Review article

Guinea pig model of tuberculosis

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Abstract

Animal models are being developed for testing different vaccine candidates as well as testing of new antitubercular since a long time. Mice, guinea pigs and rabbits are animals which are frequently used. Though each model has got its merits as well as demerits and each of them differ from human tuberculosis in one aspect or the other but none of the model completely mimics the human disease. Out of the different animal species, guinea pig model is one of the better models as it is very sensitive to *M. tuberculosis* infection but it has certain limitations like paucity of immunological reagents. However, it is the best model for tuberculosis research.

Keywords: Vaccines; Animal Models; Tuberculosis; Guinea pig

INTRODUCTION

In the 19th and 20th century, the guinea pig (Cavea porcellus) was the most popular experimental model for studying the infectious diseases like tuberculosis and diphtheria. By using guinea pig model, Robert Koch could develop the postulates of infectious disease etiology, essential for identifying the causative agent of infectious disease^[1,2]. He also used guinea pigs in his classical experiments establishing that Mycobacterium tuberculosis as the causative agent of tuberculosis. The recent taxonomical studies have changed its position from class rodents to non rodents^[3,4]. This is a better representative model of human infection than mouse^[5,6]. Sisk^[7] narrated number of similarities between humans and the guinea pigs which have direct or indirect bearing on the relevance of guinea pig as a model of infectious disease. The two species resemble with each other with regard to hormonal and immunological responses^[8-11], physiology of pulmonary tract^[12,13], corticoid resistance^[14], need for exogenous supply of Ascorbic acid^[15,16] and delayed type hypersensitivity (DTH) after exposure to infection^[11].

Tuberculosis is one of the most important bacterial disease that has been well characterized^[11, 17]. For creating this model, the animals are exposed to a small number of bacilli (10-50CFU) by aerosol route, a situation resembling human transmission. Extensive research conducted with this model indicates that the guinea pig is good model for primary human tuberculosis due to its extreme susceptibility to the infection, resembling symptoms and pathology, DTH response, good recovery after standard oral chemotherapies and excellent protection against tuberculosis (TB) after immunization with Bacille-Calmette Guerin (BCG) vaccine [11, 17-19]. Lymphadenitis is also common in guinea pigs like children infected with tuberculosis bacilli [20]. This model has also helped in understanding the effects of malnutrition which is a risk factor among the human populations. The protein malnourished guinea pigs infected with mycobacteria showed many immunological deficits like loss of protection following BCG vaccination^[21-24]. However, contrary to the humans, the liquefication and cavitation of pulmonary granulomas within infected lung tissues are rarely seen in guinea pigs^[11, 18] and the animals never show the latent form of the disease^[25,26]. During last 50 years, this model

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has facilitated in understanding the fundamental relationship between the TB bacilli and the human host. The early studies on pathogenesis of experimental tuberculosis in control and vaccinated guinea pigs showed that the previous exposure of animals to mycobacterial antigens led to delayed type of hypersensitivity resulting in lesser accumulation of the bacilli in the lung tissues^[27,28]. The interaction between tubercle bacilli and host phagocytic cells and potential role of the mycobacterial constituents as virulence factors have also been studied in guinea pig model only^[30,31].

Guinea pig model has been extensively used in testing the biological reagents as well as drugs against tuberculosis. Guinea pigs respond quite well against anti-TB antibiotics and hence widely used to test the new drugs or the drug combinations^[32,33]. With the increasing number of multi drug resistant strains of M. tuberculosis, this model will be of immense importance in search for new and efficient anti -mycobacterial drugs^[34]. PA-824, the most promising new drug candidate of recent time, was found to be efficacious in guinea pig model of low dose pulmonary infection^[35]. The tuberculosis infected guinea pig has been considered the gold standard for preclinical testing of the novel drugs and candidate vaccines^[36,37], various methods of delivery and the biological standardization of tuberculins for use in human skin testing^[38,39]. There is a need to develop improved drug regimens and preventive vaccines as the administration of present chemotherapeutic agents cause hepatotoxicity and poor patient compliance [40]. Some studies have also been conducted so as to evaluate the aerosolized administration of the anti-tubercular drugs rather than the oral route in guinea pig model^[4143]. The results from these studies indicated that this mode resulted in reducing the bacillary load in the lungs. BCG is presently the only vaccine available for prevention against tuberculosis. However it is highly controversial due to variable protective efficacy in different human populations^[44-46]. Hence this model has been tried to test various novel adjuvants and vaccine testing. Many type of vaccines have been tested in this model including non-viable bacilli [47] attenuated mycobacte- ${\rm ria}^{[48]}$, recombinant BCG strains $^{[49-53]}$, recombinant proteins /peptide vaccines $^{[54-55]}$, DNA vaccines [56-58], auxotrophs and gene-knockout mutants^[59-63] and other mycobacteria^[64]. The guinea pigs exhibit DTH response against these vaccines which is measured by skin induration test after intradermal injection^[11,65].

Besides the primary pulmonary tuberculosis, this model has helped in understanding the other forms of the disease like tuberculous pleuritis, exogenous re-infection and endogenous reactivation. Intraplural injection of either BCG^[66] or heat killed M. tuberculosis [67, 68] induced pleuritis in guinea pigs. Exogenous re-infection has also been studied in guinea pigs but the re-infection by pulmonary route with virulent M. tuberculosis of guinea pigs previously infected with non-tuberculous mycobacteria (NTM) or low virulent clinical isolates failed to exacerbate the disease like the humans [69] but showed protection^[70,71]. The re-infected animals responded better to the second challenge of virulent bacilli than those infected for the first time under uniform conditions. Further the protective effect of a prior pulmonary exposure to a low virulent isolate was greatly impaired by protein deficiency [72]. Guinea pig model of endogenous reactivation will help us in better understanding the factors associated with persistence of tuberculosis bacilli in tissues as well as the events that convert the dormant to active mycobacteria and allow their multiplication in large number. Though limited data is available on reactivation of tuberculosis in guinea pig model, Smith and Wiegeshaus^[73] has described the protocol for development of this model.

Besides the good model for testing the new candidate vaccines and new anti-tubercular drugs, this model has been helpful for elucidation of the immune response to infection and the basis for protective effects of the BCG. Number of scientists have reported that due to lack of the immunologic reagents for guinea pigs, adequate immunological studies can not in tuberculosis infected made pigs^[11,18,25,36,74]. Just to tackle this situation, numerous attempts have been made by the scientists to perform bioassays [75, 76], develop recombinant cytokines^[77-81] as well as develop antibodies^[80-82] and antiserum^[81] against these immune-mediators. Number of molecular techniques like Real time PCR for determination of cytokine and chemokine m-RNA levels[83-85], semi-quantitative PCR for RNA extraction^[86,87] and Southern and Northern blot analyses for studying gene expression^[88-90] and Microarray mRNA expression data^[91] have further helped in better understanding of the immunology of tuberculosis in M. tuberculosis infected guinea pigs .

Inspite of all the recent developments, the guinea pig models suffers from: (i) Lack of readily available immunological reagents required for qualitative and quantitative evaluation of immune responses; (ii) High cost involved in rearing guinea pigs under BSL-3 conditions; (iii) Requirement of good husbandry practices needed for rearing this species. However, the biological relevance of this species outweighs these limitations and efforts are needed to improve this model further.

REFERENCES

- Koch R. Aetiologie der Tuberculose. Berlin Klin Wochenschr. 1882; 19;221-0.
- Von Behring E, Untersuchungen UD. Zustanddekommen der Diphtherie-Immunitat bei Theiren. Dtsch Med Wochenschr. 1890; 16:1145, 288.
- 3 DErchia AM, Gissi C, Pesole G, Saccone C, Arnason U. The guinea pig is not a rodent. *Nature*. 1996; 381: 597-600.
- 4 Graur D, Hide WA, Li WH. Is the guinea pig a rodent? Nature. 1991; 351: 649-52.
- 5 Dascher CC, Hiromatsu K, Naylor JW, Brauer PP, Brown KA, Storey JR, et al. Conservation of a CD1 multigene family in the guinea pig. *J Immunol.* 1999; 163: 5478-88.
- 6 Hamuro J, Hadding U, Bitter-Suermann D. Fragments Ba and Bb derived from guinea pig factor B of the properdin system: purification, characterization, and biologic activities. J Immunol. 1978; 120: 438-44.
- 7 Sisk DB, Physiology. In: Wagner JE, Manning PJ, editors. The Biology of the Guinea Pig. New York: Academic Press, Inc. 1976; 63-98.
- 8 Ernstrom U. Hormonal influences on thymic release of lymphocytes into the blood. CIBA Found Study Group. 1970; 36: 53-65.
- 9 Hiromatsu K, Dascher CC, LeClair KP, Sugita M, Furlong ST, Brenner MB, et al. Induction of CD1-restricted immune responses in guinea pigs by immunization with mycobacterial lipid antigens. *J Immunol.* 2002; 169:330.
- Hiromatsu K, Dascher CC, Sugita M, Gingrich-Baker C, Behar SM, LeClair KP, et al. Characterization of guineapig group 1 CD1 proteins. *Immunology*. 2002; 106: 159-62.
- 11 McMurray DNGuinea pig model of tuberculosis. In: Bloom BR, editor. Tuberculosis: pathogenesis, protection, and control. Washington (DC): American Society for Microbiology; 1994.
- 12 Lechner AJ, Banchero N. Advanced pulmonary development in new born guinea pigs (Cavia porcellus). Am J Anat. 1982; 163: 235-6.
- 13 Meurs H, Santing RE, Remie R, van der Mark TW, Westerhof FJ, Zuidhof AB, et al. A guinea pig model of acute and chronic asthma using permanently instrumented

- and unrestrained animals. Nat Protocols. 2006; 1:840-7.
- 14 Claman HN. Corticosteroids and lymphoid cells. N Engl J Med. 1972; 287: 388-7.
- Collins FM, Elvehjem CA. Ascorbic acid requirement of the guinea pig using growth and tissue ascorbic acid concentrations as criteria. J Nutr. 1958; 64: 503-11.
- 16 Ganguly R, Durieux MF, Waldman RH. Macrophage function in vitamin C-deficient guinea pigs. Am J Clin Nutr. 1976; 29: 762-5.
- 17 McMurray DN, Collins FM, Dannenberg AM Jr, Smith DW. Pathogenesis of experimental tuberculosis in animal models. Curr Top Microbiol Immunol. 1996; 15: 157-9.
- 18 Orme IM. Current progress in tuberculosis vaccine development. Vaccine. 2005; 23: 2105-8.
- 19 Smith DW, McMurray DN, Wiegeshaus EH, Grover AA, Harding GE. Host-parasite relationships in experimental airborne tuberculosis. IV Early events in the course of infection in vaccinated and nonvaccinated guinea pigs. Am Rev Respir Dis. 1970; 102; 937-9.
- 20 Basaraba RJ, Dailey DD, McFarland CT, Shanley CA, Smith EE, McMurray DN, et al. Lymphadenitis as a major element of disease in the guinea pig model of tuberculosis. *Tuberculosis* (Edinb). 2006; 86:386-94.
- 21 McMurray DN, Kimbpall MS, Tetzlaff CL, Mintzer CL. Effects of protein deprivation and BCG vaccination on alveolar macrophage function in pulmonary tuberculosis. Am Rev Respir Dis. 1986; 133:1081-5.
- 22 McMurray DN, Carlomagno MA, Minttzer CL, Tetzlaff CL. Mycobacterium bovis BCG vaccine fails to protect protein deficient guinea pigs against respiratory challenge with virulent Mycobacterium tuberculosis. *Infect Immun.* 1985; 50: 555-9.
- 23 McMurray DN, Bartow RA, Mintzer CL. Protein malnutrition alters the distribution of FcgR⁺ (Tg) and FcMR⁺ (TM) T-lymphocytes in experimental pulmonary tuberculosis. *Infect Immun.* 1990; 58:563-5.
- 24 Cegielski JP, McMurray DN. The relationship between malnutrition and tuberculosis; evidence from studies in humans and experimental animals. Int J Tuberc Lung Dis. 2004; 8: 286-8.
- 25 McMurray DN. Disease model; pulmonary tuberculosis. Trends Mol Med. 2001; 7: 135-7.
- Orme IM. Preclinical testing of new vaccines for tuberculosis; a comprehensive review. *Vaccine*. 2006; 24: 2-19.
- 27 Palmer CE, Hopwood L. Effect of previous infection with unclassified mycobacteria on survival of guinea pigs challenged with virulent tubercle bacilli. *Bull Int Union Tuberc*. 1962; 32: 385-8.
- 28 Liebow AA, Burn CG, Soper WB. BCG immunization: A comparison of the effects of BCG and of heat-killed organisms on the course of a subsequent infection with virulent tubercle bacilli in the guinea pig. Am Rev Tuberc. 1940; 41: 592-4.
- Berthrong M, Hamilton MA. Tissue culture studies on resistance in tuberculosis. II. Monocytes from normal and immunized guinea pigs infected with virulent human tubercle bacilli. Am Rev Tuberc. 1959; 79; 221-1.
- 30 Middlebrook G, Dubos RJ, Pierce CH. Virulence and morphological characteristics of mammalian tubercle bacilli. J Exp Med. 1947; 86:175-4.

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- 31 Bloch H, Noll H. Studies on the virulence of tubercle bacilli. Variations in the virulence effects elicited by Tween 80 and thiosemicarbazone. Br J Exp Pathol. 1953; 97:1-16
- 32 Cohn ML, Davis CL, Middlebrook G. Chemoprophylaxis with isoniazid against aerogenic tuberculosis infection with the guinea pig. Am Rev Respir Dis. 1962; 86: 95-7.
- 33 Smith DM, Balasubramanian V, Wiegeshaus EH. A guinea pig model of experimental airborne tuberculosis for the evaluation of the response to chemotherapy: the effect on bacilli in the initial phase of treatment. *Tubercle*. 1991; 72 · 223-321.
- 34 Collins FM. Tuberculosis: the return of an old enemy. Crit Rev Microbiol. 1993; 19: 1-16.
- 35 Stover CK, Warrener P, VanDevanter DR, Sherman DR, Arain TM, Langhorne MH, et al. A small-molecule nitroimidazopyran drug candidate for the treatment of tuberculosis. *Nature*. 2000; 405: 962-6.
- 36 Gupta UD, Katoch VM, McMurray DN. Current status of TB vaccines. Vaccine. 2007; 25(19): 3742-1.
- 37 Ly LH, McMurray DN. Tuberculosis: vaccines in pipe lines. Expert Review Vaccines. 2008; 7: 635-0.
- Tolderund K, Bunch-Christensen K, Guld J. Duration of allergy and immunity in BCG-vaccinated guinea pigs. A five year study. Bull World Health Organ. 1967; 36: 759-69
- 39 Ladefoged A, Bunch-Christensen K, Guld J. Tuberculin sensitivity in guinea-pigs after vaccination with varying doses of BCG of 12 different strains. Bull World Health Organ. 1976; 53: 435-43.
- 40 OBrien RJ. The treatment of tuberculosis. In: Reichmann LB, Hershfield ES, editors. Tuberculosis: a comprehensive international approach. New York: Marcel Dekker. 1993. 207-40.
- 41 Garcia-Contreras L, Sethuraman V, Kazantseva M, Godfrey V, Hickey AJ. Evaluation of dosing regimen of respirable rifampicin biodegradable microspheres in the treatment of tuberculosis in the guinea pig. *J Antimicrob Chemother*. 2006; 58: 980-6.
- 42 Suarez S, O'Hara P, Kazantseva M, Newcomer CE, Hopfer R, McMurray DN, et al. Airways delivery of rifampicin microparticles for the treatment of tuberculosis. J Antimicrob Chemother. 2001; 48: 431-4.
- 43 Suarez S, O'Hara P, Kazantseva M, Newcomer CE, Hopfer R, McMurray DN, et al. Respirable PLGA microspheres containing rifampicin for the treatment of tuberculosis: screening in an infectious disease model. *Pharm Res.* 2001; 18: 1315-9.
- 44 Fine PEM. Variation in protection by BCG; implications of and for heterologous immunity. Lancet. 1995; 346:1339-
- 45 Brewer TF. Preventing tuberculosis with Bacillus Calmette Guerin vaccine; a meta-analysis of the literature. Clin Infect Dis. 2000; 31 (Suppl. 3); S64-7.
- 46 Roche PW, Triccas JA, Winter N. BCG vaccination against tuberculosis: past disappointments and future hopes. *Trends Microbiol.* 1995; 3: 397-1.
- 47 Fregnan GB, Smith DB. Immunogenicity and allergenicity in guinea pigs of a defatted mycobacterial vaccine and its

- fractions. Am. Rev. Respir. Dis. 1963; 87: 877-8.
- 48 Smith DW, Wiegehaus E, Harding GE. Models for potency assays of tuberculosis vaccines. Fogarty Int Cent Proc. 1972; 14: 205-8.
- Wiegeshaus EH, Smith DW. Evaluation of the protective potency of new tuberculosis vaccines. Rev Infect Dis. 1989; 11: S484-90.
- 50 Friedag BL, Melton GB, Collins F, Klinman DM, Cheever A, Stobie L, et al. CpG oligodeoxynucleotides and interleukin-12 improve the efficacy Mycobacterium bovis BCG vaccinationin mice challenged with M. tuberculosis. *Infect Immun.* 2000; 68: 2948-3.
- 51 Lyadova IV, Vordermeier HM, Eruslanov EB, Khaidukov SV, Apt AS. Intranasal BCG vaccination protects BALB/c mice against virulent Mycobacterium bovis and accelerates production of IFN-g in their lungs. Clin Exp Immunol. 2001; 126: 274-9.
- 52 Olsen AW, Van Pinxteren LAH, Okkels LM, Rasmussen PB, Andersen P. Protection of mice with a tuberculosis subunit vaccine based on a fusion protein of antigen 85B and ESAT- 6. *Infect Imun.* 2001; 69: 2773-8.
- 53 Doderty TM, Olsen AW, van Pinxteren L, Andersen P. Oral vaccination with subunit vaccines protects animals against aerosol infection with Mycobacterium tuberculosis. Infect Immun. 2002; 70: 3111-1.
- 54 Coler RN, Campos-Neto A, Ovendale P, Day FH, Fling SP, Zhu L, et al. Vaccination with the T cell antigen Mtb 8.4 protects against challenge with Mycobacterium tuberculosis. J Immunol. 2001; 166: 6227-5.
- 55 Huygen K, Content J, Denis O, Montgomery DL, Yawman AM, Deck RR, et al. Immunogenicity and protective efficacy of a tuberculosis TB DNA vaccine. *Nat Med*. 1996; 2: 893-8.
- 56 Delogu G, Li A, Repique C, Collins F, Morris SI. DNA vaccine combinations expressing either tissue plasminogen activator signal sequence fusion proteins or ubiquitin-conjugated antigens induce sustained protective immunity in a mouse model of pulmonary tuberculosis. *Infect Immun*. 2002; 70: 292-2.
- 57 Sugawara I, Yamada H, Udagawa T, Huygen K. Vaccination of guinea pigs with DNA encoding Ag85A by gene gun bombardment. *Tuberculosis* (*Edinb*). 2003; 83: 331-7.
- 58 Guleria I, Teitelbaum RA, Adam NC, Kalpana G, Jacobs Jr WR, Bloom BR. Auxotrophic vaccines for tuberculosis. *Nat Med.* 1996; 2(3): 334-7.
- 59 Jackson M, Phalen SW, Langranderie M, Ensergueix D, Chvarot P, Marchal G, et al. Persistant and protective efficacy of a Mycobacterium tuberculosis auxotroph vaccine. *Infect Immun.* 1999; 67: 2867-3.
- 60 Smith DA, Parish T, Stroker NG, Bancroft GJ. Characterization of auxotrophic mutants of Mycobacterium tuberculosis and their potential as vaccine candidates. *Infect Immun*. 2001; 69:1142-50.
- Parish T, Stoker NG. Use of a flexible cassette method to generate a double unmarked Mycobacterium tuberculosis tiyA plcABC mutant by gene replacement. *Microbiol*. 2000; 146:1969-75.
- 62 Sampson SL, Dascher CC, Sambandamurthy VK, Russell RG, Jacobs Jr WR, Bloom BR, et al. Protection elicited by

- a double leucine and pantothenate auxotroph of Mycobacterium tuberculosis in guinea pigs. *Infect Immun* . 2004; 72: 3031-7.
- 63 Sambandamurthy VK, Jacobs Jr WR. Live attenuated mutants of Mycobacterium tuberculosis as candidate vaccines against tuberculosis. 2005; 7: 955-61.
- 64 Gupta A, Mol N, Mani J, Upadhyay P, Katoch VM, Natrajan N, Gupta UD, et al. Immunogenicity and protective efficacy of M. w against M. tuberculosis in mice immunized with live M. w by aerosol or parenteral route and compared with killed M. w. Infect. Immun. (Accepted)
- 65 Gell PG, Benacerraf B. Studies on hypersensitivity. IV. The relationship between contact and delayed sensitivity: a study of the specificity of cellular immune reactions. J Exp Med. 1961; 113: 571-5.
- 66 Winstrom EH, Nilsson BS. Pleurisy induced by intrapleural BCG in immunized guinea pigs. Eur J Respir Dis. 1982; 63: 425-4.
- 67 Phalen SW, McMurray DN. T-lymphocyte response in a guinea pig model of tuberculosis pleuritis. *Infect Immun*. 1993; 61: 142-5.
- 68 Phalen SW, McMurray DN. Production of tumor necrosis factor (TNFa) in experimental tuberculous pleuritis. *J Immunol.* 1993; 150: 66A.
- 69 Nardell E, McInnis B, Thomas B, Weidhaas S, Exogenous reinfection with tuberculosis in a shelter for the homeless. N Engl J Med. 1986; 315;1570-5.
- 70 Ziegler JE, Edwards ML, Smith DW. Exogenous re-infection in experimental airborne tuberculosis. *Tubercle*. 1985; 66: 121-8.
- 71 Edwards ML, Goodrich JM, Muller D, Pollack A, Ziegler JE, Smith DW. Infection with Mycobacterium avium-intracellulare and the protective effects of Bacilli Calmette-Guerin. J Infect Dis. 1982;145: 733-1.
- 72 McMurray DN, Bartow RA, Mintzer CL. Impact of protein malnutrition on exogenous re-infection with Mycobacterium tuberculosis. *Infect Immun.* 1989; 57: 1746-9.
- 73 **Smith DW**, Wiegeshaus EH. What animal models can teach us about the pathogenesis of tuberculosis in humans. *Rev Infect Dis.* 1989; 11 (Suppl 2): S385-3.
- 74 Orme IM, McMurray DN, Belisle JT. Tuberculosis vaccine development: recent progress. *Trends Microbiol.* 2001; 9: 115-8.
- 75 Espevik T, Nissen-Meyer J. A highly sensitive cell line, WEHI 164 clone 13, for measuring cytotoxic factor/tumor necrosis factor from human monocytes. *J Immunol Meth*ods. 1986; 95: 99-5.
- 76 Yamamoto T, Jeevan A, Ohishi K, Nojima Y, Umemori K, Yamamoto S, et al. A new assay system for guinea pig interferon biological activity. J Interferon Cytokine Res. 2002; 22:793-7.
- 77 Cho H, Lasco TM, Allen SS, Yoshimura T, McMurray DN. Recombinant guinea pig tumor necrosis factor alpha stimulates the expression of interleukin 12 and the inhibition of Mycobacterium tuberculosis growth in macrophages. *Infect Immun.* 2005; 73:1367-6.
- 78 Cho H, McMurray DN. Recombinant guinea pig TNFα enhances antigen-specific type 1 T lymphocyte activation in

- guinea pig splenocytes. Tuberculosis (Edinb). 2007; 7: 87-3.
- 79 Jeevan A, McFarland CT, Yoshimura T, Skwor T, Cho H, Lasco T, et al. Production and characterization of guinea pig recombinant gamma interferon and its effect on macrophage activation. *Infect Immun.* 2006; 4: 213-4.
- 80 Lasco TM, Cassone L, Kamohara H, Yoshimura T, Mc-Murray DN. Evaluating the role of tumor necrosis factor α in experimental pulmonary tuberculosis in the guinea pig. *Tuberculosis* (Edinb). 2005; 85: 245-8.
- 81 Lyons MJ, Yoshimura T, McMurray DN. Interleukin (IL) 8 (CXCL8) induces cytokine expression and superoxide formation by guinea pig neutrophils infected with Mycobacterium tuberculosis. *Tuberculosis* (*Edinb*). 2004; 84: 283-2.
- 82 Xiong X, Morita CT, Bukowski JF, Brenner MB, Dascher CC. Identification of guinea pig γδ T cells and characterization during pulmonary tuberculosis. Vet Immunol Immunopathol. 2004; 102: 33-4.
- 83 Allen SS, McMurray DN. Coordinate cytokine gene expression in vivo following induction of tuberculous pleurisy in guinea pigs. *Infect Immun.* 2003; 71: 4271-7.
- 84 Kawahara M, Nakasone T, Honda M. Dynamics of gamma interferon, interleukin-12 (IL-12), IL-10, and transforming growth factor beta mRNA expression in primary Mycobacterium bovis BCG infection in guinea pigs measured by a real-time fluorogenic reverse transcription-PCR assay. *Infect Immun.* 2002;70: 6614-20.
- 85 **Yamada H**, Udagawa T, Mizuno S, Hiramatsu K, Sugawara I. Newly designed primer sets available for evaluating various cytokines and iNOS mRNA expression in guinea pig lung tissues by RT-PCR. *Exp Anim.* 2005; 54: 163-2.
- 86 Klunner T, Bartels T, Vordermeier M, Burger R, Schafer H. Immune reactions of CD4- and CD8-positive T cell subpopulations in spleen and lymph nodes of guinea pigs after vaccination with Bacillus Calmette Guerin. Vaccine. 2001; 19: 1968-77.
- 87 Ohtani M, Kobayashi Y, Watanabe N. Gene expression in the elicitation phase of guinea pig DTH and CHS reactions. Cytokine. 2004; 25: 246-3.
- 88 Dascher CC, Hiromatsu K, Naylor JW, Brauer PP, Brown KA, Storey JR, et al. Conservation of a CD1 multigene family in the guinea pig. *J Immunol.* 1999; 163: 5478-8.
- Jeevan A, Yoshimura T, Foster G, McMurray DN. Effect of Mycobacterium bovis BCG vaccination on interleukin 1β and RANTES mRNA expression in guinea pig cells exposed to attenuated and virulent mycobacteria. *Infect Immun*. 2002; 70: 1245-3.
- Jeevan A, Yoshimura T, Lee KE, McMurray DN. Differential expression of gamma interferon mRNA induced by attenuated and virulent Mycobacterium tuberculosis in guinea pig cells after Mycobacterium bovis BCG vaccination. *Infect Immun.* 2003; 71: 354-4.
- 91 **Tree JA**, Elmore MJ, Javed S, Williams A, Marsh PD. Development of a guinea pig immune response-related microarray and its use to define the host response following Mycobacterium bovis BCG vaccination. *Infect Immun*. 2006; 74: 1436-1.