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Retention of testicular integrity and testosterone levels upon ingestion of garlic cloves (*Allium sativum*) in the Sprague-Dawley rat

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ABSTRACT

Objective: To investigate the effects of acute and chronic aqueous garlic extract ingestion on testicular cellular integrity and serum testosterone levels.

Methods: Twenty (20) male Sprague-Dawley rats weighing an average of 120 g were used. Animals were divided into three groups. Group A served as control (10 rats for 28 and 56 d respectively), while treatment Groups B and C were given 200 mg/kg for *Allium sativum* (garlic cloves) extract for 28 and 56 d respectively.

Results: Histological analysis revealed the presence of all spermatogenic lineages, appearance of proliferative activities in the interstitial cells, as well as increased serum testosterone levels. **Conclusions:** This study confirmed proliferative and restorative potentials in both acute and chronic garlic ingestion.

1. Introduction

The use of medicinal plants for medicinal remedies has been documented since ancient times and they provide a useful source of new therapeutics[1,2].

Garlic belongs to the family Liliaceae. Its leaves collections (bulb), which can be divided into cloves, are used for culinary and medicinal purposes. It has a characteristic pungent and spicy flavor[3]. Garlic is made up of the important bioactive components such as allicin (the active compound responsible for its hot sensation), ajoene, enzymes, water, vitamin B, minerals and flavonoids[4].

Garlic has been reported to be capable of human blood pressure regulation[5,6], glucose level regulation, platelet aggregation[7], hyperlipidemia, and the reduction of atherosclerosis building up in the arterial system[8,9]. Also, it has been reported to act as an anti-viral, anti-bacterial and anti-candida agent, and it has been implicated in cancer prevention due to the presence of diallyl disulfide in its composition[10,11]. Furthermore, it is said to reduce hypercholesterolemia[12] and to boost testosterone levels[13].

However, the reported side effects of excessive garlic consumption

have been found to include halitosis (non-bacterial), nausea, emesis, diarrhoea/gastrointestinal discomfort, bleeding, sweating, *etc*. Also, the prolonged feeding of high doses of raw garlic to experimental rats will result in anaemia, weight loss and stunted growth due to red blood cells lysis[14].

United States Department of Agriculture nutrient database states that 100 g of raw garlic contains carbohydrates, dietary fibre, fat, protein, β-carotene, thiamin (Vit B1), riboflavin (Vit B2), niacin (Vit B3), pantothenic acid (Vit B5), adermin (Vit B6), folate (Vit B9), vitamin C, calcium, iron, magnesium, phosphorus, potassium, sodium, zinc, manganese and selenium[4].

While some reports show that heated garlic juice is effective in the recovery of testicular functions after experimental testicular hypogonadism[3,15,16], other studies have reported that powdered garlic preparations impair testicular and male reproductive functions[17,18]. Garlic metabolites such as diallyl trisulfide have been reported to have spermicidal effects[19]. Chronic administration of garlic powder (50 mg/d) was said to result in the inhibition of spermatogenesis in rats[17], and a study showed that 10%, 15% and 20% of the crude extracts respectively decreased serum testosterone levels[20].

The contrasting findings in the mechanisms of garlic actions on testosterone activities and testicular integrity create the need for further research[1,18,21,22]. Hence, this study was designed to evaluate the acute and chronic effects of 200 mg/kg of aqueous garlic extract

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(AGE) on the testicular weight, testosterone level and testicular cytoarchitecture.

2. Materials and methods

2.1. Experimental animals

Twenty (20) male Sprague-Dawley rats with average age of 2 months and average weight of 120 g were used for this study. Animals were kept in well ventilated cages in the animal holding of the Department of Anatomy, University of Lagos, Nigeria, under standard laboratory conditions [12 h light/dark cycle (lights on: 8:00 am), temperature at 29 °C, humidity at 50%] and given pelleted rat feed (UAC, Vital Feeds Lagos, Nigeria) and water *ad libitum*. They were cared for in accordance to the ethical regulations in the Guide for the Care and Use of Laboratory Animals[23]. The animals were allowed to acclimatize for 2 weeks before experimentation.

2.2. Plant material

Fresh garlic [*Allium sativum* (*A. sativum*)] bulbs were procured from Lawanson market, Surulere, Lagos, Nigeria. They were authenticated in Botany Department in University of Lagos, Lagos, Nigeria, where voucher specimen was deposited (Figure 1A).

2.2.1. Preparation of the aqueous extracts of each spice

A. sativum (garlic cloves) were separately sun-dried and crushed into powdered by using a blender (Figure 1B). The dried powder was weighed (10 g) and macerated in 50 mL of distilled water. Prepared extract was stored in a refrigerator and each animal was given the aqueous extract of A. sativum via gastric intubation using an oral cannula.





Figure 1. Representative image of *A. sativum* in raw form (A) and in its dried grinded (B) used in this study[24].

2.3. Experimental protocol

Rats were divided into four groups. Group A (control group) received feed pellets and water *ad libtum*. Group B received 200 mg/kg of the extract for 4 weeks (28 d). Group C received 200 mg/kg of the extract for 8 weeks (56 d). Administration was done once daily between 08:00–09:00 am.

2.4. Euthanasia of experimental animals

Experimental animals were euthanized 24 h after the last administration and final weights were recorded. Euthanization was done by giving an intraperitoneal injection of 50 mg/L of ketamine (Claris Lifesciences Ltd., India).

2.5. Histology

Incision was made in the lower abdominal wall and the testes were excised. Organ weights were taken using an analytical weighing scale (Haerus, Germany) after which organs were stored in Bouin's fluid for fixation. One half of the longitudinal sectioned testes was processed for histological studies, embedded in paraffin wax and serially sectioned using a Leica rotary microtome (Leica biosystems, Germany) set at 5 μ . Testicular tissue sections were stained using haematoxylin and eosin stain for general cytoarchitectural examination.

2.6. Statistical analysis

Data were analyzed using SPSS statistical software and Microsoft Excel. The statistical analysis was carried using repeated measures for weight changes and One-way analysis of variance for testicular weight and serum testosterone levels. Fisher's least significant difference was used for multiple mean comparison using statistica software. Data were expressed as mean±SEM. *P*<0.05 was considered significant.

3. Results

3.1. Body weight changes

An overall increase in final body weight was seen in both control and experimental groups at the end of the experiment (P<0.05) in both 28 and 56 d when compared with their respective initial body weights (Figure 2). However, at the end of the experiment, there was no statistically significant (P>0.05) final body weight change in Group B when compared with the control group. In contrast, there was a significant reduction in final body weight (P<0.0001) in Group C when compared with the control group.

3.2. Testicular weight changes

An overall increase was shown in the testicular weights in all experimental groups when compared with the control group (Figure 3). Group B animals showed statistically significant testicular weight increase when compared with the control group (P<0.05), likewise Group C animals showed statistically significant testicular weight increase (P<0.05) when compared with the control group animals. Noteworthy, Group C animals had statistically significant testicular weight increase (P<0.01) when compared with Group B animals (Figure 3).

3.3. Serum testosterone levels

Serum testosterone level was increased in the experimental Groups B and C in comparison with the control group (Figure 4). There was a statistically significant overall increase in testosterone levels in all experimental groups compared with control group (P<0.05). Indeed, *post hoc* analysis showed a statistically significant increase in testosterone levels (P<0.05) in Group B when compared with the

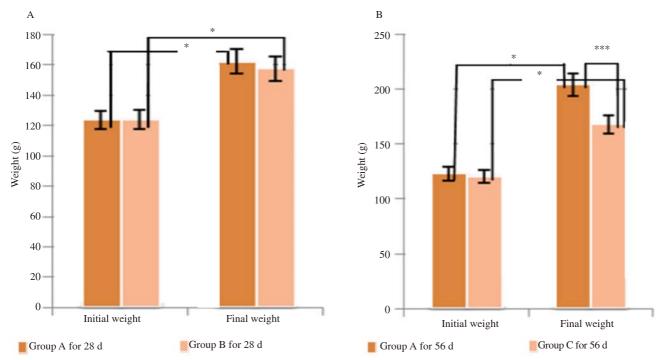


Figure 2. Mean values of initial and final body weight measurements of the experimental animals following 28 (A) and 56 (B) d of AGE treatments respectively.

*P<0.05 represents statistically significant increase in final body weights in both control (Group A) and experimental groups (B,C) in comparison with the initial body. ****P<0.001 represents statistically significant decrease in final body weight of Group C animals in comparison with the control group.

control group, likewise, there was a statistically significant increase in Group C when compared with the control group animals (P<0.05). Moreover, there was a significant increase in serum levels (P<0.01) in the Group C animals when compared with Group B animals.

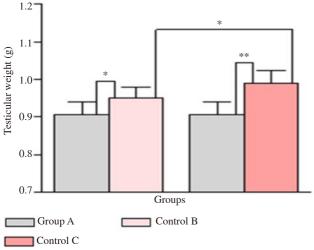


Figure 3. Mean values for testicular weights of experimental animals treated with AGE for 28 and 56 d respectively.

*P<0.05 represents significant increase in testicular weight after 28 d treatment with AGE in comparison with control group. **P<0.01 represents significant increase in testicular weight after 56 d treatment with AGE in comparison with control group. A statistically significant testicular weight increase was shown in Group C compared with Group B animals (*P<0.05).

3.4. Histological results

In correlation with serum testosterone levels, there was appearance of morphological changes in the experimental groups, an indication of proliferative activities taking place in these treatment groups (Figures 5B and 5D) when compared with the control groups (Figures 5A and 5C). The testicular cytoarchitecture of all the groups showed relatively distinct seminiferous tubules bounded together by loose interlobular connective tissue and groups of interstitial cells or leydig cells. In the experimental Group B (Figure 5B), though lumen appeared slightly wider, all cells spermatogenic lineage appeared present, also leydig cells appeared intact. Representative image of Group C animals treated for 56 d showed intact testicular histoarchitecture, with the appearance of all cells of spermatogenic lineage in the germinal epithelium. Notably, the appearance of spermatids indicated spermatogenic processes at the advanced stages of the spermatogenic cycles (Figure 5D).

4. Discussion

This study demonstrates the supportive role of garlic in body weight regulation and maintenance of testicular integrity and function.

Garlic supplementation has been implicated in body weight regulation [10,25]. This study supports the role of garlic in body weight regulation during both acute and chronic garlic intake, thus, even though the final body weight of both control and experimental animals increased when compared with their initial body weights, there was a dose dependent response in their final body weights, and this was evident by the lack of significant changes in final body weights during acute garlic ingestion when compared with the control animals and the profound decrease in final body weight during chronic garlic ingestion. The regulatory role of garlic in body weight regulation is supported by previous findings[26-28]. Furthermore, this study emphasizes the dose dependent response of garlic intake on weight regulation.

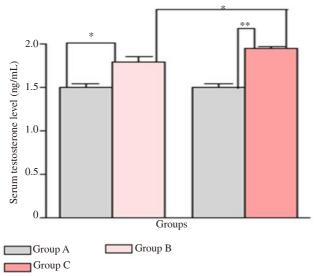


Figure 4. Mean serum testosterone levels for the AGE treated rats in comparison control group for 28 and 56 d respectively.

*P<0.05 represents significant increase in serum testosterone after 28 d treatment with AGE in comparison with control group. **P<0.01 represents significant increase in serum testosterone after 56 d treatment with AGE in comparison with control group. A statistically significant serum testosterone increase was shown in Group C compared with Group B animals (*P<0.05).

Garlic oil and the diallyl disulfide components in garlic have been implicated in weight gain regulation[25]. It has been proposed that garlic regulates body weight due to its ability to decrease lipid in adipose tissues via activation of increased levels of uncoupling proteins 1 and 2 required for fat oxidation thereby causing a decreased weight gain[29]. Another report claims that garlic possesses antiobesity properties due to its ability to mediate activation of AMP kinase which causes increased thermogenesis and decreased expression of the multiple genes involved in adipogenesis by a decrease in mRNA level in adipogenic genes in white adipose tissue[10].

This study also confirms an increase testicular weight of the animals exposed to acute and chronic garlic ingestion. Notably, a more profound increase in testicular weight was found during chronic garlic administration. The major components of garlic are zinc and selenium. These substances have been reported to play regulatory roles in the testicular activities especially steroidogenic enzymes production[4,30].

The testes are involved in spermatogenesis, a process in which spermatogenic cells are formed; testosterone (androgen) deficiency negatively affects spermatogenesis[31] by altering spermatid-Sertoli cell junctions, thus resulting in the premature detachment of round spermatids from Sertoli cells and spermatogenic epithelium[28,32]. In this study, garlic was found to play a proliferative role in testosterone secretion such that an increase in testosterone secretion is shown in both acute and chronic garlic intake. Moreover, a dose dependent activity was found such that chronic garlic intake corresponded with higher serum testosterone levels. This is contrary to some findings that report crude garlic extracts causing reduction in serum testosterone levels. The disparity in results obtained may be due to the use of varied dose of administration as well as varied duration of garlic exposure[17,20,33,34].

Testosterone plays an important role in the spermatogenic cell proliferation. The regulatory role of garlic in testosterone secretion

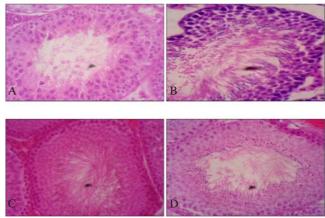


Figure 5. Representative images of haematoxylin and eosin stained testicular sections (×400).

Control sections A, B showed appearance of all spermatogenic cells in the seminiferous tubule, with intact leydig cells and clearly distinct lumen and basement membrane; C, D images represented Groups B and C given AGE for 28 and 56 d respectively. Cells have similar cytoarchitectural pattern as the control group.

was further confirmed in the histological analysis for both the control and experimental groups such that there was the appearance of intact spermatogenic cells, Leydig cells, well aligned lumen and clearly defined cellular membranes. This contradicts some findings that report degenerative changes in the testes following the administration of 1.4 and 2.4 mL of 100 mg/L garlic extract to rats[18,34], but supports some other findings that report garlic as having protective and proliferative functions in the testes[35-37]. Further studies are required to confirm the molecular mechanisms mediating these supportive and regulatory roles of garlic in testicular function.

The data from this present study confirm the regulatory role of garlic in regulating body weight, organ weight, serum testosterone levels and testicular integrity.

Conflict of interest statement

We declare that we have no conflict of interest.

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