



Molecular and phylogenetic analysis of the genus *Orthetrum* (Odonata: Anisoptera: Libellulidae) using mitochondrial COI gene

Lalţanpuii^{1,2*}, N. Senthil Kumar² and Manu Thomas Mathai¹

¹Department of Zoology, Madras Christian College, Tambaram, Chennai 600 059, India

²Department of Biotechnology, Mizoram University, Mizoram 796 004, India

Received 4 August 2014 | Revised 12 August 2014 | Accepted 19 August 2014

ABSTRACT

Molecular phylogenetic relationships among members of the odonate genus *Orthetrum* (Odonata: Anisoptera: Libellulidae) were examined using 403 bp of mitochondrial COI. The support for monophyly of the *Orthetrum* was found in some studies with unresolved complexity. The *O. sabina*, *O. serapia* and *O. trincaria* formed a separate and distinct group from the morphological analysis. We analysed the COI sequences of 22 species of *Orthetrum* using MEGA6. The p-distance between the members and the rate of transitional and transversional substitution was generated. The analysis indicated that the *Orthetrum* are monophyletic and *O. sabina* and *O. trincaria* formed a distinct and a separate group.

Key words: Molecular phylogeny; mitochondrial COI; monophyly; *Orthetrum*; p-distance.

INTRODUCTION

The genus *Orthetrum* includes about 60 species, of which one-half is distributed in tropical Africa and the rest extends across Eurasia to Australia. The genus is diverse in appearance in the Oriental region, such as red, boldly patterned and wingmarked species.¹ 14 species of *Orthetrum* are reported from India² and in Mizoram 4 species have been recorded. Mor-

phological analyses indicated that *O. trincaria* forms a distinct group with *O. sabina* and *O. serapia*.² *O. sabina* and *O. trincaria* are well known for feeding on other odonate species, sometimes of greater size than their own.³ It has been found that the African *O. trincaria* has spread out into Madagascar, Mesopotamia and Europe, while tropical Asia's most dominant species, *O. Sabina*, has migrated to northern Africa, Turkey and Europe.¹ Some support for the monophyly of *Orthetrum* have been found, but the inclusion of other species suggests some unresolved complexity.^{4,5}

Corresponding author: Lalţanpuii
 Phone: +91-9436152033
 E-mail: laltetei@yahoo.co.in

In the present study we infer a phylogeny for the genus *Orthetrum* utilising the cytochrome *c* oxidase subunit 1 (CO1) gene of the mitochondrial DNA. The mitochondrial DNA is a good choice for phylogenetic study because of its fast mutational rate which gives a significant variation between species, lack of introns, limited exposure to recombination and its haploid mode of inheritance. CO1 universal primer is very robust, enabling recovery of its 5' end from most of the animal phyla and it possesses a great phylogenetic signal.⁶

MATERIALS AND METHODS

Taxon sampling

Table 1. List of taxa for the present study.

Taxon name	Country	Genbank Accession no.
<i>Davidius lunatus</i> [Bartenev, 1914]	Republic of Korea	EU591677
<i>O. pruinosum</i> (Burmeister, 1839)	Mizoram, India	KC122236
<i>O. sabina</i> (Drury, 1770)	Mizoram, India	KC122234
<i>O. triangulare</i> (Selys, 1878)	Mizoram, India	KC287152
<i>O. glaucum</i> (Brauer, 1865)	Mizoram, India	KC122232
<i>O. triangulare</i> (Selys, 1878)	Republic of Korea	KF257074
<i>O. lineostigma</i> (Selys, 1886)	Republic of Korea	KF257071
<i>O. albistylum</i> (Selys, 1848)	Republic of Korea	KF257070
<i>O. japonicum</i> (Uhler, 1858)	Republic of Korea	KF257061
<i>O. triangulare</i> (Selys, 1878)	Japan	AB781568
<i>O. sabina</i> (Drury, 1770)	Japan	AB781554
<i>O. pruinosum</i> (Burmeister, 1839)	Japan	AB781552
<i>O. melania</i> (Selys, 1883)	Japan	AB781551
<i>O. luzonicum</i> (Brauer, 1868)	Japan	AB781544
<i>O. glaucum</i> (Brauer, 1865)	Japan	AB781542
<i>O. trinacria</i> (Selys, 1841)	Germany	KC912286
<i>O. julia falsum</i> (Longfield, 1955)	Germany	KC912281
<i>O. coeruleascens</i> (Fabricius, 1798)	Germany	KC912271
<i>O. chrysostigma</i> (Burmeister, 1839)	Germany	KC912262
<i>O. brachiale</i> (Palisot de Beauvois, 1805)	Germany	KC912258
<i>O. poecilops</i> (Ris, 1916)	Japan	AB709089
<i>O. internum</i> (McLachlan, 1894)	Japan	AB709025
<i>O. japonicum</i> (Uhler, 1858)	Japan	AB709026

The taxon sample for the present study included 18 *Orthetrum* species retrieved from GenBank (National Centre for Biotechnology Information) with the locality and the accession numbers. The 4 species generated from Mizoram were also included in the present study (Table 1). The gomphidus, *Davidius lunatus* was selected as outgroup.

MOLECULAR EVOLUTIONARY GENETIC ANALYSIS (MEGA)

Phylogenetic and molecular evolutionary analyses were conducted using MEGA version 6.⁷ All the 23 sequences of were aligned and edited using clustalW implemented in MEGA6.

Table 2. The genetic distance (p-distance) between the *Orthetrum* species.

Species	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	
<i>Orthetrum_pruinosum</i>																						
<i>Orthetrum_sabina</i>	0.142																					
<i>Orthetrum_triangulare</i>	0.084	0.158																				
<i>Othetrum_glaucum</i>	0.102	0.132	0.112																			
<i>KF257074Orthetrum_triangulare</i>	0.092	0.177	0.033	0.136																		
<i>KF257071Orthetrum_lineostigma</i>	0.090	0.180	0.107	0.121	0.114																	
<i>KF257070Orthetrum_albistylum</i>	0.111	0.138	0.120	0.094	0.138	0.134																
<i>KF257061Orthetrum_japonicum</i>	0.099	0.175	0.125	0.138	0.140	0.107	0.139															
<i>AB781568Orthetrum_triangulare</i>	0.087	0.161	0.002	0.112	0.035	0.110	0.117	0.128														
<i>giAB781554Orthetrum</i>	0.142	0.023	0.158	0.126	0.180	0.173	0.138	0.166	0.161													
<i>AB781552Orthetrum_pruinosum</i>	0.005	0.148	0.079	0.103	0.087	0.085	0.111	0.099	0.081	0.148												
<i>AB781551Orthetrum_melania</i>	0.096	0.183	0.038	0.151	0.043	0.125	0.144	0.116	0.041	0.186	0.095											
<i>AB781544Orthetrum_luzonicum</i>	0.116	0.170	0.121	0.114	0.136	0.135	0.116	0.119	0.124	0.164	0.122	0.139										
<i>AB781542Orthetrum_glaucum</i>	0.102	0.132	0.112	0.000	0.136	0.121	0.094	0.138	0.112	0.126	0.103	0.151	0.114									
<i>KC912286Orthetrum_trinacia</i>	0.114	0.147	0.147	0.136	0.169	0.153	0.154	0.149	0.150	0.147	0.114	0.178	0.160	0.136								
<i>KC912281Orthetrum_julia_falsum</i>	0.108	0.153	0.129	0.114	0.141	0.120	0.113	0.137	0.132	0.150	0.102	0.142	0.138	0.114	0.138							
<i>KC912271Orthetrum_coerulescens</i>	0.099	0.160	0.103	0.111	0.111	0.117	0.119	0.127	0.106	0.160	0.093	0.123	0.098	0.111	0.148	0.113						
<i>KC912262Orthetrum_chrysostigma</i>	0.116	0.159	0.129	0.117	0.147	0.117	0.096	0.122	0.132	0.156	0.111	0.142	0.135	0.117	0.153	0.065	0.102					
<i>KC912258Orthetrum_brachiale</i>	0.094	0.145	0.106	0.088	0.123	0.088	0.093	0.104	0.108	0.142	0.094	0.126	0.123	0.088	0.130	0.074	0.094	0.076				
<i>AB709089Orthetrum_poecilops</i>	0.119	0.150	0.105	0.114	0.113	0.123	0.107	0.107	0.108	0.141	0.113	0.119	0.145	0.114	0.172	0.128	0.116	0.113	0.098			
<i>AB709025Orthetrum_internum</i>	0.102	0.175	0.128	0.137	0.143	0.113	0.139	0.010	0.131	0.172	0.102	0.119	0.125	0.137	0.162	0.140	0.127	0.122	0.107	0.108		
<i>AB709026Orthetrum_japonicum</i>	0.117	0.176	0.118	0.129	0.127	0.133	0.122	0.133	0.121	0.179	0.111	0.127	0.145	0.129	0.174	0.118	0.108	0.117	0.108	0.110	0.133	

The evolutionary history was inferred by using the Maximum Likelihood method based on the Tamura-Nei model.¹⁰ Initial tree(s) for the heuristic search were obtained automatically by applying Neighbor-Join and BioNJ algorithms to a matrix of pairwise distances estimated using the Maximum Composite Likelihood (MCL) approach, and then selecting the topology with superior log likelihood value. The tree is drawn to scale, with branch lengths measured in the number of substitutions per site (next to the branches). The analysis involved 23 nucleotide sequences. Codon positions included were 1st+2nd+3rd+Noncoding. All positions containing gaps and missing data were eliminated. There were a total of 403 positions in the final dataset. Evolutionary analyses were conducted in MEGA6.⁷

RESULTS

The genetic distance between the 22 species of *Orthetrum* was generated (Table 2). The nucleotide distance (p-distance) between *O. lineostigma* and *O. sabina* was 0.180; *O. triangulare* and *O. sabina* was 0.180; *O. melania* and *O. sabina* was 0.183 and 0.186; *O. japonicum* and *O. sabina* was 0.179; *O. trinacria* and *O. melania* was 0.178 which were found to be highest p-distance. The p-distance between *O. pruinatum* from Mizoram and Japan was 0.005 and the p-distance between *O. trinacria* from Mizoram against Korea and Japan was 0.033 and 0.002 and between Korea and Japan was found to be 0.035. The p-distance between *O. glaucum* from Japan and Mizoram was found to be zero.

The maximum likelihood estimate of transitional substitution matrix between A/G = 5.58, T/C = 14.44, C/T = 25.64 and G/A = 11.09 (Table 3). The nucleotide frequencies are A = 31.64%, T/U = 33.56%, C = 18.89%, and G = 15.91%. The transition/transversion rate ratios are $k_1 = 1.621$ (purines) and $k_2 = 3.533$ (pyrimidines). The estimated transition/transversion bias (R) is 1.225.

The tree with the highest log likelihood (-

2745.6358) is shown (Figure 1). The Maximum Likelihood tree generated shows the monophyly of *Orthetrum* with respect to the outgroup having 2 distinct groups A and B. The Clade B consisted of *O. brachiale*, *O. julia falsum* and *O. chysostygma*, and the rest of the taxa are included in clade A. Clade A can be divided into clade A1 and A2. Within clade A1 the 2 *O. sabina* and *O. trinacria* formed a sister clade with *O. glaucum* and *O. albistylum*; these two sister clades formed a sister clade with *O. poecilop*, *O. japonicum* and *O. internum*. In clade A2 *O. coerulescens* and *O. luzonicum* formed a sister clade with 2 *O. glaucum*, 3 *O. tringulare*, *O. lineostigma* and *O. melania*.

Table 3. Maximum composite likelihood estimate of the pattern of nucleotide substitution.

	A	T	C	G
A	-	7.26	4.09	5.58
T	6.84	-	14.44	3.44
C	6.84	25.64	-	3.44
G	11.09	7.26	4.09	-

NOTE: Each entry shows the probability of substitution (r) from one base (row) to another base (column). Rates of different transitional substitutions are shown in **bold** and those of transversional substitutions are shown in *italics*.

DISCUSSIONS

The genus *Orthetrum* having more than 60 species are suggested to be monophyletic from the previous morphological and molecular studies but with a complex resolution. Morphological analyses indicated that *O. trinacria* forms a distinct group with *O. sabina* and *O. serapia*.^{1,3,4} In the present study the COI gene for *O. serapia* is not available; we analysed 22 species of *Orthetrum* using the COI gene. The phylogenetic tree showed the *Orthetrum* are monophyletic in relation to the outgroup *Davidus lunatus*. The *O. sabina* and *O. trinacria* were found to be most distantly related to the rest of the *Orthetrum* ana-

lysed having a genetic distance of approximately 18%. In the phylogenetic analysis *O. sabina* from oriental region and *O. trinacria* from Europe were forming a separate and distinct sister clade eventhough the genetic distance between them was found to be 15%. The *O. sabina* and *O. trinacria* forming a sister clade and a distinct group from the rest of the *Orthetrum* analysed is in congruent with the behaviour and morphological characters indicated by Silsby³ and Dijkstra and Kalkman.¹ The genetic distance of individuals of *O. prunosum*, *O. glaucum*, *O. triangulare* and *O. sabina* sampled from different regions (all from oriental region) were found to be very low. The nucleotide composition of *Orthetrum* COI was A + T rich, which is typical for arthropods.¹¹ The rate of transitional substitution is higher than transversional substitution in the COI sequence analysis.

The present analysis indicated that the *Orthetrum* are monophyletic and *O. sabina* and *O. trinacria* formed a distinct and a separate group. The inclusion of more species, more molecular markers and other phylogenetical analysis methods will further resolve the complexity of the genus *Orthetrum*.

ACKNOWLEDGEMENT

The authors thank Department of Biotechnology, New Delhi, India for financial assistance and computational facility through Bioinformatics Infrastructure Facility (No. BT/BI/12/060/2012(NERBIF-MUA) and State Biotech Hub (No. BT/04/NE/2009). We thank Ministry of Social Justice & Empowerment and Ministry of Tribal Affairs for funding UGC's RGNF Scheme for SC/ST [F-14-265(ST)/2007 (SA-III), March 2007].

REFERENCES

1. Dijkstra KDB & Kalkman VJ (2012). Phylogeny, classification and taxonomy of European dragonflies and damselflies (Odonata): a review. *Org Divers Evol*, **12**, 209–227.
2. Subramanian KA (2014). *A Checklist of Odonata of India* (Version 2.0.2014). Zoological Survey of India. URL: http://zsi.gov.in/check_list.html (30 July 2014).
3. Silsby J (2001). *Dragonflies of the World*. Smithsonian Institution Press. Washington, D. C, USA, pp. 1–216.
4. Ware JL, May ML & Kjer KM 2007. Phylogeny of the higher Libelluloidea (Anisoptera: Odonata): an exploration of the most speciose superfamily of dragonflies. *Mol Phylogenet Evol*, **45**, 289–310.
5. Pilgrim EM & Von Dohlen CD (2008). Phylogeny of the Sympetrinae (Odonata: Libellulidae): further evidence of the homoplasious nature of wing venation. *Syst Entomol*, **33**, 159–174.
6. Hebert PD, Cywinska A, Ball SL & Dewaard JR (2003). Biological identifications through DNA barcodes. *Proc R Soc B*, **270**, 313–321.
7. Tamura K, Stecher G, Peterson D, Filipiński A & Kumar S (2013). MEGA6: Molecular Evolutionary Genetics Analysis Version 6.0. *Mol Phylogenet Evol*, **30**, 2725–2729.
8. Nei M & Kumar S (2000). *Molecular Evolution and Phylogenetics*. Oxford University Press, New York, pp. 33–36.
9. Tamura K, Nei M, and Kumar S. (2004). Prospects for inferring very large phylogenies by using the neighbor-joining method. *Proc Natl Acad Sci (USA)*, **101**, 11030–11035.
10. Tamura K & Nei M (1993). Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. *Mol Biol Evol*, **10**, 512–526.
11. Simon C, Frati F, Beckenbach A, Crespi B, Liu H & Flook P (1994). Evolution, weighting, and phylogenetic utility of mitochondrial gene sequences and a compilation of conserved polymerase chain reaction primers. *Ann Entomol Soc Am*, **87**, 651–701.