PREVALENCE OF AFRICAN ANIMAL TRYPANOSOMIASIS AND THE ASSOCIATED BLOOD PARAMETERS AMONG TRADE RUMINANTS AT OBOLLO-AFOR MARKET

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ABSTRACT

African trypanosomiasis is a haemoprotozoan disease of animals and humans. In animals particularly, the disease directly hinders production and therefore impedes socioeconomic development of Sub-Saharan Africa. The prevalence of trypanosome infection was investigated in trade goats and sheep sold at Obollo-Afor market in Enugu State, Nigeria. Wet and thin films, haematocrit test (HCT) and buffy coat methods (BCT) were used to detect trypanosomes in jugular blood of the animals. The packed cell volume (PCV), red blood cell (RBC), and white blood cell (WBC) counts were also determined. A total of 200 goats and 100 sheep were examined within four months (October 2019 to January 2020). The prevalence rates of trypanosomes in the 200 goats and 100 sheep examined were 0(0.0 %) and 2(0.7 %) respectively. The RBC, WBC, and Hb of the two infected animals were significantly (p<0.05) lower than the uninfected. The average PCV of infected animals (20.65 ± 5.44%) was also lower than that of uninfected (34.17 ± 0.29%) although, it was not statistically significant (p>0.05).

Keywords: Trypanosomiasis, Prevalence, Haematology, Trade ruminants, Enugu State

INTRODUCTION

African animal trypanosomiasis (AAT) is a livestock disease caused by trypanosome species and frequently transmitted by the vector, tsetse flies. The representative species of the trypanosomes transmitted *en route* the infected saliva (salivaria group) of the vector, *Glossina* spp., in Sub-Saharan Africa are *Trypanosoma brucei, T. congolense and T. vivax* (Cayla *et al.,* 2019). *T. brucei* infection is zoonotic but because of preference of the vector, *Glossina* spp., to cattle for its blood

ISSN: 1597 – 3115 www.zoo-unn.org meal, the infection is less in human than in the cattle (Liana *et al.,* 2020). In addition to cattle, sheep, goats and particularly pigs have been identified as reservoirs of human infective trypanosomiasis in Sub-Saharan Africa (Desquesnes *et al.,* 2022).

The economic losses due to the infection are large and diverse. In Nigeria, there are several millions of cattle, goats, sheep, pigs, and thousands of camels and donkeys which are predisposed to trypanosome infections (Lawal-Adebowale, 2012).

Trypanosomiasis also infect livestock and wild animals in tropical and subtropical areas of the globe (Gutierrez et al., 2006; Giordani et al., 2016; Desquesnes et al., 2022). Annual losses due to trypanosomiasis is put at 5 billion US dollars (Giordani et al., 2016; Stijlemans et al., 2018). In tsetse infested areas of Sub-Saharan Africa, vast number of these animals are predisposed to this disease which hinder their efficiency and lead to reduced quantity and quality of animal products, increased management cost and gross economic losses (Matovu et al., 2020; Ngongolo et al., 2020). The predisposition of these animals are influenced by myriads of factors including demographic, environmental, entomological, livestock management, among others, which in turn influence tsetse populations and eventual prevalence of trypanosome infections (Bouyer et al., 2013; Kizza et al., 2021).

The disease can be reduced, and eventually eliminated/eradicated with the proper control measures in place (Diall *et al.*, 2017; Isaac *et al.*, 2017). Concurrent measures targeting both the parasites and the vectors have been put forward (Liana *et al.*, 2020). Ultimately cost effective control of trypanosomiasis will lead to economic boom and render better health to both livestock and humans (Matovu *et al.*, 2020).

The aim of this study was to investigate the current prevalence status and intensity of trypanosomiasis of trade goats and sheep at Obollo-Afor market in Enugu State, Nigeria. In addition, haematological parameters were also evaluated to determine the health status of both the infected and uninfected animals.

MATERIALS AND METHODS

Study Area: This study was conducted in Obollo-Afor, the headquarters of Udenu Local Government Area (LGA) of Enugu State, South East, Nigeria (Figure 1). It has an area of 271.3 km² and a population of 256,500 persons as at 2022 (City Population, 2023). The animal market is located at latitude 6^{0} 54' 56" north and longitude 7^{0} 30' 55" east (Agina *et al.*, 2021).

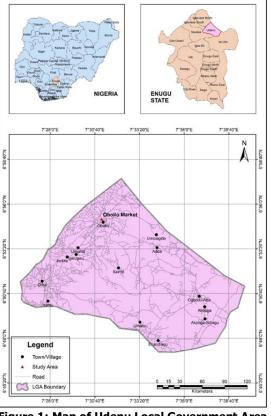


Figure 1: Map of Udenu Local Government Area showing the study area. *Source:* Soil Survey Unit, Department of Soil Science, University of Nigeria, Nsukka

Target Population: The target populations were trade goats and sheep transported from the Northern states of Nigeria to Obollo-Afor for sale. Two hundred (200) goats and one hundred (100) sheep were randomly selected for the study, between the months of October 2019 to January 2020).

Sample Collections: The animal market was visited twice every week during the four-month period of study for sample collection. Two mililitres (2 ml) of blood was collected from each animal by puncturing the jugular vein using 23-gauge needle attached to 2 ml. The collected blood sample was dispensed into a properly encrypted sodium ethylenediamine tetra-acetic acid (EDTA) bottle for subsequent transmission to the laboratory for determination of the haematological profile. The analyses were conducted at Parasitology Laboratory of the Department of Parasitology and Entomology, Faculty of Veterinary Medicine, University of Nigeria, Nsukka.

Examination of the Blood Samples for Parasites: Wet preparation, thin blood films and buffy coats were examined for the presence or absence of trypanosomes. To make a wet preparation, a drop of blood was placed on a clean glass slide and covered with 22 x 22 mm cover slip. The slides were examined using x40 objective lens of the microscope for the presence of trypanosomes. Thin blood smears were prepared, fixed in absolute methanol, Giemsa-stained and examined for trypanosomes using x100 objective of the light microscope (Gashururu et al., 2021). The trypanosomes were detected and identified by searching transects of the stained blood smear. For buffy coat examination, heparinized capillary tubes were filled with blood and then closed at one end of each, using Cristaseal (Hawksley, Sussex, United Kingdom). The capillary tubes were centrifuged in a microhaematocrit centrifuge and the junction of the buffy coat layer and the plasma of each tube were examined for trypanosomes under a microscope using x10 objective lens. Trypanosomes were identified by both size and motility under the microscope. The tubes were rotated from time to time during the examination to ensure that all sides of the tube were examined (Woo, 1970).

Assay of Haematological Parameters: The blood samples were evaluated for packed cell volume (PCV), haemoglobin (Hb) status, and red blood cells (RBC) count. Standard methods were used to evaluate the haematological parameters (Schalm *et al.*, 1975; Thrall and Weiser, 2002; Higgins *et al.*, 2008).

Data Analysis: Data was analyzed using Statistical Packages for Social Sciences (SPSS) version 23.0 (IBM Corporation, Armonk, USA). Prevalence of infection was compared using Chisquare analysis. Heamatological indices were compared by student t-test. Level of significance was set at p<0.05.

RESULTS AND DISCUSSION

Prevalence of Trypanosomiasis: This study showed that trypanosome infection was present in trade ruminants sold at Obollo-Afor market in Enugu State, Nigeria. Only 2(2%) out of 100 sheep sampled were infected by trypanosomiasis. None of the 200 goats sampled was positive for the parasite (Table 1).

Table	1: Prevalence of trypanosomiasis in							
trade	ruminants	according	to	ruminant				
type								

7.							
Ruminant	Number	Number					
Туре	examined	infected (%)					
Goat	200	0(0.0)					
Sheep	100	2(2.0)					
Total	300	2(0.7)					

Overall, prevalence of trypanosomiasis in all the 300 ruminants sampled was 2 (0.7%). Infection of the sheep was significant compared to the goats (χ 2 = 4.027, df = 1, p = 0.045). Of the two sheep infected, one was West African dwarf and the other the Yankasa breed (Table 2).

Table 2: Distribution of infectionaccording to breed of sheep

Sheep type	Number examined	Number infected	
West African	33	1(3.0)	
dwarf sheep			
Yankasa	30 1(3.3)		
Uda	37	0(0.0)	
Total	100	2(2.0)	

When compared to several prior reports of 6.9, 4.7 and 3.33% in sheep and 3.5, 4.7 and 1.2% in goats in some parts of Nigeria (Ameen et al., 2008; Ezebuiro et al., 2009; Ohaeri, 2010; Idehen et al., 2018), the results of trypanosome infections obtained for both the sheep (2%) and goats (0%) were guite low. According to Idehen et al. (2018), sheep in Plateau State had a lower prevalence of 1.1%. Reports from other regions of the world also indicated higher prevalence of trypanosome infections in sheep and goats than was seen in this study. Tadesse and Megerssa (2010) observed 2.76 and 1.70% among sheep and goats, respectively, in Guto Gidda district, East Wellega zone, western Ethiopia, and Coello-Peralta et al. (2022) recorded 20% in sheep in Ecuador, Also, Mossaad et al. (2020) reported 4% infections in the Sudanese states of Blue Nile and West Kordofan, while Maganga et al. (2020) recorded 19.2% and 7.8%

infections in sheep and goats, respectively, in South Gabon's Mongo County. Coello-Peralta et al. (2021) also reported 2% infection among 100 sheep in Ecuador just as obtained in this Several factors spanning study. through seasonal variations, number of samples assessed and environmental factors played rolls in the results obtained. Periods of study also varied. Among all the instances cited above, only Ezebuiro et al. (2009) worked on trade ruminants. While some of the studies were carried out either during the dry or rainy seasons, others were carried out all year round. The various reports obtained showed that trypanosomiasis is a tropical and subtropical animal infection.

There are myriads of other determinant factors for the level of prevalence of trypanosome infections in ruminants. These are embedded in biotic factors like the presence of the vector (Glossina) and their livestock preferences, species of trypanosomes, livestock size, treatment of the parasite, and tsetse control (Ohaeri, 2010; Liana et al., 2020; Kizza et al., 2021). There have also been reports of trypanotolerance in sheep and goats (Gutierrez et al., 2006; Malatji, 2022), but the goats are said to be more trypanotolerant than the sheep (Gutierrez et al., 2006; Maganga et al., 2020). Ecological factors (rainfall, relative humidity, and temperature) and season also influence prevalence appreciably (Malatji, 2022). This study was carried out during the dry season (October to January) which was not a thriving period for the vector, *Glossina* spp. In the study, the trade animals arrived Obollo-Afor market, in Southeastern Nigeria, from the distant northern Nigeria. Environmental factors influences the transmission of parasitic diseases (Liang et al., 2007). The few parasites diagnosed may possibly have been introduced from any cluster of sheep and goats in any endemic area of the north prior to transportation of the ruminants to Obollo-Afor market in the south. The short period of this study (4 months) and the small expanse of the study involved may have negatively influenced the result. Sample size and area covered determines whether endemic areas are included or excluded in the study (Ricciardi and Ndao, 2015). The prevalence of animal trypanosomiasis had, though, previously been reported among horses at Obollo-Afor (Agina *et al.,* 2021).

Haematological Status: The haematological profile of the infected and uninfected sheep is summarized in Table 3.

and uninfected trade sheep in the study area									
Parameters	Infected	Uninfected	Т	Р					
PCV (%)	20.65 ±	34.17 ±	-	0.176					
	5.44	0.29	3.508						
RBC	7.58 ±	10.37 ±	-	0.267					
(x10 ¹²)	2.90	1.85	1.359						
Hb (g/dl)	7.60 ±	$11.00 \pm$	-	0.209					
	3.25	1.71	1.593						

Table 3: Haematological profile of infected

The PCV, RBC and Hb values were lower in the infected sheep than the uninfected. The differences were however not significant statistically (p>0.05). Trypanosomiasis has been widely reported as a causative agent of anaemia in livestock (Kagira et al., 2008). Both significant and non-significant decreases of PCV, RBC, and Hb in trypanosome-infected livestock were previously reported (Ameen et al., 2008; Kagira et al., 2008; Tadesse and Megerssa, 2010; Fidelis Junior et al., 2016; Agina et al., 2021). Stability or fluctuation in the haematological profile of trypanosome-infected livestock is influenced by various factors, including nutrition, species of trypanosome involved, intensity of parasitaemia, and host-parasite interaction in the bloodstream (Stijlemans et al., 2018; Kasozi et al., 2021; Lelisa and Meharenet, 2021).

Conclusion: Trypanosomiasis was diagnosed in trade sheep sold at Obollo-Afor, Enugu State. The study reports the danger of trans-boundary movement of trade ruminants as a potential means of spreading trypanosome infection as well as other zoonotic livestock diseases in the area. These trade ruminants should therefore be subjected to thorough examination to ensure that they do not pose as agents of the spread of this infection or cause the economic downturn commonly associated with the infection. Further studies involving larger samples of ruminants

and a longer duration of study are therefore proposed for more insight on the trypanosomiasis infection and blood parameters in trade ruminants in southeast Nigeria.

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REFERENCES

- AGINA, O. A., IHEDIOHA, J. I., ADEYEYE, T. E., UMEAKUANA, P. U. and IDOKO, I. S. (2021). Molecular detection of *Trypanosoma* species and haematological alterations in four trypanosome-infected Nigerian horses. *Notulae Scientia Biologicae*, 13(4): 11046. <u>https://doi.org/10.15835/</u> <u>nsb13411046</u>
- AMEEN, S. A., JOSHUA, R. A., ADEDEJI, O. S., RAHEEM, A. K., AKINGBADE, A. A. and LEIGH, O. O. (2008). Preliminary studies on prevalence of ruminant trypanosomosis in Ogbomoso area of Oyo State, Nigeria. *Middle-East Journal of Scientific Research*, 3(4): 214 – 218.
- BOUYER, J., BOUYER, F., DONADEU, M., ROWAN, T. and NAPIER, G. (2013). Communityand farmer-based management of animal African trypanosomosis in cattle. *Trends in Parasitology*, 29(11): 519 – 522.
- CAYLA, M., ROJAS, F., SILVESTER, E., VENTER, F. and MATTHEWS, K. R. (2019). African trypanosomes. *Parasites and Vectors* 12: 190. <u>https://doi.org/10.1186/s1307</u> <u>1-019-3355-5</u>
- CITY POPULATION (2023). Udenu: LGA in Enugu State, Nigeria. <u>https://www.city</u> <u>population.de/en/nigeria/admin/NGA014</u> <u>enugu/</u> Accessed May 05, 2023.
- COELLO-PERALTA, R. D., SOLÓRZANO, N. C. C., MAZAMBA, M. D. L. S., ECHEVERRIA, E. O. R., VELÁSQUEZ, L. L. A. and GÓMEZ, B. J. P. (2022). Natural infection of

Trypanosoma sp. in domestic sheep from Ecuador. *Ciência Rural, Santa, Maria*, 52(2): e20210141. <u>https://doi.</u> org/10.1590/0103-8478CR20210141

- COELLO-PERALTA, R. D., SOLÓRZANO, N. C. C., PEÑAFIEL, K. A. R., MAZAMBA, M. DE L. S. and ECHEVERRIA, E. O. R. (2021). First report of sheep naturally infected with *Trypanosoma* sp. in Ecuador. *Ciencia Rural, Santa, Maria*, 51(1): e20200223. <u>https://doi.org/10.1590/01</u> 03-8478cr20200223
- DESQUESNES, M., GONZATTI, M., SAZMAND, A., THÉVENON, S., BOSSARD, G., BOULANGÉ, A., GIMONNEAU, G., TRUC, P., HERDER, S., RAVEL, S., SERENO, D., JAMONNEAU, V., JITTAPALAPONG, S., JACQUIET, P., SOLANO, P. and BERTHIER, D. (2022). A review on the diagnosis of animal trypanosomoses. *Parasites and Vectors*, 15: 64. https:// doi.org/10.1186/s13071-022-05190-1
- DIALL, O., CECCHI, G., WANDA, G., ARGILÉS-HERRERO, R., VREYSEN, M. J. B., CATTOLI, G., VILJOEN, G. J., MATTIOLI, R. and BOUYER, J. (2017). Developing a progressive control pathway for african animal trypanosomosis. *Trends in Parasitology*, 33(7): 499 – 509.
- EZEBUIRO, O. G. C., ABENGA, J. N. and EKEJINDU, G. O. C. (2009). The prevalence of trypanosome infection in trade cattle, goats and sheep slaughtered at the Kaduna abattoir. *African Journal of Clinical and Experimental Microbiology*, 10(1): 15 – 25.
- FIDELIS JUNIOR, O. L., SAMPAIO, P. H., MACHADO, R. Z., ANDRÉ, M. R., MARQUES, L. C. and CADIOLI, F. A. (2016). Evaluation of dinical signs, parasitemia, hematologic and biochemical changes in cattle experimentally infected with *Trypanosoma vivax. Revista Brasileira de Parasitologia Veterinária*, 25(1): 69 – 81.
- GASHURURU, S. R., MAINGI, N., GITHIGIA, S. M., GASANA, M. N., ODHIAMBO, P. O., GETANGE, D. O., HABIMANA, R., CECCHI, G., ZHAO, W., GASHUMBA, J. and BARGUL, J. L. (2021). Occurrence, diversity and

distribution of *Trypanosoma* infections in cattle around the Akagera National Park, Rwanda. *PLOS Neglected Tropical Diseases*, 15(12): e0009929. <u>https://</u> doi.org/10.1371/journal.pntd.0009929

- GIORDANI, F., MORRISON, L. J., ROWAN, T. G., DE KONING, H. P. and BARRETT, M. P. (2016). The animal trypanosomiases and their chemotherapy: a review. *Parasitology*, 143(14): 1862 – 1889.
- GUTIERREZ, C., CORBERA, J. A., MORALES, M. and BÜSCHER, P. (2006). Trypanosomosis in goats: current status. *Annals of the New York Academy of Sciences*, 1081(1): 300 – 310.
- HIGGINS, T., BEUTLER, E. and DOUMAS, B. T. (2008). Measurement of haemoglobin in blood. Pages 524 525. *In*: BURTIS, C. A., ASHWOOD, E. R. and BRUNS, D. E. (Eds.). *Tietz Fundamentals of Clinical Chemistry*. Sanders Elsevier, St. Louis, USA,
- IDEHEN, C. O., ISHOLA, O. O., ADEYEMI, I. G., ABONGABY, G., OLALEYE, O. O., ALUMA, A. L., OPABUNMI, R. O. and OBALOTO, O. B. (2018). Prevalence of African trypanosomosis in cattle and sheep in Bassa Local Government Area of Plateau State, Nigeria. *Sokoto Journal* of Veterinary Sciences, 16(3): 11 – 17.
- ISAAC, C., OHIOLEI, J. A., EBHODAGHE, F., IGBINOSA, I. B. and EZE, A. A. (2017). Animal African trypanosomosis in Nigeria: A long way from elimination/eradication. *Acta Tropica*, 176: 323 – 331.
- KAGIRA, J., THUITA, J., NGOTHO, M., MDACHI, R., MWANGANGI, D. and NDUNG'U, J. (2008). Haematology of experimental *Trypanosoma brucei rhodesiense* infection in vervet monkeys. *African Journal of Health Sciences*, 13(3): 59 – 65.
- KASOZI, K. I., ZIRINTUNDA, G., SSEMPIJJA, F., BUYINZA, B., ALZAHRANI, K. J., MATAMA,
 K., NAKIMBUGWE, H. N., ALKAZMI, L., ONANYANG, D., BOGERE, P., OCHIENG,
 J. J., ISLAM, S., MATOVU, W., NALUMENYA,
 D. P., BATIHA, G. E. S., OSUWAT, L. O.,
 ABDELHAMID, M., SHEN, T., OMADANG, L. and WELBURN, S. C. (2021). Epidemiology

of trypanosomiasis in wildlife – Implications for humans at the wildlife interface in Africa. *Frontiers in Veterinary Science*, 8: 621699. <u>https://</u> doi.org/10.3389/ fvets.2021.621699

- KIZZA, D., OCAIDO, M., MUGISHA, A., AZUBA, R., NALULE, S., ONYUTH, H., MUSINGUZI, S. P., OKWASIIMIRE, R. and WAISWA, C. (2021). Prevalence and risk factors for trypanosome infection in cattle from communities surrounding the Murchison Falls National Park, Uganda. *Parasites and Vectors*, 14: 513. <u>https://doi.org</u> /10.1186/s13071-021-04987-w
- LAWAL-ADEBOWALE, O. A. (2012). Dynamics of ruminant livestock management in the context of the Nigerian agricultural system. *In:* JAVED, K. (Ed.). *Livestock Production*. IntechOpen.com, London. https://doi.org/10.5772/52923
- LELISA, K. and MEHARENET, B. (2021). Anaemia associated with trypanosomes infections in cattle of West Gojjam Zone, Northwest Ethiopia. *Veterinary Medicine International*, 2021: 5531537. <u>https://doi.org/10.1155/2021/5531537</u>
- LIANA, Y. A., SHABAN, N., MLAY, G. and PHIBERT, A. (2020). African trypanosomiasis dynamics: Modelling the effects of treatment, education, and vector trapping. *International Journal of Mathematics and Mathematical Sciences*, 2020: 3690472. <u>https://doi.org/10.1155/2020/</u> <u>3690472</u>
- LIANG, S., SETO, E. Y. W., REMAIS, J. V., ZHONG, B., YANG, C., HUBBARD, A., DAVIS, G. M., GU, X., QIU, D. and SPEAR, R. C. (2007). Environmental effects on parasitic disease transmission exemplified by schistosomiasis in western China. *Proceedings of the National Academy of Sciences of the United States of America*, 104(17): 7110 – 7115.
- MAGANGA, G. D., BOUNDENGA, L., OLOGUI-MINKUE-EDZO, E. J., KOMBILA, L. B., MEBALEY, T. G. N., KUMULUNGUI, B. and MAVOUNGOU, J. F. (2020). Frequency and diversity of trypanosomes in sheep and goats from Mongo County in South

Gabon, Central Africa. *Veterinary World*, 13(11): 2502 – 2507.

- MALATJI, D. P. (2022). Breeding of African sheep reared under low-input/output smallholder production systems for trypanotolerance. *Veterinary World*, 15(4): 1031 – 1043.
- MATOVU, E., MUGASA, C. M., WAISWA, P., KITIBWA, A., BOOBO, A. and NDUNG'U, J. M. (2020). Haemoparasitic infections in cattle from a *Trypanosoma brucei rhodesiense* sleeping sickness endemic district of Eastern Uganda. *Tropical Medicine and Infectious Disease*, 5(1): 24. <u>https://doi.org/10.3390/tropicalmed</u> <u>5010024</u>
- MOSSAAD, E., ISMAIL, A. A., IBRAHIM, A. M., MUSINGUZI, P., ANGARA, T. E. E., XUAN, X., INOUE, N. and SUGANUMA, K. (2020). Prevalence of different trypanosomes in livestock in Blue Nile and West Kordofan States, Sudan. *Acta Tropica*, 203: 105302. <u>https://doi.org/10.1016/j.acta</u> <u>tropica.2019.105302</u>
- NGONGOLO, K., SHIRIMA, G., MPOLYA, E. A., ESTES, A. B., HUDSON, P. J. and GWAKISA, P. S. (2020). Influence of land cover and host species on trypanosome infection in cattle and its socioeconomic impacts to pastoralists of the Maasai Steppe, Tanzania. *Journal of Infectious Disease and Epidemiology*, 6(1): 107. <u>https://doi.org/10.23937/247</u> <u>4-3658/1510107</u>

OHAERI, C. C. (2010). Prevalence of trypanosomiasis



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in ruminants in parts of Abia State, Nigeria. *Journal of Animal and Veterinary Advances*, 9(18): 2422 – 2426.

- RICCIARDI, A. and NDAO, M. (2015). Diagnosis of parasitic infections: What's going on? *Journal of Biomolecular Screening*, 20(1): 6 – 21.
- SCHALM, O. W., JAIN, N. C. and CARROLL, E. J. (1975). *Veterinary Haematology.* 3rd Edition, Lea and Febiger, Philadelphia, USA.
- STIJLEMANS, B., DE BAETSELIER, P., MAGEZ, S., VAN GINDERACHTER, J. A. and DE TREZ, C. (2018). African trypanosomiasisassociated anemia: the contribution of the interplay between parasites and the mononuclear phagocyte system. *Frontiers of Immunology*, 9: 218. <u>https://doi.org</u> /10.3389/fimmu.2018.00218
- TADESSE, A. and MEGERSSA, G. (2010). Prevalence of trypanosomosis in small ruminants of Guto Gidda district, East Wellega zone, western Ethiopia. *Ethiopian Veterinary Journal*, 14(2): 68 – 77.
- THRALL, M. A. and WEISER, M. G. (2002). Haematology. *In*: HENDRIX, C. M. (Ed.). *Laboratory Procedures for Veterinary Technicians.* 4th Edition, Mosby Incooperated, Maryland Heights, Missouri, United States.
- WOO, P. T. K. (1970). The haematocrit centrifuge technique for the diagnosis of African trypanosomiasis. *Acta Tropica*, 27(4): 384 – 386.