical insecticides has shifted the research efforts towards development of new environmentally compatible vector control methods by using naturalistic agents. In search of new vector control strategy, science has intensified the probes to plants in recent decades, so the plant kingdom is receiving renewed attention as mosquitocides. Pavea has verified the efficacy of 56 plants species against the fourth instar larvae of *C. quinquefasciatus*[5]. Nazar *et al* have tested 100 plants for mosquito larvicidal activity[6]. Recently Scott *et al* has carried out detailed studies on different species of piper and found that phytochemistry of this genus is rich in terms of insecticidal amides[7]. Park *et al* have reported the four active isobutylamide alkaloids against *C. pipiens*, *A. aegypti* and *A. togoi* larvae from *Piper nigrum* (*P. nigrum*)[8].

*P. longum* (Indian long pepper or pippali), belonging to the family *Piperaceae* is a slender aromatic climber with red fruits. These fruits turns greyish black with pungent smell

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when dried and reported as good remedy for the treatment of gonorrhoea, menstrual pain, tuberculosis, sleeping problems, respiratory track infections, chronic gut related pain and arthritic condition[9]. Perusal of literature reveals that there have been several attempts to investigate the insecticidal activity of this plant[10,11]. As *P. longum* has proven to be an eco-friendly, our interest triggered us to explore the phyto-component responsible for the mosquito larvicidal activity of this plant against larvae of *C. quinquefasciatus*, a bancroftian vector.

## 2. Material and methods

Fruits of *P. longum* were purchased from local farmers of Mysore. All the organic solvents used in the experiments were of analytical grade and procured from sd fine chemicals, Bombay. Silica gel was obtained from Merck India limited, Bombay. Early 4th instar *C. quinquefasciatus* larvae used in the experiments were maintained in the insectarium of the Vector Biology Research Lab of Zoology department, University of Mysore, Mysore. The approval of the Institutional animal ethics committee, University of Mysore was taken for the use of mice to maintain mosquito colony.

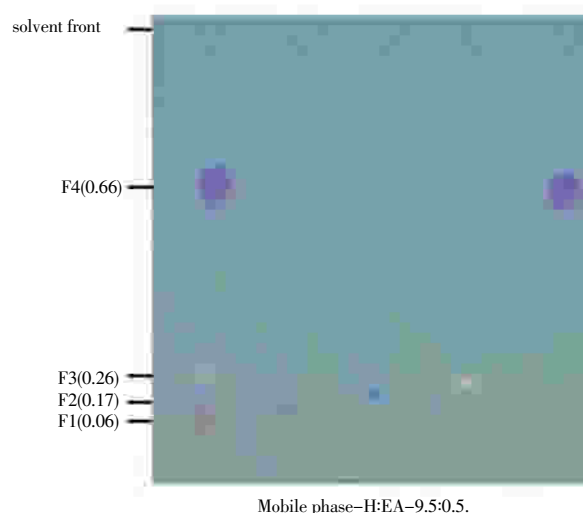
### 2.1. Phytoextracts

A soxhlet extraction was carried out with pulverised fruit material using series of organic solvents of increasing polarity viz, petroleum ether, hexane chloroform, ethylacetate, acetone and ethanol until exhaustion. Solvents were distilled in a vacuum rotary evaporator under reduced pressure of 20–22 mmHg at 35 °C and extract concentrates were further evaporated to complete dryness at room temperature. Known amount of extracts were dissolved in known amount of acetone and stocks solutions were tightly closed and stored in brown bottles at 4 °C. Surface tension of each test solution was measured using stalagmometer to ensure the complete miscibility. Each extracts were assayed for their larvicidal efficacy with variable concentrations and doses were fixed to give larval mortality ranging from 10 to 98 percent. As petroleum ether extract was found to be the most bioactive, it was selected for further analysis.

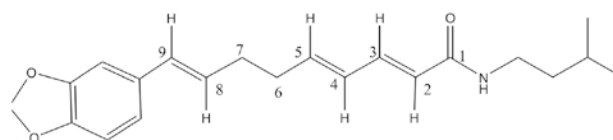
### 2.2. Isolation and purification of active component

The phytoextract of petroleum ether was dissolved in diethyl ether and subsequently washed with dilute bicarbonate, dilute base, mild acid and repeatedly with distilled water followed by neutralization at each step in a separating funnel. Then, the final diethyl ether fraction was completely dried over anhydrous calcium chloride and chromatographed over thin layer of silica gel (GF254) with standardized mobile phase of hexane and ethyl acetate in the ratio 9:1. Chromatogram was inspected under UV light at wave length of 254 as well as 366 nm and Rf values were calculated for major bands detected. 10 g of petroleum ether extract was ground with 2 g of silica gel (240–400 mesh) and loaded to the silica gel column previously equilibrated with hexane and successfully eluted with a step wise gradient of hexane and ethyl acetate (0:100; 10:90...90:10...100:0).

The 30 sub-fractions so obtained were pooled to 4 major fractions (F1, F2, F3 and F4) based on the bands appeared on TLC (Figure 1). The active F4 was found to be a complex mixture of amides. Further, the most active compound (112 mg) was obtained by repeated column chromatography. Isolated compound was dissolved in hot ethanol and passed through activated charcoal for crystallization and was subjected to RP-HPLC on a Vydac C18 column (250 mm×4.6 mm, 10 μm particle size, 300 Å pour size) in a Shimadzu LC-10 AVP system with dual wavelength detector. Purified compound was carefully evaporated to complete dryness at room temperature and subsequently characterized as pipyahyine on the basis of spectroscopic analytic data. Larval bioassays using pure compound were conducted to find out its efficacy against *C. quinquefasciatus* 4th instar larvae.



**Figure 1.** TLC profile of petroleum ether extract of *P. longum* fruits.



**Figure 2.** Structure of bioactive compound isolated from *P. longum* fruit.

### 2.3. Spectroscopic analysis

Isolated compound was analysed by IR, UV, <sup>1</sup>H, <sup>13</sup>C-NMR and mass spectra. Infrared spectra were recorded on a FT, IR, Shimadzu 8300 spectrophotometer. UV spectral data were obtained from Hitachi UV-3900 spectrophotometer. <sup>1</sup>H-NMR spectra were recorded on a Bruker 300 MHz spectrophotometer and <sup>13</sup>C-NMR spectra were on 300 MHz, JEOL-FX-90Q, FT spectrophotometer in CDCl<sub>3</sub> using TMS as internal standard.

### 2.4. Larval bioassay

Larval susceptibility tests of different solvent extracts

and isolated compound were performed employing WHO standard procedure<sup>[12]</sup>. Various concentrations of the extracts were prepared by serial dilutions of a stock solution in acetone. Batches of 25 early fourth instar larvae were released into glass beakers of 500 mL capacity containing 249 mL of dechlorinated tap water and 1.0 mL of extract. The toxicity of each extract was determined with five various concentrations ranging from 0.2 to 20.0 ppm that provided a range of 10% to 98% mortality. Control beakers contained 25 test organisms and 249 mL of tap water along with 1.0 mL acetone. Treated and control beakers were maintained at same conditions at (25±2) °C, 12 h light/dark regime. No food was provided to the larvae during the test period of 24 h till the mortality was monitored. All treatments were replicated 4 times. The larvae were considered as dead or moribund, if they were not responsive to gentle prodding with a fine needle. The results were expressed as percent mortality.

### 2.5. Statistical analysis

The observed percent mortality was adjusted for the control mortality, using Abbot's formula<sup>[13]</sup>, and then subjected to regression analysis of probit–mortality on log dosage using computerized log–probit analysis (Harvard programming; Hg1.2), providing lethal dosage of 50, 90, 95, 99 ppm as well as their 95% confidence limit. One way variance analysis (ANOVA) was performed followed by post hoc Tukey's Honestly Significant Difference Test (HSD) using SPSS software, version 11.5. *P* value less than 0.05 were considered to indicate statistical significance.

## 3. Results

The susceptibility levels of fourth instar larvae of filarial vector to different solvent extracts of *P. longum* are shown in Table 1. From the results, it appeared that all the extracts exhibited larvicidal activity at variable concentrations. Out of six solvent extracts, the effect was highly significant (*P*<0.01) in petroleum ether extracts in terms of LC<sub>50</sub> and LC<sub>90</sub>

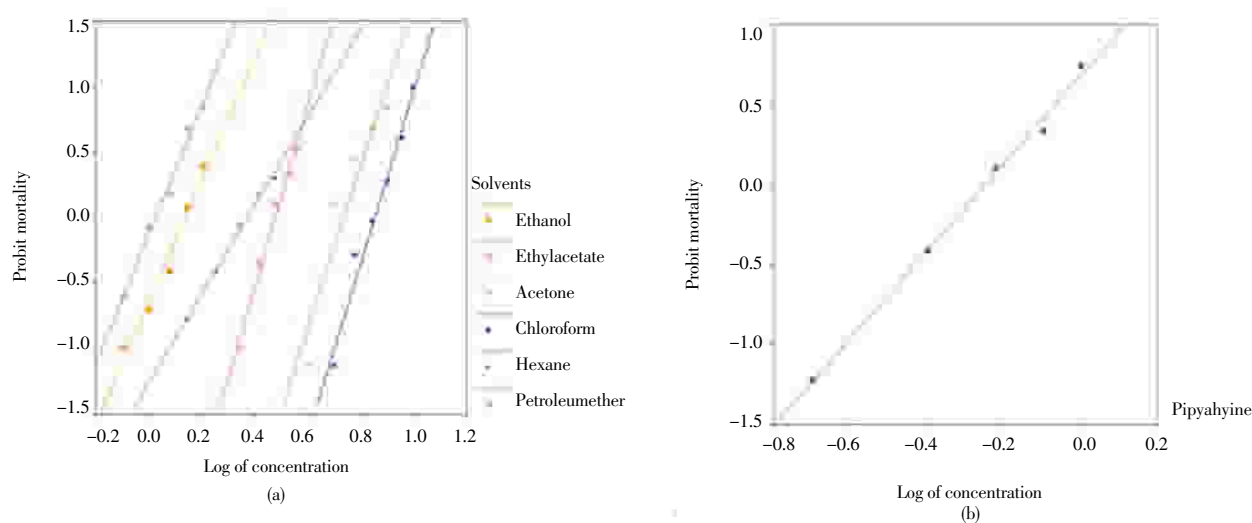
being 1.03 and 2.04 ppm respectively. Significant lethality was also exhibited by ethanol and ethyl acetate with respect to acetone with LC<sub>50</sub> being 1.37 and 2.98 respectively (*P*<0.01). However, hexane and chloroform extracts showed significant effect at 5% level with LC<sub>90</sub> being 5.66 and 11.24 ppm respectively (*P*<0.05), whereas, the acetone extract required higher concentrations (21.31 ppm) than all other extracts to produce 90% larval mortality. The slopes of regression lines for different phytoextracts were computed and provided in Figure 3a.

### 3.1. Bioactive principle

Purified phytoisolate was characterized as pipyahyine and its larval efficacy against mosquitoes are emphasized in Table 2. The compound exhibited larval toxicity with LC<sub>50</sub> being 0.58 and 1.88 ppm respectively. *Culex* immatures produced 77% mortality at concentration of 1.2 ppm whereas 11% mortality was observed in 0.2 ppm. However, the dose–mortality responses are depicted in Table 2 and Figure 3b.

Structure of pipyahyine is provided in Figure 2, which was slowly solidified as white needles (112 mg/10g). IUPAC: 9–Benzol[1,3]dioxol–5–yl–nona–2,4,8–trienoic acid (3–methyl–butyl)–amide IR (Neat): 1400–1600 (C=C aromatic aliphatic), 1650 (C=O), 3330 cm<sup>-1</sup> (–NH); <sup>1</sup>H NMR (CDCl<sub>3</sub>): 1.01 (d, 6H, isopropyl 2CH<sub>3</sub>), 1.5 (q, 2H, CH<sub>2</sub> adjacent to isopropyl group), 1.8 (m, 1H, isopropyl C–H), 2.0 (t, 4H, C<sub>7</sub> & C<sub>6</sub>–H), 2.95 (t, 2H, amide CH<sub>2</sub>), 5.5 (q, 1H, C<sub>5</sub>–H), 5.9 (s, 2H, dioxole–H), 6.0 (q, 1H, C<sub>8</sub>–H), 6.2 (t, 1H, C<sub>4</sub>–H), 6.4 (d, 1H, C<sub>9</sub>–H), 6.55 (d, 1H, C<sub>2</sub>–H), 6.8–7.0 (m, 6H, Ar–H), 7.3 (t, 1H, C<sub>3</sub>–H), 7.9 (bs, 1H, CONH); C<sub>13</sub>NMR (CDCl<sub>3</sub>) 22.0 (q for 2CH<sub>3</sub>), 25.6 (d for isopropyl C–H), 33.0 (t for C<sub>7</sub>), 33.4 (t for C<sub>6</sub>) 40.3 (t for amide CH<sub>2</sub>), 40.8 (t for CH<sub>2</sub> adjacent to isopropyl group), 91.3 (t for dioxole CH<sub>2</sub>), 112.8 (d for Ar–CH), 115.0 (d for Ar–CH), 119.5 (d for Ar–CH), 123.8 (d for C<sub>2</sub>), 126.3 (d for C<sub>6</sub>), 127.6 (d for C<sub>9</sub>), 128.2 (s for Ar–C), 128.7 (d for C<sub>6</sub>), 130.3 (d for C<sub>5</sub>), 142.7 (d for C<sub>3</sub>), 146.8 (s for Ar–C), 147.5 (s for Ar–C), 166.5 (s for CO group).

EI–MS: m/z 341 (M<sup>+</sup>, 65). Anal. Calcd. for C<sub>21</sub>H<sub>27</sub>NO<sub>3</sub> (341): C, 73.87; H, 7.97; N, 4.10. Found: C, 73.67; H, 7.83; N, 4.19%.



**Figure 3.** a: Log concentration–percent mortality relationship of *C. quinquefasciatus* larvae to *P. longum* fruit extracts. b: Log concentration–percent mortality relationship of *C. quinquefasciatus* larvae to pipyahyine, an active compound obtained from *P. longum* fruit.

**Table 1**Larvicidal efficacy of a few solvent extracts of *P. longum* fruits against *C. quinquefasciatus*.

Solvent extracts	LC <sub>50</sub> (ppm)(LCL–UCL)	LC <sub>90</sub> (ppm)(LCL–UCL)	Slope±SE	χ <sup>2</sup>	P value
Petroleum ether	1.03(0.98–1.10)	2.04(1.82–2.39)	4.36±0.40	2.76	0.0001**
Hexane	2.35(2.17–2.58)	5.67(4.49–8.61)	3.35±0.50	0.39	0.02*
Chloroform	7.08(6.79–7.36)	11.24(10.44–2.45)	6.37±0.57	3.48	0.04
Acetone	15.64(15.24–16.16)	21.31(20.61–26.84)	8.77±0.48	7.09	0.788
Ethyl acetate	2.98(1.24–1.39)	4.59(2.10–2.74)	6.84±0.79	1.21	0.001**
Ethanol	1.37(1.29–1.47)	2.55(2.18–3.28)	6.98±0.50	2.24	0.001**

LCL: Lower confidence limit; UCL: Upper confidence limit. \*P&lt;0.05 significant difference at 5%; \*\*P&lt;0.01 Significant difference at 1%.

**Table 2**Mosquito larvicidal effects of a purified compound from petroleum ether extract of *P. longum* fruits.

Test compound	Concentration (ppm)	Percent mortality	LC <sub>50</sub> (ppm) (LCL–UCL)	LC <sub>90</sub> (ppm) (LCL–UCL)	Slope±SE	χ <sup>2</sup>
Control	0.0	0.0	–	–	–	–
Pipyahyine	0.2	11.1	0.58(0.52–0.65)	1.88(1.57–2.58)	2.50±0.25	0.63
	0.4	34.8	–	–	–	–
	0.6	54.3	–	–	–	–
	0.8	63.0	–	–	–	–
	1.2	77.0	–	–	–	–

LCL: Lower confidence limit; UCL: Upper confidence limit.

#### 4. Discussion

It is very important to monitor the insecticide susceptibility status of mosquito vectors at different areas in view of the resurgence of various communicable diseases. All the organic components of the tested extracts revealed larvicidal activity against the filarial vector. High significant ( $P<0.01$ ) results were exhibited by both polar (ethanol) and non polar (petroleum ether) phases of the extracts. Our results are in consonance with the studies conducted by Moawed, against *C. pipiens* using petroleum ether and ethanol extracts of *P. nigrum*[14]. Literature survey has revealed bioassay experiments for exploring the insecticidal activity on mosquito vectors from many piper species. But most of the studies with piper species on mosquitoes are concentrated on *P. nigrum*, the common pepper[14–19]. However, considerable works have also been done with several other plants of the same species. For instance, three plants namely *P. logum*, *P. ribesoides* and *P. sarmentosum* have shown adulticidal activity against *Stegomyia aegypti* at Thailand[10]. The ethanol extracts of *P. beetle* has successfully killed the larvae of 4 mosquito vectors *A. aegypti*, *C. quinquefasciatus*, *A. dirus* and *Monsonia uniformis*[20]. It was noted that most of the compounds responsible for insecticidal activity from these species are amides[7]. Four amide compounds namely pipnoohine, pipyahyine, piptigine and pipwaquarine possessing larvicidal toxicity against *A. aegypti* have been reported from *P. nigrum* fruits in separate studies by Siddiqui *et al*[16–18]. Rasheed *et al* have isolated 14 amide components from seeds of *P. nigrum*, all of which showed insecticidal activity against 4th instar larvae of *A. stephensi* and *A. aegypti*[19]. Findings from the present study too strongly support such observations, as major bioactive fraction (80%) obtained from this plant was a mixture of amides. Many derivatives of piperamides have been synthesized in search of new insecticides and some of them

are in the way of successes[21–23]. The literature offers limited publications on the isolation of bioactive components from *P. longum*. Lee and Yang *et al* have isolated piperonaline, an alkaloid as mosquito larvicide from methanol extract of this plant[24,25]. But the authors failed to find such a compound in present investigation.

By repeated chromatographic resolution, the active isobutyl amide was characterized as pipyahyine (Figure 2) with molecular formula  $C_{24}H_{33}NO_5$ . This has nine unsaturations among which five are accounted by aromatic ring. The IR spectrum showed absorption for a secondary amide and its NMR spectra supported the presence of isopentyl group and signals for a benzo (1, 3) dioxy moiety conjugated with double bond. These spectral peaks matched exactly with that of pipyahyine, isolated from the whole fruits of *P. nigrum* by Siddiqui *et al*[16]. Further, they have also tested its larvicidal efficacy against *A. aegypti* with LC<sub>50</sub> value of 30.0 ppm. In contrast to that, the larvicidal effect was 29.5 times more effective against *C. quinquefasciatus* with LC<sub>50</sub> being 0.58 ppm with the same compound from *P. longum* in the present investigation. Moreover, the compound yield (11.5 mg/10 g) from *P. nigrum* was comparatively very less than that of *P. longum* (112 mg/g). The difference in the activity and components may be due to the nature and nurture of the plants and test organisms used in the experiments. It is important to note that the bioactivity of the extracts mainly depend on the chemistry of photoproduct, number of compounds present, nature of the solvent and procedure followed for extraction[26]. Our earlier studies have illustrated a promising synergistic activity of this plant and propoxur with synergistic factor 4.13[27]. Piperonyl butoxide, the well known synergist derived from Piper species is being used extensively to reduce the dosage of synthetic chemical and to overcome resistance problem in vectors[28]. In this regard, our present work seems to be different and quantitative as it involves simple procedure to isolate more



quantity of larvicide/synergist from this local medicinal plant. The larvicidal activity of this plant is encouraging and might form an eco-friendly and human compatible arsenal in vector management programme. Further, the biochemical mechanisms and mode of action are under investigation and warrants extensive study.

### Conflict of interest statement

We declare that we have no conflict of interest.

### Acknowledgements

Authors are thankful to Department of Science and Technology, New Delhi, India for providing financial assistance under women scientist scheme - B.

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