

---

**SALINITY TOLERANCE OF LARVAE OF AFRICAN CATFISH *Clarias gariepinus* (♀) X *Heterobranchus bidorsalis* (♂) HYBRID**

---

<sup>1</sup>GABRIEL, Ugwemorubong Ujagwung, <sup>2</sup>GBULUBO, Awotongha Jacob and <sup>1</sup>DEEKAE, Suanu

<sup>1</sup>Department of Fisheries and Aquatic Environment, Rivers State University of Science and Technology, PMB 5080, Port Harcourt, Rivers State, Nigeria.

<sup>2</sup>African Regional Aquaculture Centre, Nigeria Institute of Oceanography and Marine Research, Aluu, Port Harcourt, Rivers State, Nigeria.

**Corresponding Author:** Gabriel, U. U. Department of Fisheries and Aquatic Environment, Faculty of Agriculture, Rivers State University of Science and Technology, PMB 5080, Port Harcourt, Nigeria.

**Email:** [ugwemg@yahoo.com](mailto:ugwemg@yahoo.com) **Phone:** +234 8032720882

---

**ABSTRACT**

*Fourteen and twenty one day larvae were exposed to abrupt stepwise change in salinity (2, 4, 6, 8, 10 and 12 ppt) for 96 hours to determine mortality, median lethal mortality, MLS and median lethal time, MLT. The fourteen day-old fry that were exposed to 0 – 6 ppt recorded 90%, 87.5% 77.5% and 10% survival at the end of the 96 hours test period. Those that were subjected to 8 ppt and 10 ppt had 100% mortality within 48 and 12 hours, respectively. The median lethal salinity at 96 hours (MLS-96) was 4.4 ppt which was about half the value for 6 ppt. The cumulative mortality of the graded salinities did not differ in 4 and 8 ppt but differed in 6 and 10 ppt, whereas the time intervals differed at 24 and 36 hours but did not differ at 48 – 96 hours,  $p < 0.05$ . The 21 day-old fingerlings transferred into 8, 10 and 12ppt, recorded 100% mortality with 12, 3 and 2 hours, respectively. Those transferred into 0, 4 and 6 ppt had 100%, 80% and 10% survival, respectively at the end of 96 hours test period. The median lethal salinity (MLS-96) for 6 and 12 hours were the same 7.19 ppt, while that for 48 hours was 4.79 ppt. The median lethal time (MLT<sub>50</sub>) for 8 ppt was 8.55 hours, which was seven and half times the value for 4 ppt. The cumulative mortality the various salinity concentrations were not different in 10 and 12 ppt, but differ in 4 – 8 ppt,  $p < 0.05$ . Mortality at various time intervals did not differ at 36 – 96 hours, but did at 3 and 6 hours,  $p < 0.05$ .*

**Keywords:** Catfish hybrid, Post larvae, Median lethal salinity, Median lethal time

---

**INTRODUCTION**

Clariid catfishes occur both in south-east Asia and in Africa and the highest generic diversity is on the African continent where some 14 genera have been reported (Teugels, 1984) against two in south-west Asia. In both continents, the clariids are of great economic important as food fish and for several years, they have been used in local fish culture where they proved to be fast growing protein source. They exhibit many qualities that make them suitable for culture.

These include their hardy nature, disease resistance, tolerance to poor water quality, consumer acceptability, high fecundity, nutritional efficiency and attainment of large size within a short time (Hecht *et al.*, 1988; Haylor, 1989; Salami and Fagbenro, 1993). Salinity is an important factor affecting the distribution of aquatic organisms. Depending on the concentration, it is known to affect the survival of the fish and other aquatic organisms by interacting with temperature, dissolved oxygen and other environmental factors (Eddy,

1981). The effects of physico-chemical parameters on cultured fish are of paramount importance and should be assessed in order to evaluate the optimum requirements and tolerances. The maximum and minimum levels of various environmental parameters such as: temperature, salinity, pH, dissolved oxygen (DO) concentrations, and ammonia among others are important in formulating culture practices of the organisms.

The osmotic pressure of water increases with increasing salinity (Boyd, 1982). Salinity therefore has been observed to influence growth, survival and production potentials of fish by its effect on osmoregulatory and metabolic activities of aquatic animals. At unfavourable concentrations, it can cause stress or even death of fish depending on the species (Eddy, 1981). Most estuarine and marine fish species which are euryhaline have optimal salinity range within which the scope for growth and other biological activities are optimal. Hence fish could be classified on the basis of their ability to tolerate environment differing in salinity. Thus, those that can tolerate wide range of salinity are termed euryhaline, while those with narrow range of salinity are stenohaline. Salinity tolerance tests with fish have been performed using different saline solution. Salinity tolerance tests with fish have been performed using different saline solutions for example, sodium chloride (Matern, 2001), diluted seawater (Chen and Chen, 2000) and synthetic seawater (Patridge and Jenkins, 2002). Study conducted indicated that mature catfish, *C. gariepinus* (0.6 – 1.5 kg) tolerated 10 ppt (25.6% seawater) for 100 hours with no sign of stress and with acclimatization the fish was able to tolerate 20ppt salinity (51.3%) seawater (Clay, 1977). Chervinski (1984) in his study of the salinity tolerance of young catfish (*C. gariepinus*) indicated that 95% of those on direct transfer tolerated 25% seawater (9.5%) but that no fish survived 30% seawater (11.7%), even though gradual increase. The medium lethal salinity (MLS<sub>50</sub>) was 8 ppt using abrupt transfer and 10 ppt by gradual transfer (Iyagi, 1986). Britz and Hecht (1989) monitored the survival and growth of larvae of *C. gariepinus* between 0 and 5 ppt salinity.

The fingerlings of *H. bidorsalis* were found to resist changes from freshwater (0 ppt) up to 10 ppt (isosmotic) salinity without mortality (Fagbenro *et al.*, 1993). The present study was conducted to assess the tolerance of the post fry to salinity ranges which many occur in the natural habitat.

## MATERIALS AND METHODS

**Hybrid Procurement:** Brood *C. gariepinus*, ♀ and *H. bidorsalis*, ♂ used for the study were obtained from a private fish farm at Aluu. The study was carried out in the hatchery and laboratory of African Regional Aquaculture Centre (ARAC) at Aluu, Port Harcourt from September to December, 2007. Gravid females of *C. gariepinus* (400 – 800 grams) were injected with ovaprim, a synthetic analogue of gonadotropin releasing agent. Dosage was calculated based on the manufacturer's recommendation of 0.5 ml/kg body weight of fish and administered intramuscularly in the dorsal muscle mass as described by Viveen *et al.* (1985). Ovulated eggs were stripped from the induced females into plastic bowl. Brood males of *H. bidorsalis* (0.6 – 1.3kg) were sacrificed and the testes dissected out. Incisions were made along the edges of the testes using a clean razor blade in order to extract the milt. The milt was squeezed out into a 0.9% saline solution. This was gently and thoroughly mixed. Fertilization was effected with addition of freshwater to the mixture of the eggs and milt, and by gently stirring with a plastic spoon.

The fertilized eggs were incubated in trays (30 x 45 cm<sup>2</sup>) in a rectangular concrete tank containing freshwater. The hatched larvae were nursed in the concrete tank. The larvae were fed with *artemia* after the absorption of the yolk sacs up to the seventh day. The fry was then fed with ground and sieved Coppens feed (crude protein 45%) at 5% body weight for another week. Part of this was used for the salinity tolerance test for the fourteen day-old fry. The leftover in the concrete tank was fed with the same feed for another one week. These were used for the salinity tolerance test for the twenty one day-old fingerlings.

**Saline Solution:** Six salinity levels were prepared by weighting 2, 4, 6, 8, 10 and 12 g of sodium chloride with (Laptop balance Yamato LE 180, Yamato Scientific Company Limited, Tokyo, Japan) and dissolving each in a litre of freshwater to obtain 2, 4, 6, 8, 10 and 12 ppt, respectively. Freshwater was used as control (0ppt).

**Salinity Tolerance test:** Fingerlings each of fourteen day-old ( $1.22 \pm 0.10$  cm) and twenty one day-old ( $2.02 \pm 0.2$  cm) were introduced into each of the following test solutions 4, 6, 8, 10 and 12 ppt. The tolerance test was conducted in a five litre plastic aquarium containing three litre of the test solution. Each treatment level was replicated three times. The aquaria were covered with mosquito netting to prevent the fish from jumping out. Freshwater (0 ppt) was used as control. The fingerlings were not fed throughout the experimental period. Mortality was monitored at hourly interval for 96 hours. Dead fingerlings were removed, counted and recorded. Water temperature, dissolved oxygen and pH were determined twice a day. The test solution in each aquarium was changed daily. A fingerling was considered dead when opercular movement stopped and the fingerling did not react to gently prodding with a glass rod.

**Water Quality:** Mercury in glass thermometer was used to measure the water temperature during the salinity tolerance test for the fry and fingerlings of the African catfish hybrid. Hydrogen ion concentration (pH) was determined with handheld pH meter (Hannah Instruments Portugal, HA Model 191) which was calibrated against standard buffer solutions with pH values of 4, 7 and 10. Calibrations were carried done at each pH reading. Dissolved Oxygen was done with the Winkler method (APHA, 1985). Salinity was determined by a refractometer (ATAGOS/MILLE). The refractometer was first standardized with water from ARAC borehole to get reading of 0 ppt, before salinity of test solutions were determined.

**Data Analysis:** Probit analysis model (Finney, 1984) was used to determine the median lethal salinities (MLS) and median lethal times MLTs). Mortality at the various concentrations and duration were subjected to ANOVA and differences among means separated by Duncan multiple range test (Wahua, 1999). All analysis were done using Statistical Package for the Social Sciences, SPSS version 15 for Window.

## RESULTS AND DISCUSSION

The water quality variables in the test aquaria were not different for the various salinity levels in the two life stages (Tables 1 and 2). The fourteen day-old fry that exposed to 0 – 6 ppt had 90%, 87.5% and 10% survival at the end of the 96 hour test period. Those subjected to 8 ppt and 10 ppt had 100% dead within 48 and 12 hours, respectively. The median lethal salinity at 96 hour ( $MLS_{50}$ ) was 4.4 ppt which was about half the value for 6 hour (Table 3). However, the mortality at the exposure duration 48 – 96 hour was higher ( $p < 0.05$ ) from 3 – 36 hour for 14 day-old, whereas for 21 day-old mortality at 12 – 96 hour was similar but greater ( $p < 0.05$ ) than those at 3 – 6 hour (Table 3). The median lethal time ( $MLT_{50}$ ) at 10 ppt was 11.65 hours which was about half the value for 6 ppt (Table 4). The percentage mortality of the 14 day-old larvae in this study declined with increased in salinity (Table 5). However, the survival at 48 – 96 hours were higher ( $p < 0.05$ ) compared to 3 – 36 hours (Table 5). For the 21 day-old, the 48 hour  $MLS_{50}$  was 4.79 (Table 6) and  $MLT_{50}$  at 8 ppt, 8.55 hour (Table 7). Percentage mortality increased with increase in the test salinity and at the exposure duration, and was similar at 12 – 96 hour and greater ( $p < 0.05$ ) than those at 3 – 6 hour (Table 8).

Salinity is one of the most important environmental factors exerting selective pressures on aquatic organisms and that organisms respond to varying salinity by either spending their life cycle in a single habitat where salinity is stable or variable; while others undergo ontogenic migrations with successive stages based on salinity regimes (Varsamos *et al.*, 2005).

The ability of each ontogenic stage to cope with salinity depends on the capacity to osmoregulate. Brett (1979) observed that highest growth rates of various fish species relative to salinity clustered around  $0 - 10 \pm 2.00$  or  $28 - 35$  ppt. These clusters correspond roughly to three ecological groupings: freshwater, stenohaline anadromous species; euryhaline and stenohaline marine species. The 21 day-old larva had higher survival when compared with the 14 day-old at  $0 - 6$  ppt; however, at 8 and 10 ppt where 100% mortality was recorded in the 21 day-old it occurred at one-fourth and one-third respectively the time for that in the 14 day-old indicating the ontogenetic variation in salinity tolerance as reported in a number of fish species with increase in age (Murashige *et al.*, 1991; Haddy and Pankhurst, 2004; Moustakas, *et al.*, 2004). However, the more rapid rate of death at 8 and 10 ppt in the 21 day-old is difficult to explain. The increased mortality ( $p < 0.05$ ) of the larvae in this study with increase in salinity was due to efflux of water out of the fish which has also been reported in larvae (Fashina-Bombata and Busari, 2003). The mortality at the exposure duration 48 – 96 hour was higher ( $p < 0.05$ ) from 3 – 36 hour for 14 day-old, whereas for 21 day-old mortality at 12 – 96 hour was similar but greater ( $p < 0.05$ ) than those at 3 – 6 hour and may suggest at that the larvae were able to attain internal homeostasis hence the rate of death did not differ as the exposure progressed with time.

Teleost embryos have developed strategies for coping with salinity challenge which includes formation of impermeable chorion and ion pump such as chloride cells, CC on the epithelium of embryos (Lin *et al.* 1999; Kaneko *et al.*, 2002). CC also played important roles in hydromineral and ionic homeostasis before tissues and organs for osmoregulation are developed (Ayson *et al.*, 1994). Besides, the 21 day-old may have developed tissues/organs in even miniature forms that have helped them cope better than the 14-day old. In some species such as *Mugil cephalus* (Lee and Menu, 1981), grouper- *Epinepalus coioides* (Yeh *et al.*, 1995) and black porgy- *Acanthopagrus schlegelii* (Chu *et al.*, 1999) salinity tolerance seem to

improve with age. The tolerance range of the larvae,  $0 - 6$  ppt with the optimum  $2 - 4$  and  $2 - 6$  ppt for 14 and 21 day-old, respectively indicated that the tolerance ranges were very narrow. The 96 hour  $MLS_{50}$  for the 21 day-old was 4.43 ppt; 48 hour  $MLS_{50}$  for 14 and 21 day-old were 4.52 and 4.79 ppt, a difference of 0.27 ppt suggesting the value was very narrow. Although the difference between the  $MLS_{50}$ , that between the  $MLT_{50}$  for 14 and 21 day-old larva (6.35 hour) at 8 ppt clearly showed that the older larvae was more tolerant. Decreasing tolerance with age is characteristic of freshwater stenohaline fishes possibly due to the physiological high energy cost of maintaining internal hydromineral and ionic balance in the face of increasing salinity (Kilambi, 1980). Hence several studies have confirmed that the early life stages of *Clarias and Heterobranchus* sp. hardly survive and grow for a long period beyond salinity above 10 ppt (Iyaji, 1986; Fagbenro *et al.*, 1993). Under salinity challenge it appears the body fluid osmolality of the larvae are raised relative to that of the external medium. Thus their tolerance is probably limited by the maximum osmotic pressure of the body fluid in which the cells can function (Maceina *et al.*, 1980).

The fingerlings of the parent stocks, *H. bidorsalis* and *C. gariepinus* had been classified as freshwater stenohaline fishes (Fagbenro *et al.*, 1993; Oladosu *et al.*, 1999) which is true of larvae from their hybrid. A study by Odiete and Jacob (1985) indicated that the skin of the parents stock, *Clarias* sp. was non-keratinized with 25 – 32 layers of cells which make it permeable to  $Na^+$  and  $Cl^-$  compared with that of another catfish, *Chrysichthys* sp. having keratinized layers that are impermeable to water and electrolytes. Hence, the fish tolerates wide salinity ranges even as was evidenced in fingerling *C. nigrodigitatus* that tolerated up to 22 ppt salinity (Anyanwu, 1991) unlike the clariids. The nature and functions of the integument of the larvae in this study may be similar to that of the parents and hence their response.

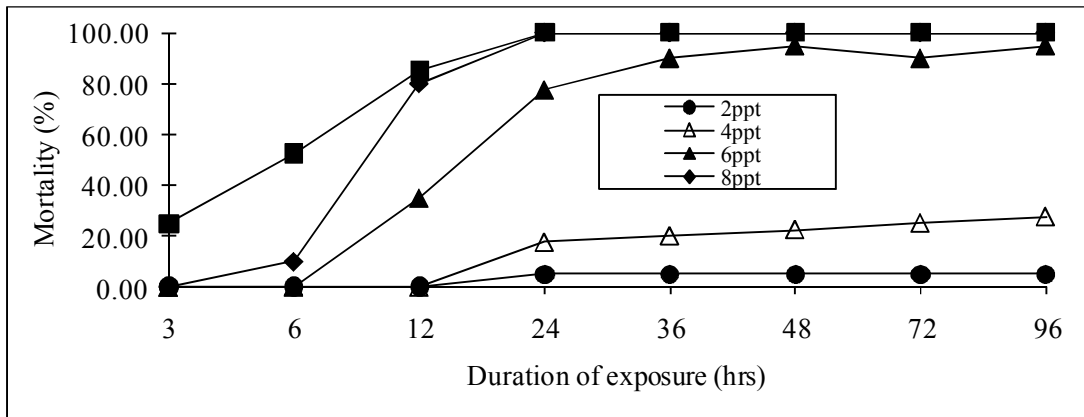
Laboratory studies on the effects of changing salinity on the early life stages of a number of freshwater fish species are meant to

**Table 1: Water quality variables of the various test salinities for 14 day old *C. gariepinus* larvae**

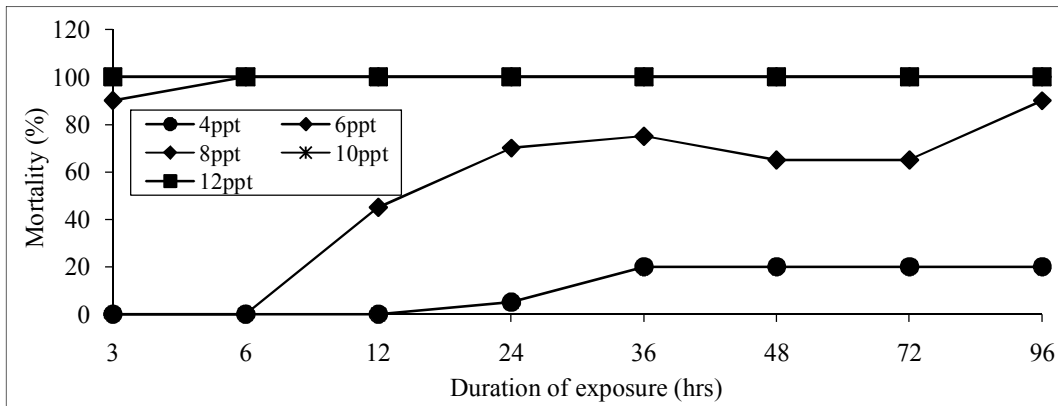
Salinity (‰)	Temperature (°C)		Dissolved oxygen (mg/l)		pH	
	Range	Mean	Range	Mean	Range	Mean
0	25.5-27.5	26.6±0.74	5.2-6.2	5.8±0.37	5.6-6.2	5.8±0.26
2	26.0-27.5	26.7±0.47	5.8-6.2	5.9±0.16	5.8-6.2	6.0±0.14
4	25.5-27.5	26.7±0.69	6.0-6.2	6.1±0.10	5.8-6.4	6.2±0.21
6	26.5-27.5	27.0±0.40	6.2-6.4	6.3±0.10	5.8-6.4	6.1±0.19
8	26.5-27.5	27.2±0.38	5.8-6.4	6.1±0.22	5.8-6.4	6.2±0.23
10	26.0-27.0	26.5±0.34	6.0-6.4	6.1±0.16	5.8-6.2	5.1±0.16

**Table 2: Water quality parameters of the various test salinities for 21 day old *C. gariepinus* larva**

Salinity (‰)	Temperature (°C)		Dissolved oxygen (mg/l)		pH	
	Range	Mean	Range	Mean	Range	Mean
0	26.0-27.0	26.5±0.46	6.0-6.2	6.1±0.10	5.6-6.2	5.9±0.22
4	25.5-27.0	26.1±0.68	5.8-6.2	6.1±0.17	5.8-6.2	6.1±0.17
6	25.5-27.0	26.5±0.76	5.8-6.2	5.9±0.16	5.6-5.8	5.7±0.10
8	26.5-27.0	26.2±0.34	5.6-6.2	5.8±0.21	5.4-6.2	5.8±0.29
10	26.5-27.0	26.5±0.44	6.2-6.8	6.4±0.24	6.2-6.8	6.4±0.024
12	25.0-27.0	25.8 ±0.68	6.2-6.6	6.4 ±0.14	6.2-6.6	6.4 ±0.16



**Figure 1: Mortality (%) of 14-day old larvae of *Clarias gariepinus* exposed graded levels of salinity under laboratory conditions**



**Figure 2: Mortality (%) of 21-day old larvae of *Clarias gariepinus* exposed graded levels of salinity under laboratory conditions**

**Table 3: Median Lethal salinity, MLS and associated 95% lower and upper confidence bounds (limits) of 14 day- old hybrid fry (*C. gariepinus* ♂ x *H. bidorsalis* ♀)**

Duration (hours)	MLS <sub>5</sub>	MLS <sub>50</sub>	MLS <sub>85</sub>	MLS <sub>90</sub>	MLS <sub>95</sub>	MLS <sub>99</sub>
<b>6</b>	7.85 (6.68-8.41)	9.80 (9.41-10.34)	11.02 (10.45-11.25)	11.31 (10.67-12.72)	11.74 (10.99-13.43)	12.55 (11.58-14.77)
<b>12</b>	5.19 (4.30-5.75)	7.38 (6.99-7.77)	8.75 (8.30-9.43)	9.08 (8.58-9.85)	9.56 (8.98-10.49)	10.47 (9.72-11.70)
<b>24</b>	2.96	5.36	6.87	7.23	7.76	8.76
<b>48</b>	2.06 (-2.02-3.30)	4.52 (241.6) (3.28-5.93)	6.07 (4.97-9.23)	6.44 (5.26-10.12)	6.98 (5.67-11.47)	8.00 (6.38-14.04)
<b>72</b>	2.00 (-1.42-3.17)	4.48 (3.36-5.71)	6.03 (5.00-8.68)	6.40 (5.30-9.12)	6.98 (5.67-11.47)	7.97 (6.44-12.98)
<b>96</b>	1.94 (-0.95-3.05)	4.43 (1.43-5.52)	6.00 (5.04-8.23)	6.37 (5.34-8.95)	6.92 (5.16-10.05)	7.95 (6.5042.14)

**Table 4: Median lethal time, MLT and associated 95% lower and upper confidence bounds (limits) of 14 day- old hybrid fry (*C. gariepinus* ♂ x *H. bidorsalis* ♀)**

Duration (hours)	MLT <sub>5</sub>	MLT <sub>50</sub>	MLT <sub>85</sub>	MLT <sub>90</sub>	MLT <sub>95</sub>	MLT <sub>99</sub>
<b>2</b>	38.7 (13.55-58.96)	172.99 (124.97-394.41)	257.82 (177.34-640.69)	277.88 (189.59-699.06)	307.61 (207.71-785.64)	363.39 (241.60-948.12)
<b>4</b>	24.60 (12.30-41.53)	132.77 (106.97-201.84)	200.93 (153.81-329.80)	217.05 (164.92-360.23)	240.94 (181.32-405.40)	285.76 (211.99-490.22)
<b>6</b>	21.4	29.99	62.47	70.15	81.53	102.89
<b>8</b>	12.02 (9.34-13.89)	17.48 (15.77-19.37)	20.92 (19.07-23.57)	21.74 (19.79-24.63)	22.94 (20.84-26.21)	25.20 (22.73-29.25)
<b>10</b>	8.74	15.77	11.65	13.49	13.92	14.57

**Table 5: Cumulative mortality associated 95% upper and lower confidence bounds of 14d-old larvae of catfish hybrid exposed to 9a) graded salinity at (b) various durations**

(a) Salinity (‰)	Cumulative mortality	95%confidence bounds		(b) Duration	Cumulative mortality	95%confidence bounds	
		Upper	Lower			Upper	Lower
4	2.97 <sup>c</sup>	1.98	3.96	3	1.20 <sup>d</sup>	-0.10	2.50
6	1.47 <sup>b</sup>	0.47	2.47	6	1.26 <sup>d</sup>	0.02	2.48
8	3.50 <sup>c</sup>	2.52	4.49	12	2.60 <sup>c</sup>	1.30	3.90
10	8.03 <sup>a</sup>	7.04	9.03	24	5.60 <sup>e</sup>	4.34	6.84
4	2.97 <sup>c</sup>	1.98	3.96	36	4.05 <sup>b</sup>	2.79	5.31
6	1.47 <sup>b</sup>	0.47	2.47	48	6.00 <sup>a</sup>	4.74	7.26
8	3.50 <sup>c</sup>	2.52	4.49	72	6.45 <sup>a</sup>	5.19	7.71
10	8.03 <sup>a</sup>	7.04	9.03	96	6.50 <sup>a</sup>	5.24	7.76

Means with similar alphabets in the same are not significantly different ( $p > 0.05$ ) for salinity and duration respectively.

**Table 6: Median Lethal salinity, MLS and associated 95% lower and upper confidence bounds (limits) of 21 d- old hybrid fry (*C. gariepinus* ♂ x *H. bidorsalis* ♀)**

Duration (h)	MLS <sub>5</sub>	MLS <sub>50</sub>	MLS <sub>85</sub>	MLS <sub>90</sub>	MLS <sub>95</sub>	MLS <sub>99</sub>
6	5.72 (4.91-6.18)	7.18 (6.83-7.51)	8.09 (7.73-8.64)	8.31 (8.19-8.94)	8.6 (8.19-9.38)	9.24 (8.68-10.24)
12	4.74 (3.80-5.35)	7.19 (6.78-7.60)	8.73 (8.24-9.4)	9.09 (6.55-9.91)	9.63 (9.00-0.61)	10.64 (9.84-11.96)
24	4.53 (3.35-5.00)	5.66 (5.29-5.93)	6.38 (5.09-6.905)	6.55 (6.23-7.24)	6.80 (6.42-7.68)	7.27 (6.77-8.59)
48	2.25 (2.42-3.72)	4.79 (4.45-5.12)	5.79 (5.40-6.34)	5.99 (5.59-6.65)	6.33 (5.80-7.13)	6.97 (6.39-8.04)
72*						
96 *						

\*Percentage responding in all salinities is the same.

**Table 7: Median Lethal Time, MLT and associated 95% lower and upper confidence bounds (limits) of 21 day-old hybrid fry (*C. gariepinus* ♂ x *H. bidorsalis* ♀)**

Salinity (‰)	MLT <sub>5</sub>	MLT <sub>50</sub>	MLT <sub>85</sub>	MLT <sub>90</sub>	MLT <sub>95</sub>	MLT <sub>99</sub>
<b>4</b>	19.05 (9.15 -26.34)	65.06 (59.57 -71.53)	96.06 (85.72-105.62)	100.92 (91.67-133.92)	111.08 (100.41-126.29)	130 (116.67-149.60)
<b>6</b>	29.79 (18.97-52.57)	34.10 (24.05 -87.83)	74.36 (47.11-423.31)	83.88 (53.72-512.87)	98.00 (62.09-164.46)	124.47 (78.21-898.21)
<b>8</b>	-11.13	8.55	18.6	21.40	25.45	33.01
<b>10*</b>						
<b>12*</b>						

\*Percentage responding in all salinities is the same.

**Table 8: Cumulative mortality associated 95% upper and lower confidence bounds of 14 day old larvae of catfish hybrid exposed to 9a) graded salinity at (b) various durations**

(c)	Salinity (‰)	Cumulative mortality	95% confidence bounds		(d) Duration	Cumulative mortality	95%confidence bounds	
			Upper	Lower			Upper	Lower
<b>4</b>		1.00d	0.75	1.25	3	3.67e	3.38	3.95
<b>6</b>		5.56c	5.32	5.81	6	4.83d	4.55	5.12
<b>8</b>		8.88b	8.63	9.12	12	5.50b	5.21	5.79
<b>10</b>		10.00a	9.75	10.25	24	6.25	5.96	6.54
<b>12</b>		10.00a	9.75	10.25	36	6.50	6.21	6.79
<b>4</b>		1.00d	0.75	1.25	48	6.83a	6.55	7.12
<b>6</b>		5.56c	5.32	5.81	72	6.83a	6.55	7.12
<b>8</b>		8.88b	8.63	9.12	96	6.83a	6.55	7.12

Means with similar alphabets in the same are not significantly different ( $p > 0.05$ ) for salinity and duration respectively



assess the possibility of introducing such species into non-natural ecosystems especially with the salinization of rivers and wetlands which is a major environmental concern in many parts of the world (Williams, 1987). A most common approach to laboratory measure of salinity tolerance is the concentration that is lethal to 50% of the individuals (the LC<sub>50</sub>) over a period of time usually 48 – 98 hours. Kefford *et al.* (2004) noted that the values are usually determined from three broad methods: direct transfer (direct LC<sub>50</sub>), slow acclimation (slow LC<sub>50</sub>) and early life stage or early LC<sub>50</sub> which gives very large differences in the LC<sub>50</sub>. Besides, the studies are conducted without other stressors, quite contrary to what obtain in natural ecosystems. The authors observed that “survival in laboratory experiment may not predict survival in nature, let alone the maximum salinity that could support a self sustaining population” Besides, although laboratory measurements of sublethal effects are possible, it is difficult to relate to the salinity at which a species can maintain a self sustaining population in nature (Brinkhurst *et al.*, 1983). Hence, although the larvae in this study had optimum salinity tolerance range at 0 – 4 ppt, in the natural environment with the vagaries in physico-chemical and other environmental factors the optimum range may be greatly influenced. The farmer intending to introduce the larvae to the optimum salinity in the natural environment should carry out a field validation study as suggested by Connell *et al.* (1999) so as avoid heavy losses that may discourage such attempts and the associated benefits.

## REFERENCES

- ANYANWU, P. E. (1991). Influence of salinity on survival of fingerlings of the estuarine catfish *Chrysichthys nigrodigitatus* (Lap.). *Aquaculture*, 99: 157 – 165.
- APHA (1985). *Standard Methods for the Examination of Water and Waste Water*, 15<sup>th</sup> Edition, American Public Health Association (APHA), Washington DC, USA.
- AYSON, F. G., KANEKO, T., HASEGAWA, S. and HIRANO, T. (1994). Development of mitochondrion-rich cells in the yolk-sac membrane of embryos and larvae of tilapia, *Oreochromis mossambicus*, in freshwater and seawater. *Journal of Experimental Zoology*, 270: 129 – 135.
- BOYD, C. E. (1982). Water management for pond fish culture. Pages 1 – 231. *In: Development of Aquaculture and Fisheries Science*, Volume 9, Elsevier Scientific Publications, Amsterdam.
- BRETT, J. R. (1979). Environmental factors and growth. Pages 599 – 675. *In: HOAR, W. S. and RANDALL, D. J. (Eds). Fish Physiology*, Volume, 8, Academic Press, New York.
- BRINKHURST, R. O., CHAPMAN, P. M. and FARRELL, M. A. (1983). A comparative study of respiration rates of some aquatic oligochaetes in relation to sublethal stress. *International Review of Hydrobiology*, 68: 683 – 699.
- BRITZ, P. J. and HECHT, T. (1989). Effect of salinity on the growth and survival of African sharp tooth catfish, *Clarias gariepinus* larvae. *Journal of Applied Ichthyology*, 5: 194 – 202.
- CHEN, J. C. and CHEN, W. C. (2000). Salinity tolerance of *Haliotis diversicolor supertexta* at different salinity and temperature levels. *Aquaculture*, 18: 191 – 203.
- CHERVINSKI, J. (1984). Salinity tolerance of young *Clarias lazera* (Burch.). *Journal Fish Biology*, 25(2): 147 – 149.
- CHU, Y. T., YEH, S. L., HSEU, J. R. and TING, Y. Y. (1999). Effect of salinity on egg development of black porgy, *Acanthopagrus schlegeli*. *Journal of Fisheries Society of Taiwan*, 26(1): 35 – 41.
- CLAY, D. (1977). Preliminary observations on salinity tolerance of *Clarias gariepinus* from Israel. *Israeli Journal of Aquaculture (Bamidgeh)*, 29: 102 – 109.
- CONNELL, D., LAM, P., RICHARDSON, B. and WU, R. (1999). *Introduction to Ecotoxicology*. Blackwell Science, Oxford.
- EDDY, F. B. (1981). Effect of salinity on osmotic and ionic regulation in fish. Pages 77 – 102. *In: PICKERING, A. D. (Ed.) Stress and Fish*. Academic Press, New York.

- FAGBENRO, O. A., ADEDIRE, C. O., OWOSENI, E. A. and AYOTUNDE, E. O. (1993). Studies on the biology and aquaculture potential of feral catfish, *Heterobranchus bidorsalis* (Geoffrey St. Hilaire) Clariidae. *Tropical Zoology*, 6: 67 – 79.
- FASHINA-BOMBATA, H. A. and BUSARI, A. N. (2003). Influence of salinity on the developmental stages of African catfish, *Heterobranchus longifilis* (Val.). *Aquaculture*, 224: 213 – 222.
- FINNEY, D. J. (1984). *Probit Analysis*. 3<sup>rd</sup> Edition, Cambridge University Press, New York.
- HADDY, J. A. and PANKHURST, N. W. (2000). The effect of salinity on reproductive development, plasma steroid levels, fertilization and egg survival in black bream, *Acanthopagrus butcheri*. *Aquaculture*, 188: 115 – 131.
- HAYLOR, G. S. (1989). The case of the African catfish, *Clarias gariepinus* (Burh. 1822) Clariidae: a comparison of the relative merits of tilapine fishes, especially *Oreochromis niloticus* (L.) and *Clarias gariepinus* for African aquaculture. *Aquaculture and Fisheries Management*, 20(3): 279 – 285.
- HECHT, T., UYS, W. and BRITZ, P. J. (1988). The culture of the African sharp tooth catfish, *Clarias gariepinus* in South Africa. Pages 1 – 136. In: *South Africa National Scientific Programme Report, Number 153*, Foundation for Research Development, Pretoria, South Africa.
- IYAJI, F. O. (1986). *The Effects of Environmental Factors with Special Reference to pH, Salinity and Dissolved Oxygen on Survival of Clarias gariepinus (Burchell 1822) Fry and Fingerlings*. M. Tech. Thesis, Rivers University of Science and Technology, Port Harcourt, Nigeria.
- KANEKO, T., SHIRAISHI, K., KATOH, F., HASEGANA, S. and HIROI, S. (2002). Chloride cells during early life stages of fish and their functional differentiation. *Fishery Science*, 68: 1 – 9.
- KEFFORD, B. J., PAPAS, P. J., METLING, L. and NCEGEGODA, D. (2004). Do laboratory tolerances of freshwater animals correspond with field salinity? *Environmental Pollution*, 129: 355 – 362.
- KILAMBI, R. V. (1980). Food consumption, growth and survival of grass carp, *Ctenopharyngodon idella* at four salinities. *Journal of Fish Biology*, 17: 613 – 618.
- LEE, C. S. and MENU, B. (1981). Effect of salinity on egg development and hatching in grey mullet, *Mugil cephalus*. *Journal Fish Biology*, 19: 179 – 188.
- LIN, H. C., HWANG, P. P. and LEE, T. H. (1999). Mitochondria-rich cells in the embryonic sac of *Oreochromis niloticus*. *Journal of Fish Biology*, 54: 648 – 655.
- MACEINA, M. J. NORDLE, F. G. and SHIREMAN, J. V. (1980). The influence of salinity on oxygen consumption and plasma electrolytes in grass carp, *Ctenopharyngodon idella*. *Journal of Fish Biology*, 16: 613 – 619.
- MATERN, S. A. (2001). Using temperature and salinity tolerance to predict the success of the *Shimiofuri gobi*, a recent invader into California. *Transactions of American Fisheries Society*, 130: 592 – 599.
- MOUSTAKAS, C. T., WANATABE, W. O. and COPELAND, K. A. (2004). Combined effects of photoperiod on growth, survival and osmoregulation of larval, Southern flounder, *Paralichthys lethostigma*. *Aquaculture*, 229: 179 – 182.
- MURASHIGE, R., BASS, P., WALLACE, L., MOLNAR, A., EASTMAN, B., SATO, V., TAMARU, C. and LEE, C. S. (1991). The effect of salinity on the survival and growth of striped mullet (*Mugil cephalus*) in the laboratory. *Aquaculture*, 96: 249 – 254.
- ODIETE, S. S. and JACOB, S. S. (1985). Osmotic responses of species of *Clarias* and *Chrysichthys* in freshwater and diluted seawater. *Biologia Africana*, 2(2):
- OLADOSU, G. A., BUSARI, A. N., UKA, A., OLADOSU, O. O. and AYINLA, O. A. (1999). Influence of salinity on the developmental stages in African catfish (*Clarias gariepinus*). *Journal of Applied Science and Environmental Management*, 2(1): 29 – 34.

**Salinity tolerance of larvae of *Clarias gariepinus* (♀) x *Heterobranchus bidorsalis* (♂) 1664 hybrid**

- PATRIGDE, G. J. and JENKINS, G. I. (2002). The effect of salinity on growth and survival of juvenile black bream, *Acanthopagrus butcheri*. *Aquaculture*, 210: 219 – 230.
- SALAMI, A. A. and FAGBENRO, O. A. (1993). The production and growth of *Clarias gariepinus* in ponds. *Israeli Journal of Aquaculture (Bamidgeh)*, 45(1): 18 – 25.
- TEUGELS, G. G. (1984). Clariidae. Pages 66 – 101. In: DAGET, J., GOOSE, J. P. and THYSVANDEN AUDENAERDE, D. F. E. (Eds.). *Checklist of the Freshwater Fishes of Africa*. CLOFFA II.
- VARSAMOS, S., NEBEL, C. and CHARMANTIER, G. (2005). Ontogeny of postembryonic fish: a review. *Comparative Biochemistry and Physiology A*, 141(4): 401 – 429.
- VIVEEN, W. J. A. R., RICHTER, C. J. J., Van DODT, P. G. W. J., JANSEEN, J. A. L. and HUISMAN, E. A. (1985). *Practical Manual for the Culture of the African Catfish (Clarias gariepinus)*. Ministry for Development Co-operation, Section for Research and Technology, The Hague, The Netherlands.
- WAHUA, T. A. T. (1999). *Applied Statistics for Scientific Studies*. Afrilink Publishers, Aba, Nigeria.
- WILLIAMS, W. D. (1987). Salinization of rivers and streams: an important environmental hazard. *Ambio*, 16: 313 – 327.
- YEH, S. L., CHU, Y. T., HSEU, J. R. and TING, Y. Y. (1995). Effect of salinity on development of the grouper, *Epinephelus coioides* embryos before and post-organogenesis. *Journal of Taiwan Fishery Research*, 3: 133 – 142.