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Original article

Adenosine deaminase isoenzymes estimation – as a diagnostic tool for tuberculous pleural effusions

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Abstract

Objective: To assess the efficacy of ADA isoenzyme estimation over that of total ADA level in pleural fluid and serum as a more efficient diagnostic indicator in tuberculous pleural effusions in high prevalence country like India. **Methods:** The efficacy was analysed in total thirty four patients of pleural effusions. Total ADA was estimated by Guitsi and Galanti Calorimetric method and ADA isoenzymes with and without EHNA[Erythro-9-(2-hydroxy-3-nonyl) adenine] a potent ADA₁ inhibitor using the same method. **Results:** The results demonstrated a statistically significant values of ADA₂ in serum (P < 0.001), pleural fluid (P = 0.000) and significant value for the ratio of pleural fluid ADA₂/serum ADA₂(P < 0.001) and pleural fluid ADA₁/ADA₂(P < 0.005). The sensitivity and specificity values of pleural fluid ADA₂ is 81.8% and 91.6% (cut off value 60 IU/L for Tuberculous effusions), serum ADA₂ 95.4% and 66% (cut off value 70 IU/L for tuberculous effusions). ADA₂ is an isoenzyme, which is significantly raised in tuberculous pleural effusions both in the serum and pleural fluid. **Conclusion:** Estimation of ADA isoenzymes is redundant as a diagnostic aid over total ADA estimation in view of the limited improvements both in specificity and sensitivity patterns and also in term of cost-benefit ratio.

Keywords: Adenosine deaminase; Isoenzymes; Tuberculous pleural effusions

INTRODUCTION

Tuberculous pleural effusion is a global problem and its special reference to the high prevalence country like India cannot be ignored. Tuberculous pleural effusions pose a diagnostic problem for the clinician before starting the specific therapy.

Adenosine deaminase (EC 3. 5. 4. 4) called ADA^[1], catalyses the pathway from adenosine and deoxyadenosine to inosine and deoxyinosine. ADA

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exists in 3 molecular masses or isoenzymes ADA₁, ADA_{1+CP}, ADA₂. These isoenzymes are coded by different genes. ADA₁ is monomeric protein with a molecular mass of about 35 kDa (gene assignment chromosome 20); ADA_{1+CP} (molecular mass about 280 kDa) is composed of two ADA₁ molecules (dimer) connected via a connecting protein (CP)-a glycoprotein (gene assignment, chrome 2 and 6). The third isoenzyme ADA₂, appears to be coded by separate gene locus of unknown chromosomal position ($\approx 100~{\rm kDa})^{[2]}$. The isoenzymes are divided based on the different kinetic properties.

Putting one of the properties to use i. e. selective inhibition of ADA₁ by Erythro-9 (2-hydroxy 3-nonyl) adenine (EHNA), an electrophoretic technique was developed by Buel E^[3] and later on modified by Ungerer *et al*. He studied using this principle and esti-

mated that ADA₂ is the predominant (mean 88 %) contributor of total ADA in tuberculous pleural effusions and ADA₁ and ADA_{1 + CP} in parainfective etiologies (mean 70 %). ADA₂ activity principally reflects that of monocytes and macrophage turnover and has been evaluated as a diagnostic tool in the tuberculous pleural effusions. These studies were done in view of the varied cut-off points and positive predictive values, sensitivities and specificities and associated limitations of ADA estimation in diagnosing tuberculous pleural effusions [4-20] and varying positive predictive value with age of patients^[21]. The estimation of ADA isoenzymes has been proved as an efficient biochemical parameter for the diagnosis of tuberculous pleural effusions than total ADA levels^[22-27].

With different studies taking different combinations of obtained parameters for diagnosing the tuberculous pleural effusions, the present study tries to evaluate the utility of ADA isoenzymes in high prevalence country, India.

MATERIALS AND METHODS

Study design

Twenty two confirmed patients of tuberculous pleural effusions (median age of 35.5 years, range 18-62) were evaluated, 12 patients of confirmed malignant pleural effusions were taken as the comparative group as they constitute an important etiological cause of pleural effusions after tuberculosis (median age 54.7, range 28-74) The patients were confirmed by standard diagnostic criteria like cultures for tuberculous mycobacterium of pleural fluid and pleural biopsy for histopathological examination and culture.

Properties of ADA₁ and ADA₂ isoenzymes [28-31] were shown in Table 1. Total ADA was estimated by Guitsi and Galanti Calorimetirc method [32]. To estimate the levels of ADA₁ and ADA₂ isoenzymes, the ADA activity was measured by the same above technique with and without Erythro-9 (2-hydroxy 3-nonyl) adenine (EHNA). EHNA is a potent inhibitor of only ADA₁ isoenzyme and a concentration of 200 μ mol/L was used in the reaction [29]. In its presence, only the ADA₂ isoenzyme is active. The ADA₁ activity is then calculated by subtracting the ADA₂ isoenzyme from the total activity (this method cannot differentiate between ADA₁ and ADA_{1+GP}).

Statistical analysis

The results were analysed using the non-parametric Wilcoxon Mann-Whitney U-test. The results of the

parameters were then evaluated by looking at sensitivity, specificity, positive predictive value (PPV), negative predictive value (NPV), percentage of false positivities and percentage of false negativities.

RESULTS

The mean values of the total ADA, ADA₂ both in pleural fluid and serum were significantly higher in tuberculous pleural effusions in comparison to malignant effusions (P < 0.001). Similarly the ratio of ADA₂ in pleural fluid and serum is found statistically significant (P < 0.005) in tuberculous pleural effusion (Table 2).

The sensitivity, specificity, positive predictive value, negative predictive values of different parameters is shown in Table 3. There is no marked variation in the sensitivity and specificity values of pleural fluid ADA, over that of the total ADA estimated when a higher cut off value is taken; in our study the values for total pleural fluid ADA were estimated when the cut off value of 70 IU/L is taken. This higher cut off value is taken, when compared to the previous studies to get a comparative estimation of the efficacy of isoenzyme estimation over that of the total ADA. Even though mycobacterium culture and histopathology of pleural biopsy is the specific test it has widely different and low sensitivities. Sensitivities of the other parameters using the isoenzymes were found to be higher than the total ADA even though a higher cut off value is taken for it (Figure 1).

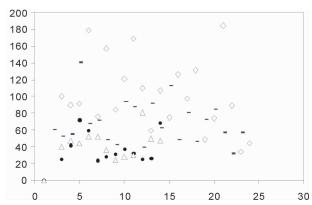


Figure 1 Distribution of ADA₂ in pleural fluid and serum of tuberculous and malignant pleural effusions.

- · Serum ADA2 of malignant pleural effusions
- ♦ Pleural fluid ADA₂ in tuberculous pleural effusions
- △ Pleural fluid ADA₂ in malignant pleural effusions
- Serum ADA2 in tuberculous pleural effusions

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Table 1 Properties of ADA_1 and ADA_1 isoenzymes [28-31].

Properties	ADA_1	ADA_2	
Site of distribution	> 90 % of Adenosine deaminase activity in human cells and tissues	Specially seen in spleen with 2 % of Adenosine deaminase activity and monocytes (18 %)	
Molecular weight	ADA ₁ -35 kDa ADA _{1 + CP} -280 kDa	100 kDa	
Optimum pH	5.5-8.0	6.5	
Km for Adenosine	50 μmol/L	2 000 μmol/L	
Deaminating activity for 2' deoxyadenosine versus adenosine	0.8 (higher)	0.2 (lower)	
Inhibition by EHNA	Inhibited	Insensitive for inhibition	
Gene assignment	$\mbox{ADA}_1\mbox{-chromosome}$ 20 $\mbox{ADA}_{1+\mbox{CP}}\mbox{-chromosome}$ 2 and 6	${\rm ADA_2 ext{-}coded}$ by separate gene locus of unknown chromosomal position	

Table 2 Mean values of the different parameters in tuberculous and malignant pleural effusions (values in IU/L).

Parameter	Malignant	Tuberculous	P
Serum ADA ₁	6.23	7.70	< 0.790
Serum ADA ₂	39.16	68.79	< 0.001 *
Total Serum ADA	45.38	76.50	< 0.001 *
Pleural fluid ADA ₁	10.99	14.82	< 0.423
Pleural fluid ADA ₂	44.25	102.09	0.000 *
Total Pleural fluid ADA	55.41	116.91	0.000 *
P/S ratio of ADA ₁	2.04	3.08	< 0.632
P/S ratio of ADA ₂	1.02	1.50	< 0.001 *
Serum ADA ₁ /ADA ₂	0.17	0.13	< 0.136
Pleural fluid ADA ₁ /ADA ₂	0.27	0.18	< 0.005 *

^{* =} significant and others not significant

P/S ratio = pleural / serum ratio

Table 3 Yield of different parameters in diagnosing tuberculous pleural effusion, which were found to be statistically significant.

	Total ADA	Pl. fluid ADA ₂	Serum ADA ₂	P/S ratio of ADA ₂	Pl. fluid ratio of ADA ₁ /ADA ₂
Sensitivity	86.36 %	81.8 %	95.4 %	95.45 %	80.0 %
Specificity	91.66 %	91.6 %	66.0 %	75.00 %	83.3 %
P P V	95.00 %	94.7 %	84.0 %	87.50 %	88.8 %
N P V	78.57 %	73.3 %	66.6 %	60.00 %	71.4 %
% of F N	13.63	18.00	4.50	4.50	20.00
% of F P	8.33	8.30	25.00	25.00	16.60
Cut-off value for tuberculosis	≥70 IU/L	≥60 IU/L	≥40 IU/L	≥1.00	≤0.20

⁽ Pl. fluid = Pleural fluid, P P V = positive predictive value, N P V = Negative predictive value, % of F N = percentage of false negatives, % of F P = percentage of false positives)

DISCUSSION

The diagnosis of tuberculous pleural effusion is best made on the conventional diagnostic techniques of pleural biopsy (culture and histopathology). ADA estimation has over taken other markers in the march as the cheap and easily performed tests with wide acceptability in India and other countries, amply demonstrated by a number of studies with wide range of specificities and sensitivities, and wide availability of the test in many laboratories.

The greatest ADA activity was found in lymphocytes and monocytes^[28]. ADA is the major isoenzyme originating from lymphocytes or neutrophils and ADA₂ is an isoenzymes unique to monocyte/macrophage cell lineage^[22,29] with the studies showing the ADA₂ is specific marker in the diagnosis of tuberculous pleural effusion than ADA^[22-24,26,27].

All the pleural biochemical factors are taken as a ratio in comparison with serum so as to negate any confounding factor affecting the marker; therefore we estimated the serum levels of isoenzymes.

The sensitivity and specificity values have varied widely with different cut off values adopted for isoenzyme parameters (in pleural fluid and serum). The standard deviation from mean in case of isoenzymes was similar as in case of the total ADA levels, but did not offer any advantage over that of the total ADA estimation.

ADA2 is the major contributor of the total ADA activity in tuberculous pleural effusions [22,29] and ADA, activity is unique to monocyte/macrophage lineage^[28]. In the study it was found that in some cases the contribution of ADA2 to the total activity of ADA in pleural fluid and serum as 100 %, with nil contribution by ADA, isoenzyme. Even though the correlation of ADA cell lineage is not probed in our study it poses the question - don't lymphocytes contribute in any way to the total ADA activity as seen in some cases, but ADA is raised in all lymphocytic pleural effusions. So what is the reason for the total absence of ADA₁ in few cases of tuberculous pleural effusion, which is predominant lymphocyte pleural effusion. The reason for this can be deduced from the explanations that have come out of the previous studies^[8,17]. The ADA₁ contribution in the tuberculous effusions may be related to the subsets of T lymphocytes and their maturity, and this may dictate the level of ADA_1 in tuberculous effusion, with the rest being contributed by ADA_2 from the monocyte/macrophage. It has also been shown in the previous studies that the ADA in the tuberculous effusion correlated to the monocyte/macrophage cell lineage^[25].

To conclude the study showed that ADA_2 is the predominant isoenzyme both in serum and pleural fluid, which is significantly raised in tuberculous pleural effusion. But the advantage in terms of sensitivity and specificity and in cost-benefit ratio in a developing country like India makes the estimation of ADA isoenzymes redundant and should not be done routinely.

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REFERENCES

- 1 Conway EJ. The deaminases of adenosine and adenylic acid in blood and tissues. *Biochem J* 1939; 33: 479-492.
- 2 Hirschhorn R, Ratech H. Isoenzymes of adenosine deaminase -Isoenzymes. Curr Top Biol Med Res 1980; 4: 131-157.
- 3 Buel E, Mac-Quarrie R. Purification of ADA of human RBC. Prep Biochem 1981; 11: 863-880.
- 4 Banales JL, Pineda PR, Fitzgerald JM, Rubio H, Sel man M, SalazarLezama M. ADA in diagnosis of TB effusion, a report of 218 patients and review of literature. *Chest* 1991; 99: 355-357.
- Martiz FJ. ADA estimation in differential diagnosis of pleural effusions. S Afr Med J 1982; 62: 556-558.
- Ocavna I, Martinez-Vazques JM, Sergera RM, Fernandez De Sevilla, Capdevila J. ADA in pleural fluids: test for diagnosis of Tuberculous pleural effusions. *Chest* 1983; 84: 51-53.
- 7 Bovornkitti S, Pushpakom R, Marnnetra N, Nana A, Charoenratanakul S. ADA and lymphocytic populations. *Chest* 1991; 99: 789-790.
- Baganha MF, Pego A, Lima MA, Gaspar EV, Cordeiro AR. Serum and pleural ADA: correlation with lymphocytic populations. Chest 1990; 97: 605-610.
- 9 **Gibbon M**, Labonte R, Laverack G. Evaluation community capacity. *Chest* 1991; 99: 1555-1556.

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- 10 Raj B, Vanlerberghe V, Perez D, Lefevre P, Ceballos E, Bandera D, et al. Achieving sustainability of communitybased dengue control in Santiago de Cuba. *Ind J Chest dis* and Alli Sci 1985; 27(2): 76-80.
- Rajendra-Prasad, Tripathi RP, Mukerji PK, Singh M, Srivastava VML. ADA activity in pleural fluid. *Ind J Chest dis and Alli Sci* 1992; 34(3): 123-126.
- 12 Gupta DK, Suri JC, Goel A. Efficacy of ADA in diagnosis of pleural effusions. *Ind J Chest Dis and Alli Sci* 1990; 32 (4): 205-208.
- 13 Maldhure BR, Bedarkar SP, Kulkarnt HR, Paplnwar SP. Pleural biopsy and ADA in pleural fluid for the diagnosis of TB pleural effusions. *Ind J Tub* 1994; 41: 161-165.
- 14 Nagaraja MV, Shokan PK, Hande HM. ADA in pleural effusions. *JAPI* 1992; 40(3): 157-158.
- Gilhotra R, Sehgal S, Jindal SK. Pleural biopsy and ADA enzyme in pleural effusions of different etiologies. *Lung India* 1989; 7(3): 122-124.
- 16 Gholani DR, Parikh FS, Hakim AS, Pai-Dhungat JV. Diagnostic significance of immunoglobulin and ADA in pleural effusions. JAPI 1999; 47: 787-790.
- 17 Ocana I, Martinez-vazquez JM, Ribera E, Sequra RM, Pascual C. ADA activity in the diagnosis of lymphocytic pleural effusions of TB, neoplastic and lymphomatous origin. *Tubercle* 1986; 97: 141-143.
- 18 Perez-Vidal R, Aran X. High ADA activity level in pleural effusion. Chest 1986; 90(4): 625.
- 19 Strankinga WFM, Nauta JJP, Straub JP, Stam J. ADA activity in TB pleural effusions, a diagnostic test. *Tubercle* 1987; 68: 137-140.
- 20 Sinha PK. Diagnosing TB pleural effusions, comparative sensitivity of Mycobacteruim Culture, Histopathology and ADA activity. *JAPI* 1985; 33(10): 644-645.
- 21 Hamada T, Sanaka M, Hata E, Hasegawa T. Pleural ADA levels in TB pleurisy- its diagnostic performance under the different prevalences in different age of population. *Japa*nese Journal of Thoracic and Cardiovascular Surgery 1998; 46(1): 51-57.

- 22 Ungerer JPJ, Oosthuizen HM, Retief JH, Bissbort SH. Significance of ADA activity and its isoenzymes in TB pleural effusions. Chest 1994; 106; 333-337.
- Valdes L, Jose E. San, Alvarez D, Valle JM. ADA isoenzyme analysis in pleural effusions, diagnostic role and relevance to the origin of increased ADA in TB pleurisy. *Euro Respi J* 1996; 9 (4): 747-751.
- 24 Valdes L, San-Jose E, Alvarez D, Valle JM. ADA isoenzymes ADA₁ and ADA₂ diagnostic and biological role. Southeast Asian J Trop Med Public Health 1992;23(2):200-206.
- 25 Perez-Rodriguez E, Perez-Walton IJ, Sanchez-Hernandez JJ, Pallares E, Rubi J, Jimenez-Castro D, et al. ADA₁/ADA_p ratio in pleural TB, an excellent diagnostic parameter in pleural fluid. Respiratory Medicine 1999; 93: 816-821.
- 26 Gorguner M, Cerci M, Gorguner I. Determination of adenosine deaminase activity and its isoenzymes for diagnosis of pleural effusions. *Respirology* 2000; 5(4): 321-324.
- 27 Shibagaki T, Hasegawa Y, Saito H, Yamori S, Shimokata K. Adenosine deaminase isoenzymes in tuberculous pleural effusions. *J Lab Clin Med* 1996; 127(4): 348-352.
- 28 Ungerer JPJ, Oosthuzer HM, Bissbort SH. Serum Adenosine Deaminase Isoenzymes and diagnostic application. *Clin Chem* 1992; 3(2): 1322-1326.
- 29 Ratech H, Hirschhorn R. Serum ADA in normals and a patient with ADA deficient severe combined immunodeficiency. Euro Respi J 1996; 9: 632-633.
- 30 Schrades WP, Pollara B, Meuwissen HJ. Characterization of residual ADA activity in spleen of patients with combined immuno deficiency disease and ADA deficiency. *Proc Natl Aca Sci USA* 1978; 75: 446-450.
- 31 Ratech H, Thorbecke GJ, Hirshhorn R. Comparision and possible homology of isoenzymes of adenosine deaminase in apes and humans. *Enzyme* 1981; 26: 74-84.
- 32 Giusti G, Galanti B. Colorometric method. In: Bergmeyer HU, ed. Methods of enzymatic analysis. 3rd ed. Weinheim: Verlag Chemie; 1984. 315-323.

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