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Prevalence and clinical significance of antiphospholipid antibodies among hospitalized COVID-19 patients

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ABSTRACT

Objective: To describe the prevalence of antiphospholipid antibodies in coronavirus disease-19 (COVID-19) and to find potential associations between antiphospholipid antibody positivity and clinical outcomes.

Methods: From September to November 2020, clinical and laboratory data were collected from 50 COVID-19 patients hospitalized at Saiful Anwar General Hospital in Malang, Indonesia. Antiphospholipid antibodies were measured by finding IgM anti- β 2 glycoprotein, lupus anticoagulant, and IgM/IgG anticardiolipin. Clinical characteristics, thrombotic events, ICU admission, and mortality during hospitalization were recorded. Disease severity was defined by the Guidelines for the Prevention and Control of COVID-19, Indonesia.

Results: Among 50 patients, 5 patients (10.0%) were positive for antiphospholipid antibodies: 4 patients (80.0%) had IgM anti- β 2 glycoprotein and 1 patient had IgG anti-cardiolipin (20.0%) and IgM anti-cardiolipin (20.0%), none of lupus anticoagulant was detected. Antiphospholipid antibodies were associated with anosmia (*OR* 8.1; 95% *CI* 1.1-57.9; *P*=0.018), nausea and vomiting (*OR* 12.4; 95% *CI* 1.2-122.6; *P*=0.010), diarrhea (*OR* 9.8; 95% *CI* 1.3-70.9; *P*=0.010), cardiovascular disease (*OR* 1.4; 95% *CI* 1.0-1.9; *P*=0.001), chronic kidney disease (*OR* 12.0; 95% *CI* 2.0-423.7; *P*=0.001), moderate (*OR* 0.11; 95% *CI* 0.01-1.10; *P*=0.031) and severe (*OR* 18.5; 95% *CI* 1.8-188.4; *P*=0.002) disease severity, and in-hospital mortality (*OR* 8.1; 95% *CI* 1.1-57.9; *P*=0.018). However, there is no correlation between the presence of antiphospholipid antibody and ICU admission.

Conclusions: In summary, the prevalence of antiphospholipid antibodies in COVID-19 patients is low, mainly against IgM anticardiolipin, and is associated with an acute coronary syndrome, gastrointestinal manifestations, moderate and severe disease severity, and increased risk of mortality.

KEYWORDS: Antiphospholipid antibodies; COVID-19; COVID-19 disease severity; Mortality

1. Introduction

In January 2020, a pneumonia outbreak caused by a novel coronavirus known as severe acute respiratory coronavirus-2 (SARS-CoV-2) was first reported in Wuhan, China[1]. The World Health Organization (WHO) proclaimed the coronavirus disease 2019 (COVID-19) as a global health emergency since the confirmed infected cases spread throughout the globe[2]. In the meantime, the infection has rapidly spread, resulting in a pandemic that has affected more than 4.8 million individuals worldwide. In May 2021, the most significant numbers of new cases in South-East Asia were reported from Indonesia (26 908 new cases; 212.2 new cases per

Significance

Antiphospholipid antibody is associated with thrombotic manifestations. Evidence suggests that COVID-19 increases the risk of thromboembolic event, which leads to increased mortality in hospitalized COVID-19 patients. However, there are few studies on the prevalence and clinical outcomes of COVID-19 patients. This study shows the low prevalence of antiphospholipid antibodies in COVID-19, and its association with acute coronary syndrome, gastrointestinal manifestations, moderate and severe disease severity, and increased risk of mortality.

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100 000; an 8% increase); the enormous numbers of new deaths were also reported (1 125 deaths; 0.4 news per 100 000)[3]. Similarly, venous thromboembolism (VTE) has become a major concern in COVID-19. A recent meta-analysis study showed an increased risk of mortality associated with high thromboembolism events in COVID-19[4].

COVID-19 infection was associated with alterations in coagulation markers in the early reports[5,6]. A recent study shows that COVID-19 appears to have many coagulation abnormalities, including a significant increase in fibrin/fibrinogen breakdown products (i.e. D-dimers) and prolonged activated partial thromboplastin time (aPTT). Although the elevated D-dimer value is consistent with continued activation of the coagulation and fibrinolysis cascade, the combination of prolonged aPTT and arteriovenous thrombosis is unexpected. We have to remind clinicians of antiphospholipid syndrome (APS)-like clinical conditions[6,7]. Several studies have attempted to explain the pathogenesis of thrombosis, one of which is the appearance of antiphospholipid (aPL) antibodies[8]. According to several recent studies, there is limited evidence on aPL antibodies in COVID-19 disease, and it is not clear whether they are incidental phenomena or whether they are related to any hemostatic abnormalities reported in COVID-19[9,10].

There are many ways to drive virus-induced autoimmunity. The production of aPL antibodies in patients infected with SARS-CoV-2 can be calculated by two possible pathogenesis: neoepitope formation and molecular simulation[11,12]. These antibodies in COVID-19 mainly target β 2GP1 and anticardiolipin (aCL)[9,13]. However, several studies report that there are limited data on the occurrence of anti-phospholipid syndrome during SARS-CoV-2 infection. Positive antiphospholipid antibodies have been identified in a small number of patients. Their association with COVID-19

thrombotic events and clinical outcomes remains unclear[8,9,13]. Therefore, this study aims to discover the correlation between antiphospholipid antibodies and coagulation dysfunction, clinical manifestations, disease severity, and mortality in hospitalized patients with COVID-19, especially in Indonesia.

2. Subjects and methods

2.1. Study design and participants

This is a descriptive single-center cross-sectional study considering COVID-19 patients admitted to Saiful Anwar General Hospital in Malang, East Java, Indonesia, between September and November 2020. This study has been approved by the Malang Saiful Anwar General Hospital, Indonesia Ethics Committee (ethics number 400/194/K.3/302/2020). We randomly selected 50 confirmed COVID-19 cases from the general and intensive care unit (ICU). Our inclusion criteria were confirmed COVID-19 cases according to the WHO definition: real-time reverse transcription-polymerase chain reaction (RT-PCR) SARS-CoV-2 test results were positive, and respiratory tract specimens collected from nasopharyngeal swabs[14]. We excluded patients initially treated with heparin to reduce the interpretation bias of lupus anticoagulant (Figure 1).

2.2. Data collection

We collected patient demographic data, clinical characteristics, laboratory data, and clinical outcomes. Data were collected from the patients' medical records, including signs and symptoms, comorbidities, admission to ICU, and in-hospital mortality.



Figure 1. The study flowchart.

COVID-19 severity is classified as mild, moderate, and severe based on the Guidelines for the Prevention and Control of COVID-19 in Indonesia^[15]. Data were gathered comprehensively through a medical records review and communication with attending doctors and other health workers to fill in the missing data.

Thrombotic events, such as cerebrovascular accident (CVA), acute coronary syndrome (ACS), deep vein thrombosis (DVT), and pulmonary embolism, were defined as the presence of arterial or venous thromboembolism (PE). The appropriate examination to evaluate subjects who were clinically suspected with thrombotic events to establish the diagnosis, such as radiology examination, electrocardiography (ECG) and cardiac enzyme panel were performed.

Laboratory examinations, such as complete blood count, coagulation test, liver and renal function, were all measured for firsttime patients admitted to the hospital (Table 1). The antiphospholipid antibodies, including lupus anticoagulant, IgM/IgG anticardiolipin, IgM anti- β 2-glycoprotein, were examined by ELISA (Orgentec Diagnostika GmbH).

2.3. Statistical analysis

Categorical variables were measured using percentages and frequency rates. Continuous variables with normal distribution were presented as mean \pm SD and not-normal variables were reported

Table 1. Laboratory	findings of	f hospitalized	l patients with	COVID-19.
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as interquartile range (IQR). A *t*-test was employed to compare normally distributed data; otherwise, the Mann-Whitney test was utilized. The *Chi*-square test was used to compare proportions for categorical variables, and the Fisher exact test was utilized when data was insufficient. In studying the parameters associated with the occurrence of aPL antibodies, bivariate analysis was used. Variables with a *P*-value <0.05 according to a bivariate analysis were included in the multivariate analysis. All statistical analyses were performed using SPSS version 25.

3. Results

3.1. Baseline characteristics of COVID–19 patients with and without antiphospholipid antibodies

We recruited 50 patients diagnosed with COVID-19 according to the inclusion and exclusion criteria, 27 men (54.0%) and 23 women (46.0%), with an average age of (53.4 \pm 14.4) years. The most common early signs and symptoms are dry cough (64.0%), shortness of breath (62.0%) and fever (60.0%). We found that the most common comorbidity in patients was diabetes (50.0%), followed by cardiovascular disease (34.0%), hypertension (32.0%) and chronic kidney disease (16.0%). We classify patients into mild (12.0%), moderate (64.0%), and severe (24.0%) based on the severity

$ \begin{array}{ c c c c c c } \mbox{Periadrameters} & Peri$	<u> </u>						
Laboratorium parametersNorma valueAir partents (n=20)antibody (n=5)antibody (n=45) $b2$ values i Complete blood countsHemoglobin, g/dL*, mean ± SD13.4+17.712.25 ± 2.2412.24 ± 1.6012.25 ± 2.330.100.99WBC, x10 ³ /L, median (IQR)*4.3-10.39.29 (6.39-12.91)12.95 (7.21-18.52)9.08 (6.03-12.31)-1.120.27PLT, x10 ³ /L*142.424304.86 ± 135.36377.00 ± 128.75296.84 ± 136.58-1.250.21NLR, median (IQR)*<<.1.35.46 (3.55-9.60)3.80 (2.47-15.22)5.53 (3.53-9.37)-2.430.80Hemostatic functionsPPT, s, median (IQR)*9.4+11.31.22 (10.37-11.6)7.65 (11.25-18.90)1.00 (10.30-10.80)-2.020.04INR, median (IQR)*<.1.50.13 (0.99+1.12)0.75 (1.08-1.82)0.10 (0.99-1.09)-1.890.05APTT, s, median (IQR)*<.4.6-30.68.42 (25.47-33.9)11.50 (26.5-38.00)8.30 (25.45-33.75)-0.760.44Fibrinogen, mg/L, median (IQR)*0.53.12 (0.70-3.82)2.13 (0.85-2.97)3.91 (0.63-4.54)-0.140.88Liver function testALT, U/L, median (IQR)*0-4162.00 (21.00-83.00)36.71 (21.00-57.71)71.00 (21.00-92.00)-1.060.28AST, U/L, median (IQR)*0-4039.71 (24.00-63.71)41.36 (22.00-63.35)38.71 (25.00-63.71)-0.400.68Renal function test-1.20.58 (0.75-1.33)0.43 (1.21-1.64)0.56 (0.74+1.30) <t< th=""><th>Laboratorium parameters</th><th>Normal value</th><th>All patients $(n-50)$</th><th>Positive antiphospholipid</th><th>t/Z volues</th><th>D</th></t<>	Laboratorium parameters	Normal value	All patients $(n-50)$	Positive antiphospholipid	t/Z volues	D	
Complete blood counts I3.4.17.7 12.25 ± 2.24 12.24 ± 1.60 12.25 ± 2.33 0.10 0.99 WBC, ×10 ³ /L, median (IQR) [#] 4.3.10.3 9.29 (6.39-12.91) 12.95 (7.21-18.52) 9.08 (6.03-12.31) -1.12 0.07 PLT, ×10 ³ /L 142-424 304.86 ± 135.36 377.00 ± 128.75 296.84 ± 136.58 -1.25 0.21 NLR, median (IQR) [#] <3.13 5.46 (3.55-9.60) 3.80 (2.47-15.22) 5.53 (3.59-3.7) -2.43 0.80 Hemostatic functions 7.55 (11.25-18.90) 1.00 (10.30-10.80) -2.02 0.04 NR, median (IQR) [#] <1.5 0.13 (0.99-1.12) 0.75 (1.08-1.82) 0.10 (0.99-1.09) -1.89 0.05 APTT, s, median (IQR) [#] 8.42 (25.47-33.9) 11.50 (26.5-38.00) 8.30 (25.45-33.75) -0.76 0.46 D-Dimer, mg/L, median (IQR) [#] 3.12 (0.02-38.20) 2.13 (0.85-2.97) 3.91 (0.63-4.54) 0.86 Liver function test 3.41 (2.00 (2.100-83.00) 3.671 (2.100-57.71) 71.00 (21.00-92.00)	Laboratorium parameters		An patients (n=50)	antibody (n=5)	antibody (n=45)		1
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Complete blood counts						
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Hemoglobin, g/dL [*] , mean ± SD	13.4-17.7	12.25 ± 2.24	12.24 ± 1.60	12.25 ± 2.33	0.10	0.99
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	WBC, $\times 10^{3}$ /L, median (IQR) [#]	4.3-10.3	9.29 (6.39-12.91)	12.95 (7.21-18.52)	9.08 (6.03-12.31)	-1.12	0.27
	PLT, $\times 10^{3}/L^{*}$	142-424	304.86 ± 135.36	377.00 ± 128.75	296.84 ± 136.58	-1.25	0.21
	NLR, median (IQR) [#]	<3.13	5.46 (3.55-9.60)	3.80 (2.47-15.22)	5.53 (3.53-9.37)	-2.43	0.80
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Hemostatic functions						
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	PPT, s, median (IQR) [#]	9.4-11.3	1.22 (10.37-11.6)	7.65 (11.25-18.90)	1.00 (10.30-10.80)	-2.02	0.04
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	INR, median (IQR) [#]	<1.5	0.13 (0.99-1.12)	0.75 (1.08-1.82)	0.10 (0.99-1.09)	-1.89	0.05
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	APTT, s, median (IQR) [#]	24.6-30.6	8.42 (25.47-33.9)	11.50 (26.5-38.00)	8.30 (25.45-33.75)	-0.76	0.44
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Fibrinogen, mg/dL, median (IQR) [#]	154.3-397.9	113.25 (281.60-394.85)	202.00 (290.85-492.85)	110.90 (281.60-392.50)	-0.72	0.46
	D-Dimer, mg/L, median (IQR) [#]	0.5	3.12 (0.70-3.82)	2.13 (0.85-2.97)	3.91 (0.63-4.54)	-0.14	0.88
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	Liver function test						
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	ALT, U/L, median (IQR) [#]	0-41	62.00 (21.00-83.00)	36.71 (21.00-57.71)	71.00 (21.00-92.00)	-1.06	0.28
Renal function testUrea, mg/dL, median (IQR)#16.6-48.528.04 (18.87-46.91) $83.60 (17.90-101.50)$ 28.27 (18.65-46.91) -0.60 0.54 Creatinine, mg/dL, median (IQR)#<1.2	AST, U/L, median (IQR) [#]	0-40	39.71 (24.00-63.71)	41.36 (22.00-63.35)	38.71 (25.00-63.71)	-0.40	0.68
$ \begin{array}{c ccccc} Urea, mg/dL, median (IQR)^{\#} & 16.6-48.5 & 28.04 (18.87-46.91) & 83.60 (17.90-101.50) & 28.27 (18.65-46.91) & -0.60 & 0.54 \\ \hline Creatinine, mg/dL, median (IQR)^{\#} & <1.2 & 0.58 (0.75-1.33) & 0.43 (1.21-1.64) & 0.56 (0.74-1.30) & -1.57 & 0.11 \\ eGFR, mL/min/1.73 m^2, median & 60 & 33.25 (62.25-95.5) & 30.17 (45.50-75.67) & 32.50 (65.00-97.50) & -1.51 & 0.13 \\ (IQR)^{\#} & & & & & & & & & & \\ Inflammatory markers & & & & & & & & & & \\ LDH, U/L, median (IQR)^{\#} & 240-480 & 270.50 (555.00-825.50) & 249.30 (604.20-853.50) & 276.00 (552.00-828.00) & -0.37 & 0.70 \\ Ferritin, ng/mL, median (IQR)^{\#} & 30-400 & 2570.97 (524.72-1565.75) & 567.70 (775.30-1343.00) & 1086.10 (479.65-1565.75) & -0.24 & 0.80 \\ CRP, mg/dL, median (IQR)^{\#} & <0.3 & 7.18 (0.83-8.01) & 6.85 (1.14-7.99) & 8.01 (0.76-8.76) & -0.43 & 0.66 \\ \end{array}$	Renal function test						
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	Urea, mg/dL, median (IQR) [#]	16.6-48.5	28.04 (18.87-46.91)	83.60 (17.90-101.50)	28.27 (18.65-46.91)	-0.60	0.54
eGFR, mL/min/1.73 m², median 60 33.25 (62.25-95.5) 30.17 (45.50-75.67) 32.50 (65.00-97.50) -1.51 0.13 (IQR) [#]	Creatinine, mg/dL, median (IQR)#	<1.2	0.58 (0.75-1.33)	0.43 (1.21-1.64)	0.56 (0.74-1.30)	-1.57	0.11
(IQR) [#] Inflammatory markers LDH, U/L, median (IQR) [#] 240-480 270.50 (555.00-825.50) 249.30 (604.20-853.50) 276.00 (552.00-828.00) -0.37 0.70 Ferritin, ng/mL, median (IQR) [#] 30-400 2570.97 (524.72-1565.75) 567.70 (775.30-1343.00) 1086.10 (479.65-1565.75) -0.24 0.80 CRP, mg/dL, median (IQR) [#] <0.3	eGFR, mL/min/1.73 m ² , median	60	33.25 (62.25-95.5)	30.17 (45.50-75.67)	32.50 (65.00-97.50)	-1.51	0.13
Inflammatory markers LDH, U/L, median (IQR) [#] 240-480 270.50 (555.00-825.50) 249.30 (604.20-853.50) 276.00 (552.00-828.00) -0.37 0.70 Ferritin, ng/mL, median (IQR) [#] 30-400 2570.97 (524.72-1565.75) 567.70 (775.30-1343.00) 1086.10 (479.65-1565.75) -0.24 0.80 CRP, mg/dL, median (IQR) [#] <0.3	$(IQR)^{\#}$						
LDH, U/L, median (IQR) [#] 240-480 270.50 (555.00-825.50) 249.30 (604.20-853.50) 276.00 (552.00-828.00) -0.37 0.70 Ferritin, ng/mL, median (IQR) [#] 30-400 2570.97 (524.72-1565.75) 567.70 (775.30-1343.00) 1086.10 (479.65-1565.75) -0.24 0.80 CRP, mg/dL, median (IQR) [#] <0.3	Inflammatory markers						
Ferritin, ng/mL, median (IQR) [#] 30-400 2570.97 (524.72-1565.75) 567.70 (775.30-1343.00) 1086.10 (479.65-1565.75) -0.24 0.80 CRP, mg/dL, median (IQR) [#] <0.3	LDH, U/L, median (IQR) [#]	240-480	270.50 (555.00-825.50)	249.30 (604.20-853.50)	276.00 (552.00-828.00)	-0.37	0.70
CRP, mg/dL, median (IQR) [#] <0.3 7.18 (0.83-8.01) 6.85 (1.14-7.99) 8.01 (0.76-8.76) -0.43 0.66	Ferritin, ng/mL, median (IQR) [#]	30-400	2570.97 (524.72-1565.75)	567.70 (775.30-1343.00)	1086.10 (479.65-1565.75)	-0.24	0.80
	CRP, mg/dL, median (IQR) [#]	< 0.3	7.18 (0.83-8.01)	6.85 (1.14-7.99)	8.01 (0.76-8.76)	-0.43	0.66

Note: data were presented in n (%) or mean \pm SD; *t*-test for mean \pm SD, labeled with^{*}; Mann-Whitney test for nonparametric test, labeled with^{*}; eGFR measure based on CKD-EPI equation; WBC: white blood cell; PLT: platelet; NLR: neutrophil/lymphocyte ratio; PPT: plasma prothrombin time; aPTT: activated partial thromboplastin time; INR: international normalizing ratio; ALT: alanine aminotransferase; AST: aspartate aminotransferase; eGFR: estimated glomerular filtration rate; CKD-EPI: chronic kidney disease epidemiology collaboration; LDH: lactate dehydrogenase; CRP: C-reactive protein.

of COVID-19. Several manifestations of thrombosis have been reported: acute coronary syndrome (6.0%), cerebrovascular accident (6.0%), and deep vein thrombosis (4.0%) (Table 2). The laboratory profile is shown in Table 1. In addition, 9 patients (18.0%) were admitted to the ICU ward due to mechanical ventilators, 40 patients (80.0%) were discharged, and 10 patients (10.0%) died (Table 2).

3.2. Prevalence rates of aPL antibodies in COVID-19 patients

In our finding, from 50 patients with COVID-19, five patients (10.0%) had at least one circulating antiphospholipid antibody. The most frequently detected aPL antibodies were IgM anticardiolipin (80.0%) following by IgG anticardiolipin (20.0%), and IgM anti- β 2-glycoprotein (20.0%). Three patients are positive for IgM anticardiolipin, one patient is positive for IgG anticardiolipin, and one patient is positive for both IgM anticardiolipin and IgM anti- β 2-glycoprotein. No lupus anticoagulant was detected in this study.

3.3. Correlation of the clinical parameters in COVID-19 patients with antiphospholipid antibodies

The incidence of nausea and vomiting (80.0% vs. 24.4%, P=0.010), diarrhea (60.0% vs. 13.3%, P=0.010) and anosmia (60.0% vs. 15.6%, P=0.018) in COVID-19 patients with positive aPL antibodies statistically significantly higher than negative aPL antibody group, followed by fever, dry cough, shortness of breath, headache and chest pain, but there was no statistical difference (20.0%-80.0%, P<0.05). Interestingly, we found that all COVID-19 patients with aPL antibodies have cardiovascular disease (100.0% vs. 26.7%, P=0.001). In addition, There is a statistically significant higher prevalence of chronic kidney disease in aPL antibodies group as compared to negative aPL group (P=0.005) (Table 2). Multivariate analysis showed that the presence of aPL antibodies was significantly associated with a higher risk of nausea and vomiting (OR 12.4; 95% CI 1.2-122.6), diarrhea (OR 9.8; 95% CI 1.3-70.9), and anosmia (OR8.1; 95% CI 1.1-57.9). Similarly, the risk of cardiovascular disease

Table 2. Comparison	between baseline character	istics and clinical outcome	s of COVID-19 patien	ts with and without and	iphos	pholi	pids antib	oody
								~

a	All COVID-19 patients	All COVID-19 patients Positive antiphospholipid Negative antiphospholipid		. 2 .	_	
Characteristics	(n=50)	antibody (<i>n</i> =5)	antibody (<i>n</i> =45)	t/χ^2 values	P	
Age (years) (mean \pm SD) [*]	53.4 ± 14.4	58.0 ± 11.9	52.9 ± 14.7	26.21	0.462	
Sex, n (%)				0.43	0.508	
Female	23 (46.0)	3 (60.0)	20 (44.4)			
Male	27 (54.0)	2 (40.0)	25 (55.6)			
Sign and symptoms, n (%)						
Fever	30 (60.0)	4 (80.0)	26 (57.8)	0.92	0.336	
Dry cough	32 (64.0)	4 (80.0)	28 (62.2)	0.61	0.432	
Shortness of breath	31 (62.0)	2 (40.0)	29 (64.4)	1.14	0.285	
Diarrhea	9 (18.0)	3 (60.0)	6 (13.3)	6.64	0.010	
Nausea vomiting	15 (30.0)	4 (80.0)	11 (24.4)	6.61	0.010	
Headache	10 (20.0)	2 (40.0)	8 (17.8)	1.38	0.239	
Anosmia	11 (22.0)	3 (60.0)	7 (15.6)	5.55	0.018	
Hemiparesis	3 (6.0)	0 (0.0)	3 (6.7)	0.35	0.552	
Chest pain	4 (8.0)	1 (20.0)	3 (6.7)	1.07	0.297	
Comorbidities, n (%)						
Diabetes mellitus#	25 (50.0)	4 (80.0)	21 (46.7)	2.00	0.157	
Hypertension	16 (32.0)	2 (40.0)	14 (31.1)	0.16	0.686	
Malignancy	2 (4.0)	0 (0.0)	2 (4.4)	0.23	0.630	
Cardiovascular disease	17 (34.0)	5 (100.0)	12 (26.7)	10.78	0.001	
Chronic kidney disease	8 (16.0)	3 (60.0)	5 (11.1)	8.00	0.005	
HIV infection	2 (4.0)	1 (20.0)	1 (2.2)	3.70	0.054	
COVID-19 severity, n (%)						
Mild	6 (12.0)	0 (0.0)	6 (13.3)	0.75	0.384	
Moderate	32 (64.0)	1 (20.0)	31 (68.9)	4.66	0.031	
Severe	12 (24.0)	4 (80.0)	8 (17.8)	9.55	0.002	
Thrombosis events, n (%)						
ACS	3 (6.0)	2 (40.0)	1 (2.2)	11.38	0.001	
CVA	3 (6.0)	0 (0.0)	3 (6.7)	0.35	0.552	
DVT	2 (4.0)	1 (20.0)	1 (2.2)	3.70	0.054	
Admission, n (%)				1.82	0.177	
Non-ICU	41 (82.0)	3 (60.0)	38 (84.4)			
ICU	9 (18.0)	2 (40.0)	7 (15.6)			
In-hospital mortality, n (%)				5.55	0.18	
Survivor	40 (80.0)	2 (40.0)	38 (84.4)			
Non-survivor	10 (20.0)	3 (60.0)	7 (15.6)			

Note: data were presented in n (%) unless stated otherwise, *Chi*-square test (χ^2 values) for percentages; *t*-test for mean \pm SD, labeled with^{*}; HIV: human immunodeficiency virus; ICU: intensive care unit; COVID-19: coronavirus disease 2019; ACS: acute coronary syndrome; CVA: cerebrovascular accident; DVT: deep vein thrombosis.

is higher (*OR* 1.4; 95% *CI* 1.0-1.9). Chronic kidney disease (*OR* 12.0; 95% *CI* 1.6-90.1) might be affected due to the presence of aPL antibodies (Table 3).

Table 3. Clinical parameters affected by antiphospholipid antibody positivity.

Clinical manifestations	OR (95% CI)	Р
Diarrhea	9.8 (1.3-70.9)	0.010
Nausea and vomiting	12.4 (1.2-122.6)	0.010
Anosmia	8.1 (1.1- 57.9)	0.018
Cardiovascular disease	1.4 (1.0-1.9)	0.001
Chronic kidney disease	12.0 (1.6-90.1)	0.005
Moderate degree of COVID-19	0.11 (0.01-1.10)	0.031
Severe Degree of COVID-19	18.5 (1.8-188.4)	0.002
Acute coronary syndrome	29.3 (2.0-423.7)	0.001
Mortality in COVID-19	8.1 (1.1-57.9)	0.018

Note: data were presented in n (%) or mean \pm SD; OR: odds ratio; CI: confidence interval; COVID-19: coronavirus disease 2019.

3.4. Clinical outcomes of COVID-19 patients with antiphospholipid antibodies

Circulating aPL antibodies may aggravate the severity of COVID-19 disease. Our research shows that COVID-19 patients with aPL antibodies have a higher incidence of severe patients (80.0% vs.17.8%, P=0.002), and a lower incidence of moderate patients (20.0% vs.68.9%, P=0.031). COVID-19 without aPL antibody is classified as mild. We recorded thrombotic events that occurred during hospitalization. Our results show that the incidence of acute corona syndrome in COVID-19 patients with aPL antibodies is significantly higher than that of patients without aPL antibodies (Table 2). Approximately 40.0% of COVID-19 patients with aPL antibodies are admitted to the ICU ward, but this is not significantly different from COVID-19 patients with aPL antibodies (P>0.05). We also found that COVID-19 patients with aPL antibodies have a higher mortality rate (60.0% vs. 15.6; P=0.018).

We performed multivariate analysis to measure the association of aPL antibodies with disease severity, thrombotic events, and mortality. We found that there were associations of aPL antibodies positivity in COVID-19 patients with a higher risk of severe degree (*OR* 18.5; 95% *CI* 1.8-188.4), lower risk of moderate degree (*OR* 0.11; 95% *CI* 0.01-1.10), higher risk of acute coronary syndrome events (*OR* 29.3; 95% *CI* 2.0-423.7), and higher risk of mortality (*OR* 8.1; 95% *CI* 1.1-57.9).

4. Discussion

This descriptive single-center cross-sectional study was conducted in Malang, Indonesia. This study evaluated the prevalence of aPL antibodies in COVID-19 patients and its association with this patient subgroup's clinical characteristics and clinical outcomes. Recent other single-center studies from China, Italy, and Mexico indicate that the prevalence of aPL antibodies in COVID-19 patients is low[9,13,16]. Similar to our findings, we studied the low incidence of aPL antibodies in COVID-19 patients, and the most commonly detected aPL antibodies are anticardiolipin and anti-β2-glycoprotein. We also found that the presence of aPL antibody is significantly related to the severity of COVID-19. The pathophysiology of the hypercoagulable state of COVID-19 is still unclear, but it is proven that most severe and critically ill patients have coagulopathy[17]. However, in this study, no lupus anticoagulant was detected in all of our patients. In contrast with another recent study, it shows a more tremendous amount of lupus anticoagulant in COVID-19 patients but not in critically ill patients[18]. They are potentially biased due to heparin administration for hypercoagulable state prophylaxis[19].

Infection-induced aPL antibody production has been widely acknowledged[20]. The association between aPL antibody detection and SARS-CoV-2 viral infection remains unclear. The S1 may induce the generation of aPL antibodies, and the S2 subunits of S protein in the SARS-CoV-2 virus might produce a phospholipidlike epitope that stimulates the production of aPL antibodies[11]. A previous meta-analysis study reported a very high prevalence of aPL antibodies in human immunodeficiency virus (HIV), Epstein-Barr virus (EBV), and hepatitis C virus (HCV), as well as gastrointestinal manifestation with COVID-19 infection[21,22]. Our findings show that gastrointestinal manifestations (diarrhea, nausea, and vomiting) have a significant association with the presence of aPL antibodies in serum.

Interestingly, the latest study found that IgA anti-β2-glycoprotein serum was the most common aPL antibody isotype in COVID-19. However, aPL antibodies may be temporary and disappear within a few weeks. As COVID-19 primarily affects the lung and intestines, preferential IgA isotype production could be linked to the breakdown of mucosal immune tolerance[13].

The circulating aPL antibody in COVID-19 has been considered to be one of the mechanisms leading to pro-inflammatory, hypercoagulable state and thrombotic events[23]. In this study, we found that aPL antibodies are present in many moderate and severe patients, which are associated with the risk of cardiovascular disease and chronic kidney disease as comorbidities. Our study also reported an increased risk of thrombotic events. Approximately 40.0% of COVID-19 patients with aPL antibodies have the acute coronary syndrome, which is associated with a higher mortality rate. Although the aPL antibody in COVID-19 is related to disease severity and mortality, our findings and previous studies[8,13,19] found no significant association between aPL antibody and ICU admission. These findings suggest that aPL antibodies may be a marker of disease severity, thrombotic events, and mortality. Unfortunately, we cannot carry out long-term follow-up. There is still a lack of published evidence regarding the correlation between the presence of aPL antibodies and the clinical outcome of COVID-19. A prospective study of aPL antibodies in COVID-19 patients is urgently needed to investigate.

In conclusion, this study is a descriptive single-centered crosssectional study that is conducted in Malang, Indonesia. The purpose is to evaluate clinical characteristics, disease severity, and clinical outcome of COVID-19 patients with aPL antibody. The COVID-19 pandemic is an emerging global health problem that has high mortality rates. A vast population in Indonesia and the high number of new cases and death rates due to COVID-19 are emerging problems for the local government. We reported that a slight prevalence of aPL antibodies in COVID-19 had been associated with specific gastrointestinal manifestations. In addition, it has an increased risk of acute coronary syndrome and an increased mortality rate. Further studies using a larger scale and multi-center are needed to see the prevalence of clinical survival of COVID-19 patients that have aPL antibodies for its application to alertness, management and therapy of COVID-19.

Conflict of interest statement

The authors affirm no conflict of interests in this study.

Authors' contributions

C.S.W., H.S., and K.H. designed, directed and supervised the project. T.W.I.D. and M.Z.P. processed the experimental data, performed the analysis, and wrote the manuscript in consultation with C.S.W., H.S., and K.H. P.A.R. helped supervise the project and contributed to the draft interpretation of the results. I.A.W., K.D.H., and E.S.D. collected the data and performed the laboratorium analysis.

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