



## The Impact of Low Level X-rays on Biosynthesis of Gold Nanoparticles by Actinobacteria

Faranak Saghatchi <sup>a,\*</sup>, Gholam Ali Jafari <sup>b</sup>, Jafar Taran <sup>b</sup>, Zohre Farahmandkia <sup>b</sup>

<sup>a</sup> Department of Radiology, School of Paramedical Sciences, Zanjan University of Medical Sciences, Zanjan, Iran.

<sup>b</sup> Department of Environmental Engineering, School of Public Health, Zanjan University of Medical Sciences, Zanjan, Iran.

\*Corresponding author. E-mail address: saghatchif@zums.ac.ir

### ARTICLE INFO

#### Article history:

Received August 23, 2016

Accepted November 8, 2016

#### Article Type:

Original Article

#### Keywords:

Gold Nanoparticles  
Biosynthesis  
Radiation Hormesis  
Actinomycetals

### ABSTRACT

**Background:** Gold nanoparticles (GNPs) play an important role in medical, health and environmental applications. All kinds of microorganisms were found to be able to synthesize GNPs. The optimization of laboratory conditions for achieving more economical benefits of mass production has been studied widely.

**Methods:** This study assesses the enhancing effect of low-level X-rays on the biosynthesis of GNPs by Actinomycetals. The isolated Actinomycetes were grown aerobically in MGYB broth media. The harvested bacteria were suspended in 50 mL aqueous HAuCl<sub>4</sub> in 12 Erlenmeyer flask. Each group contained 4 flasks. 2 groups of samples were irradiated by 30 mGy and 5 mGy X-rays respectively. The third group as control remained without irradiation. The solutions were shake- incubated for 120 h.

**Results:** After 5 days, the color of first group samples changed from milky to purple, while the color changing occurred after 10 days in the 2nd group samples and the control samples. The UV-vis absorption spectrometry of the irradiated aqueous medium by 30 mGy X-rays confirmed the formation of GNPs.

**Conclusion:** The findings showed that 30 mGy X-rays stimulated the microorganism to form GNPs in a half time in comparison to other groups.

## 1. Introduction

Nanotechnology and Nano sciences are gaining most attention due to their expected impact on many important fields such as medicine, health, and environment [1]. GNPs are the most known multi-functional nanoparticles for medical applications [2-5]. One of the major fields of application of nanoparticles in agriculture and the environment is detection and removal the chemical pollutants found in water, soil and air [6, 7].

Synthesis of metal nanoparticles apply various strategies such as physico-chemical methods and biological synthesis [8, 9]. Recent methods use biological processes or enzymatic reactions to reduce metal ions using special enzymes. This method has been successfully used in production of small nanoparticles on a large scale. Biological synthesis of nanoparticles is more reliable, eco-friendly and economical in terms of energy consumption [10].

**To cite:** Saghatchi F, Jafari GhA, Taran J, Farahmandkia Z. The Impact of Low Level X-rays on Biosynthesis of Gold Nanoparticles by Actinobacteria. *J Hum Environ Health Promot.* 2016; 2(1):32- 38.

Different types of microorganisms can produce gold nanoparticles in the form of intra or extra cellular [11-13]. The whole range of bacteria as well as Actinobacteria due to the ease of genetic manipulation, methods for industrialization and utilization of genetic engineering have attracted the most attention [11, 14-16]. Actinomycetes are a group of aerobic gram-positive bacteria, usually immovable and disciplinary. Actinomycetes can often be easily cultured in conventional environments. Actinomycetes in the soil remain mostly saprophytic and the pathogenic species are relatively low. Actinomycetes isolated from soil produce a variety of different antibiotics some of which are of great value such as streptomycin and neomycin due to their medical significance [17].

Since the cellular mechanisms leading to the recovery of gold ions and the formation of gold nanoparticles are not well known, there is the potential for manipulation of key parameters that control cell growth and other activities to achieve optimal production of gold nanoparticles [18-21].

X-rays were discovered in 1895 by German physicist Wilhelm Conrad Roentgen. X-rays are electromagnetic waves, short wavelength and high energy photons [22]. The radiation energy is transferred through the atoms and lead to the ionization in mater. Ionizing radiation has many Short-term and late hazardous effects [23].

However, over the past few decades, some scientists report of positive effects of low-level ionizing radiation named *HORMESIS* [24-27]. In 1980, Professor T.D Lucky reported the stimulating effects of ionizing radiation, both in animals and plants [28]. After the first report of this case, more than 3,000 articles were published about the positive effects of low- level ionizing radiation [29, 30]. Radiation Hormesis refers to generally positive physiological effects of low - radiation dose in the range of 1-50 cGy [26].

Several studies have demonstrated metabolism promotion in microorganisms, plants, invertebrates and laboratory animals by low-level ionizing radiation. These studies also have reported the effect of ionizing radiation on stimulation of respiratory system, boosting metabolism, cell resistance and extending life span

[25, 31-34]. In various surveys many factors have been studied to increase production of nanoparticles by microorganisms [1, 13, 35-37].

So far, no studies have been conducted on the impact of ionizing radiation on optimization of biosynthesis of nanoparticles by microorganisms.

This study is the first research on this issue.

## 2. Materials and Methods

### 2.1. Materials

The absorption spectra of the samples were taken using a UV-vis spectrophotometer .

HAuCl<sub>4</sub> was obtained from Sigma-Aldrich, USA. All other chemicals such as agar, yeast extract, peptone, casein, starch, malt extract, nystatin, glucose, CaCO<sub>3</sub>, FeSO<sub>4</sub>, K<sub>2</sub>HPO<sub>4</sub>, KNO<sub>3</sub>, MgSO<sub>4</sub> and NaCl were purchased from Merck, Germany. Freshly- prepared double distilled water was used throughout the experimental work.

### 2.2. Isolation and growth of Actinomycetes

The soil samples were collected from Angoran Lead mine in Zanjan. Five grams of sample was diluted by 9mL of distilled water. Then, 1ml of the aqueous solution was diluted again by 9 ml of distilled water and was shaken for 30 min. The aqueous solution was cultured in starch casein agar medium (starch 10 g, casein 0.3 g, CaCO<sub>3</sub> 0.02 g, FeSO<sub>4</sub> 0.01 g, K<sub>2</sub>HPO<sub>4</sub> 2 g, KNO<sub>3</sub> 2 g, MgSO<sub>4</sub> 0.05 g, NaCl 2 g, and agar 15 g in 1,000 mL sterile distilled water at pH 7). To minimize the fungal and bacterial growth, nystatin (100 mg/L) was added to the cultures. The plates were then incubated at 28°C for 7 days. After an initial period of bacterial growth in culture media, different colonies were obtained. In order to identify the bacteria genus, the colonies were identified based on typical properties such as chalky appearance surface color and the colonies smell like the smell of soil. Then the isolated colonies were cultured in medium containing starch casein agar plates and incubated for 7 days at 28 °C. The Actinomycetes colonies were identified morphologically according to the procedures described in Bergey's manual of

determinative bacteriology. Fig. 1 shows the plates containing single clone of bacteria isolated from Angoran Mine. The samples were gram stain. The purple color of gram positive Actinomycetes were observed through a light microscope (Fig. 2). The



**Fig. 1:** Plates containing single clone of bacteria isolated from Angoran Mine.

### 2.3. Irradiation and biosynthesis of GNPs

The cultures were then centrifuged at 6000 rpm for 20 min. The harvested bacteria were washed with distilled water under sterile condition. About 2 g of wet weight bacteria was suspended in 50 mL of 1 mM aqueous  $\text{HAuCl}_4$  in each 12 Erlenmeyer flasks in 3 groups. Then the first group of samples (4 flasks) was irradiated to 30 mGy X-rays, by 200Kv Shimadzu X-ray tube while the second group (4 flasks) was irradiated to 5mGy X-rays. The control group (4 flasks) remained without radiation. All solutions were then shake- incubated (200 rpm) at 25°C for 120 h.

### 2.4. Characterization of Gold Nanoparticles

The preliminary detection of the formation of GNPs was carried out by observing the color change of the solutions after irradiation to 30 mGy X-rays. The absorption spectra of the samples were taken using a UV-vis spectrophotometer from 300 to 800 nm (International MAX-USA, Genesys E5).

## 3. Results and Discussion

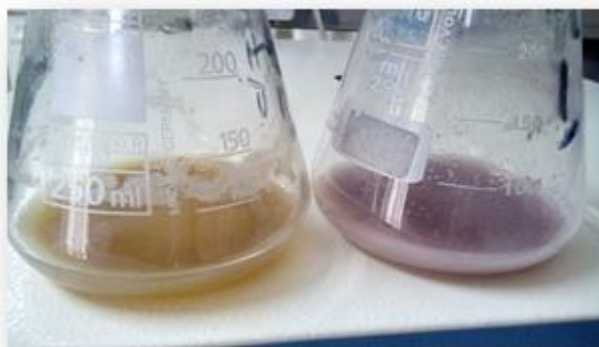
isolated Actinomycetes were grown aerobically in 500 mL of MGYB broth media (malt extract 3 g, glucose 10 g, yeast cultures were incubated at 28 °C for 72 h with extract 3 g and peptone 5 g per in one liter of distilled water and pH 7. The continuous shaking (200 rpm).



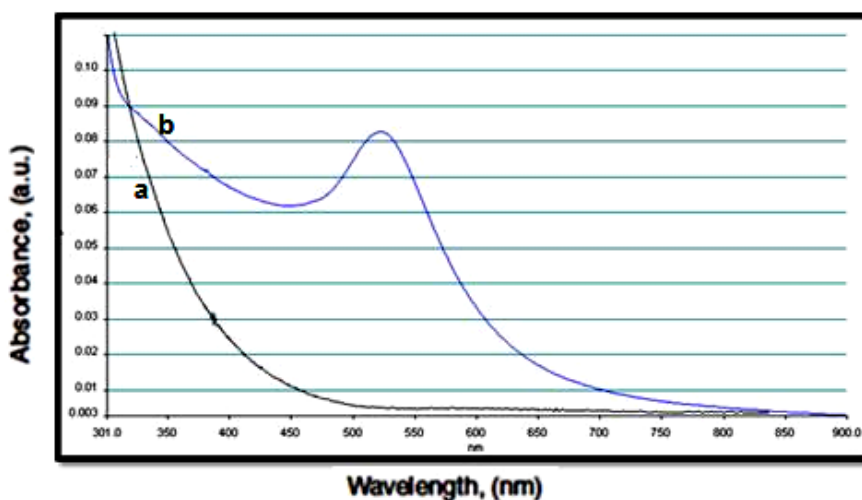
**Fig. 2:** Microscopic view of microorganisms isolated from Angoran Mine.

After 5 days the suspended bacteria to aqueous  $\text{HAuCl}_4$  which were irradiated to 30 mGy X-rays changed their color from milky to purple (Fig. 3).

This was the result of reduction of gold ions to GNPs. The UV-vis spectroscopy indicated that the samples had the maximum absorption at 540 nm ( $\lambda$  max of GNPs) attributable to the Surface Plasmon Resonance band (SPR) of gold nanoparticles [38]. Figure 4 shows the UV-vis spectra recorded from the irradiated bacteria to 30 mGy X-ray. The 5 mGy irradiated and control flasks remained unchanged. Only after 10 days of incubation the color change was observed in the control and 2nd group of samples. UV-vis spectroscopy demonstrated the reduction of the gold ions to GNPs in the second group of samples and the control group as well. Gold nanoparticles have a broad range of applications in a variety of fields such as medical diagnostics and treatment, industry and the environment [4]. The range of applications for gold nanoparticles is growing rapidly. Gold nanoparticles are used to detect biomarkers in the diagnosis of heart diseases, cancers, and infectious agents [16]. One of the major fields of application in agriculture and the environment is detection and cleanup of chemical pollutants found in water, soil and air [7].



**Fig. 3:** The Color change in Actinomycetes irradiated to X-rays after 120 h incubation.



**Fig. 4:** UV- vis spectra recorded from the samples (a) before and (b) after synthesis of gold nanoparticles.

There are some methods to synthesize nano sized particles such as chemical synthesis and solid state processes. Biological synthesis of nanoparticles known as green synthesis is more reliable and eco-friendly. According to the effectiveness and flexibility, biological production systems are of particular value. Some single-celled organisms such as bacteria and algae are capable of synthesizing biocompatible nano particles [14].

In this regard manipulation of key parameters that control cell growth could lead us to achieve optimal production of gold nanoparticles. X-rays are electromagnetic waves short wavelength and high energy photons. Several studies, has demonstrated metabolism promotion in

microorganisms, plants, invertebrates and laboratory animals by low-level ionizing radiation [25]. The goal of this study is the Impact of low level X-rays in the formation of GNPs by microorganisms. The colonies of microorganisms were collected from Lead mine in North West of Iran and characterized by microscopic and macroscopic studies as Actinomycetals. Different types of microorganisms as well as Actinobacteria can produce gold nanoparticles when exposed to  $\text{HAuCl}_4$  solution [11, 14]. In various surveys many factors been studied to increase production of nanoparticles by microorganisms by changing the culture medium or the physical parameters [36, 37]. Our results emphasize the Hormetic effect of Ionizing radiation in microorganisms. Reduction

of the aqueous gold ions during exposure to 30 mGy X-rays may be easily followed by UV-vis spectroscopy. The earlier color change and UV-vis spectroscopy results in irradiated samples in first group reveals that X-rays stimulates the formation of gold nanoparticles by microorganisms. The late color change in 2th groups of samples could result of low intensity of X-rays that wouldn't impose sufficient stimulation to formation of GNPs in 120h. In future it is predictable that complementary studies in biosynthesis of GNPs by microorganisms under influence of Ionizing radiation and optimization of other laboratory conditions could be more applicative and affordable .

#### 4. Conclusion

The findings showed that X-ray radiation stimulates the Actinobacteria to form gold nanoparticles.

#### References

1. Manivasagan P, Venkatesan J, Sivakumar K, Kim SK. Actinobacteria Mediated Synthesis of Nanoparticles and Their Biological Properties: A Review. *Crit Rev Microbiol*. 2014(0): 1-13.
2. Duguet E, Vasseur S, Mornet S, Devoisselle JM. Magnetic Nanoparticles and Their Applications in Medicine. *Future Med*. 2006; 1(2).
3. Yallapu MM, Othman SF, Curtis ET, Gupta BK, Jaggi M, Chauhan SC. Multi-Functional Magnetic Nanoparticles for Magnetic Resonance Imaging and Cancer Therapy. *Biomater*. 2011; 32(7): 1890-905.
4. Farokhzad OC, Multifunctional Nanoparticles for Medical Application. *7<sup>th</sup> International Nanomedicine and Drug Delivery Symposium*; 2009.
5. Zazo H, Colino CI, Lanao JM. Current Applications of Nanoparticles in Infectious Diseases. *J Control Release*. 2016; 224: 86-102.
6. Wong MS, Alvarez PJ, Fang YI, Akçin N, Nutt MO, Miller JT, et al. Cleaner Water Using Bimetallic Nanoparticle Catalysts. *J Chem Technol Biotechnol*. 2009; 84(2): 158-66.
7. Sharma VK, Filip J, Zboril R, Varma RS. Natural Inorganic Nanoparticles—Formation, Fate, and Toxicity in the Environment. *Chem Soc Rev*. 2015; 44(23): 8410-23.
8. Samal AK, Polavarapu L, Rodal-Cedeira S, Liz-Marzán LM, Pérez-Juste J, Pastoriza-Santos I. Size Tunable Au@ Ag Core–Shell Nanoparticles: Synthesis and Surface-Enhanced Raman Scattering Properties. *Langmuir*. 2013; 29(48): 15076-82.
9. Cao Y, Zheng R, Ji X, Liu H, Xie R, Yang W. Syntheses and Characterization of Nearly Monodispersed, Size-Tunable Silver Nanoparticles Over a Wide Size Range of 7–200 nm by Tannic Acid Reduction. *Langmuir*. 2014; 30(13): 3876-82.
10. Mohanpuria P, Rana NK, Yadav SK. Biosynthesis of Nanoparticles: Technological Concepts and Future Applications. *J Nanopart Res*. 2008; 10(3): 507-17.
11. Zonooz NF, Salouti M, Shapouri R, Nasseryan J. Biosynthesis of Gold Nanoparticles by Streptomyces Sp. ERI-3 Supernatant and Process Optimization for Enhanced Production. *J Cluster Sci*. 2012; 23(2): 375-82.
12. Sastry M, Ahmad A, Islam Khan M, Kumar R. Biosynthesis of Metal Nanoparticles Using Fungi and Actinomycete. *Curr Sci*. 2003; 85(2): 162-70.
13. Durán N, Marcato PD, Durán M, Yadav A, Gade A, Rai M. Mechanistic Aspects in the Biogenic Synthesis of Extracellular Metal Nanoparticles by Peptides, Bacteria, Fungi, and Plants. *Appl Microbiol Biotechnol*. 2011; 90(5): 1609-24.
14. Hulkoti NI, Taranath T. Biosynthesis of Nanoparticles Using Microbes—A Review. *Colloids Surf B Biointerfaces*. 2014; 121: 474-83.
15. Khadivi DF, Dehnad A, Saluoti M, Babaei H, Zereshki NL. Biosynthesis of Gold Nanoparticles

- by *Rhodococcus* Species Isolated from Ahar Copper Mine (Iran). *SID*. 2010; 3(2): 37-44.
16. Golinska P, Wypij M, Ingle AP, Gupta I, Dahm H, Rai M. Biogenic Synthesis of Metal Nanoparticles from Actinomycetes: Biomedical Applications and Cytotoxicity. *Appl Microbiol Biotechnol*. 2014; 98(19): 8083-97.
  17. Korzybski T, Kowszyk-Gindifer Z, Kurylowicz W. Antibiotics: Origin, Nature and Properties. *Elsevier*. 2013.
  18. Saurav K, Kannabiran K. Diversity and Optimization of Process Parameters for the Growth of *Streptomyces VITSVK9* spp Isolated From Bay of Bengal, India. *J Nat Environ Sci*. 2010; 1(2): 56-65.
  19. Kulkarni N, Muddapur U. Biosynthesis of Metal Nanoparticles: A Review. *J Nanotechnol*. 2014.
  20. Singh S, Vidyarthi AS, Nigam VK, Dev A. Extracellular Facile Biosynthesis, Characterization and Stability of Gold Nanoparticles by *Bacillus licheniformis*. *Artif Cells Nanomed Biotechnol*. 2014; 42(1): 6-12.
  21. Zhang X, Yan S, Tyagi R, Surampalli R. Synthesis of Nanoparticles by Microorganisms and Their Application in Enhancing Microbiological Reaction Rates. *Chemosphere*. 2011; 82(4): 489-94.
  22. Als-Nielsen J, McMorrow D. Elements of modern X-ray Physics. *John Wiley & Sons*; 2011.
  23. Hall EJ, Giaccia AJ. Radiobiology for the Radiologist. *Lippincott Williams & Wilkins*. 2006.
  24. Luckey T. Radiation Hormesis Overview. *Radiat Prot Manage*. 1999; 16:22-34.
  25. Feinendegen L. Evidence for Beneficial Low Level Radiation Effects and Radiation Hormesis. *Br J Radiol*. 2014.
  26. Mortazavi SJ, Ikushima T, Mozdarani H. An Introduction to Radiation Hormesis. Available from: URL: <http://www.wangelfire.com/mo/radioadaptive/inthor.html>. Accessed. 2004; 18(05).
  27. Liu S-Z. Biological Effects of Low Level Exposures to Ionizing Radiation: Theory and Practice. *Hum Exp Toxicol*. 2010; 29(4): 275-81.
  28. Luckey TD. Radiation Hormesis. *Boston; CRC Press*; 1991.
  29. Thompson RE. Epidemiological Evidence for Possible Radiation Hormesis from Radon Exposure: A Case-Control Study Conducted in Worcester, MA. *Dose Response*. 2011; 9(1): 10-26.
  30. Doss M, Little MP, Orton CG. Point/Counterpoint: Low-Dose Radiation is Beneficial, not Harmful. *Med Phys*. 2014; 41(7).
  31. Robertson KL, Mostaghim A, Cuomo CA, Soto CM, Lebedev N, Bailey RF, et al. Adaptation of the Black Yeast *Wangiella dermatitidis* to Ionizing Radiation: Molecular and Cellular Mechanisms. *Plos*. 2012.
  32. Bryan R, Jiang Z, Friedman M, Dadachova E. The Effects of Gamma Radiation, UV and Visible Light on ATP Levels in Yeast Cells Depend on Cellular Melanization. *Fungal Biol*. 2011; 115(10): 945-9.
  33. Ristow M, Zarse K. How Increased Oxidative Stress Promotes Longevity and Metabolic Health: The Concept of Mitochondrial Hormesis (Mitohormesis). *Exp Gerontol*. 2010; 45(6): 410-8.
  34. Ristow M, Schmeisser S. Extending Life Span by Increasing Oxidative Stress. *Free Radic Biol Med*. 2011; 51(2): 327-36.
  35. Bharde A, Kulkarni A, Rao M, Prabhune A, Sastry M. Bacterial Enzyme Mediated Biosynthesis of Gold Nanoparticles. *J Nanosci Nanotechnol*. 2007; 7(12): 4369-77.
  36. Nayak RR, Pradhan N, Behera D, Pradhan KM, Mishra S, Sukla LB, et al. Green Synthesis of Silver Nanoparticle by *Penicillium Purpurogenum*

NPMF: the Process and Optimization. *J Nanopart Res.* 2011; 13(8): 3129-37.

37. Shakouri V, Salouti M, Mohammadi B, Zonooz NF. Procedure Optimization for Increasing Biosynthesis Rate of Gold Nanoparticles by *Aspergillus Flavus* Supernatant. *J Synthesis and Reactivity in Inorganic, Metal-Organic, and Nano-Metal Chemistry.* 2016; 46(10).

38. Lee KS, El-Sayed MA. Gold and Silver Nanoparticles in Sensing and Imaging: Sensitivity of Plasmon Response to Size, Shape, and Metal Composition. *J Phys Chem B.* 2006; 110(39): 19220-5.