COMU Journal of Marine Sciences and Fisheries

Journal Home-Page: http://jmsf.dergi.comu.edu.tr Online Submission: http://dergipark.org.tr/jmsf

RESEARCH ARTICLE

Seasonal Changes in Proximate and Bioactive Compounds of Brown and Red Seaweeds from İskenderun Bay, the North-Eastern Mediterranean Sea

İbrahim Gür¹, Sevim Polat^{2*}

¹District Directorate of Agriculture and Forestry, Gelibolu, Çanakkale, Türkiye ²Department of Marine Science, Faculty of Fisheries, Cukurova University, 01330 Balcalı, Adana, Türkiye

Received: 15.03.2023 /Accepted: 02.06.2023 / Published online: 04.07.2023

https://orcid.org/0000-0002-6941-8785 https://orcid.org/0000-0002-4756-1177

Key words:

Seaweed Proximate composition Bioactive compounds Phenolics Flavonoids **Abstract:** Proximate and bioactive compounds (total phenolic, flavonoid, chlorophyll-*a* and total carotenoid contents) of three brown seaweeds (*Dictyota dichotoma, Padina pavonica, Stypopodium schimperi*) and a red seaweed (*Jania rubens*) from the north-eastern Mediterranean Sea (İskenderun Bay) were investigated seasonally at three sampling sites. Seasonal variations were found for all of the parameters studied. The highest ash content was in *J. rubens* (77.7%) in the spring. The results showed that *J. rubens* is a rich source with respect to mineral content. *D. dichotoma* had the highest crude protein content, whereas *S. schimperi* contained the most lipids. Phenolics ranged between 34.6 - 107.0 mg GAE/g dw. The highest total phenolics were found in *S. schimperi* in the summer, and the lowest in *P. pavonica* in the spring. The flavonoid contents (9.05-10.6 mg QE/g dw) were higher in brown seaweeds than that in the red seaweed. Moreover, chlorophyll-*a* and carotenoids levels were highest in *D. dichotoma* (4.53 and 2.83 mg/g, respectively) during the autumn. The results revealed that the biochemical composition of the examined seaweeds showed significant changes depending on the species, location and seasons.

Anahtar kelimeler:

Makroalg Temel besin maddesi kompozisyonu Biyoaktif bileşikler Fenolik Flavonoid

İskenderun Körfezi'ndeki (Kuzeydoğu Akdeniz) Kahverengi ve Kırmızı Makroalglerin Temel Besin Maddesi ve Biyoaktif Bileşiklerindeki Mevsimsel Değişimler

Öz: Kuzeydoğu Akdeniz'de (İskenderun Körfezi) dağılım gösteren üç kahverengi (*Dictyota dichotoma, Padina pavonica, Stypopodium schimperi*) ve bir kırmızı makroalgin (*Jania rubens*) temel besin maddesi ve biyoaktif bileşikleri (toplam fenolik, flavonoid, klorofil-*a* ve toplam karotenoid içerikleri) üç örnekleme istasyonunda mevsimsel olarak incelenmiştir. İncelenen tüm parametrelerin mevsimsel değişimler gösterdiği belirlenmiştir. En yüksek kül içeriği ilkbaharda *J. rubens* türünde (%77.7) bulunmuştur. Sonuçlar, bu türün zengin bir mineral kaynağı olduğunu göstermiştir. *D. dichotoma* en yüksek ham protein içeriğine, *S. schimperi* ise en fazla lipit içeriğine sahip tür olmuştur. Makroalglerde fenolik madde içeriği 34.6 ile 107.0 mg GAE/g kuru ağ. arasında değişmiştir. Toplam fenolik madde miktarı yazın *S. schimperi* türünde en yüksek düzeye ulaşırken, en düşük değer ilkbaharda *P. pavonica* türünde bulunmuştur. Flavonoid içerikleri (9.05-10.6 mg QE/g kuru ağ.) kahverengi deniz yosunlarında kırmızı deniz yosunundan daha yüksek bulunmuştur. Klorofil-*a* ve karotenoid içeriği ise *D. dichotoma* türünde sonbahar mevsiminde en yüksek düzeylerde (sırasıyla 4.53 ve 2.83 mg/g) bulunmuştur. Sonuçlar, incelenen makroalg türlerinin biyokimyasal kompozisyonunun türe, lokaliteye ve mevsimlere bağlı olarak belirgin değişimler gösterebileceğini ortaya koymuştur.

*Corresponding author: sevcan@cu.edu.tr



How to cite this article: Gür, 1., & Polat, S. (2023). Seasonal changes in proximate and bioactive compounds of Brown and Red Seaweeds from İskenderun Bay, the North-Eastern Mediterranean Sea. COMU J. Mar. Sci. Fish, 6(1): 33-43. doi:10.46384/jmsf.1265503

Introduction

Seaweeds are divided into three groups depending on pigmentation, namely red, green and brown algae. As primary producers, they provide food and shelter for many organisms living in the coastal waters. They are utilized as food for humans, feed for animals and as a source of various chemicals (Nedumaran and Arulbalachandran, 2015). They have been traditionally consumed by humans and used to extract ingredients. Seaweeds contain protein, lipids, dietary fibre, carotenoids, minerals and vitamins (Kumar et al., 2008; Peñalver et al., 2020). Many species red and brown seaweeds are utilized to produce alginate, agar and carrageenan, which are used as thickeners in foods, cosmetics and medicine. Moreover, seaweeds are particularly rich in biologically active compounds, i.e., functional foods such as phenolic and flavonoids, which have antioxidant activities (Machu et al., 2015; Yılmaz et al., 2021). The antioxidant activity of seaweeds has been studied and strong relations between phenolic compounds and antioxidant activities were found. Wang et al. (2009) showed that the antioxidant activity of red seaweed was closely correlated with its extracted phenolics. Antioxidants are known to have a protective effect since they can defend the human body against damage by free radicals (Kalasariya et al., 2021). Free radicals are associated with human diseases, including cardiovascular disease, cancer, diabetes, hypertension, ischemia, ageing, and Alzheimer's disease (Chauhan and Chauhan, 2006; Matanjun et al., 2008).

Although Turkey is surrounded by the Black Sea, the Mediterranean Sea, the Sea of Marmara and the Aegean Sea with >8000 km of coastline, the bioactive compounds of seaweeds have still not been adequately studied. The previous studies in Turkish coastal waters are generally concentrated on the taxonomy and distribution of seaweeds (Aysel et al., 2006a; Aysel et al., 2006b; Taşkın, 2014). Data on the chemical composition of seaweeds from the coasts of Turkey have increased in recent vears but are still limited (Polat and Özoğul, 2008; Polat et al., 2012; Turan et al., 2015; Güner and Yavasoglu, 2018; Caf et al., 2019; Saygılı et al., 2022). Moreover, there are only few studies on seasonal variations in the biochemical contents of seaweeds on our coasts (Polat and Özoğul, 2013; İrkin and Erduğan, 2016; İrkin and Erduğan, 2017; Yeşilova et al., 2017). This study is aimed to investigate the proximate composition, total phenolic, flavonoid and pigment contents of seaweeds seasonally collected from İskenderun Bay on the north-eastern Mediterranean coast of Turkey.

Material and Methods

Collection of samples

Samples of three brown seaweeds (*Padina pavonica* (Linnaeus) Thivy, *Stypopodium schimperi* (Kützing) Verlaque & Boudouresque and *Dictyota dichotoma* (Hudson) J.V.Lamouroux), and one red seaweed (*Jania rubens*) (Linnaeus) J.V. Lamouroux were collected from three localities along the coast of İskenderun Bay (Fig. 1). The samples were collected seasonally over one year (between 2013-2014); in the spring, summer, autumn, and winter. Samples were taken from depths of up to 3 m.

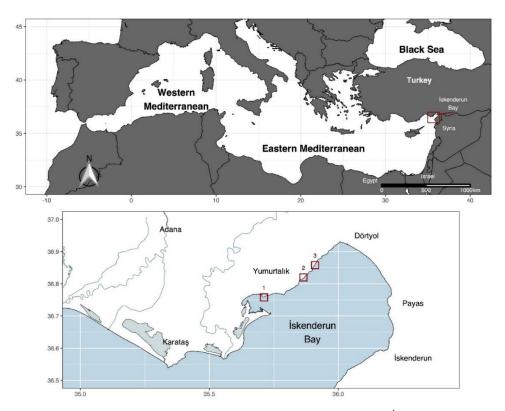


Figure 1. The study area and location of seaweed sampling sites in İskenderun Bay

Some species were only found in some seasons and locations. *P. pavonica* was not found in the winter, *S. schimperi* in the autumn and the winter, and *D. dichotoma* in the spring. Fresh samples of four selected seaweed species were washed with local seawater and immediately transported to the laboratory in an icebox with ambient

seawater. Samples were gently washed with distilled water to remove epiphytes, salts, epifauna and sand. Samples were stored at -80 °C until further analysis. The temperature and salinity were measured with a YSI model SCT probe during the sampling periods, and the values for each sampling site are shown in Table 1.

				-	
	Samplin g sites	Spring	Summer	Autumn	Winter
	1	27.9	29.2	24.1	15.3
Temperature (°C)	2		31.6	25.4	17.6
	3	27.4	30.9	24.7	16.7
Salinity (‰)	1	35.5	37.2	37.4	37.2
	2		35.5	36.4	38
	3	34.8	37.1	36.5	37.3

Table 1. Seasonal variations of temperature and salinity during seaweed sampling

Proximate composition analysis

Moisture content was determined using the AOAC method 950.46 (1990), and crude ash content was analyzed using the AOAC method 938.08 (1998a). Lipid analysis was carried out according to the method of Bligh and Dyer (1959). Samples were homogenized with a 1:2 mixture of chloroform and methanol and left in a dark place overnight after adding CaCl₂. The chloroform layer was removed and vaporized in an evaporator and finally in an oven at 60°C for one hour. Total crude protein was determined using the AOAC method 955.04 (1998b). Seaweed samples were placed in Kjeldahl tubes, two Kjeldahl tablets and 20 mL H₂SO₄ were added to the incinerator, and the sample inside the tubes was burned at 420 °C. After digestion, 75 mL of water was added to the sample tube. Distillation was performed for 6 min with 40% NaOH by placing 25 mL of 40% boric acid (H₃BO₃) solution. Then, the distillate was titrated with 0.1 M HCl, and the amount of protein was found by recording the HCl consumed. The percentage of protein was determined with a conversion factor of 6.25.

Total phenolic content (TPC)

TPC of seaweed extracts was determined using the Folin-Ciocalteu method (Gámez-Meza *et al.*, 1999). The gallic acid was used as the standard, and the total phenol content was expressed as milligrams of GA equivalents per gram of extract (mg GAE/g dw).

Total flavonoid content (TFC)

The TFC of seaweeds were determined using the method described by Chang *et al.* (2002). The amount of TFC was determined from a standard calibration curve and expressed as mg QE/g dw (QE, quercetin equivalents) per gram of seaweed extract.

Chlorophyll-a

Chlorophyll-*a* was estimated according to the method described by Arnon (1949) and Thirumaran *et al.* (2009) with minor modifications. Ground seaweed (500 mg) was extracted with 10 ml of 80 % acetone (Merck, 99 %) and centrifuged at 3000 rpm for 15 min. The absorbance was measured at 645 and 663 nm. The content of chlorophyll-*a* was calculated based on the equation given by Thirumaran *et al.* (2009).

Total carotenoid

The total carotenoid level of seaweeds was determined spectrophotometrically using the same extract for chlorophyll-a estimation. The total carotenoid content was calculated from absorbance values at 480 nm using the equation of Kirk and Allen (1965).

Statistical analysis

The general linear models were used to compare biochemical compounds (ash, moisture, crude protein and lipid) among seasons and sampling sites. The models were separately fitted for each compound and species and then checked against the violations of normality and homogeneity using the Levene and Shapiro-Wilk tests on model residuals. Normality was met with all models. When heterogeneity was observed, a heteroscedasticitycorrected coefficient covariance matrix was calculated using the White Adjustment argument in the "car" package (Fox and Weisberg, 2011). After linear models were fitted, multiple comparisons were made using general linear hypothesis tests with Tukey's adjustment using the "multcomp" package (Hothorn et al., 2008). In case of a violation of the homogeneity of the variance, the heteroskedasticity-consistent covariance matrix was estimated using the "sandwich" package (Zeileis, 2004). All statistical analyses used the R 3.4.4 statistical language (R Core Team, 2018).

Results and Discussion

The results of the four seaweed species' moisture, crude ash, lipid, total crude protein, total phenolic, flavonoid and pigment contents are shown in Tables 2-5. Ash content in seaweeds was generally higher than those in most land plants (Rupérez, 2002; Radha, 2018). The highest ash content was in *J. rubens* (77.7 %) collected from Site (St) 2 in the spring, whereas the lowest ash content was found in *D. dichotoma* (3.12 %) collected from St. 2 in the winter (Tables 2, 5). The high ash level of *J. rubens* shows that this species is rich in minerals. A similar high ash value was found in *Corallina officinalis* (77.8 %) by Marsham *et*

al. (2007). The ash contents of all seaweed species showed significant differences between seasons and sampling sites (p<0.01, p<0.05), respectively. Marsham *et al.* (2007) reported that high ash indicates of a reasonable amount of mineral content. The maximum moisture was found in *D. dichotoma* (80.74 %) collected from St.1 in the winter and minimum content in *J. rubens* (3.15 %) collected from St.2 in the spring. The moisture content differed significantly (p<0.01, p<0.05) between seasons and sampling sites in all seaweed species. The moisture and ash composition of seaweeds differs depending on the species, geographical location and seasons.

D. dichotoma	Sampling sites	Summer	Autumn	Winter
	1	$3.32\pm 0.29^{Aa^{**}}$	n.a	$3.12\pm0.89^{\rm Aa}$
Ash (%)	2	n.a.	5.07 ± 0.10^{Ab}	$3.12\pm0.19^{\rm Aa}$
	3	$4.98\pm0.16^{\rm Ba}$	$5.98\pm0.78^{\rm Bb}$	5.49 ± 0.89^{Ba}
Moisture (%)	1	$75.67 \pm 1.09^{bc^*}$	n.a	$80.74\pm3.67^{\rm c}$
	2	n.a.	72.87 ± 1.68^{ab}	$79.87 \pm 1.10^{\rm c}$
	3	69.42 ± 0.35^{ab}	70.25 ± 1.11^{ab}	$67.81\pm4.16^{\rm a}$
Lipid (%)	1	$3.74\pm0.01^{\rm b}$	n.a	$0.90\pm0.40^{\rm a}$
	2	n.a.	$3.19\pm0.32^{\rm b}$	$1.02\pm0.32^{\rm a}$
	3	$5.13\pm0.17^{\rm c}$	$3.72\pm0.08^{\rm b}$	2.72 ± 0.31^{ab}
	1	4.42 ± 0.29	n.a	5.16 ± 0.38
Crude protein (%)	2	n.a.	5.56 ± 1.67	4.66 ± 0.89
	3	6.15 ± 0.47	6.15 ± 3.33	4.92 ± 1.27
Phenolic content (mg GAE/g dw)	1	65.85 ± 3.95	n.a	47.06 ± 2.82
	2	n.a.	50.58 ± 3.03	46.48 ± 3.11
	3	52.70 ± 3.16	51.20 ± 3.07	46.28 ± 2.77
T 1 1	1	9.23 ± 0.55	n.a	9.47 ± 0.61
Flavonoid (mg QE/g dw)	2	n.a.	9.16 ± 0.55	9.44 ± 0.56
	3	9.16 ± 0.54	9.05 ± 0.54	9.63 ± 0.57
Chlorophyll- <i>a</i> (mg/g)	1	1.43	n.a	3.51
	2	n.a.	4.50	3.69
	3	2.98	4.53	4.09
T 10 11	1	1.05	n.a	2.04
Total Carotenoid	2	n.a.	2.46	2.23
(mg/g)	3	2.24	2.83	2.53

Table 2. The proximate composition, total phenolics, flavonoids and pigments of D. dichotoma

The values are expressed as means \pm standard deviation, n = 3, n.a.: sample was not found, *Different superscript letters in the same variable indicate significant differences, **Lower case letters indicate significant differences between seasons, and caps indicate the differences between sampling sites when only two main effects are significant at 0.05 significance level.

The lipid contents of the seaweeds varied between 0.25 - 6.35%. The highest lipid content was found in *S. schimperi* collected from St.3 in the summer, while the lowest content (0.25%) was in *J. rubens* collected from St.3 in the autumn (Tables 4, 5). The differences in the

lipid contents of all seaweed species were statistically significant (p<0.05) among seasons. However, the difference between sampling sites was only significant in *D. dichotoma* and *S. schimperi*. Similarly, Polat and Özoğul (2013) found the highest lipid in *S. schimperi* in

the summer, ranging between 2.03% - 2.16%. In another study, Polat and Özoğul (2009) reported the lowest total lipid content for *J. rubens*, as 0.12%. Parthiban *et al.* (2013) reported that the lipid contents of some seaweeds collected from Tuticorin varied from 3.15 to 5.30%. The

results of this study were consistent with those of Nelson *et al.*, (2002) and Kostetsky *et al.* (2004) and showed that the lipid concentration of seaweeds may vary according to species, seasons and sampling sites.

P. pavonica	Sampling sites	Spring	Summer	Autumn
	1	$22.68\pm1^{bd*}$	25.14 ± 0.2^{cd}	n.a.
Ash (%)	2	$18.17\pm0.23^{\rm a}$	24.11 ± 0.60^{cd}	22.20 ± 1.99^{bc}
	3	19.97 ± 0.71^{ab}	$32.00\pm2.42^{\text{e}}$	$25.75\pm0.78^{\rm d}$
Moisture (%)	1	$32.74\pm0.82^{\rm a}$	$56.41\pm0.31^{\text{e}}$	n.a.
	2	53.67 ± 0.28^{cd}	$55.24 \pm 1.33^{\text{de}}$	$58.30\pm2.77^{\text{e}}$
	3	53.63 ± 0.28^{cd}	$43.14\pm4.04^{\text{b}}$	$52.10 \pm 1.25^{\circ}$
Lipid (%)	1	$0.34\pm0.05^{\rm a}$	2.16 ± 0.06^{d}	n.a.
	2	0.83 ± 0.15^{abc}	$1.80\pm\!\!0.47^{abcd}$	$1.20\pm0.00^{\text{b}}$
	3	1.36 ± 0.14^{bc}	1.76 ± 0.20^{bcd}	$1.27\pm0.01^{\circ}$
	1	$3.35\pm0.15^{\rm a}$	4.03 ± 0.11^{b}	n.a.
Crude protein (%)	2	$3.46\pm0.14^{\rm a}$	$5.30\pm0.21^{\rm b}$	$5.31\pm0.60^{\text{b}}$
	3	$3.39\pm0.58^{\rm a}$	$4.53\pm0.07^{\rm b}$	$5.69 \pm 1.28^{\text{b}}$
	1	34.61 ± 2.07	51.86 ± 2.01	n.a.
Phenolic content (mg GAE/g dw)	2	n.a.	53.21 ± 3.19	49.44 ± 2.96
(ing OTIL/g uw)	3	42.50 ± 2.55	51.49 ± 3.02	61.41 ± 3.68
	1	10.07 ± 0.78	10.10 ± 0.31	n.a
Flavonoid (mg QE/g dw)	2	n.a.	10.49 ± 0.62	10.00 ± 0.97
	3	10.47 ± 0.62	10.09 ± 0.60	10.00 ± 0.54
Chlorophyll- <i>a</i> (mg/g)	1	1.40	1.45	n.a.
	2	0.75	0.91	1.34
	3	1.11	1.12	1.62
T + 1 C + 1	1	0.92	0.91	n.a.
Total Carotenoid (mg/g)	2	0.47	0.34	0.76
(1118/8/	3	0.79	0.94	0.96

Table 3. The proximate compositon, total phenolics, flavonoids and pigments of P. pavonica

The values are expressed as means \pm standard deviation, n = 3, n.a.: sample was not found. *Different superscript letters in the same variable indicate significant differences, **Lower case letters indicate significant differences between seasons, and caps indicate the differences between sampling sites when only two main effects are significant at 0.05 significance level.

Crude protein ranged from 2.94 to 6.15%. The maximum crude protein was found in the brown seaweed *D. dichotoma* collected from St.3, during the summer and autumn (6.15%), followed by *S. schimperi* (6.05%) from St.1 in the spring. The minimum crude protein (2.94%) was in the *S. schimperi* collected from St.3 in the spring. The differences in crude protein were significant (p<0.05) among seasons in all species except *D. dichotoma*. However, the difference was not significant between the sampling sites in all species except *S. schimperi*. Similar results were reported by Tabarsa *et al.* (2012), who found

relatively high crude protein in the brown seaweed *D. dichotoma* from the coastal area of Kuvehei (Iran). Polat and Özoğul (2013) found lower crude protein values ranging from 2.37 % to 2.68 % in *S. schimperi* than those reported in the present study. Higher crude protein levels (9.47 and 14.7%) were also reported by Parthiban *et al.* (2013) for some seaweed species collected from the Tuticorin and Mandapam coasts (India). The protein contents of the other species, *J. rubens* and *P. pavonica*, in the present study were higher than those previously found by Polat and Özoğul (2013). Factors such as the

physiological status of seaweed species and life cycle may have contributed to these results. In addition to speciesspecific differences, water quality, the season and the geographic area could influence the lipid and crude protein content of seaweeds (Fleurence, 1999; Haroon, 2000; Ratana-arporn and Chirapart, 2006).

S. schimperi	Sampling sites	Spring	Summer	
	1	$5.42 \pm 0.53^{bc^{\ast}}$	$3.96\pm0.06^{\rm a}$	
Ash (%)	2	n.a.	$6.09\pm0.24^{\rm c}$	
	3	5.71 ± 0.25^{bc}	5.18 ± 0.10^{b}	
Moisture (%)	1	$63.72 \pm 3.09^{Ca^{**}}$	74.95 ±0.15 ^{Cb}	
	2	n.a.	$66.26 \pm 0.92^{\text{Bb}}$	
	3	$61.21\pm0.77^{\mathrm{Aa}}$	69.49 ± 0.20^{Ab}	
Lipid (%)	1	$3.28\pm0.20^{\mathrm{Aa}}$	$4.73\pm0,14^{\rm Ab}$	
	2	n.a.	$3.04 \pm 1.82^{\text{Bb}}$	
	3	$5.18\pm0.08^{\text{Ca}}$	$6.35\pm0.19^{\text{Cb}}$	
	1	$6.05\pm0.23^{\rm b}$	$3.60\pm0.06^{\rm a}$	
Crude protein (%)	2	n.a.	$3.96\pm0.20^{\rm a}$	
	3	$2.94\pm0.22^{\rm a}$	$3.44\pm0.04^{\rm a}$	
Phenolic content (mg GAE/g dw)	1	92.85 ± 5.57	102.84 ± 5.23	
	2	n.a.	106.05 ± 6.36	
(ing GAL/g uw)	3	52.90 ± 3.17	100.61 ± 6.03	
	1	10.41 ± 0.62	10.62 ± 0.51	
Flavonoid (mg QE/g dw)	2	n.a.	10.50 ± 0.72	
(Ing QE/g uw)	3	10.55 ± 0.63	10.61 ± 0.66	
	1	3.68	2.49	
Chlorophyll- <i>a</i> (mg/g)	2	n.a.	1.59	
(1116/5)	3	3.04	1.18	
T . 10	1	2.61	2.73	
Total Carotenoid	2	n.a.	2.11	
(mg/g)	3	2.33	1.95	

Table 4. The proximate composition, total phenolics, flavonoids and pigments of S. schimperi

The values are expressed as means \pm standard deviation, n = 3, n.a.: sample was not found, *Different superscript letters in the same variable indicate significant differences, **Lower case letters indicate significant differences between seasons, and caps indicate the differences between sampling sites when only two main effects are significant at 0.05 significance level.

TPC of the seaweed species varied from 34.6 to 106.05 mg GAE/g dw. *S. schimperi* collected from St.2 in the summer showed the highest TPC, followed by *J. rubens* collected from St.1 in the autumn. The lowest TPC was found in *P. pavonica* collected from St.1 in the spring (Table 3). Kumar *et al.* (2011) reported phenolic contents in three green algae species *Caulerpa veravelensis*, *Caulerpa racemosa* and *Caulerpa scalpelliformis* as 32.57, 61.69 and 36.00 mg/g dw, respectively. Connan *et al.*

(2007) reported that total phenolic contents in seaweeds may vary according to the difference between day and night temperatures and light intensity during the day. In the present study, the concentration of total phenolics in *S. schimperi* for the summer period (100.61-106.05 mg/g dw) was higher than all seaweed species sampled and those recorded in previous studies. *S. schimperi* seems to be a good antioxidant source for human health.

J. rubens	Sampling sites	Spring	Summer	Autumn	Winter
	1	n.a.	n.a.	$59.24 \pm 0.64^{\mathrm{A}^{**}}$	n.a.
Ash (%)	2	$77.73\pm0.20^{\rm d}$	$72.62 \pm 1.48^{\rm c}$	58.56 ± 0.43^{bA}	$51.26\pm2.85^{\mathrm{a}}$
	3	n.a.	n.a.	$70.35\pm4.22^{\mathrm{B}}$	n.a.
Moisture (%)	1	n.a.	n.a.	$26.84\pm0.92^{\rm B}$	n.a.
	2	$3.15\pm0.43^{\rm a}$	$8.68 \pm 1.56^{\text{b}}$	$27.38\pm0.25^{\rm cB}$	$3.69 \pm 1.24^{\rm a}$
	3	n.a.	n.a.	$6.31\pm4.22^{\rm A}$	n.a.
Lipid (%)	1	n.a.	n.a.	0.39 ± 0.08^{ab}	n.a.
	2	$0.42\pm0.39^{ab^\ast}$	$0.78\pm0.10^{\rm b}$	0.61 ± 0.25^{ab}	$0.26\pm0.01^{\rm a}$
	3	n.a.	n.a.	0.25 ± 0.00^{ab}	n.a.
Crude protein (%)	1	n.a.	n.a.	$3.70\pm0.04^{\rm a}$	n.a.
	2	4.64 ± 0.14^{b}	4.69 ± 0.45^{ab}	$3.74\pm0.06^{\rm a}$	$3.36\pm0.21^{\rm a}$
	3	n.a.	n.a.	$3.77\pm0.20^{\rm a}$	n.a.
Phenolic content (mg GAE/g dw)	1	n.a.	n.a.	70.35 ± 4.22	n.a.
	2	41.14 ± 2.46	52.65 ± 3.17	51.49 ± 3.08	40.67 ± 2.44
	3	n.a.	n.a.	68.03 ± 5.03	n.a.
Flavonoid (mg QE/g dw)	1	n.a.	n.a.	1.72 ± 0.10	n.a.
	2	1.80 ± 0.10	1.80 ± 0.10	1.75 ± 0.10	1.39 ± 0.08
	3	n.a.	n.a.	1.74 ± 0.07	n.a.
Chlorophyll- <i>a</i> (mg/g)	1	n.a.	n.a.	0.66	n.a.
	2	0.47	0.46	0.55	2.24
	3	n.a.	n.a.	0.37	n.a.
	1	n.a.	n.a.	0.19	n.a.
Total Carotenoid	2	0.17	0.15	0.16	1.06
(mg/g)	3	n.a.	n.a.	0.14	n.a.

Table 5. The proximate compositon, total phenolics, flavonoids and pigments of J. rubens.

The values are expressed as means \pm standard deviation, n = 3, n.a.: sample was not found, *Different superscript letters in the same variable indicate significant differences, **Lower case letters indicate significant differences between seasons, and caps indicate the differences between sampling sites when only two main effects are significant at 0.05 significance level.

The flavonoid contents of seaweed species ranged from 1.39 to 10.62 mg QE/g dw. The maximum flavonoid content was found in S. schimperi collected from St.1 in the summer, followed by P. pavonica, collected from St.2 in the summer (Table 3, 4). The minimum flavonoid content was recorded in J. rubens collected from St.2 in the winter (Table 5). Sahayaraj et al. (2014) reported a similar flavonoid content for P. pavonica (11.53 mg/g dw). Similar to results obtained for S. schimperi and P. pavonica in the present study, Marinho et al. (2019) reported that the total flavonoid content of brown seaweed, Saccharina latissima reached a maximum level in the summer. However, the highest flavonoid value (4.83 mg RE g⁻¹ dm) reported by Marinho et al. (2019) for S. latissima was lower than those found for seaweed species except J. rubens in this study. The primary photosynthetic pigment, chlorophyll-a content, varied between 0.37 and 4.53 mg/g, with the lowest in J. rubens and the highest in D. dichotoma (Tables 2, 5). Both seaweeds were collected from St.3 in the autumn. This result shows that the contents of chlorophyll-a may show species-specific Palanivelu et al. differences. (2012)reported comparatively lower chlorophyll-a content in J. rubens (0.07 mg/g ww). Chakraborty and Bhattacharya (2012) found chlorophyll-a content of D. dichotoma as 1.38 mg/g from the Gulf of Kutch (India). These results are consistent with the results of the present study. The total carotenoid contents of seaweed species ranged from 0.14 to 2.83 mg/g. The highest carotenoid level was found in D. dichotoma collected from St.3 in autumn, while the lowest value was recorded for J. rubens collected from the same site in the autumn (Tables 2, 5). Similar to the results of the present study, Sukalyan and Santra (2008) found the highest carotene level in Dictyota ceylanica Kützing. Moreover, Etemadian et al. (2017) stated that there was more carotene in brown algae, as was found in this study. The carotenoid levels of seaweed species found in the present study were relatively higher than those reported by Chinnadurai *et al.* (2013) for six seaweed species (0.26 -0.63 mg/g). However, the carotenoid contents of the seaweed species in the present study were lower than those of eight different seaweeds whose carotenoid levels were found between 18.85-29.02 mg/g in Sunderban (India) (Sukalyan and Santra, 2008). Godinez-Ortega *et al.* (2008) expressed that chlorophyll-*a* and carotenoid contents of seaweeds could vary according to light intensity. Similarly, Necchi and Zucchi (2001) highlighted that environmental conditions such as temperature, light intensity and period of light might affect the pigment content.

The biochemical analyses showed marked variations among species. Although comparatively low crude protein, lipid, flavonoid and pigment levels were observed, the highest ash levels were found in J. rubens, a heavily calcified alga. Renaud and Luong-Van (2006) stated that calcified seaweeds were rich in ash but low in nutrients. This suggests that red seaweed J. rubens may be a good source of minerals (Dixit and Reddy, 2017). On the other hand, brown seaweed S. schimperi contained the highest amount of TPC and TFC, which may have potential important roles in promoting human health due to their antioxidant, anti-aging and anti-carcinogenic properties. Moreover, seaweed carotenoids are strong antioxidants that prevent cardiovascular and neurodegenerative diseases and cancer (Boominathan and Mahesh, 2015). In this study, D. dichotoma showed the highest chlorophyll-a and carotene content, while red seaweed J. rubens had the lowest pigment levels.

Conclusion

Apart from their direct consumption, the substances extracted from seaweeds can be used in many applications, such as antioxidant and antibacterial agents in the food industry and for human health. The biochemical composition of the sampled seaweeds has the potential as antioxidant and mineral sources for functional uses, but differences were observed depending on the species, geographical location and season. Among the investigated species, *S. schimperi* and *J. rubens* seem to be the best source of phenolic and minerals, respectively. More studies are needed to evaluate the nutritional value and functional properties of these seaweeds as food supplements and for other industrial uses.

Acknowledgement

This study was produced from İbrahim Gür's MSc Thesis. The authors are grateful to the Scientific Research Project Fund of Cukurova University for their support of Research Project (SÜF2013YL4). A part of this study was presented at the FABA 2014 Symposium held in Trabzon, Türkiye.

Conflict of Interest

The authors declare that they have no conflicts of interest.

Author Contributions

Sevim Polat and İbrahim Gür planned and designed the research. İbrahim Gür performed the sample collection and chemical analysis. All authors contributed to the writing of the final manuscript.

Ethics Approval

This article does not contain any studies with human or animal subjects.

References

- AOAC (1990). Official Methods of Analysis of AOAC International.15th ed. Association of Official Analytical Chemists, Method 950.46, Moisture in Meat. Washington, DC: USA.
- AOAC (1998a). Official Methods of Analysis of AOAC International. Association of Official Analytical Chemists. Method 938.08, Ash of seafood: Fish and other marine products. Gaithersburg, MD: USA.
- AOAC (1998b). Official Methods of Analysis of AOAC International. Association of Official Analytical Chemists. Method 955.04, Nitrogen (Total) in Seafood. Fish and Other Marine Products. Gaithersburg, MD: USA.
- Arnon, D.I. (1949). Copper enzymes in isolated chloroplast, polyphenol oxidase in *Beta Vulgarise*, *Plant Physiology*, 2, 1-15.
- Aysel, V., Erdugan, H., & Okudan, E.Ş. (2006a). Marine algae and seagrasses of Adana (Mediterranean, Turkey), Journal of Black Sea/Mediterranean Environment, 12, 35-57.
- Aysel, V., Erdugan, H., & Okudan, E.Ş. (2006b). Marine algae and seagrasses of Hatay Mediterranean, Turkey), *Journal of Black Sea/Mediterranean Environment*, 12, 159-179.
- Bligh, E.G., & Dyer, W.J. (1959). A rapid method of total lipid extraction and purification, *Canadian Journal Biochemistry Physiology*, 37, 911-917.
- Boominathan, M., & Mahesh, A. (2015). Seaweed carotenoids for cancer therapeutics. In: *Handbook of Anticancer Drugs from Marine Origin* (edited by S.K. Kim) (pp.185-203). Cham, Switzerland: Springer.
- Caf, F., Şen Özdemir, N., Yılmaz, Ö., Durucan, F., & Ak, İ. (2019). Fatty acid and lipophilic vitamin composition of seaweeds from Antalya and Çanakkale (Turkey), *Grasas Y Aceites 70 (3)*, 1-7. doi:10.3989/gya.0704182
- Chakraborty, S., & Bhattacharya, T. (2012). Nutrient composition of marine benthic algae found in the Gulf of Kutch coastline, Gujarat India, *Journal Algal Biomass*, *3*, 32-38.
- Chauhan, V., & Chauhan, A. (2006). Oxidative stress in Alzheimer's disease, *Pathophysiology*, 13, 195-208. doi.org/10.1016/j.pathophys.2006.05.004

- Chang, C.C., Yang, M.H., Wen, H.M., & Chern, J.C. (2002). Estimation of total flavonoid content in propolis by two complementary colorimetric methods, *Journal of Food Drug Analysis*, 10, 178-182. doi.org/10.38212/2224-6614.2748
- Chinnadurai, S., Karthik, G., Chermapandi P., & Hemalatha, A. (2013). Estimation of Major Pigment Content in Seaweeds Collected from Pondicherry Coast, *The Experiment*, 9 (1), 522-525.
- Connan, S., Deslandes, E., & Gall, E.A. (2007). Influence of day–night and tidal cycles on phenol content and antioxidant capacity in three temperate intertidal brown seaweeds, *Journal of Experimental Marine Biology* and Ecology, 349, 359-369. doi.org/10.1016/j.jembe.2007.05.028
- Dixit, D., & Reddy, C. R. K. (2017). Non-Targeted Secondary Metabolite Profile Study for Deciphering the Cosmeceutical Potential of Red Marine Macro Alga Jania rubens-An LCMS-Based Approach" Cosmetics, 4(4), 45. doi.org/10.3390/cosmetics4040045
- Etemadian, Y., Shabanpour, B., Ghaemi, V., & Kordjazi.M. (2017). Compare the chlorophyll amount in three brown algae species of the Persian Gulf by using three solvents and applying two formulas, *International Journal of Biochemistry*, *Biophysics & Molecular Biology*, 2 (6), 77-79. doi: 10.11648/j.ijbbmb.20170206.14
- Fleurence, J. (1999). Seaweed proteins: biochemical, nutritional aspects and potential uses, *Trends in Food Science Technology*, 10, 25-28. doi.org/10.1016/S0924-2244(99)00015-1
- Fox, J., & Weisberg, S. (2011). An R Companion to Applied Regression. Second Edition. Thousand Oaks CA:Sage,<u>http://socserv.socsci.mcmaster.ca/jfox/Books/</u>
- Gámez-Meza, N., Noriega-Rodriguez, J.A., Medina-Juarez, L.A. et al. (1999). Antioxidant activity in soybean oil of extracts from Thompson grape bagasse, *Journal of American Oil Chemists' Society*, 76, 1445-1447. doi.org/10.1007/s11746-999-0182-4
- Godinez-Ortega, J.L., Snooeijis, P., Robledo, D., Freile-Pelegrin, Y., & Pedersen M. (2008). Growth and pigment composition in the red alga *Halymenia floresii* cultured under different light qualities, *Journal of Applied Phycology*, 20, 253-260. doi.org/10.1007%2Fs10811-007-9241-0
- Güner, A., & Yavasoglu, N.K. (2018). Evaluation of antioxidant, antimicrobial and antimutagenic activity with irritation effects of *Ceramium rubrum* (Red Algae) extract, *International Journal of Secondary Metabolites*, 5, 279-287. doi.org/10.21448/ijsm.432654
- Haroon, A., Szaniawska, A., Normant, M., & Janas, U. (2000). The biochemical composition of *Enteromorpha* spp. from the Gulf of Gdansk coast on the southern Baltic Sea, *Oceanologia*, 116/117, 513-516.

- Hothorn, T., Bretz, F., & Westfall, P. (2008). Simultaneous inference in general parametric Models, *Biometrical Journal*, 50, 346-363. doi.org/10.1002/bimj.200810425
- İrkin, L.C., & Erdugan, H. (2016). Seasonal variation in ash, lipid and protein contents of *Scytosiphon lomentaria* Lyngbye and *Palisada perforata* Bory de Saint-Vincent along Çanakkale Strait (Dardanelles), Turkey, *Marine Science and Technology Bulletin*, 4 (2), 1-4.
- Irkin, L.C., & Erduğan, H. (2017). Investigation of seasonal variations in biochemical composition of some red algae distributed in the Strait of Çanakkale (Dardanelles), Turkey, Archive of Applied Science Research, 9, 1-8.
- Kalasariya, H.S., Yadav, V.K., Yadav, K.K., Tirth, V., Algahtani, A., Islam, S., Gupta, N., & Jeon, B-H. (2021). Seaweed-based molecules and their potential biological activities: An eco-sustainablecosmetics, *Molecules*, 26, 5313. doi.org/10.3390/molecules26175313
- Kirk, J.T.O., & Allen, R.L. (1965). Dependence of chloroplast pigments synthesis on protein synthetic effect on actilion, *Biochemical and Biophysical Research*, 27, 523-530.
- Kostetsky, E.Y., Goncharova, S.N., Sanina, N.M., & Shnyrov, V.L. (2004). Season influence on lipid composition of marine macrophytes, *Botanica Marina*, 47, 134-139. doi.org/10.1515/BOT.2004.013
- Kumar, C.S., Ganesan, P., Suresh, P.V., & Bhaskar N. (2008). Seaweeds as a source of nutritionally beneficial compounds - A review, *Journal of Food Science and Technology*, 45, 1-13.
- Kumar, M., Gupta, V., Kumari, P., Reddy, C.R.K., & Jha, B. (2011). Assessment of nutrient composition and antioxidant potential of Caulerpaceae seaweeds, *Journal of Food Composition Analysis*, 24, 270-278. doi.org/10.1016/j.jfca.2010.07.007
- Machu, L., Misurcova, L., Ambrozova, J.V., Orsavova, J., Mlcek, J., Sochor, J., & Jurikova, T. (2015). Phenolic content and antioxidant capacity in algal food products, *Molecules*, 20, 1118-1133. doi.org/10.3390/molecules20011118
- Marinho, G.S., Sørensen, A.D.M., Safafar, H., Pedersen, A.H., & Holdt, S.L. (2019). Antioxidant content and activity of the seaweed Saccharina latissima: A seasonal perspective. *Journal of Applied Phycology*, 31, 1343–1354. doi: 10.1007/s10811-018-1650-8
- Marsham, S., Scott, G.W., & Tobin, M.L. (2007). Comparison of nutritive chemistry of a range of temperate seaweeds, *Food Chemistry*, 100, 1331-1336. doi.org/10.1016/j.foodchem.2005.11.029

- Matanjun, P., Mohamed, S., Mustapha, N.M., Muhammad, K., & Ming, C.H. (2008). Antioxidant activities and phenolics content of eight species of seaweeds from north Borneo, *Journal of Applied Phycology*, 20, 367-373.doi:10.1007/s10811-007-9264-6
- Necchi, O., & Zucchi, M.R. (2001). Effects of temperature, irradiance and photoperiod on growth and pigment content in some freshwater red algae in culture, *Phycological Research*, 49, 103-114. doi.org/10.1111/j.1440-1835.2001.tb00240.x
- Nedumaran, T., & Arulbalachandran, D. (2015). Seaweeds: A Promising Source for Sustainable Development. In: Thangavel P, Sridevi G (Eds.) Environmental Sustainability (pp. 65-88).
- Nelson, M.M., Phleger, C.F., & Nichols, P.D. (2002). Seasonal lipid composition in macroalgae of the Northeastern Pacific Ocean, *Botanica Marina*, 45, 58-65. doi.org/10.1515/BOT.2002.007
- Palanivelu, A., Darsis, A., & Arunkumar, K. (2012). Nutraceutical values of seaweeds found along the Coast of Thondi (Palk Bay, India) with specific investigation on fatty acids methyl esters through GC/MS, *Journal of Green Bioenergy*, 1, 3-18.
- Peñalver, R., Lorenzo, J.M., Ros, G., Amarowicz, R., Pateiro, M., & Nieto, G. (2020). Seaweeds as a functional ingredient for a healthy diet, *Marine Drugs*, 18, 1-27. doi.org/10.3390/md18060301
- Parthiban, C., Saranya, C., Gırija, K., Hemalatha, A., Suresh, M., & Anantharaman, P. (2013). Biochemical composition of some selected seaweeds from Tuticorin coast, *Advances in Applied Science Research*, 4, 362-366.
- Polat, S., & Özoğul, Y. (2008). Biochemical composition of some red and Brown macroalgae from the Northeastern Mediterranean Sea. *International Journal* of Food Science and Nutrition, 59, 566-572. doi.org/10.1080/09637480701446524
- Polat, S., & Özoğul, Y. (2009). Fatty acid, mineral and proximate composition of some seaweeds from the northeastern Mediterranean coast, *Italian Journal Food Science*, 21, 317-324.
- Polat, S., Özoğul Y., & Küley Boğa, E. (2012). Protein, lipid and fatty acid composition of some brown and red seaweeds from the coast of İskenderun Bay (Northeastern Mediterranean), *Journal of FisheriesSciences.com*, 6, 107-113. doi: 10.3153/jfscom.2012014
- Polat, S., & Özoğul, Y. (2013). Seasonal proximate and fatty acid variations of some seaweeds from the northeastern Mediterranean coast, *Oceanologia*, 55, 375-391. doi.org/10.5697/oc.55-2.375
- R Core Team (2018). R: A language and environment for statistical computing. R Foundation for Statistical Computing Vienna Austria. https://www.R-project.org/

- Radha, P. (2018). Proximate analysis and mineral composition of seaweeds of Manamelkudi coast, Pudukkottai District, India, *International Journal of Current Microbiology and Applied Sciences*, 7(8): 3121-3128. doi.org/10.20546/ijcmas.2018.708.333
- Ratana-arporn, P.A., & Chirapart, A. (2006). Nutritional evaluation of tropical green seaweeds *Caulerpa lentillifera* and *Ulva reticulate, Kasetsart Journal Natural Science, 40*, 75-83.
- Renaud, S.M., & Luong-Van, J.T. (2006). Seasonal variation in the chemical composition of tropical Australian marine macroalgae, *Journal of Applied Phycology*, 18, 381-387. doi.org/10.1007/s10811-006-9034-x
- Rupérez, P. (2002). Mineral content of edible marine seaweeds, *Food Chemistry*, 79, 23-26. doi.org/10.1016/S0308-8146(02)00171-1
- Sahayaraj, K., Asharaja, A.C., Rajesh, S., & Martin Rathi, J.A. (2014). Qualitative and quantitative profiles of secondary metabolites of chosen Chlorophyta and Ochrophyta from Gulf of Mannor, *Cahiers de Biologie Marine*, 55, 69-76.
- Saygılı, E. I., Naz, M., Okudan, E. S., Çetin, Z., Benlier, N., Öğüt, E., Güngör, M., Bakır, S.B., Karadeniz, P.G., Veziroglu, S., Gülses, A., Depci, T., Aktas, O.C., & Sayın, S. (2022). The determination of the molecular weight profiles and biochemical compositions eight macroalgae species from Turkey, *International Aquatic Research*, 14(2), 117-125. doi:10.22034/iar.2022.1949245.1226
- Sukalyan, C., & Santra, S.C. (2008). Biochemical composition of eight benthic algae collected from Sunderban, *Indian Journal of Marine Sciences*, 37, 329-332.
- Tabarsa, M., Rezai, M., Ramezanpour, Z., Waaland, J.R., & Rabirei, R. (2012). Fatty acids, amino acids, mineral contents and proximate composition of some brown seaweeds, *Journal of Phycology*, 48, 285-292. doi.org/10.1111/j.1529-8817.2012.01122.x
- Taşkın, E. (2014). Comparison of the brown algal diversity between four sea coasts of Turkey, *Journal of Academic Documents for Fisheries and Aquaculture. 3*, 145-150.
- Turan, F., Özgun, S., Sayın, S., & Özyılmaz, G. (2015). Biochemical composition of some red and green seaweeds from Iskenderun Bay, the northeastern Mediterranean coast of Turkey, *Journal of Black Sea/Mediterranean Environment*, 21, 239-249.
- Thirumaran, G., Manivannan, K., Karthikai, D.G., Ananthamaran, P., & Balasubramanian, T. (2009). Photosynthetic pigments of different colour strains of the cultured seaweeds *Kappaphycus alvarezii* (Doty) ex P.Silva in Valler Estuary, *Academic Journal of Plant Science*, 2, 150-153.

- Wang, B.G., Zhang, W.W., Duan, X.J., & Li, X.M. (2009). In vitro antioxidative activities of extract and semipurified fractions of the marine red alga, *Rhodomela confervoides* (Rhodomelaceae), *Food Chemistry*, 113, 1101-1105. doi.org/10.1016/j.foodchem.2008.08.078
- Yeşilova, K., Balkis, N., & Taşkın, E. (2017). Seasonal investigation of the protein, carbohydrate and lipid contens of dominant macroalgae on the western coast of the Black Sea. *Fresenius Environmental Bulletin*, 26 (1), 46-55.
- Yılmaz, M., Türker, G., & Ak, İ. (2021). The Effect of Different Solvents on Antioxidant Properties of Gongolaria barbata (Phaeophyceae). Çanakkale Onsekiz Mart University Journal of Marine Sciences and Fisheries, 4(2), 197-201. doi:10.46384/jmsf.1021387
- Zeileis, A. (2004). Econometric computing with HC and HAC covariance matrix estimators, *Journal of Statistical Software*, 11, 1-17. doi.org/10.18637/jss.v011.i10