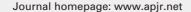


Review Article

Asian Pacific Journal of Reproduction





doi: 10.4103/2305-0500.288583

Effects of nitric oxide on reproductive organs and related physiological processes Ayoob Rostamzadeh¹, Reza Ahmadi², Mahdi Heydari¹, Amir Raoofi^{3⊠}

¹Student Research Committee, Department of Anatomical Sciences, Faculty of Medicine, Iran University of Medical Sciences, Tehran, Iran ²Clinical Biochemistry Research Center, Basic Health Sciences Institute, Shahrekord University of Medical Sciences, Shahrekord, Iran ³Leishmaniasis Research Center, Department of Anatomy, Sabzevar University of Medical Sciences, Sabzevar, Iran

ABSTRACT

Nitric oxide (NO), a member of the reactive nitrogen species family, plays a role in several physiologic processes, including vasculogenesis and angiogenesis, growth and puberty, and senescence and apoptosis. NO plays an important role in the production of ovarian steroids, ovulation, and follicular apoptosis. In other words, increased activity of nitric oxide synthase (NOS) leads to an increased amount of NO, which triggers production of prostaglandins and inflammatory cascades which facilitate follicular rupture and atresia. NO concentration elevation inhibits steroid synthesis in luteal and granulosa cells. Since NO is a major paracrine mediator of various biological processes, as well as a key factor in both the reproductive cycle and embryo implantation, oversynthesis of NO in the uterus results in toxicity and inflammation in epithelial cells and immunorejection of implantation. In the male physiological system, NO synthesized by NOS plays a major role in erectile function and androgen secretion, as well as semen parameters, and oocyte junction to the sperm. Furthermore, this supposedly simple molecule is involved in a number of other functions, such as germ cell evolution, connections between sertoli cells and germ cells in the blood-testis barrier, homodynamic contraction, and germ cell apoptosis. Moreover, NO is considered a key factor in male fertility due to its widespread distribution in both normal and diseased testis tissue. The difference of expression level of NOS in normal and pathological states is a probable cause of fertility destructive processes.

KEYWORDS: Reproduction; Nitric oxide; Nitric oxide synthase; Cell signaling; Oxidants

1. Introduction

Nitric oxide (NO) is a short half-time gas (about a few seconds) which has diverse biochemical and physiological potentials. This molecule was initially discovered in 1978 and nominated as the molecule of the year in the 1992[1,2]. This molecule is an inner cell and intra-cell messenger which plays a key role in the maintenance

of body hemostasis[2]. NO usually accomplishes its purpose through synthesizing cyclic guanosine monophosphate. In the production of NO from the L-arginine, its synthesis is mediated by nitric oxide synthase (NOS). This enzyme exists in three isoforms: nervous, endothelial, and induction[1]. NO cascade activates different pathways in the various tissues, e.g., they emerge as a vasodilator factor and a known endothelium-derived relaxing factor in the cardiovascular system[2]. However, in the nervous system, NO is considered as a neurotransmitter. Also, in some cases, it is involved in neutrophil-induced cell toxicity, platelet aggregation, blood flow, synaptic transmission, and long-term memory loss[3,4]. In addition to aforementioned functions, NO involves in ovulation, menstruation, and sperm capacity and motility[5,6]. NO is an important paracrine messenger, which participates in several physiological and pathophysiological events in the elementary and endocrine organs[7]. Furthermore, NO has some roles in immune system, such as antiviral and antimicrobial effects, excitation and suppression of the immune system, and cytoprotection[8].

2. NO sources in the body

2.1. NO sources

In mammals, NO can be generated by three isoforms of NO synthase including nervous NOS (nNOS), endothelial NOS (eNOS), and inducible NOS (iNOS). Mitochondrial NOS has been

For reprints contact: reprints@medknow.com

 $^{^{\}boxtimes}\ensuremath{\mathsf{To}}$ whom correspondance may be addressed. E-mail: amirrezaraoofi@yahoo.com

This is an open access article distributed under the terms of the Creative Commons Attribution-Non Commercial-Share Alike 4.0 License, which allows others to remix, tweak and buid upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

^{©2020} Asian Pacific Journal of Reproduction Produced by Wolters Kluwer- Medknow. All rights reserved.

How to cite this article: Rostamzadeh A, Ahmadi R, Heydari M, Raoofi A. Effects of nitric oxide on reproductive organs and related physiological processes. *Asian Pac J Reprod* 2020; 9(4): 159-165.

Article history: Received: 23 March 2020; Revision: 10 May 2020; Accepted: 25 May 2020; Available online: 4 July 2020

discovered recently, which is specifically found in mitochondrion[9]. Furthermore, iNOS and eNOS are localized in different reproductive tissues such as granulosa, theca cells and cytoplasm of oocytes. Most apoptotic cascades result in producing reactive nitrogen species and reactive oxygen species[9].

2.2. Mechanisms of nitric oxide synthesis

All isoforms of NOS utilize L-arginine as the substrate, and molecular oxygen and reduced nicotinamide-adeninedinucleotide phosphate (NADPH) as co-substrates. A functional NOS transfers electrons from NADPH to the haem in the aminoterminal oxygenase domain[4,10]. At the haem site, the electrons are used to reduce and activate oxygen (O₂) and to oxidize L-arginine to L-citrulline and NO[11]. To synthesize nitric oxide, the NOS enzyme must pass two key steps, including hydroxylating L-arginine to Nv-hydroxy-L-arginine and also oxidizing Nv-hydroxy-L-arginine to L-citrulline and NO[12-14]. Binding to calcium via calmodulin in nNOS and eNOS is done by using an increase in intracellular calcium ion (Ca²⁺). When calmodulin affinity to NOS increases, it triggers the transport of electrons from NADPH in the reductase domain to the haem in the oxygenase domain. Due to the presence of different amino acids in the iNOS structure, calmodulin is able to bind at extremely low intracellular Ca²⁺ concentrations approximately 40 nM[15,16].

The ovary is an organ that undergoes major structural and functional changes during the reproductive cycle^[17]. Luteolysis is the structural and functional degradation of the corpus luteum which occurs at this cycle and indicates a decrease in cellular function. Analysis of corpus luteum in healthy ovaries is accompanied by increased production of reactive oxygen species such as O_2 and H_2O_2 . One of the results of the free radical production in ovarian tissue has been the lipid peroxidation in the plasma membrane of the corpus luteum, which may lead to loss of gonadotropin receptors, reduce adenylyl cyclase–cyclic AMP cyclic adenosine monophosphate (cAMP) formation, and finally reduce steroidization of the corpus luteum following its destruction[18].

Previous studies have shown that NO plays an important role in maintaining the physiological balance of organs including the ovary[19]. Motta *et al*[20] reported a direct link between a destruction of the corpus luteum and an increase in ovarian prostaglandin F2. They stated that corpus luteum depletion was directly related to decreased ovarian glutathione production. Also, their study showed that increased NO through the mechanism of ovarian glutathione depletion could increase the oxidizing effect of oxidase substances and lead to destruction of the corpus luteum[21]. Another study from the same group showed the effect of NO on the ovary and stated that corpus luteum depletion may increase as a result of increased lipid oxidation by using *L*-NG-nitro-arginine methyl ester hydrochloride (*L*-NAME) to prevent the production of intracellular NO in the ovary[21,22].

3. NO in the female genital system

3.1. Oocyte

Full comprehension of the meiosis cell division program has not yet been accomplished. However, more evidence indicates NO involvement in meiosis administration. NO possesses an essential physiologic role in oocyte maturation and fertilization, embryo development, and fetal implantation[23-25]. eNOS and iNOS are reported to be synthesized in the mammalian oocyte, and their presence has been confirmed during follicular maturity. NO synthesis inhibition in the in vivo maturation of oocytes causes a reduction in the number of blastocysts and increases apoptosis in the fetus. Alternatively, high levels of NO cause disruption in meiosis development and fetus development in the cow, along with a delay in the restart (or resumption) of meiosis[25]. Goud et al demonstrated NO is a crucial factor in the maintenance of oocyte quality[26]. An investigation of earlier literature reveals the binary role of NO in oocyte maturity. Moreover, Bu et al described how the concentration dependence of NO has effects on oocyte maturity in the rat. For instance, NO produced from eNOS activity triggers oocyte maturity in the cumulus cells. Nevertheless, a higher concentration of NO causes meiosis arrest of oocyte. Before ovulation, NO reduction following the sudden increase in luteinizing hormone production (luteinizing hormone surge) can be act as a key agent in the restart (or resumption) of meiosis[27]. Recent evidence has validated that oocytes are able to block meiosis in the diplotene phase by expressing iNOS, which produces a sufficient amount of NO[28]. Abbasi et al reported that a cAMP cascade is the inducer platform of NO which starts meiosis in the rat oocyte, though cyclic guanosine monophosphate cascades support the inhibitory effort of NO[29] (Figure 1).

3.2. Follicular growth and maturation

Pituitary gonadotropin is a key regulator of the final steps of follicular evolution, and current data emphasize the balance between autocrine and/or paracrine factors in normal follicular growth. NO presence in the follicular liquid has been proven in several animal species. NOS expression in the follicles exhibits an inner ovarian scheme to synthesize NO and to manage follicular growth. NO is synthesized by several ovarian cell types, as well as ovarian arteries. The ovarian macrophages remove apoptotic cells during specific phases of tissue remodeling, as opposed to the removal of foreign debris macrophages, which is another source of NO in ovarian tissue[6]. These macrophage-rich regions of the ovary, such as the theca layer, corpus luteum, and atretic follicles, are involved in the phagocytosis of atretic granulosa cells and apoptotic luteal cells[30,31].

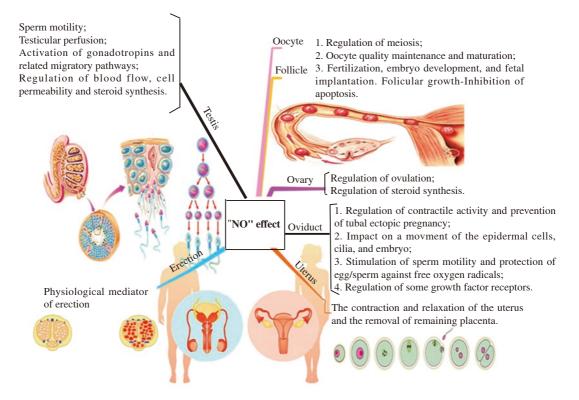


Figure 1. Role of nitric oxide (NO) in different parts of male (left side) and female (right side) genital system.

3.3. Ovary

The iNOS and eNOS have several roles in the ovulation process[6]. NO synthesis increases with follicle growth, and this NO increase is related to increased estrogen. There are similar changes in the amount of NO in circulation with the growth of follicles in women undergoing in vitro fertilization. NO is continuously treated with the gonadotropin releasing hormone and human chorionic gonadotropic hormone, and other hormones, such as luteinizing hormone, follicle stimulating hormone, and progesterone have been observed[5]. The use of NOS inhibitors intraperitoneally inhibits ovulation in rats, which is evidence of the role of NO in ovulation processes[30]. These findings suggest that NO is involved in the regulation of ovarian functions[31]. Stimulation of the ovary with gonadotropins increases the expression of both iNOS and eNOS genes and this suggests that both NO isoforms play a role in the ovulation process. Inhibition of iNOS by using specific inhibitors of NG-Methyl-L-arginine and aminoguanidine inhibitors leads to dose-dependent inhibition of ovulation in rats, which indicates the role of iNOS in the ovulation process[30]. During follicular growth, eNOS is expressed in theca cells and in the granulosa cells of the follicle wall, and after ovulation, eNOS is expressed in the yellow body. In the immature ovary and during follicular development, iNOS expression occurs in the theca cells and stroma cells, and after ovulation, iNOS is expressed in the outer layers of the yellow body. Estimating the quantity of iNOS suggests that unlike eNOS,

iNOS concentration does not change during follicular growth[6]. Since theca cells, luteal granulosic cells, and yellow body cells are involved in steroidogenesis, it can be concluded that NO also plays a role in regulating steroid synthesis. In most organs of the body, iNOS is expressed only in response to an immune stimulation, such as infection or trauma, and the physiological association of iNOS expression in the natural ovaries at all stages is still uncertain. It is possible that the reduction of expression or non-expression is mainly due to the presence of macrophages and interleukin-1 β in the ovary, and it is also possible that NO derived from iNOS acts as a monitoring/growth support molecule. Interleukin-1 β is a stimulant for NO synthesis in humans and cows' fallopian tubes[6]. On the other hand, glucose increases in the middle of the menstrual cycle, and glucose stimulates NO synthesis. Therefore, it is possible that NO and glucose can interact as follicles or inducers and facilitate follicular growth pathways[6]. Sugino et al reported a relationship between the concentration of NO in follicular fluid and apoptosis, where small follicles show more apoptosis, in contrast to large and moderate-sized follicles. However, the concentration of NO (nitrite/ nitrate), arginine, and citrulline is not different in these follicles[32]. Furthermore, the concentration of NO is increased in human follicular fluid, and this increase is directly related to follicle volume and estradiol concentration. Taken together, these observations suggest that the local production of NO causes follicular growth and inhibits apoptosis[33].

3.4. Oviduct (fallopian tube)

In the uterine tube, increased contraction due to endothelin in the presence of L-NAME, an NO synthesis inhibitor, was the first evidence of the role of NO in the regulation of the functions of the uterine tube[5]. NO regulates contractile activity in the human fallopian tube by using prostaglandins, prostacyclin and cAMP, so it even prevents tubal ectopic pregnancy[34]. Various studies confirm the presence of calcium-dependent, as well as NOS-dependent, calcium-shaped forms in the rat, cow, and human fallopian tube, and immunohistochemical studies confirm the presence of eNOS in the epithelial cells of the uterine tube[35,36]. Although the activity of NOS in the uterus during the proestrus phase is relatively less than in the other stages of the estrous cycle, the distribution of calciumdependent NOS in the isthmus, fimbriae, and ampulla of uterine tube is the same[36]. It appears that the release of NO could stimulate sperm motility and protect the egg and sperm from damage caused by free oxygen radicals[6]. Also, NO may affect the movements of the epidermal cells of the uterine tube. NO has been shown to regulate some growth factor receptors, such as the epidermal growth factor, binding proteins and integrin[37]. In contrast to the physiological state, NO synthesis in the uterine tube can be increased under certain pathological conditions, such as infection or endometriosis that leads to reduced fertility through a destructive or toxic effect on sperm cells as well as oocytes. In addition, the increased production of NO may affect the movements of cilia and thus embryo transfer, and the final result can be miscarriage[6].

3.5. Uterus

NO regulates the contraction of smooth muscle cells and uterus dilation during pregnancy; therefore, the role of NO in the regulation of pathophysiology and uterine biology has been considered to be significant[38]. The presence of NOS in the glandular epithelium, endometrial stromal cells, smooth muscle cells, and mast cells indicates the role of NO in the control of the functions of the uterus. In addition, local NO synthesis in the uterus may be important for regulating myometrial activity, such as contraction and relaxation of the uterus[39]. Although smooth muscle cells express eNOS[40], the myometrium is one of the rare parts of uterine tissue which iNOS expresses in non-provocative implications[41], including 1) facilitating the uterine traction for removal of the remaining placenta of the uterus after parturition with nitroglycerin, 2) preventing preterm delivery and prolonging pregnancy with nitroglycerin, and 3) reducing the power of uterine contractions induced by oxytocin with amyl nitrate that this issue indicates the role of NO in regulating and controlling uterine contractions during pregnancy[6]. A study conducted by Bansal et al showed that the expression of iNOS rather than eNOS and nNOS in humans in preterm labor was the highest. It is possible that the increase in NOS activity during pregnancy due to positive regulation of cytokines and reduction afterwards during childbirth is highly related to inhibitory cytokines^[42]. Interaction between cyclooxygenase, NO, and cytokine has been proven in the uterus of the mouse, and these factors may regulate the function of the uterus during pregnancy[43]. Ovarian hormones also induce iNOS expression in the uterus and may regulate the function of the uterus. However, the role of eNOS in epithelial cells and endometrial stroma is still unclear. But, it is possible that continuous production of NO through the synthesis of prostaglandin and through binding proteins facilitates processes such as menstruation and implantation. A NO derivative of eNOS which is the result of activating the production of guaniline cyclase solution or the decomposition of cyclooxygenase functions as an inhibitor of endometrial platelet aggregation[6]. Buhimschi et al showed that the cervix expresses all three NOS isoforms. In addition, the expression of iNOS increases during normal and preterm labor in the cervix and decreases in the uterus, and nNOS, which is not expressed in the uterus during pregnancy, increases during childbirth in the cervix[41]. During childbirth, regardless of the presence of iNOS and nNOS, there is no significant change in the expression of the eNOS gene. These findings indicate that NOS activity in the uterus and cervix has a different function and concentration during childbirth and may play a role in reconstructing the connective tissue during cavity preparation. The physiological and biological relevance of NO during pregnancy and childbirth shows that the NO synthesis inhibitor (meaning L-NAME) prolongs the delivery time and also reduces the opening of external orifice of the uterus (external os) and vagina[41,44].

3.6. Placenta and preeclampsia

Both *eNOS* and *iNOS* genes are expressed in the placenta, and eNOS expression in the fetal and placental vessels occurs in patients whose preeclampsia increases[45]. Placental vessels are more important in the pathophysiology of preeclampsia, and it seems eNOS is important in conditions such as preeclampsia. NO biosynthesis increases with advancing gestation during normal pregnancy and decreases in preeclampsia, and eNOS is expressed in human placental syncytiotrophoblasts and extravillous trophoblasts[43,45]. In fact, it may increase *eNOS* gene expression in the fetal and placental vessels in patients with preeclampsia who have an adaptive response to low perfusion and hypoxia. The study conducted by Buhimschi *et al* indicated that administration of *L*-NAME to pregnant mice leads to a condition similar to preeclampsia[41].

4. NO in the male reproductive system

4.1. Testis

NO has also been identified in the testicular vasculature endothelium, and its mechanism of action has been partially specified. Accordingly, it is seen that NO can be effective in testicular perfusion, activation of gonadotropins and its migratory pathway to the Leydig cells of the testes, and affect androgen displacement[39]. NO in the testes is involved in regulating blood flow, cell permeability, and contractile function of myofibroblasts, as well as in regulating steroid synthesis. Also, NO regulates sperm motility, as in a low NO concentration increases sperm motility and a medium (or high) concentration reduces sperm motility. In human semen (seminal fluid), a positive correlation between the concentration of NO and the percentage of sperm without mobility was observed[46]. Under physiological conditions, NO is produced in a small amount, and it causes the free radicals to scavenge, which is one of the main causes of decreasing sperm motility. In contrast, excessive NO production under pathological conditions, such as infection, varicocele, or diabetes mellitus, can cause sperm toxicity and also reduce sperm motility through the formation of peroxynitrite. It is likely that the sperm ejaculated into the female reproductive tract can lead to an immune response, which induces iNOS activity and produces a large amount of NO, which can cause the non-maturation and capacity of sperm[46]. Hence, the presence of endogenous NO inhibitor in seminal plasma can play a physiological role in inhibiting NOS activity and maintaining NO in low concentrations to prevent toxic injuries to sperm and Leydig cells or to suppress hypermotility of the sperm associated with the capacitybuilding process[47] (Figure 1).

4.2. Erection

In humans, NOS activity in the pelvic mesh, cavernous sinus nerves in penile tissue, dorsal branches of the penis, and deep arteries of the sinus cavernous is observed[6]. NOS activity in the rat penile neurons (which autonomically innervated corpus cavernosum and entered the glans cavernous tissue) and in the neural network in the adventitia layer of the penile vessels, shows that NO is a physiological mediator of erectile function. Apart from expression in the nerves, eNOS is abundantly expressed in the penile endothelium and the corpus cavernosum of the endothelium sinusoidum[40]. Different findings indicate that all three iNOS, nNOS, and eNOS genes are expressed in smooth muscle cells of the cavernous sinus of the penis. Administration of anti-androgenic drug to healthy rats reduces the expression of nNOS and eNOS genes and reduces erection[48]. Ignarro *et al* also showed that the electrical stimulation of the isolated cell line from the Corpus cavernosum of the rabbit secretes NO into the intestine. Based on these findings, they suggested that erection is caused by NO and occurs in response to a nonadrenergic-noncholinergic neurotransmitter. In addition, direct injection of *L*-NAME into periventricular nucleuses leads to inhibition of apomorphine and oxytocin (as an erection inducer)[6].

5. Conclusion and future perspectives

NO is a lipophilic molecule with a short half-life and is synthesized by many organs in the body. It is also introduced as an intra-cell messenger directing many physiologic cascades. Among other things, NO associates with cell growth, apoptosis, and reproduction signal transduction, as well as regulation of blood flow in sexual organs, regulation of vascular tonicity, genital tracts formation, and defense mechanisms. NO reacts with oxygen-active species, thiol groups, and proteins. Considering its concentration and site of action, it is able to protect or poison cells. NO is a nitrogen-active species which plays a role in most physiologic processes, such as vasculogenesis and angiogenesis, growth and puberty, senescence and apoptosis. In the male physiologic system, NO, which is synthesized by NOS, plays a significant role in physiological cascades, such as erectile function and androgen secretion, as well as sperm motility, maturity, quality, and capacity, and oocyte binding to the sperm. Furthermore, this supposedly simple molecule is involved in other functions, such as germ cell evolution, connections between Sertoli cells and germ cells in the blood-testis barrier, homodynamic contraction, and germ cell apoptosis. Moreover, NO is considered as a key factor in male fertility due to its widespread distribution in both normal and diseased testis tissue. The expression levels of eNOS and iNOS are different in the normal or pathological state, and overexpression of these two isoforms is a probable cause of fertility destructive processes, including low sperm motility and viability, disruption of testis tissue, induction of apoptosis in the germ cells, and, literally, perturbation of spermatogenesis. NO is an important factor in fabricating ovarian steroids, ovulation, and follicular apoptosis. iNOS is the major isoform in the ovulation process. In other words, increased activity of iNOS leads to an increased amount of NO, which triggers production of prostaglandins and runs inflammatory cascades which can cause follicular rupture and atresia. NO concentration elevation avoids steroid synthesis in the luteal and granulosa cells. Since NO is a major paracrine mediator of various biological processes and plays a key role in both the reproductive cycle and embryo implantation, oversynthesis of NO in the uterus results in toxicity and inflammation in epithelial cells and immunorejection of implantation.

Currently, the number of infertile couples has increased

dramatically as different forms include decreased fertilization rates, increased levels of abortion and high level of morbidity. Increased NO level is one of the main chemical and physiopathological factors in this regard.

Due to different roles of NO and a variety of functions in the molecular signaling of male and female reproductive system, scientific community needs new technology and synthetic materials to detect, recognize, and control its level. Paradoxical role of NO in both pathologic and physiologic processes depends on general state of body and oxidant-antioxidant balance system, thus using the antioxidants acts to improve levels of NO.

Conflict of interest statement

There is no conflict of interest to declare.

Acknowledgments

The authors would like to express their gratitude to the School of Medicine, Sabzevar University of Medical Sciences, Sabzevar, Iran for their support. Also, The authors appreciate of the Research Council of Shahrekord University of Medical Sciences.

Authors' contributions

Ayoob Rostamzadeh designed this study and Amir Raoofi drafted the manuscript. Reza Ahmadi and Mahdi Heydari helped to draft the manuscript. All authors read and approved the final manuscript.

References

- [1] Tuteja N, Chandra M, Tuteja R, Misra MK. Nitric oxide as a unique bioactive signaling messenger in physiology and pathophysiology. *Biomed Res Int* 2004; 2004(4): 227-237.
- [2] Zhao Y, Vanhoutte PM, Leung SW. Vascular nitric oxide: Beyond eNOS. J Pharmacol Sci 2015; 129(2): 83-94.
- [3] Ying L, Hofseth LJ. An emerging role for endothelial nitric oxide synthase in chronic inflammation and cancer. *Cancer Res* 2007; 67(4): 1407-1410.
- [4] Forstermann U, Sessa WC. Nitric oxide synthases: Regulation and function. *Eur Heart J* 2012; **33**(7): 829-837.
- [5] Wang J, He Q, Yan X, Cai Y, Chen J. Effect of exogenous nitric oxide on sperm motility *in vitro*. *Biol Res* 2014; **47**(1): 44.
- [6] Li J, Zhang W, Zhu S, Shi F. Nitric oxide synthase is involved in follicular development via the PI3K/AKT/FoxO3a pathway in neonatal and immature rats. Animals 2020; 10(2): 248.
- [7] Lanas A. Role of nitric oxide in the gastrointestinal tract. Arthritis Res Ther 2008; 10(Suppl 2): S4.

- [8] Cuzzocrea S, Mazzon E, Calabro G, Dugo L, De Sarro A, Van De Loo FA, et al. Inducible nitric oxide synthase — knockout mice exhibit resistance to pleurisy and lung injury caused by carrageenan. *Am J Respir Crit Care Med* 2000; **162**(5): 1859-1866.
- [9] Tejero J, Shiva S, Gladwin MT. Sources of vascular nitric oxide and reactive oxygen species and their regulation. *Physiol Rev* 2019; **99**(1): 311-379.
- [10]Stuehr DJ. Enzymes of the *L*-arginine to nitric oxide pathway. *J Nutrit* 2004; **134**(10): 2748S-2751S.
- [11]Feng MS, Guo P, Jiang LX, Shi JB, Yao QZ. Synthesis of novel methotrexate derivatives with inhibition activity of nitric oxide synthase. *Chin Chem Lett* 2009; 20(2): 178-180.
- [12]Mukherjee P, Cinelli MA, Kang S, Silverman RB. Development of nitric oxide synthase inhibitors for neurodegeneration and neuropathic pain. *Chem Soc Rev* 2014; 43(19): 6814-6838.
- [13]Jansen Labby K, Li H, Roman LJ, Martásek P, Poulos TL, Silverman RB. Methylated Nω-hydroxy-*L*-arginine analogues as mechanistic probes for the second step of the nitric oxide synthase-catalyzed reaction. *Biochemistry* 2013; **52**(18): 3062-3073.
- [14]Rath M, Müller I, Kropf P, Closs EI, Munder M. Metabolism *via* arginase or nitric oxide synthase: Two competing arginine pathways in macrophages. *Frontiers Immunol* 2014; 27(5): 532.
- [15]Pavanelli WR, Gutierrez FRS, da Silva JJN, Costa IC, Watanabe MAE. The effects of nitric oxide on the immune response during giardiasis. *Braz J Infect Dis* 2010; **14**(6): 606-612.
- [16]Spratt DE, Newman E, Mosher J, Ghosh DK, Salerno JC, Guillemette JG. Binding and activation of nitric oxide synthase isozymes by calmodulin EF hand pairs. *FEBS J* 2006; **273**(8): 1759-1771.
- [17]Rojas J, Chávez-Castillo M, Olivar LC, Calvo M, Mejías J, Rojas M, et al. Physiologic course of female reproductive function: A molecular look into the prologue of life. *J Pregnancy* 2015; doi: 10.1155/2015/715735.
- [18]Lu J, Wang Z, Cao J, Chen Y, Dong Y. A novel and compact review on the role of oxidative stress in female reproduction. *Reprod Biol Endocrinol* 2018; 16(1): 80.
- [19]Devine PJ, Perreault SD, Luderer U. Roles of reactive oxygen species and antioxidants in ovarian toxicity. *Biol Reprod* 2012; 86(2): 27.
- [20]Motta AB, Estevez A. Dual effects of NO in functional and regressing rat corpus luteum. *Mol Hum Rep* 2001; 7(1): 43-47.
- [21]Agarwal A, Aponte-Mellado A, Premkumar BJ, Shaman A, Gupta S. The effects of oxidative stress on female reproduction: A review. *Reprod Biol Endocrinol* 2012; **10**(1): 49.
- [22]Premkumar BJ, Aponte A, Shaman A, Agarwal A. Reactive oxygen species and female infertility. Syst Biol Fre Rad Antiox 2014; 3: 2743-2772.
- [23]Chen K, Pittman RN, Popel AS. Nitric oxide in the vasculature: Where does it come from and where does it go? A quantitative perspective. *Antioxid Redox Signal* 2008; **10**(7): 1185-1198.
- [24]Shiva S, Huang Z, Grubina R, Sun J, Ringwood LA, MacArthur PH, et al. Deoxymyoglobin is a nitrite reductase that generates nitric oxide and regulates mitochondrial respiration. *Circ Res* 2007; **100**(5): 654-661.
- [25]Basini G, Grasselli F. Nitric oxide in follicle development and oocyte competence. *Reproduction* 2015; **150**(1): R1-R9.

- [26]Goud PT, Goud AP, Najafi T, Gonik B, Diamond MP, Saed GM, et al. Direct real-time measurement of intraoocyte nitric oxide concentration *in vivo*. *PloS One* 2014; 9(6): e98720.
- [27]Bu S, Xia G, Tao Y, Lei L, Zhou B. Dual effects of nitric oxide on meiotic maturation of mouse cumulus cellenclosed oocytes *in vitro*. *Mol Cell Endocrinol* 2003; **207**(1): 21-30.
- [28]Tripathi A, Kumar K, Chaube SK. Meiotic cell cycle arrest in mammalian oocytes. J Cell Physiol 2010; 223(3): 592-600.
- [29] Abbasi M, Akbari M, Amidi F, Kashani IR, Mahmoudi R, Sobhani A, et al. Nitric oxide acts through different signaling pathways in maturation of cumulus cell-enclosed mouse oocytes. *Daru J Pharm Sci* 2015; **17**(1): 48-52.
- [30]Sela-Abramovich S, Galiani D, Nevo N, Dekel N. Inhibition of rat oocyte maturation and ovulation by nitric oxide: Mechanism of action. *Biol Reprod* 2008; 78(6): 1111-1118.
- [31]Basini G, Grasselli F. Nitric oxide in follicle development and oocyte competence. *Reproduction* 2015; 150(1): R1-R9.
- [32]Jee BC, Kim SH, Moon SY. The role of nitric oxide on apoptosis in human luteinized granulosa cells. *Gynecol Obstet Invest* 2003; 56(3): 143-147.
- [33]Da Broi MG, Giorgi VS, Wang F, Keefe DL, Albertini D, Navarro PA. Influence of follicular fluid and cumulus cells on oocyte quality: Clinical implications. J Assist Reprod Genet 2018; 35(5): 735-751.
- [34]Shaw JL, Dey SK, Critchley HO, Horne AW. Current knowledge of the aetiology of human tubal ectopic pregnancy. *Hum Reprod Update* 2010; 16(4): 432-444.
- [35]Hu J, Ma S, Zou S, Li X, Cui P, Weijdegård B, et al. The regulation of nitric oxide synthase isoform expression in mouse and human fallopian tubes: Potential insights for ectopic pregnancy. *Int J Mol Sci* 2015; **16**(1): 49-67.
- [36]Zhan X, Li D, Johns RA. Expression of endothelial nitric oxide synthase in ciliated epithelia of rats. J Histochem Cytochem 2003; 51(1): 81-87.
- [37]Zamberlam G, Sahmi F, Price CA. Nitric oxide synthase activity is critical for the preovulatory epidermal growth factor-like cascade induced by luteinizing hormone in bovine granulosa cells. *Free Radical Biol Med* 2014; **74**: 237-244.

- [38]Norman JE, Bollapragada S, Yuan M, Nelson SM. Inflammatory pathways in the mechanism of parturition. *BMC Pregnancy Childb* 2007; 7(Suppl 1): S7.
- [39]Kong L, Wei Q, Fedail JS, Shi F, Nagaoka K, Watanabe G. Effects of thyroid hormones on the antioxidative status in the uterus of young adult rats. *J Reprod Dev* 2015; 61(3): 219-227.
- [40]Favini R, Aldieri E, Revelli A, Bosia A, Massobrio M, Ghigo D. Nitric oxide synthesis in human nonpregnant myometrium and uterine myomas. *Fertil Steril* 2003; **79**(Suppl 1): 749-753.
- [41]Buhimschi I, Ali M, Jain V, Chwalisz K, Garfield RE. Differential regulation of nitric oxide in the rat uterus and cervix during pregnancy and labour. *Hum Reprod* 1996; **11**(8): 1755-1766.
- [42]Bansal RK, Goldsmith PC, He Y, Zaloudek CJ, Ecker JL, Riemer RK. A decline in myometrial nitric oxide synthase expression is associated with labor and delivery. *J Clin Invest* 1997; **99**(10): 2502-2508.
- [43]Cella M, Farina MG, Dominguez Rubio AP, Di Girolamo G, Ribeiro ML, Franchi AM. Dual effect of nitric oxide on uterine prostaglandin synthesis in a murine model of preterm labour. *Brit J Pharmacol* 2010; 161(4): 844-855.
- [44]Dymara-Konopka W, Laskowska M. The role of nitric oxide, ADMA, and homocysteine in the etiopathogenesis of preeclampsia. *Int J Mol Sci* 2019; **20**(11): 2757.
- [45]Aouache R, Biquard L, Vaiman D, Miralles F. Oxidative stress in preeclampsia and placental diseases. *Int J Mol Sci* 2018; **19**(5): 1496.
- [46]Francavilla F, Santucci R, Macerola B, Ruvolo G, Romano R. Nitric oxide synthase inhibition in human sperm affects sperm-oocyte fusion but not zona pellucida binding. *Biol Reprod* 2000; 63(2): 425-429.
- [47]Doshi SB, Khullar K, Sharma RK, Agarwal A. Role of reactive nitrogen species in male infertility. *Reprod Biol Endocrinol* 2012; 10(1): 109.
- [48]Traish AM, Goldstein I, Kim NN. Testosterone and erectile function: From basic research to a new clinical paradigm for managing men with androgen insufficiency and erectile dysfunction. *Eur Urol* 2007; 52(1): 54-70.