



A continuous publication, open access, peer-reviewed journal

ACCESS ONLINE

REVIEW

The multiple faces of ketamine in anaesthesia and analgesia

Silvia Natoli¹

¹Department of Clinical Science and Translational Medicine and Unit of Pain Therapy, Polyclinic of Tor Vergata, University of Rome, Tor Vergata, Rome, Italy

Abstract

Objective: Ketamine is an anaesthetic agent with a unique dissociative profile and pharmacological effects ranging from the induction and maintenance of anaesthesia to analgesia and sedation, depending on the dose. This article provides information for the clinical use of ketamine in anaesthesia, in both conventional and special circumstances.

Methods: This is a non-systematic review of the literature, through a PubMed search up to February 2021.

Results: With a favourable pharmacokinetic profile, ketamine is used in hospital and prehospital settings for emergency situations. It is suitable for patients with many heart conditions and, unlike other anaesthetics, its potential for cardiorespiratory depression is low. Furthermore, it may be used when venous access is difficult as it may be administered through various routes. Ketamine is the anaesthetic of choice for patients with bronchospasm thanks to its bronchodilatory and antiinflammatory properties.

Conclusion: With a favourable pharmacokinetic profile, ketamine is used in hospital and prehospital settings for emergency situations and is suitable for patients with many cardiac and respiratory conditions.

Keywords: anaesthesia, dissociative profile, ketamine.

Citation

Natoli S. The multiple faces of ketamine in anaesthesia and analgesia. Drugs in Context 2021; 10: 2020-12-8. DOI: 10.7573/dic.2020-12-8

Introduction

Ketamine is an anaesthetic agent with a unique dissociative profile, with pharmacological effects ranging from the induction and maintenance of anaesthesia to analgesia and sedation, depending on the dose. Additional effects include bronchodilation, stimulation of the sympathetic nervous system, catalepsy and psychiatric effects, including rapid and sustained antidepressant activity.^{1–4} Although these activities may be valuable in anaesthesia representing interesting advantages for special patient subgroups, such as those with respiratory or cardiovascular conditions, a possible psychotropic activity and other central effects have limited the use of ketamine as an anaesthetic in clinical practice.

This non-systematic review of the literature presents useful information for the clinical use of ketamine in anaesthesia, in both conventional and special circumstances.

Methods

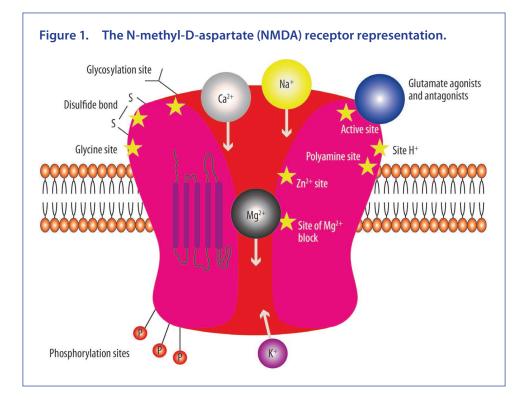
For this review of the literature, a non-systematic search was performed in PubMed using the following keywords to retrieve

pharmacological data: "ketamine", "NMDA", "GABA", "receptor", "pharmacodynamics", "pharmacokinetics", "pain", "central nervous system". A search with the keywords "ketamine", "anesthesia", "administration route", "bronchodilation", "hemodynamics", "congenital heart disease", "burn", "pain", "emergence phenomena" and "adverse event" was performed to review clinical uses. Articles in English, published up to February 2021, were included.

Review

Pharmacological aspects

Since its synthesis in 1962, several clinical applications have been described for the phencyclidine derivative ketamine, including anaesthesia, pain management and psychiatry.⁵ Many therapeutic activities of ketamine have been linked to its antagonism to the N-methyl-D-aspartate (NMDA) receptor (NMDAR) (Figure 1).⁶ In the early 1980s, it was discovered that ketamine blocks the NMDAR by binding to a specific site (referred to as the phencyclidine (PCP) site) in an noncompetitive way.⁷ As PCP is localized inside the NMDA



channel, it can be reached and bound to only when the receptor is activated.⁸ Furthermore, the ability of ketamine to bind to and dissociate from the PCP binding site in vivo is dictated by the degree of activation of the receptor, which depends on glutamate release in the synapses, cell membrane depolarization and the levels of other modulatory factors (Figure 2).⁹ The affinity of ketamine to the NMDAR is similar to that of other non-competitive NMDA antagonists. In rodents, higheraffinity NMDA antagonists determine neurotoxicity, neuronal vacuolization and neurodegeneration, although this has not been demonstrated in primates.^{10,11} High-affinity compounds, which are administered in the therapeutic dose range, determine learning and memory impairment, sedation, ataxia, and psychotomimetic effects, such as hallucinations in humans, whereas low-affinity blockers, such as memantine, seem to have a better therapeutic index and an activity similar to that of magnesium, which is an endogenous NMDA channel blocker.^{10,12}

The effects of ketamine on the central nervous system (CNS) seem to go beyond NMDAR blocking; several molecular targets and neurophysiological properties are known, although many mechanisms of action remain to be understood. Ketamine interacts with opioid receptors^{13,14} and blocks monoaminergic reuptake¹⁵ and muscarinic receptors^{16,17} as well as voltage-sensitive ion channels (Table 1).^{18,19}

It is well known that the affinity of ketamine to opioid receptors may be relevant at high doses and related to its anaesthetic activity, whilst low doses (≤ 0.3 mg/kg intravenous (i.v.)) have an analgesic effect.^{20–22} Topically administered ketamine displays local anaesthetic properties due to the ability to block the conductance of ion channels.²³ Due to the lipid solubility of ketamine and its relatively low protein binding (about 20–50%), a considerably large volume of distribution (3–5 L/kg) is attained after either an i.v. or an intramuscular (i.m.) bolus dose.²⁴ In addition, ketamine quickly crosses the blood–brain barrier, and concentrations in the cerebrospinal fluid may be four- to five-fold higher than in plasma.² Due to these pharmacokinetic features, the analgesic effect of ketamine has a rapid onset.²⁵

Ketamine is mainly metabolized in the liver and several metabolites have been identified. Although there are some dissonant results regarding the contribution of enzymes to clinical ketamine metabolism, CYP2B6, CYP3A4 and CYP2C9 contribute to the production of norketamine via ring hydroxylation and N-demethylation pathways.²⁶ This primary metabolite is pharmacologically active, with 30% of the anaesthetic and analgesic potency compared with the parent compound, and is further metabolized to 4-, 5- and 6-hydroxynorketamine by CYP2B6 and CYP2A6.²⁷ Children are known to require relatively higher doses of ketamine compared with adults although the pharmacokinetics were found to be similar.²⁸ It is possible that pharmacokinetic modelling may not apply to the paediatric population as analyses were scaled to a standardized 70 kg patient. Thus, dosing by titration to effect is advisable in children whilst dosing by body weight may not be reliable. Elderly patients behave as poor metabolizers, hence a lower dosing is recommended.²⁹ In addition, the effect of metabolic enzyme variants or sex on pharmacokinetics is still unknown.³⁰

Ketamine can be administered through various routes. The more conventional route is i.v. but i.m. injection can be used

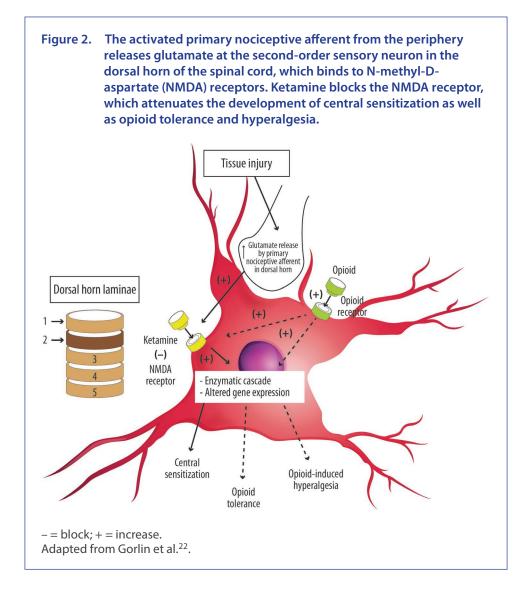


Table 1. Pharmacological actions of ketamine.

Molecular target	Mechanism	Potency (µM)	References
NMDA receptor (PCP site)	Antagonism	K _i 0.4–46 IC ₅₀ 1.6–6.2	6,7,9,138
μ-Opioid receptors	Agonism	K _i 27	13,14,139
δ-Opioid receptor	Agonism	K _i 101	13,139
k-Opioid receptor	Agonism	K _i 85	13
Sigma receptor	Agonism	K _i 66	13
Noradrenaline transporter	Inhibition of reuptake	K _i 67	14,140
Dopamine transporter	Inhibition of reuptake	K, 162	140
Serotonin transporter binding	Inhibition of reuptake	K, 67	140
Muscarinic, nicotinic receptor	Antagonism	IC ₅₀ : >50 or 100	16,17
Voltage-dependent Na ²⁺ , Ca ²⁺ channels	Block	K _i 67	19,21
Dopamine D ₂	Partial agonism	K _i 0.5	141,142
Serotonin $5HT_2$	Antagonism	K _i 15	141

when venous access is not available, retaining a satisfactory bioavailability (93%).²⁴

Ketamine is used in children and adults as an anaesthetic agent for diagnostic and surgical procedures as either i.v. infusion, i.v. injection or i.m. injection. It is often used for short procedures, but it can be used for longer procedures with additional doses or by i.v. infusion. If skeletal muscle relaxation is desired, a muscle relaxant can be used together with ketamine. Ketamine is recommended for the induction of anaesthesia prior to the administration of other general anaesthetic agents and to supplement other anaesthetic agents. To induce anaesthesia, an infusion of 0.5-2 mg/kg is typically administered with a duration of action of ~5–10 min. Anaesthesia may be maintained using a microdrip infusion of $10-45 \mu$ g/kg/min.³¹

The role of NMDAR in the CNS

Some considerations on the role of NMDA in the CNS may be useful to understand the anaesthetic activity of ketamine. NMDARs are widely expressed in the CNS and play critical roles in excitatory synaptic transmission, excitotoxicity and plasticity by participating in neuronal regeneration and circuit formation, coordinating functional circuits and controlling dendritic growth. Hence, NMDARs are essential to memory, to learning and during development. Excitotoxic events involving NMDARs have also been linked to degenerative diseases such as Alzheimer disease and Huntington disease.

The blockade of NMDARs is neuroprotective in animal models of both stroke and seizure but the therapeutic use of NMDA antagonists has failed in humans due to the development of severe side-effects because NMDARs are essential to physiological neuronal function.^{32,33}

An important element is that the NMDAR hypofunction produced by any mechanism can be psychotogenic, possibly resulting in dopaminergic hyperactivity and behavioural changes characteristic of schizophrenia. However, the mechanism linking NMDAR blockade by ketamine and psychosis remains to be established.³²

The role of NMDAR in pain

Peripheral nociceptor activation by high-energy stimuli can evoke pain. On peripheral nociceptor activation, glutamate is released in the dorsal horns of the spinal cord and binds to postsynaptic glutamate ionotropic AMPA and kainate subtype receptors that generate excitatory postsynaptic currents. The summation of multiple sub-threshold excitatory postsynaptic currents in the postsynaptic neuron induces firing of the action potential and transmission of the pain message to second-order nociceptive projecting neurons. However, pain responses are enhanced following repetitive stimulation of nociceptive afferents, leading to the well-known 'wind-up' phenomenon, a progressive increase of nociceptive response to each successive stimulus.^{34,35} Wind-up is a form of shortlasting synaptic plasticity that leads to nociceptive pathway

potentiation. In humans, pain wind-up results from a temporal summation of either subjective pain intensity or nociceptive flexor reflexes evoked by repetitive noxious stimuli and can be prevented by low-dose ketamine-blocking NMDARs. Wind-up can only be evoked if small-calibre afferents are involved, 35,36 eliciting the release of substance P and calcitonin generelated peptide along with glutamate in the dorsal horn.^{37–41} The presence of both glutamate and neuropeptides in the synaptic cleft induces a relatively more prolonged postsynaptic depolarization compared to glutamate alone. Under these conditions, the normally inactive NMDA glutamate receptor unblocks and lets calcium in. Indeed, at normal resting membrane potentials, NMDAR is blocked by magnesium ions in a voltage-dependent manner. Sustained membrane depolarization can reduce the blockage because a lower electrical gradient may force magnesium into the channel. Thus, a prolonged and lasting synaptic activity can amplify current flow through open NMDA channels. When nociceptive inputs are intense and prolonged (i.e. induce high-frequency nociceptor firing), NMDAR is involved, inducing wind-up in second-order nociceptive neurons and leading to short-term central sensitization.⁴² In the context of inflammation or tissue injury, this short-living synaptic plasticity induces secondary hyperalgesia, which may be an excessive response to nociceptive stimuli out of the primary injury site or stimulation site that can endure until healing occurs and inflammation fades. However, repetitive primary afferent stimulation at frequencies much higher than those that evoke wind-up may induce long-term potentiation (LTP) of the nociceptive pathway.⁴³ LTP is a well-described event in several neural networks and is the neural basis of processes such as learning and memory. LTP mechanisms include the NMDA-mediated elevation of cytosolic Ca²⁺ in the postsynaptic neuron and subsequent downstream activation of signalling pathways and second messenger systems such as kinases (such as MAPK, PKA, PKC, PI3K and Src) as well as the release of nitric oxide by Ca²⁺-activated neuronal nitric oxide synthase and the release of prostaglandins by cyclooxygenase enzymes, which may further increase the excitability of these neurons in the long term.^{44,45} Together, these downstream effects of NMDA activation result in the amplification of pain messages.⁴⁶

Under normal conditions, high-frequency firing able to induce LTP does not usually occur in nociceptive C-fibres. However, bursts of ectopic activity recorded from nociceptive primary afferents in pain patients with nerve injury can be sufficient to trigger sustained NMDA activation and LTP, which is the neural basis of chronic pain within the spinal cord.⁴⁷

Central sensitization is a major pathophysiological event common to inflammatory and neuropathic pain. It is important to understand that central sensitization is a physiological and reversible adaptive mechanism during inflammatory pain, whereas it is a pathological, hardly reversible and maladaptive event when neuropathic pain occurs. However, the diverse events that converge onto the mechanism of NMDAR-mediated pronociceptive plasticity and central sensitization can potentially lead to chronic pain regardless of the trigger. When peripheral tissue damage occurs, the subsequent inflammatory process induces changes in peripheral nociceptive endings, resulting in peripheral activation and sensitization and a further increased firing rate that leads to rapid-onset homosynaptic and heterosynaptic facilitation in the dorsal horn of the spinal cord in a short-term wind-up-like manner. However, for some reason, the physiological short-term synaptic potentiation may turn into a longer-term LTP-like increase in synaptic strength and maintenance of central sensitization and hence into chronic pain, which most frequently occurs as a result of maladaptive repair of the injured nervous system in neuropathic pain.⁴⁸

Clinically, manifestations of central sensitization are hyperalgesia (an increased pain response to mild noxious stimuli) and allodynia (abnormal pain caused by normally innocuous stimuli) and reduction in opioid responsiveness or opioid-induced hyperalgesia, in both neuropathic and inflammatory pain. This state can be prevented or reduced by i.v. infusion of low-dose ketamine either in the setting of acute inflammatory pain^{49–54} or in chronic pain conditions, including osteoarthritic and rheumatoid pain, neuropathic pain, fibromyalgia, irritable bowel syndrome and migraine.^{55–66}

Based on this rationale, ketamine – the most potent of all NMDA antagonists currently available for use in humans – has been used in various pain states, including acute, chronic and neuropathic pain (Figure 2).

Ketamine advantages in anaesthesia

Ketamine has many attributes yet many drawbacks. It affects the CNS by producing a unique dissociative state wherein a patient's eyes are open but disconnected from the surroundings, in a cataleptic condition with strong analgesia and sedation.⁶⁷

Ketamine has undoubted advantages for anaesthetists (Table 2). Its unique pharmacokinetics properties that provide a high bioavailability (between 100% when administered by the i.v. route and 93% for i.m. administration) have led to its prevalent use in hospital and prehospital environments for emergencies. Ketamine can also be applied via rapid-sequence induction, producing dissociative anaesthesia ~1–2 min after administration.

Unlike other general anaesthetic agents, ketamine shows no direct interaction with GABA receptors at clinically relevant concentrations. Indeed, subanaesthetic doses of ketamine do not bind to GABA-A receptors in the human brain,⁶⁸ and anaesthetic concentrations of ketamine do not alter GABA-A receptor function *in vitro*.⁶⁹ Thus, at least at subanaesthetic doses, the effect of ketamine on GABA-A receptor activity might only be indirect. As a consequence, relevant cardiorespiratory depression after induction is unlikely, especially if ketamine is given slowly or as monotherapy.⁷⁰

More recent literature highlights that ketamine may produce a dose-dependent GABA release in specific brain cortical areas, thus altering the overall glutamate–GABA balance. Another work suggests that ketamine may decrease GABA release by blocking NMDAR located on GABAergic interneurons, thus increasing cortical excitability.⁷¹ These latter mechanisms may underlie the antidepressant effects of ketamine in treatment-resistant depression.⁷¹

The possibility to administer ketamine by different routes has prompted its use when venous access is difficult such as in trauma patients with hypovolemic shock. Indeed, the low cardiorespiratory depressant effects and sympathomimetic effects of ketamine render this drug a valid alternative to other anaesthetic agents in trauma patients as well as in septic shock patients, as several clinical reports have indicated that ketamine produced either no change or a slight increase in arterial pressure and heart rate.^{1,72} Indeed, it has been observed that ketamine can improve the blood gas and pulmonary function index of patients with acute lung injury caused by mechanical ventilation.⁷³ With a wide variation in individual response, ketamine leads to increased blood pressure, stroke volume and

Characteristic	Advantage	References
Dissociative sedation	Strong sedation and analgesia, useful for emergency in uncooperative subjects	
High bioavailability	Rapid action, for emergency in prehospital or hospital setting	108
No direct interaction with GABA receptors	Cardiorespiratory depression is unlikely	
Multiple administration routes	Useful when venous access is difficult	108
Bronchodilatory activity	Anaesthesia in patients with bronchospasm	29
Preserves haemodynamic stability	Anaesthesia in patients with congenital heart disease, shocked and hypotensive subjects	76,115,119
Analgesic activity	Useful for postoperative pain control	129–132

heart rate, and maintains systemic vascular resistance. These effects are commonly observed at a maximum of ~2 min after the injection and resolve over 15–20 min. However, severe hypotension after a ketamine bolus dose has been described as the loss of sympathoadrenal activity that accompanies the loss of consciousness.⁷⁴

The haemodynamic changes induced by ketamine make it suitable for patients with congenital heart defects and other cardiac conditions.^{75,76} However, it must be mentioned that ketamine is contraindicated in patients with serious myocardial disease or serious heart failure and when blood hypertension or increased myocardial oxygen consumption may be dangerous.³¹

Due to its bronchodilatory properties, ketamine is the anaesthetic of choice for patients with bronchospasm^{29,77,78} and has also been successfully used as a medication in the treatment of status asthmaticus.⁷⁹ Joint to anticholinergic and spasmolytic actions, ketamine may have some antiinflammatory effects, which may contribute to its efficacy in asthma patients.^{80–82} Other mechanisms are the inhibition of catecholamine uptake and L-type Ca²⁺ channel blocking,^{83–86} although the precise mechanism by which ketamine induces airway muscle relaxation is still to be elucidated.

Drawbacks of ketamine in anaesthesia

NMDAR occupancy is related to the potential of ketamine to produce adverse symptoms.⁸⁷ Emergence phenomena, delusions, hallucinations, delirium and confusion, sometimes described as 'out of body' and 'near-death experiences', are amongst the adverse effects related to the use of ketamine.⁸⁸ These events are more common in patients older than 16 years, in women, during shorter operative procedures and when large doses of ketamine are administered quickly.^{89,90} Benzodiazepines effectively prevent these disturbing psychotic phenomena. Midazolam reduced the incidence of unpleasant dreams when compared with diazepam (number needed to treat of 6).^{91,92} Propofol, lorazepam and diazepam are also effective.⁹³ In clinical practice, and in the opinion of some authors, the regular coadministration of benzodiazepines needs to be increased.⁹⁴ Finally, a recent trial (n=100) reported that a positive persuasion may reduce unpleasant sensations.95

Intracranial pressure is increased during ketamine use. Cerebral blood flow is increased secondary to a decrease in cerebral vascular resistance. Hence, ketamine should be avoided in patients with intracranial disease or abnormal cerebral blood flow.⁹⁶

Psychotic symptoms similar to schizophrenia have been described in association with ketamine activity; these need to be managed to reduce any undesired effects.^{87,97} Moreover, semantic and episodic memory may be impaired by subanaesthetic doses of ketamine.^{98,99}

Other adverse effects have been described after ketamine administration. These include nausea and vomiting in 5–15% of patients¹⁰⁰ and hypersalivation, which can be anticipated

by atropine.¹⁰¹ Limb purposeless movements and clonus have been occasionally reported.¹⁰⁰

Anaesthetic effects

During the ketamine-induced dissociative state, patients may appear awake with preserved airway reflexes and respiratory drive, but they are unable to respond to sensory input.^{102,103}

Clinical use of ketamine

In 1966, the anaesthetic effects of ketamine were reported for the first time in 130 patients aged 6 weeks to 86 years undergoing a total of 133 surgical procedures.¹⁰⁴ Ketamine produced profound and rapid analgesia with a unique state of altered consciousness; its duration of effect was limited but could be safely prolonged with repeated administration.¹⁰⁵

A well-established use of ketamine is anaesthesia induction in the emergency setting in shocked or hypotensive patients.¹⁰⁴ Ketamine was used for anaesthetic induction and maintenance in patients with cardiac tamponade and restrictive pericarditis.¹⁰⁶ A study showed that ketamine was as safe and effective as etomidate for endotracheal intubation in critically ill patients with sepsis.¹⁰⁷

Ketamine is considered the i.v. anaesthesia induction agent of choice in patients with active bronchospasm because of its bronchodilating properties and allowing the use of high oxygen concentrations.¹⁰⁸

Ketamine is the anaesthetic drug of choice for the induction of patients with congenital heart disease with a right to left shunt because it increases systemic vascular resistance, resulting in a reduced right to left shunt.¹⁰⁹ In a study, i.v. or i.m. ketamine as induction agents did not significantly affect the proportion of SaO₂ in patients with Fallot's tetralogy.¹⁰⁹ Finally, ketamine preserves intraoperative and postoperative haemodynamic stability in patients with congenital heart disease and is an alternative to sevoflurane.^{76,110}

Ketamine has a major role in repeated anaesthesia for burn dressings and for sedation during excision and grafting, both in adults and in children.^{111–113} The major advantage of ketamine in patients with burns is that it usually preserves airway and spontaneous respiratory function whilst providing good sedation and analgesia.^{111,114} In addition, the venous access may be difficult in patients with extensive burns and ketamine i.m. administration may be useful.¹¹⁵ Ketamine can be used for burn dressings in adults and children in combination with midazolam/dexmedetomidine or with propofol to obtain effective sedoanalgesia without any significant side effects.^{111,116}

Low-dose ketamine alone (5–25 mg/kg/min infusion) can be used for sedation and analgesia during local or regional anaesthetic procedures.^{106,117} Low doses (i.v. 0.5 mg/kg) may be combined with i.v. diazepam or midazolam for local and regional anaesthesia techniques, including spinal anaesthesia in adults and children.¹⁰⁶ Prophylactic i.v. ketamine 0.5 mg/kg before neuraxial blockade decreases the incidence of shivering, improves the haemodynamic profile, provides good sedation and prevents recall.¹¹⁸ Ketamine 1 mg/kg i.v. given before spinal anaesthesia results in good haemodynamic stability in elderly patients undergoing transurethral resection of the prostate.¹¹⁹

Several authors found that ketamine reduced postoperative pain.^{120–123} Perioperative low-dose ketamine was found to improve postoperative analgesia following caesarean delivery with general anaesthesia. In a randomized study on 52 women, a ketamine bolus of 0.5 mg/kg i.v. was administered at the time of induction of general anaesthesia. After induction, a ketamine infusion of 0.25 mg/kg/h was started and discontinued at the end of surgery.¹²⁴ Low-dose regimens (0.25–0.5 mg/kg i.v. as an initial bolus followed by 50–500 µg/kg/h) have been proposed for postoperative analgesia and for the reduction of exogenous opioid-induced hyperalgesia.¹²⁵ In a recent review on postoperative pain, it was found that ketamine administered in addition to opioids for i.v. patient-controlled analgesia significantly reduced pain scores, cumulative morphine consumption and postoperative desaturation in patients undergoing thoracic surgery.¹²⁶

Post-tonsillectomy pain was controlled by low-dose ketamine 0.5 mg/kg i.v./subcutaneous at the end of surgery.¹²⁷ After tonsillectomy, ketamine 0.5 mg/kg added to fentanyl 1 mg/kg improved analgesia without delaying hospital discharge in a study on 60 children.¹²⁸

In a prospective randomized study on 30 patients undergoing coronary artery bypass grafting surgery, the combination of ketamine compared with propofol, with midazolam and fentanyl for the induction of anaesthesia provided better haemodynamic stability during induction and until the end of sternotomy.¹²⁹

A systematic review with a meta-analysis of 12 randomized clinical trials, which included patients undergoing major or minor surgery, assessed the effectiveness of ketamine in reducing morphine consumption and pain intensity scores after remifentanil-based general anaesthesia.¹³⁰ Ketamine reduced the use of morphine in the first 24 postoperative hours whilst postoperative pain intensity was improved in the first 2 hours in the minor and major surgery groups. In addition, ketamine significantly reduced pain intensity in the first 24 hours in the minor surgery group. Patients administered with ketamine and undergoing major surgery had a longer time to the first rescue analgesia.¹³⁰

Use of ketamine in children

Traditionally, ketamine is considered the agent of choice in children.¹⁰² It is suitable for use in paediatrics for analgesia, procedural sedation and anaesthesia, overcoming some barriers to achieving adequate paediatric analgesia, such as a culture of underdosing and difficulty obtaining i.v. access, thanks to the possibility of using a number of routes of administration and the pharmacokinetic profile.¹⁰²

No major side effects were reported when ketamine was used in 164 awake non-trapped children with blunt trauma for procedural sedation and analgesia.¹³¹

In general anaesthesia, ketamine was safely used in addition to fentanyl or other anaesthesia induction, improving analgesia and intubation conditions and preserving haemodynamics in children with congenital heart or oncological disease.^{110,128,132,133} Ketamine 0.25 mg/kg reduced sevofluraneinduced postoperative agitation (*n*=60), whilst propofol 1 mg/kg was not effective.¹³⁴ Less agitation compared with saline was also reported by two other studies.^{135,136}

Ketamine in combination with propofol ensured stable haemodynamics, with reduced incidence of adverse events compared to single agents, during anaesthesia induction in 120 children subjected to short-term elective and urgent interventions.¹³⁷

Finally, it may be mentioned that the incidence of psychotic phenomena at awakening is lower in children than in adults.^{31,137}

Conclusion

Ketamine can be considered as the most versatile drug for anaesthesia. It can be used solely or in combination with other coadjuvant drugs, increasing their efficacy. Ketamine has been widely used in several clinical settings due to its specific properties, including its neuroprotective and anti-inflammatory effects. In addition, subanaesthetic regimens of ketamine represent a great clinical advantage. With a favourable pharmacokinetic profile, ketamine is used in hospital and prehospital settings for emergency situations. It is suitable for patients with many cardiac conditions.^{75,76} It may be used when venous access is difficult as it may be administered through different routes.³¹ Finally, it is the anaesthetic of choice for patients with bronchospasm thanks to its bronchodilatory and anti-inflammatory properties.²⁹

Contributions: SN searched the literature, evaluated it, and prepared the manuscript. The author meets the International Committee of Medical Journal Editors (ICMJE) criteria for authorship for this article, takes responsibility for the integrity of the work as a whole, and has given her approval for this version to be published.

Disclosure and potential conflicts of interest: In the past 2 years, SN received honoraria for an advisory role for Grunenthal Italia, Sandoz and Angelini Spa; she also received honoraria to participate in a speakers' bureau for Mylan. The International Committee of Medical Journal Editors (ICMJE) Potential Conflicts of Interests form for the authors is available for download at: https://www.drugsincontext.com/wp-content/uploads/2021/03/dic.2020-12-8-COI.pdf

Acknowledgements: Editorial assistance was provided by Laura Brogelli and Aashni Shah (Polistudium SRL, Milan, Italy). Unconditioned support to this assistance was provided by Minerva Medica and Molteni Farmaceutici.

Funding declaration: Minerva Medica and Molteni Farmaceutici (Firenze, Italy) funded editorial assistance to the article.

Copyright: Copyright © 2021 Natoli S. Published by *Drugs in Context* under Creative Commons License Deed CC BY NC ND 4.0 which allows anyone to copy, distribute, and transmit the article provided it is properly attributed in the manner specified below. No commercial use without permission.

Correct attribution: Copyright © 2021 Natoli S. https://doi.org/10.7573/dic.2020-12-8. Published by *Drugs in Context* under Creative Commons License Deed CC BY NC ND 4.0.

Article URL: https://www.drugsincontext.com/the-multiple-faces-of-ketamine-in-anaesthesia-and-analgesia

Correspondence: Silvia Natoli, Roma Tor Vergata University, Clinical Science and Translational Medicine Department, Via Montpellier 1, 00133 Roma, Italy. Email: silvia.natoli@uniroma2.it

Provenance: Submitted; externally peer reviewed.

Submitted: 17 December 2020; Accepted: 8 March 2021; Publication date: 23 April 2021.

Drugs in Context is published by BioExcel Publishing Ltd. Registered office: Plaza Building, Lee High Road, London, England, SE13 5PT.

BioExcel Publishing Limited is registered in England Number 10038393. VAT GB 252 7720 07.

For all manuscript and submissions enquiries, contact the Editorial office editorial@drugsincontext.com

For all permissions, rights and reprints, contact David Hughes david.hughes@bioexcelpublishing.com

References

- 1. Domino EF, Chodoff P, Corssen G. Pharmacologic effects of CI-581, a new dissociative anesthetic, in man. *Clin Pharmacol Ther*. 1965;6:279–291. https://doi.org/10.1002/cpt196563279
- 2. Zanos P, Moaddel R, Morris PJ, et al. Ketamine and ketamine metabolite pharmacology: insights into therapeutic mechanisms. *Pharmacol Rev.* 2018;70(3):621–660. https://doi.org/10.1124/pr.117.015198
- 3. Dincer B, Halici Z, Cadirci E. Investigation of the role of stimulation and blockade of 5-HT7 receptors in ketamine anesthesia. *J Mol Neurosci*. 2020. https://doi.org/10.1007/s12031-020-01732-3
- 4. Mihaljević S, Pavlović M, Reiner K, Ćaćić M. Therapeutic mechanisms of ketamine. *Psychiatr Danub*. 2020;32(3–4):325–333. https://doi.org/10.24869/psyd.2020.325
- 5. Li L, Vlisides PE. Ketamine: 50 years of modulating the mind. *Front Hum Neurosci*. 2016;10:612. https://doi.org/10.3389/fnhum.2016.00612
- 6. Lodge D. The history of the pharmacology and cloning of iono-tropic glutamate receptors and the development of idiosyncratic nomenclature. *Neuropharmacology*. 2009;56:6–21. https://doi.org/10.1016/j.neuropharm.2008.08.006
- Zukin SR, Fitz-Syage ML, Nichtenhauser R, Zukin RS. Specific binding of [3H]phencyclidine in rat central nervous tissue: further characterization and technical considerations. *Brain Res.* 1983;258(2):277–284. https://doi.org/10.1016/0006-8993(83)91151-4
- 8. MacDonald JF, Bartlett MC, Mody I, et al. Actions of ketamine, phencyclidine and MK-801 on NMDA receptor currents in cultured mouse hippocampal neurones. *J Physiol*. 1991;432:483–508. https://doi.org/10.1113/jphysiol.1991.sp018396
- 9. Parsons CG, Danysz W, Quack G. Glutamate in CNS disorders as a target for drug development: an update. *Drug News Perspect*. 1998;11(9):523–569. https://doi.org/10.1358/dnp.1998.11.9.863689
- 10. Danysz W, Zajaczkowski W, Parsons CG. Modulation of learning processes by ionotropic glutamate receptor ligands. *Behav Pharmacol.* 1995;6(5):455–474. https://doi.org/10.1097/00008877-199508000-00007
- 11. Low SJ, Roland CL. Review of NMDA antagonist-induced neurotoxicity and implications for clinical development. *Int J Clin Pharmacol Ther.* 2004;42(1):1–14. https://doi.org/10.5414/CPP42001
- 12. Parsons CG, Danysz W, Quack G. Memantine is a clinically well tolerated N-methyl-D-aspartate (NMDA) receptor antagonist a review of preclinical data. *Neuropharmacology*. 1999;38(6):735–767. https://doi.org/10.1016/S0028-3908(99)00019-2
- 13. Smith DJ, Bouchal RL, deSanctis CA, et al. Properties of the interaction between ketamine and opiate binding sites in vivo and in vitro. *Neuropharmacology*. 1987;26(9):1253–1260. https://doi.org/10.1016/0028-3908(87)90084-0
- 14. Hustveit O, Maurset A, Oye I. Interaction of the chiral forms of ketamine with opioid, phencyclidine, sigma and muscarinic receptors. *Pharmacol Toxicol*. 1995;77(6):355–359. https://doi.org/10.1111/j.1600-0773.1995.tb01041.x
- 15. Crisp T, Perrotti JM, Smith DL, et al. The local monoaminergic dependency of spinal ketamine. *Eur J Pharmacol*. 1991;194(2–3):167–172. https://doi.org/10.1016/0014-2999(91)90101-U

- 16. Mimura M, Namiki A, Kishi R, et al. Central cholinergic action produces antagonism to ketamine anesthesia. *Acta Anaesthesiol Scand*. 1992;36(5): 460–462. https://doi.org/10.1111/j.1399-6576.1992.tb03497.x
- 17. Durieux ME. Inhibition by ketamine of muscarinic acetylcholine receptor function. *Anesth Analg*. 1995;81(1):57–62. https://doi.org/10.1213/00000539-199507000-00012
- 18. Baum VC, Tecson ME. Ketamine inhibits transsarcolemmal calcium entry in guinea pig myocardium: direct evidence by single cell voltage clamp. *Anesth Analg.* 1991;73(6):804–807. https://doi.org/10.1213/00000539-199112000-00022
- 19. Yamakage M, Hirshman CA, Croxton TL. Inhibitory effects of thiopental, ketamine, and propofol on voltage-dependent Ca2+ channels in porcine tracheal smooth muscle cells. *Anesthesiology*. 1995;83(6):1274–1282. https://doi.org/10.1097/00000542-199512000-00018
- 20. Smith B, Pinnock C, Fischer B, et al. Unilateral analgesia following injection of fentanyl into the lumbosacral plexus. *Lancet*. 1987;1(8548):1497–1498. https://doi.org/10.1016/S0140-6736(87)92254-9
- 21. Eide PK, Stubhaug A. Relief of glossopharyngeal neuralgia by ketamine-induced N-methyl-aspartate receptor blockade. *Neurosurgery*. 1997;41(2):505–508. https://doi.org/10.1097/00006123-199708000-00043#
- 22. Gorlin AW, Rosenfeld DM, Ramakrishna H. Intravenous sub-anesthetic ketamine for perioperative analgesia. *J Anaesthesiol Clin Pharmacol*. 2016;32(2):160–167. https://doi.org/10.4103/0970-9185.182085
- 23. Dowdy EG, Kaya K, Gocho Y. Some pharmacologic similarities of ketamine, lidocaine, and procaine. *Anesth Analg.* 1973;52(5):839–842. https://doi.org/10.1213/00000539-197309000-00039
- 24. Schuttler J, Zsigmond EK, White PF. Ketamine and its isomers. In: White PF, editor. *Textbook of Intravenous Anesthesia*. Media, PA: Williams & Wilkins, 1997; 171–188.
- 25. Niesters M, Martini C, Dahan A. Ketamine for chronic pain: risks and benefits. *Br J Clin Pharmacol*. 2013;72:357–367. https://doi.org/10.1111/bcp.12094
- 26. Reves JG, Glass PS, Lubarsky DA, McEvoy MD. Intravenous nonopioid anesthetics. In: Miller RD, Fleisher LA, Johns RA, et al., editors. *Miller's anesthesia*. 6th ed. Philadelphia: Elsevier; 2005; 317–378.
- 27. Desta Z, Moaddel R, Ogburn ET, et al. Stereoselective and regiospecific hydroxylation of ketamine and norketamine. *Xenobiotica*. 2012;42(11):1076–1087. https://doi.org/10.3109/00498254.2012.685777
- 28. Grant IS, Nimmo WS, McNicol LR, Clements JA. Ketamine disposition in children and adults. *Br J Anaesth*. 1983;55:1107–1111. https://doi.org/10.1093/bja/55.11.1107
- 29. Sun LH, Fan YY, Wang X, Zheng HB. Pharmacodynamic elucidation of glutamate & dopamine in ketamine-induced anaesthesia. *Chem Biol Interact*. 2020;327:109164. https://doi.org/10.1016/j.cbi.2020.109164
- Kamp J, Van Velzen M, Olofsen E, et al. Pharmacokinetic and pharmacodynamic considerations for NMDA-receptor antagonist ketamine in the treatment of chronic neuropathic pain: an update of the most recent literature. *Expert Opin Drug Metab Toxicol*. 2019;15(12):1033–1041. https://doi.org/10.1080/17425255.2019.1689958
- 31. EMC: Ketamine SPC. https://www.medicines.org.uk/emc/product/6935/smpc#gref
- 32. Cull-Candy S, Brickley S, Farrant M. NMDA receptor subunits: diversity, development and disease. *Curr Opin Neurobiol.* 2001;11(3):327–335. https://doi.org/10.1016/S0959-4388(00)00215-4
- 33. Sepulveda FJ, Bustos FJ, Inostroza E, et al. Differential roles of NMDA receptor subtypes NR2A and NR2B in dendritic branch development and requirement of RasGRF1. *J Neurophysiol*. 2010;103(4):1758–1770. https://doi.org/10.1152/jn.00823.2009
- 34. Mendell LM. Physiological properties of unmyelinated fiber projection to the spinal cord. *Exp Neurol*. 1966;16(3):316–332. https://doi.org/10.1016/0014-4886(66)90068-9
- 35. Eide PK. Wind-up and the NMDA receptor complex from a clinical perspective. *Eur J Pain*. 2000;4(1):5–15. https://doi.org/10.1053/eujp.1999.0154
- 36. Herrero JF, Laird JM, López-García JA. Wind-up of spinal cord neurons and pain sensation: much ado about something? *Prog Neurobiol*. 2000;61(2):169–203. https://doi.org/10.1016/S0301-0082(99)00051-9
- 37. De Biasi S, Rustioni A. Glutamate and substance P coexist in primary afferent terminals in the superficial laminae of spinal cord. *Proc Natl Acad Sci USA*. 1988;85(20):7820–7824. https://doi.org/10.1073/pnas.85.20.7820
- 38. Skilling SR, Smullin DH, Beitz AJ, Larson AA. Extracellular amino acid concentrations in the dorsal spinal cord of freely moving rats following veratridine and nociceptive stimulation. J Neurochem. 1988;51(1):127–132. https://doi.org/10.1111/j.1471-4159.1988.tb04845.x
- 39. Merighi A, Polak JM, Theodosis DT. Ultrastructural visualization of glutamate and aspartate immunoreactivities in the rat dorsal horn, with special reference to the co-localization of glutamate, substance P and calcitonin-gene related peptide. *Neuroscience*. 1991;40(1):67–80. https://doi.org/10.1016/0306-4522(91)90175-N
- 40. Jeftinija S, Jeftinija K, Liu F, et al. Excitatory amino acids are released from rat primary afferent neurons in vitro. *Neurosci Lett*. 1991;125(2):191–194. https://doi.org/10.1016/0304-3940(91)90025-O
- 41. Duggan AW, Furmidge LJ. Probing the brain and spinal cord with neuropeptides in pathways related to pain and other functions. *Front Neuroendocrinol*. 1994;15(3):275–300. https://doi.org/10.1006/frne.1994.1011

- 42. Li P, Wilding TJ, Kim SJ, et al. Kainate-receptor-mediated sensory synaptic transmission in mammalian spinal cord. *Nature*. 1999;397(6715):161–164. https://doi.org/10.1038/16469
- 43. Drdla R, Sandkühler J. Long-term potentiation at C-fibre synapses by low-level presynaptic activity in vivo. *Mol Pain*. 2008;4:18. https://doi.org/10.1186/1744-8069-4-18
- 44. Latremoliere A, Woolf CJ. Central sensitization: a generator of pain hypersensitivity by central neural plasticity. *J Pain*. 2009;10(9):895–926. https://doi.org/10.1016/j.jpain.2009.06.012
- 45. Ji RR, Woolf CJ. Neuronal plasticity and signal transduction in nociceptive neurons: implications for the initiation and maintenance of pathological pain. *Neurobiol Dis*. 2001;8(1):1–10. https://doi.org/10.1006/nbdi.2000.0360
- 46. Petrenko AB, Yamakura T, Baba H, Shimoji K. The role of N-methyl-d-aspartate (NMDA) receptors in pain: a review. *Anesth Analg.* 2003;97:1108–1116. https://doi.org/10.1213/01.ANE.0000081061.12235.55
- 47. Ochoa J, Torebjörk HE, Culp WJ, Schady W. Abnormal spontaneous activity in single sensory nerve fibers in humans. *Muscle Nerve*. 1982;5(9S):S74–S77.
- 48. Costigan M, Scholz J, Woolf CJ. Neuropathic pain: a maladaptive response of the nervous system to damage. *Annu Rev Neurosci*. 2009;32:1–32. https://doi.org/10.1146/annurev.neuro.051508.135531
- 49. Park KM, Max MB, Robinovitz E, et al. Effects of intravenous ketamine, alfentanil, or placebo on pain, pinprick hyperalgesia, and allodynia produced by intradermal capsaicin in human subjects. *Pain*. 1995;63(2):163–172. https://doi.org/10.1016/0304-3959(95)00029-R
- 50. Andersen OK, Felsby S, Nicolaisen L, et al. The effect of ketamine on stimulation of primary and secondary hyperalgesic areas induced by capsaicin a double-blind, placebo-controlled, human experimental study. *Pain*. 1996;66(1):51–62. https://doi.org/10.1016/0304-3959(96)02995-8
- 51. Gottrup H, Hansen PO, Arendt-Nielsen L, Jensen TS. Differential effects of systemically administered ketamine and lidocaine on dynamic and static hyperalgesia induced by intradermal capsaicin in humans. *Br J Anaesth*. 2000;84(2):155–162. https://doi.org/10.1093/oxfordjournals.bja.a013396
- 52. Koppert W, Dern SK, Sittl R, et al. A new model of electrically evoked pain and hyperalgesia in human skin: the effects of intravenous alfentanil, S(+)-ketamine, and lidocaine. *Anesthesiology*. 2001;95(2):395–402. https://doi.org/10.1097/00000542-200108000-00022
- 53. Willert RP, Woolf CJ, Hobson AR, et al. The development and maintenance of human visceral pain hypersensitivity is dependent on the N-methyl-D-aspartate receptor. *Gastroenterology*. 2004;126(3):683–692. https://doi.org/10.1053/j.gastro.2003.11.047
- 54. Chizh BA, Headley PM. NMDA antagonists and neuropathic pain multiple drug targets and multiple uses. *Curr Pharm Des*. 2005;11(23):2977–2994. https://doi.org/10.2174/1381612054865082
- 55. Morris VH, Cruwys SC, Kidd BL. Characterisation of capsaicin-induced mechanical hyperalgesia as a marker for altered nociceptive processing in patients with rheumatoid arthritis. *Pain*. 1997;71(2): 179–186. https://doi.org/10.1016/S0304-3959(97)03361-7
- 56. Sörensen J, Graven-Nielsen T, Henriksson KG, et al. Hyperexcitability in fibromyalgia. J Rheumatol. 1998;25(1):152–155.
- 57. Attal N, Bouhassira D. Mechanisms of pain in peripheral neuropathy. *Acta Neurol Scand Suppl*. 1999;173:12–24; discussion 48–52. https://doi.org/10.1111/j.1600-0404.1999.tb07386.x
- 58. Petersen KL, Fields HL, Brennum J, et al. Capsaicin evoked pain and allodynia in post-herpetic neuralgia. *Pain*. 2000;88(2):125–133. https://doi.org/10.1016/S0304-3959(00)00311-0
- 59. Farrell M, Gibson S, McMeeken J, Helme R. Pain and hyperalgesia in osteoarthritis of the hands. J Rheumatol. 2000;27(2):441–447.
- 60. Burstein R. Deconstructing migraine headache into peripheral and central sensitization. *Pain*. 2001;89(2–3):107–110. https://doi.org/10.1016/S0304-3959(00)00478-4
- 61. Staud R, Smitherman ML. Peripheral and central sensitization in fibromyalgia: pathogenetic role. *Curr Pain Headache Rep.* 2002;6(4):259–266. https://doi.org/10.1007/s11916-002-0046-1
- 62. Price DD, Verne GN. Does the spinothalamic tract to ventroposterior lateral thalamus and somatosensory cortex have roles in both pain sensation and pain-related emotions? *J Pain*. 2002;3(2):105–108; discussion 113–114. https://doi.org/10.1054/jpai.2002.122950
- 63. Graven-Nielsen T, Aspegren Kendall S, Henriksson KG, et al. Ketamine reduces muscle pain, temporal summation, and referred pain in fibromyalgia patients. *Pain*. 2000;85(3):483–491. https://doi.org/10.1016/S0304-3959(99)00308-5
- 64. Schaible HG, Ebersberger A, Von Banchet GS. Mechanisms of pain in arthritis. *Ann NY Acad Sci*. 2002;966:343–354. https://doi.org/10.1111/j.1749-6632.2002.tb04234.x
- 65. Jensen TS, Baron R. Translation of symptoms and signs into mechanisms in neuropathic pain. *Pain*. 2003;102(1–2):1–8. https://doi.org/10.1016/s0304-3959(03)00006-x
- 66. Desmeules JA, Cedraschi C, Rapiti E, et al. Neurophysiologic evidence for a central sensitization in patients with fibromyalgia. *Arthritis Rheum*. 2003;48(5):1420–1429. https://doi.org/10.1002/art.10893

- 67. Kurdi MS, Theerth KA, Deva RS. Ketamine: current applications in anesthesia, pain, and critical care. *Anesth Essays Res.* 2014;8(3):283–290. https://doi.org/10.4103/0259-1162.143110
- 68. Salmi E, Långsjö JW, Aalto S, et al. Subanesthetic ketamine does not affect 11C- flumazenil binding in humans. *Anesth Analg.* 2005;101:722–725. https://doi.org/10.1213/01.ANE.0000156951.83242.8D
- 69. Flood P, Krasowski MD. Intravenous anesthetics differentially modulate ligand-gated ion channels. *Anesthesiology*. 2000;92:1418–1425. https://doi.org/10.1097/00000542-200005000-00033
- 70. Craven R. Ketamine. Anaesthesia. 2007;62(Suppl. 1):48–53. https://doi.org/10.1111/j.1365-2044.2007.05298.x
- 71. Pham TH, Gardier AM. Fast-acting antidepressant activity of ketamine: highlights on brain serotonin, glutamate, and GABA neurotransmission in preclinical studies. *Pharmacol Ther.* 2019;199:58–90. https://doi.org/10.1016/j.pharmthera.2019.02.017
- 72. White PF, Way WL, Trevor AJ. Ketamine its pharmacology and therapeutic uses. *Anesthesiology*. 1982;56(2):119–136. https://doi.org/10.1097/00000542-198202000-00007
- 73. Wang WF, Liu S, Xu B. A study of the protective effect and mechanism of ketamine on acute lung injury induced by mechanical ventilation. *Eur Rev Med Pharmacol Sci.* 2017;21(6):1362–1367.
- 74. Hoffman WE, Pelligrino D, Werner C, Kochs E, Albrecht RF, Schulte am Esch J. Ketamine decreases plasma catecholamines and improves outcome from incomplete cerebral ischemia in rats. *Anesthesiology*. 1992;76(5):755–762. https://doi.org/10.1097/00000542-199205000-00014
- 75. Morray JP, Lynn AM, Stamm SJ, Herndon PS, Kawabori I, Stevenson JG. Hemodynamic effects of ketamine in children with congenital heart disease. *Anesth Analg.* 1984;63:895–899.
- 76. Goyal R, Singh S, Bangi A, Singh SK. Case series: Dexmedetomidine and ketamine for anesthesia in patients with uncorrected congenital cyanotic heart disease presenting for non-cardiac surgery. *J Anaesthesiol Clin Pharmacol*. 2013;29:543–546. https://doi.org/10.4103/0970-9185.119142
- 77. Lam E, Rochani A, Kaushal G, et al. Pharmacokinetics of ketamine at dissociative doses in an adult patient with refractory status asthmaticus receiving extracorporeal membrane oxygenation therapy. *Clin Ther*. 2019;41(5):994–999. https://doi.org/10.1016/j.clinthera.2019.03.005
- 78. Goyal S, Agrawal A. Ketamine in status asthmaticus: a review. *Indian J Crit Care Med*. 2013;17(3):154–161. https://doi.org/10.4103/0972-5229.117048.
- 79. Sarma VJ. Use of ketamine in acute severe asthma. *Acta Anaesthesiologica Scandinavica*. 1992;36:106–107. https://doi.org/10.1111/j.1399-6576.1992.tb03432.x
- 80. Wilson LE, Hatch DJ, Rehder K. Mechanisms of the relaxant action of ketamine on isolated porcine trachealis muscle. *Br J Anaesth*. 1993;71(4):544–550. https://doi.org/10.1093/bja/71.4.544
- 81. Hirota K, Sato T, Rabito SF, et al. Relaxant effect of ketamine and its isomers on histamine-induced contraction of tracheal smooth muscle. *Br J Anaesth*. 1996;76:266–270. https://doi.org/10.1093/bja/76.2.266
- 82. Schmidt H, Ebeling D, Bauer H, et al. Ketamine attenuates endotoxin-induced leukocyte adherence in rat mesenteric venules. *Crit Care Med.* 1995;23:2008–2014. https://doi.org/10.1097/00003246-199512000-00009
- 83. Zhao Y, Sun L. Antidepressants modulate the in vitro inhibitory effects of propofol and ketamine on norepinephrine and serotonin transporter function. *J Clin Neurosci.* 2008;15(11):1264–1269. https://doi.org/10.1016/j.jocn.2007.11.007
- 84. Pabelick CM, Jones KA, Street K, et al. Calcium concentration-dependent mechanisms through which ketamine relaxes canine airway smooth muscle. *Anesthesiology*. 1997;86:1104–1111. https://doi.org/10.1097/00000542-199705000-00014
- 85. Jung I, Jung SH. Vasorelaxant mechanisms of ketamine in rabbit renal artery. *Korean J Anesthesiol*. 2012;63(6):533–539. https://doi.org/10.4097/kjae.2012.63.6.533
- 86. Gateau O, Bourgain JL, Gaudy JH, Benveniste J. Effects of ketamine on isolated human bronchial preparations. *Br J Anaesth*. 1989;63(6):692–695. https://doi.org/10.1093/bja/63.6.692
- Stone JM, Erlandsson K, Arstad E, et al. Relationship between ketamine-induced psychotic symptoms and NMDA receptor occupancy: a [(123)I]CNS-1261 SPET study. *Psychopharmacology*. 2008;197(3):401–408. https://doi.org/10.1007/s00213-007-1047-x
- 88. Treston G, Bell A, Cardwell R, et al. What is the nature of the emergence phenomenon when using intravenous or intramuscular ketamine for paediatric procedural sedation? *Emerg Med Australas*. 2009;21(4):315–322. https://doi.org/10.1111/j.1742-6723.2009.01203.x
- 89. Mort TC. Preoxygenation in critically ill patients requiring emergency tracheal intubation. *Crit Care Med*. 2005;33:2672–2675. https://doi.org/10.1097/01.CCM.0000187131.67594.9E
- 90. Green SM, Johnson NE. Ketamine sedation for pediatric procedures: part 2. Review and implications. *Ann Emerg Med.* 1990;19:1033–1046. https://doi.org/10.1016/S0196-0644(05)82569-7
- 91. Sener S, Eken C, Schultz CH, et al. Ketamine with and without midazolam for emergency department sedation in adults: a randomized controlled trial. *Ann Emerg Med*. 2011;57(2):109–114.e2. https://doi.org/10.1016/j.annemergmed.2010.09.010

- 92. Cartwright PD, Pingel SM. Midazolam and diazepam in ketamine anaesthesia. *Anaesthesia*. 1984;39:439–442. https://doi.org/10.1111/j.1365-2044.1984.tb07312.x
- 93. Lilburn JK, Dundee JW, Nair SG, et al. Ketamine sequelae. Evaluation of the ability of various premedicants to attenuate its psychic actions. *Anaesthesia*. 1978;33(4):307–311. https://doi.org/10.1111/j.1365-2044.1978.tb12412.x
- 94. Marland S, Ellerton J, Andolfatto G, et al. Ketamine: use in anesthesia. *CNS Neurosci Ther.* 2013;19(6):381–389. https://doi.org/10.1111/cns.12072
- 95. Cheong SH, Lee KM, Lim SH, et al. Brief report: the effect of suggestion on unpleasant dreams induced by ketamine administration. *Anesth Analg.* 2011;112(5):1082–1085. https://doi.org/10.1213/ANE.0b013e31820eeb0e
- 96. Gardner AE, Olson BE, Lichtiger M. Cerebrospinal-fluid pressure during dissociation anesthesia with ketamine. *Anesthesiology*. 1971;35:226–228. https://doi.org/10.1097/00000542-197108000-00029
- 97. Yadav M, Parle M, Jindal DK, Dhingra S. Protective effects of stigmasterol against ketamine-induced psychotic symptoms: possible behavioral, biochemical and histopathological changes in mice. *Pharmacol Rep.* 2018;70(3):591–599. https://doi.org/10.1016/j.pharep.2018.01.001
- 98. Fletcher PC, Honey GD. Schizophrenia, ketamine and cannabis: evidence of overlapping memory deficits. *Trends Cogn Sci.* 2006;10:167–174. https://doi.org/10.1016/j.tics.2006.02.008
- 99. Morgan CJ, Mofeez A, Brandner B, Bromley L, Curran HV. Ketamine impairs response inhibition and is positively reinforcing in healthy volunteers: a dose-response study. *Psychopharmacology*. 2004;172(3):298–308. https://doi.org/10.1007/s00213-003-1656-y
- 100. Strayer RJ, Nelson LS. Adverse events associated with ketamine for procedural sedation in adults. *Am J Emerg Med.* 2008;26(9):985–1028. https://doi.org/10.1016/j.ajem.2007.12.005
- 101. Kye YC, Rhee JE, Kim K, et al. Clinical effects of adjunctive atropine during ketamine sedation in pediatric emergency patients. *Am J Emerg Med*. 2012;30(9):1981–1985. https://doi.org/10.1016/j.ajem.2012.04.030
- 102. Gao M, Rejaei D, Liu H. Ketamine use in current clinical practice. *Acta Pharmacol Sin*. 2016;37(7):865–872. https://doi.org/10.1038/aps.2016.5
- 103. Acevedo-Diaz EE, Cavanaugh GW, Greenstein D, et al. Comprehensive assessment of side effects associated with a single dose of ketamine in treatment-resistant depression. *J Affect Disord*. 2020;263:568–575. https://doi.org/10.1016/j.jad.2019.11.028
- 104. Morris C, Perris A, Klein J, Mahoney P. Anaesthesia in haemodynamically compromised emergency patients: does ketamine represent the best choice of induction agent? *Anaesthesia*. 2009;64:532–539. https://doi.org/10.1111/j.1365-2044.2008.05835.x
- 105. Corssen G, Domino EF. Dissociative anesthesia: further pharmacologic studies and first clinical experience with the phencyclidine derivative CI-581. *Anesth Analg.* 1966;45(1):29–40. https://doi.org/10.1213/00000539-196601000-00007
- Reves JG, Glass PS, Lubarsky DA, McEvoy MD, Ruiz RM. Intravenous anaesthetics. In: Miller RD, ed. Miller's Anaesthesia.
 7th ed. Philadelphia, USA: Churchill Livingstone; 2010; 719–771. https://drive.google.com/file/d/0B81TNkQfgbpENFlr S2tDN3IZbWc/view
- 107. Jabre P, Combes X, Lapostolle F, et al. Etomidate versus ketamine for rapid sequence intubation in acutely ill patients: a multicentre randomised controlled trial. *Lancet*. 2009;374:293–300. https://doi.org/10.1016/S0140-6736(09)60949-1
- 108. Stoelting RK, Hillier SC. Nonbarbiturate intravenous anaesthetic drugs. In: Stoelting RK, Hillier SC, editors. *Pharmacology and Physiology in Anaesthetic Practice*. 4th ed. Philadelphia: Lippincott Williams and Wilkin; 2006; 155–178.
- 109. Tavakollian AR, Allahyary E. The comparison of the effect of three anesthetic induction regimens on the arterial oxygen saturation in children with tetralogy of fallot undergoing cardiac surgery. *Iran Red Crescent Med J.* 2011;13:702–706.
- 110. Sungur Ulke Z, Kartal U, Orhan Sungur M, et al. Comparison of sevoflurane and ketamine for anesthetic induction in children with congenital heart disease. *Paediatr Anaesth*. 2008;18:715–721. https://doi.org/10.1111/j.1460-9592.2008.02637.x
- 111. Gündüz M, Sakalli S, Güneş Y, et al. Comparison of effects of ketamine, ketamine-dexmedetomidine and ketamine-midazolam on dressing changes of burn patients. J Anaesthesiol Clin Pharmacol. 2011;27:220–224. https://doi.org/10.4103/0970-9185.81823
- 112. Kolawole IK. Ketamine hydrochloride: a useful but frequently misused drug. *Niger J Surg Res.* 2001;3:118–125 https://doi.org/10.4314/njsr.v3i3.12232
- 113. O'Hara D, Ganeshalingam K, Gerrish H, Richardson P. A 2 year experience of nurse led conscious sedation in paediatric burns. *Burns*. 2014;40:48–53. https://doi.org/10.1016/j.burns.2013.08.021
- 114. Owens VF, Palmieri TL, Comroe CM, et al. Ketamine: a safe and effective agent for painful procedures in the pediatric burn patient. *J Burn Care Res*. 2006;27:211–216. https://doi.org/10.1097/01.BCR.0000204310.67594.A1
- 115. Dundee JW, Wyant GM. Intravenous Anasethesia. New York: Churchill Livingstone; 1988; 135–159.
- 116. Samad MA, Islam MS, Ahmed M, Maruf AA. Evaluation of ketofol (ketamine-propofol combination) as total intravenous anaesthetic for burn dressing in adult patient. *J Armed Forces Med Coll Bangladesh*. 2012;8:20–24. https://doi.org/10.3329/jafmc.v8i1.13534

- 117. Tobin HA. Low-dose ketamine and diazepam. Use as an adjunct to local anesthesia in an office operating room. *Arch Otolaryngol*. 1982;108(7):439–440. https://doi.org/10.1001/archotol.1982.00790550043011
- 118. Wason R, Jain N, Gupta P, Gogia AR. Randomized double-blind comparison of prophylactic ketamine, clonidine and tramadol for the control of shivering under neuraxial anaesthesia. *Indian J Anaesth*. 2012;56:370–375. https://doi.org/10.4103/0019-5049.100821
- 119. Ozkan F, Kaya Z, Suren M. The effect of intravenous ketamine in prevention of hypotension during spinal anaesthesia in patients with benign prostatic hyperplasia. *Nobel Medicus*. 2011;7:82–88.
- 120. Radvansky BM, Shah K, Parikh A, et al. Role of ketamine in acute postoperative pain management: a narrative review. *Biomed Res Int*. 2015;2015:749837. https://doi.org/10.1155/2015/749837
- 121. Elia N, Tramer MR. Ketamine and postoperative pain a quantitative systematic review of randomised trials. *Pain*. 2005;113(1–2): 61–70. https://doi.org/10.1016/j.pain.2004.09.036
- 122. Bell RF, Dahl JB, Moore RA, Kalso E. Peri-operative ketamine for acute post-operative pain: a quantitative and qualitative systematic review (Cochrane review). *Acta Anaesthesiol Scand*. 2005;49(10):1405–1428. https://doi.org/10.1111/j.1399-6576.2005.00814.x
- 123. Laskowski K, Stirling A, McKay WP, Lim HJ. A systematic review of intravenous ketamine for postoperative analgesia. *Can J Anaesth*. 2011;58(10):911–923. https://doi.org/10.1007/s12630-011-9560-0
- 124. Haliloglu M, Ozdemir M, Uzture N, et al. Perioperative low-dose ketamine improves postoperative analgesia following Cesarean delivery with general anesthesia. *J Matern Fetal Neonatal Med*. 2016;29(6):962–966. https://doi.org/10.3109/14767058.2015.1027190
- 125. Berti M, Baciarello M, Troglio R, Fanelli G. Clinical uses of low-dose ketamine in patients undergoing surgery. *Curr Drug Targets*. 2009;10:707–715. https://doi.org/10.2174/138945009788982496
- 126. Costantini R, Affaitati G, Fabrizio A, Giamberardino MA. Controlling pain in the post-operative setting. *Int J Clin Pharmacol Ther.* 2011;49(2):116–127. https://doi.org/10.5414/CP201401
- 127. Javid MJ, Hajijafari M, Hajipour A, et al. Evaluation of a low dose ketamine in post tonsillectomy pain relief: a randomized trial comparing intravenous and subcutaneous ketamine in pediatrics. *Anesth Pain Med*. 2012;2:85–89. https://doi.org/10.5812/aapm.4399
- 128. Elshammaa N, Chidambaran V, Housny W, et al. Ketamine as an adjunct to fentanyl improves postoperative analgesia and hastens discharge in children following tonsillectomy a prospective, double-blinded, randomized study. *Paediatr Anaesth*. 2011;21(10):1009–1014. https://doi.org/10.1111/j.1460-9592.2011.03604.x
- 129. Basagan-Mogo E, Goren S, Korfali G, et al. Induction of anesthesia in coronary artery bypass graft surgery: the hemodynamic and analgesic effects of ketamine. *Clinics*. 2010;65(2):133. https://doi.org/10.1590/S1807-59322010000200003
- 130. García-Henares JF, Moral-Munoz JA, Salazar A, Del Pozo E. Effects of ketamine on postoperative pain after remifentanil-based anesthesia for major and minor surgery in adults: a systematic review and meta-analysis. *Front Pharmacol*. 2018;9:921. https://doi.org/10.3389/fphar.2018.00921
- 131. Bredmose PP, Grier G, Davies GE, Lockey DJ. Pre-hospital use of ketamine in paediatric trauma. *Acta Anaesthesiol Scand*. 2009;53(4):543–545. https://doi.org/10.1111/j.1399-6576.2008.01852.x
- 132. Kim KS, Kwak HJ, Min SK, et al. The effect of ketamine on tracheal intubating conditions without neuromuscular blockade during sevoflurane induction in children. *J Anesth*. 2011;25(2):195–199. https://doi.org/10.1007/s00540-011-1092-9
- 133. Aouad MT, Moussa AR, Dagher CM, et al. Addition of ketamine to propofol for initiation of procedural anesthesia in children reduces propofol consumption and preserves hemodynamic stability. *Acta Anaesthesiol Scand*. 2008;52:561–565. https://doi.org/10.1111/j.1399-6576.2008.01584.x
- 134. Tsai PS, Hsu YW, Lin CS, et al. Ketamine but not propofol provides additional effects on attenuating sevoflurane-induced emergence agitation in midazolam premedicated pediatric patients. *Paediatr Anaesth*. 2008;18:1114–1115. https://doi.org/10.1111/j.1460-9592.2008.02593.x
- 135. Abu-Shahwan I, Chowdary K. Ketamine is effective in decreasing the incidence of emergence agitation in children undergoing dental repair under sevoflurane general anesthesia. *Paediatr Anaesth*. 2007;17:846–850. https://doi.org/10.1111/j.1460-9592.2007.02298.x
- 136. Dalens BJ, Pinard AM, Létourneau DR, et al. Prevention of emergence agitation after sevoflurane anesthesia for pediatric cerebral magnetic resonance imaging by small doses of ketamine or nalbuphine administered just before discontinuing anesthesia. *Anesth Analg.* 2006;102(4):1056–1061. https://doi.org/10.1213/01.ane.0000200282.38041.1f
- 137. Berlinskiĭ VV, Zhdanov GG, Mushkin VV, et al. Combined anesthesia using Diprivan and ketamine in pediatric surgery. *Anesteziol Reanimatol*. 2000;3:10–12.
- 138. Fisher K, Coderre TJ, Hagen NA. Targeting the N-methyl-D-aspartate receptor for chronic pain management. Preclinical animal studies, recent clinical experience and future research directions. *J Pain Symptom Manage*. 2000;20(5):358–373. https://doi.org/10.1016/S0885-3924(00)00213-X

- 139. Kohrs R, Durieux ME. Ketamine: teaching an old drug new tricks. *Anesth Analg*. 1998;87(5):1186–1193. https://doi.org/10.1097/00000539-199811000-00039
- 140. Nishimura M, Sato K, Okada T, et al. Ketamine inhibits monoamine transporters expressed in human embryonic kidney 293 cells. *Anesthesiology*. 1998;88(3):768–774. https://doi.org/10.1097/00000542-199803000-00029
- 141. Kapur S, Seeman P. NMDA receptor antagonists ketamine and PCP have direct effects on the dopamine D(2) and serotonin 5-HT(2)receptors-implications for models of schizophrenia. *Mol Psychiatry*. 2002;7(8):837–844. https://doi.org/10.1038/sj.mp.4001093
- 142. Irnaten M, Wang J, Chang KS, et al. Ketamine inhibits sodium currents in identified cardiac parasympathetic neurons in nucleus ambiguus. *Anesthesiology*. 2002;96(3):659–666. https://doi.org/10.1097/00000542-200203000-00024