



DOI Number: 10.5958/2277-940X.2015.00006.6

## Seroprevalence of Leptospirosis in Clinically Ailing Bovine

Jignesh M. Patel<sup>1\*</sup>, Priti D. Vihol<sup>1</sup>, Jeetendra K. Raval<sup>1</sup>, Kartik M. Patel<sup>2</sup>, Naresh F. Chaudhari<sup>3</sup>, Puspa H. Rathod<sup>4</sup> and Jatin H. Patel<sup>5</sup>

<sup>1</sup>Department of Veterinary Pathology, Vanbandhu College of Veterinary Sciences & A. H. Navasari Agricultural University, Navsari, Gujarat, INDIA

<sup>3</sup>Department of Veterinary Gynaecology and Obstetrics, Vanbandhu College of Veterinary Sciences & A. H., Navasari Agricultural University, Navsari, Gujarat, INDIA

<sup>4</sup>Department of Teaching Veterinary Clinical Complex, Vanbandhu College of Veterinary Sciences & A. H., Navasari Agricultural University, Navsari, Gujarat, INDIA

<sup>5</sup>Department of Veterinary Pharmacology and Toxicology, Vanbandhu College of Veterinary Sciences & A. H. Navasari Agricultural University, Navsari, Gujarat, INDIA

<sup>2</sup>State Department of Animal Husbandry, Patan District, Gujarat, INDIA

\* Corresponding author: JM Patel; Email: dr.jams@rediffmail.com

Received: 19 January, 2015

Accepted: 23 March, 2015

### ABSTRACT

The purpose of this study was to carried out seroepidemiological study of leptospirosis in clinically ailing cattle and buffaloes of South Gujarat where cases of leptospirosis in human is increase every monsoon since last decade. To determine clinical pattern of bovine Leptospirosis in this area a total of 130 serum samples of cattle and buffaloes were collected randomly from different age groups of cattle and buffaloes of either sex reared in this area and tested by microscopic agglutination test (MAT) using different serovars of *Leptospira* spp. The prevalence of leptospiral antibodies was detected in 15.84 and 17.24 % in clinically ailing cattle and buffaloes, respectively. In clinically ailing seropositive cattle history of mastitis/agalactia/oligolactia was recorded in maximum number of cases (24.00 %) followed by abortion (20.00 %), fever (14.81%), repeat breeding (11.76%) and anorexia (5.88%) in different combinations. In seropositive clinically ailing buffaloes the clinical signs included mastitis (30.00%), fever (16.66 %) and abortion (14.28%). In clinically ailing cattle highest prevalence was noted in animals above 4 years of age (20.00%) followed by 1-4 years of age (4.76%) whereas in clinically ailing buffalo seropositivity was recorded only in above 4 years of age (20.00%). The most prevalent serovar pomona has been reported from different clinical conditions in both cattle and buffaloes. Based on present and few past investigation reports the sero-epidemiological condition of bovine leptospirosis that need due attention as South Gujarat region is endemic zone for leptospirosis.

**Keywords:** Bovine, Clinical cases, Leptospirosis, MAT, Seroprevalence

Leptospirosis is an economically important worldwide zoonotic bacterial infection of livestock caused by pathogenic spirochetes *Leptospira*. The disease causes a wide spectrum of clinical manifestations in animals and is responsible for economical losses due its reproductive impacts like mastitis, repeat breeding, early embryonic death and abortion (Vinetz, 2001). The clinical signs associated with bovine leptospirosis are variable and depend upon the infecting serovar as well as the susceptibility of the individual animals and so its presentation may varies from

subclinical infection to a severe illness (Adler and de la Pena Moctezuma, 2010). Clinically bovine Leptospirosis is difficult to diagnose because the expressions are non-specific and may be easily confused with other infectious diseases (Ellis, 1984). So to know the spectrum of clinical sign of bovine leptospirosis along with infecting serovars as well as keeping in view the sporadic reports of leptospirosis in human (Shivakumar, 2008) and in animal (Balakrishnan *et al.*, 2011) from South Gujarat region the present seroprevalence study of leptospirosis has been carried out.

**Table 1.** Prevalence of leptospirosis in clinically ailing cattle (N=101) and buffaloes (N=29).

Particulars	Cattle		Buffaloes	
	Total cases	Seropositive cases	Total cases	Seropositive cases
A. Overall prevalence				
Total Clinically ailing animals	101	16 (15.84%)	29	5 (17.24%)
Mastitis/Agalactia/ Oligolactia	25	6 (24.00%)	10	3 (30.00%)
Abortion	15	3(20.00%)	7	1(14.28%)
Repet breeder	17	2(11.76%)	2	0 (00%)
Fever	27	4(14.81%)	6	1(16.66%)
Anorexia	17	1(5.88%)	4	0(00.00%)
	$\chi^2 = 9.49^{NS}(P<0.05)$		$\chi^2 = 9.49^{NS}(P<0.05)$	
B. Age wise prevalence in clinically ailing animals				
<1 year	5	0 (00%)	1	0(00%)
1-4 years	21	1(4.76%)	3	0(00%)
>4 years	75	15(20.00%)	25	5 (20.00%)
	$\chi^2 = 5.99^{NS}(P<0.05)$		$\chi^2 = 5.99^{NS}(P<0.05)$	
C. Sex wise prevalence in clinically ailing animals				
Male	5	1(20.00%)	1	0(00.00 %)
Female	96	15(15.63%)	28	5(17.85%)
	$\chi^2 = 3.84^{NS}(P<0.05)$		$\chi^2 = 3.84^{NS}(P<0.05)$	

**Note:** <sup>NS</sup>-Non significant at P < 0.05

## MATERIALS AND METHODS

### Collection of blood and serum samples

A total of 130 blood/serum samples were collected randomly from clinically ailing (cattle= 101; buffaloes=29) bovines reared in villages of various districts (Navsari, Surat, Tapi, Valsad) of South Gujarat having history of anorexia, fever, abortion, repeat breeding and mastitis. The age and sex of individual animals were also recorded during sample collection.

Whole blood samples were collected from jugular vein directly in cattle and or during slaughter in some of the buffaloes in sterile 9.0 ml plain vacutainers. To obtain serum whole blood was kept in slanting position in 9.0 ml plain vacutainers until serum oozed out from clotted blood. The 9.0 ml plain vacutainers were centrifuged at 7000 rpm for 10 minutes. The straw coloured serum was collected into of 1.5 ml sterile cryovials and stored at - 20°C for carrying out Microscopic Agglutination Test (MAT).

### Microscopic Agglutination Test (MAT)

All the sera collected were tested for antibodies against live antigens of *Leptospira* sp. (serovars Pyrogenes, Australis, Bankinang, Grippytyphosa, Patoc, Pomona, Icterohaemorrhagiae, Hebdomadis, Canicola, Hardjo, Bellum, Bataviae, Tarassovi, Shermani, Kaup, Hurstbridge and Javanica) following MAT at Leptospirosis Reference Laboratory, Government Medical College, Surat (Vijayachari *et al.*, 2001) and Project Directorate on Animal Disease Monitoring and Surveillance (PD-ADMAS), Bengaluru using standard procedure (WHO-OIE, 2013).

### Warthin-Starry and H&E Staining Method

Tissues were preserved in 10 per cent neutral buffered formalin for at least 24-48 hours for Warthin-starry special staining technique for demonstration of leptospires in microsections (5 μ). Briefly, the steps followed were (i) Deparaffinize and hydrate the tissue section in distilled

**Table 2.** Distribution of *Leptospira* serovars among clinically ailing cattle and buffaloes.

Different Serovars of leptospira	Clinical expressions										
	Mastitis		Abortion		Repeat breeder		Fever		Anorexia		
	C	B	C	B	C	B	C	B	C	B	
Pyrogen	1	-	-	-	-	-	-	-	-	-	-
Australis	1	-	-	-	1	-	-	-	-	-	-
Autumnalis/Bankinang	-	-	-	-	-	-	1	-	-	-	-
Grippotyphosa	-	-	-	-	-	-	-	-	-	-	-
Patoc	-	-	-	-	2	-	-	-	-	-	-
Pomona	3	1	2	1	2	-	4	1	-	-	-
Icterohaemorrhagiae	1	-	1	-	-	-	2	-	-	-	-
Hebdomadis	-	-	1	-	-	-	2	-	-	-	-
Canicola	3	1	1	-	-	-	2	-	-	-	-
Hardjo	3	-	3	-	-	-	2	-	-	-	-
Bellum	-	-	-	-	-	-	-	-	-	-	-
Bataviae	-	1	-	-	-	-	-	-	-	-	-
Sejroe	-	-	-	-	-	-	-	-	-	-	-
Tarassovi	-	-	-	-	-	-	-	-	-	-	-
Shermani	-	-	-	-	-	-	-	-	-	-	-
Kaup	-	-	-	-	-	-	-	-	-	-	-
Hurstbridge	-	-	-	-	1	-	-	-	1	-	-
Javanica	-	-	-	-	1	-	-	-	1	-	-

C=Cattle, B=Buffalo

water. (ii) Impregnate in 1% silver nitrate solution at 43°C for 30 minutes. (iii) Then place slides on staining rack using glass rods and flood with the developing solution. Allow to develop yellow brown colouration. (iv) Wash quickly and thoroughly in hot tap water to stop reaction. (v) Dehydrate and clear the slide through 95% isopropyl alcohol, absolute isopropyl alcohol and xylene, 2 changes each for 2 minutes. (vi) And finally mount the slide with DPX (Prophet *et al.*, 1994). Hematoxyline and Eosin Staining method (H&E) was performed on paraffin embedded microsections (5 µ) of tissue according to Luna (1968).

### Statistical Analysis

Chi-square test was used according to WEB AGRI STAT PACKAGE software developed by Jangam and Wadekar, ICAR research complex, Goa for statistical analysis of data (Jangam and Wadekar, 2012).

### RESULTS AND DISCUSSION

The prevalence of leptospiral antibodies was detected in 15.84 and 17.24 % clinically ailing cattle and buffaloes, respectively. In clinically ailing seropositive cattle history of mastitis/ agalactia/ oligolactia was recorded in maximum number of cases (24.00%) followed by abortion (20.00%) fever (14.81%) repeat breeding (11.76%) and anorexia (5.88%) in different combinations. In seropositive clinically ailing buffaloes the clinical signs included mastitis (30.00%), fever (16.66%) and abortion (14.28%) (table 1).

Clinically ailing seropositive cases were having history of mastitis/ agalactia/ oligolactia in maximum number both in cattle (24.00%) and buffaloes (30.00%) and supported the observations of earlier worker (Sakhaee *et al.*, 2007). Our present findings related to reproductive (abortion, repeat breeding, mastitis/agalactia/oligolactia) and systemic ailments (fever and anorexia) were also in agreement with

such observations made in cattle and buffaloes in past (Mariya *et al.*, 2007; Balakrishnan *et al.*, 2011).

Reproductive problems seen in cattle and buffaloes in the present study could be due to localization of leptospire in reproductive tract/uterus and supported earlier observations (Ellis and Michna, 1977; Ellis *et al.*, 1982). Further, Ellis and Michna (1977) mentioned that leptospire is demonstrated in placenta for 14-60 days and renal tubules up to 174 days. Ellis *et al.* (1982) isolated leptospire from bovine uterus having the history of impaired fertility. To support these conjuncture in the present study further studies are needed because morbid material (liver, kidneys, lung and spleen) collected from slaughter house did not revealed any leptospira like structure in microsections stained by H&E and Warthin Starry special staining technique. However, Vegad and Katiyar (2001) have mentioned about occult form of disease which is characterized by absence of clinical symptoms. Eaglesome *et al.* (1992) mentioned that subclinical/occult disease is the main form occurring in areas where leptospirosis is endemic.

In respect of age among clinically ailing cattle highest prevalence was noted in animals above 4 years of age (20.00%) followed by 1-4 years of age (4.76%) whereas in clinically ailing buffaloes seropositivity was recorded only in those above 4 years of age (20.00%). Clinical expressions among cattle were seen in more number of males (20.00%) than in females (15.63%). Conversely in buffaloes, females showed higher prevalence (17.85%) in comparison to males (0.0%).

Agrawal *et al.* (2005) reported that frequency of leptospirosis increase with increasing age of the animals. Ramin and Azizzadeh (2013) in their latest study could not observe any sex bias in respect of seropositivity. Contrary to the observations of Balakrishnan *et al.* (2011) noted significantly higher seropositivity in males than females in cattle. In brief, there is absence of unanimity about sex bias in bovine leptospirosis.

Serovars reported from clinically ailing cattle comprised of Pyrogen, Australis, Pomona, Icterohaemorrhagiae, Canicola, Hardjo, Hebdomadis, Patoc, Hurstbridge, Javanica and Autumnalis. On the other hand in buffaloes, serovars Pomona, Canicola and Bataviae were detected (Table 2).

The reported serovars in this study were in agreement with the observations made in past in ailing cattle and

buffaloes from India (Sachan *et al.*, 2011; Mariya *et al.*, 2007). From Iran, Bahari *et al.* (2011), Sakhaee *et al.* (2007) and Hamali *et al.* (2012) also reported serovars like Canicola, Icterohaemorrhagiae, Pomona and Hardjo from cattle having history of mastitis, abortion, jaundice and haemoglobinuria. Further Hamali *et al.* (2012) mentioned that Pomona and Canicola serovars were involved in equal number of abortion cases while Bahari *et al.* (2011) indicated that Canicola alone was predominant in cattle having history of abortion. In cases of mastitis and abortion serovar Hardjo was mostly involved (Durfee and Allen, 1980).

Based on the world literature available on distribution of various leptospiral serovar from different countries it has been concluded that serovar distribution in clinical cases varies from countries to countries and from areas to areas in the same countries (Patel, 2014). These variations could be due to the fact that the survival of leptospire outside their hosts depends on a complex interaction involving climatic variables, soil salinity and other factors (Radostits *et al.*, 2009).

#### ACKNOWLEDGEMENTS

The authors are highly thankful to the Dean & Principal, Vanbandhu College of Veterinary science & Animal Husbandry, Navsari Agricultural University, Navsari for providing facilities and fund. Authors also wish to thank Dr. Sumaiya Mulla, Professor & Head, Department of Microbiology, Government Medical College, Surat and The director, National Institute of Veterinary Epidemiology and Disease Informatics (NIVEDI), Indian Council of Agricultural Research, formerly PD\_ADMAS, Hebbal, Bengaluru, Karnataka, India for their help in screening serum samples.

#### REFERENCES

- Adler, B. and de la Pena Moctezuma, A. 2010. Leptospira and Leptospirosis. *Vet. Microbiol.*, **140**: 287-296.
- Agrawal, R. Kumar, M. Kumar, M. and Srivastava, S.K. 2005. Epidemiological pattern of leptospirosis in livestock of Uttaranchal state. *Indian J. Comp. Microbiol. Immunol. Infect. Dis.*, **26**(2):109-113.
- Bahari, A., Abdollahpour, G., Sadeghi-Nasab, A., Sattari Tabrizi, S., Yavari, M and Dadmehr, B. 2011. A serological survey on leptospirosis in aborted dairy cattle in industrial farms of Hamedan suburb, Iran. *Iranian J. Vet. Res.*, **12**(4): 337-339.

- Balakrishnan, G., Roy, G.P., Govindarajan, R., Ramaswamy, V. and Murali Manohar, B. 2011. Seropidemiological studies on leptospirosis among bovines in organized farm. *Int. J. Agro Vet. Med. Sci.*, **5**(6): 511-519.
- Durfee, P.T. and Allen, J.D. 1980. Serological titres of dairy cows over a 63-week period following natural infection with *Leptospira interrogans* serovar Hardjo. *Aust. Vet. J.*, **56**: 574-579.
- Eaglesome, M.D., Garcia, M.M. and Steward, R.B. 1992. Microbial agent associated with bovine genital tract infections and semen. Part-II. *Haemophilus somnus*, *Mycoplasma* spp and *Ureaplasma* Spp., *Chlamydia*; Pathogens and semen contaminants; treatments of bull semen with antimicrobial agent. *Vet. Bull.*, **62**: 751-753.
- Ellis, W.A. 1984. Bovine leptospirosis in the tropics: prevalence, pathogenesis and control. *Prev. Vet. Med.*, **2**: 411-421.
- Ellis, W.A. and Michna, S.W. 1977. Bovine leptospirosis: experimental infection of pregnant heifers with a strain belonging to the Hebdomadis serogroup. *Res. Vet. Sci.*, **22**(2): 229-236.
- Ellis, W.A., Neill, S.D., O'Brien, J.J., Cassells, J.A. and Hanna, J. 1982. Bovine leptospirosis: Microbiological and serological findings in normal fetuses removed from the uteri after slaughter. *Vet. Rec.*, **110**(9): 192-194.
- Hamali, H., Jafari Joozani, R. and Abdollahpour, G.R. 2012. Serodiagnosis and molecular survey on leptospiral abortions in the dairy cattle of Tabriz. *Iranian J. Vet. Res.*, **13**(2): 120-125.
- Jangam, A.K. and Wadekar, P. 2012. Web Agri State Package. ICAR research complex for Goa. <http://www.icargoa.res.in/wasp/index.php>
- Luna, A. G. 1968. Manual of histological staining methods of the Armed Forces Institute of Pathology, 3rd edition Mc Graw Hill book Co, London. 124-125.
- Mariya, R., Srivastava, S.K. and Thangapandian, E. 2007. Seroprevalence of leptospiral antibodies in Bovine. *Indian Vet. J.*, **84**: 547 – 548.
- Patel, J.M. 2014. Pathoepidemiological and molecular studies in on leptospirosis in bovines of South Gujarat. PhD thesis, Navsari Agricultural University, Gujarat, India.
- Prophet, E.B., Mills, B., Arrington, J.B. and Sobin, L.H. 1994. Armed forces Institute of Pathology- Laboratory Methods in Histotechnology. 214-215.
- Radostits, O.M., Gay, C.C., Hinchcliff, K.W. and Constable, P.D. 2009. Veterinary Medicine, A text book of the diseases of cattle, horse, sheep, pigs, and goat. 10th edition, Saunders Elsevier. 1094-1110.
- Ramin, A.G. and Azizzadeh, F. 2013. Seropidemiological Detection of Antibodies against *Leptospira* spp. using Microscopic Agglutination Test in Urmia Cows and Sheep. *Acta. Vet- Beograd.*, **63**(1): 53-61.
- Sachan, N., Binita N., Singh, V.P., Sabrinath, T., Chaudhary, P. and Agarwal, R.K. 2011. Seroprevalence of leptospirosis in the animals in Rohilkhand region of Uttar Pradesh, India. *Int. J. Agro Vet. Med. Sci.*, **5**(6): 511-519.
- Sakhaee, E., Abdollahpour, Gh. R., Bolourchi, M., Hasani Tabatabayi, A.M. and Sattari Tabrizi, S. 2007. Serologic and bacteriologic diagnosis of bovine leptospirosis in Tehran suburb dairy farms. *Iranian J. Vet. Res.*, **8**(4): 325-332.
- Shivakumar, S. 2008. Leptospirosis - Current Scenario In India, Article 15. *Medicine Update*, **18**: 799 – 809.
- Vegad, J.L. and Katiyar, A.K. 2001. A Textbook of Veterinary Special Pathology, Infectious Diseases of Livestock and Poultry. 1st Edn. International Book Distributing Co. (Publishing Division). pp:340-346.
- Vijayachari, P., Suganan, A.P. and Sehgal, S.C. 2001. Role of microscopic agglutination test (MAT) as a diagnostic tool during acute stage of leptospirosis in low and high endemic areas. *Indian J. Med. Res.*, **114**: 99-106.
- Vinetz, J.M. 2001. Leptospirosis. *Curr. Opin. Infect. Dis.*, **14**: 527-538. <http://dx.doi.org/10.1097/00001432-200110000-00005>
- World Organization for Animal Health (Office International des Épizooties -OIE). 2013. Manual of Diagnostic Tests and Vaccines for Terrestrial Animals. OIE, Paris; Chapter 2.1.9: 251-264.