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Research Article

Preparation and characterization of a new adsorbent from raphia taedigera seed

Emmanuel Folorunso Olasehinde^{1,a}, Segun Michael Abegunde^{*2,b}

¹Department of Chemistry, Federal University of Technology, Akure, Nigeria

²Department of Science Technology, Federal Polytechnic, Ado-Ekiti, Ekiti State, Nigeria

Article Info

Abstract

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carbonization;

activation;

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There has been widespread research and engagement in recent years on the use of plant material in preparing porous carbon for wastewater treatment. In this study, new biochars (BCs) impregnated with 0.1 M sulphuric acid (RTA) and 0.1 M sodium hydroxide (RTB) were prepared. The BCs and raw seed [RTR] were characterized using scanning electron microscopy (SEM), Fouriertransform infrared (FTIR) spectrophotometer, thermogravimetric/Differential thermal analyser. [TGA/DTA) to determine their morphology, spectral bands and thermal behaviour respectively. RTR showed a scattered aggregated surface microstructure, while both RTA and RTB gave aggregated rough surfaces with high number of pores. The FTIR analysis showed significant changes in term of band shift, bands disappearances, and new band formations due to pyrolysis and activation of the BCs. The results of the TGA/DTA revealed that RTA and RTB have better thermal stability than RTR, with the maximum degradation temperature of RTR, RTA, and RTB occurred at 380°C, 425°C and 410°C respectively. Adsorption efficiencies ranged from 81.671 to 99.743% for performance evaluation on the BCs using different adsorbent dosages and the initial concentrations of methylene blue solutions. These preliminary investigations suggest that biochars prepared from Raphia taedigera seed can be used for methylene blue removal from wastewater.

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1. Introduction

In recent years, heavy metals and textile dyes have been considered the most ubiquitous contaminants of water worldwide. Water pollution is a major concern in our environment, and so providing a solution to this issue will be of obvious benefit to humans [1]. Over the years, adsorption has been recognized as one of the most reliable and efficient methods of removing organic and inorganic pollutants from industrial wastewater. Activated carbon (AC) has been undoubtedly recognized as the most common and widely employed adsorbent for wastewater treatment, given its simplicity of use, low cost, renewable nature, availability, and non-toxicity; its porous structure over large surface areas makes it a material of choice as an effective adsorbent [2 - 4]. With the introduction of stricter environmental regulations, the demand for porous carbons has progressively increased [5, 6]. Recently, there have been reports on several

*Corresponding author: abegundes@gmail.com

^a orcid.org/0000-0002-3909-1283; ^b orcid.org/0000-0002-6688-5230

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treatment processes by which to remove contaminants from wastewater, including degradation by photocatalytic processes [7–12]; the Fenton process [13]; photo-Fenton processes [14]; sono-chemical degradation [15]; chemical coagulation/flocculation, ozonation, cloud point extraction, oxidation, nano-filtration, chemical precipitation, ion exchange, reverse osmosis, and ultra-filtration [16-20]; and modified processes such as those combining ultrasound with adsorption-desorption processes [21-24]. However, among these, adsorption techniques have gained favour, owing to their simplicity, costeffectiveness, and efficiency in removing pollutants that are too stable for conventional methods [8, 25-29]. Recently, the idea of using locally sourced materials to prepare adsorbent materials for water and effluent treatment has gained popularity among environmentalists and material engineers. Given the exorbitant cost of commercial AC, there is a need to develop a low-cost and high-performance adsorbent from readily available and locally sourced alternative materials. Several agricultural waste products have been utilized [30], including cotton stalks [31], cereal by-product [6], bamboo [32], acorn shells [33], groundnut shells [34, 35], bagasse [36], cow bone powder [37], palm oil stones [38], coconut shells [39], tobacco residues [40], olive stones [41], apricot stones [42], nutshells [43], walnut shells [41], pine cones [44], and cherry stones [45].

Little information is currently available on *Raphia taedigera*, despite having been in existence for centuries. The plant remains quantitatively and qualitatively untapped in terms of their potential applications. *R taedigera* is native to Brazil, Cameroon, Costa Rica, Nicaragua, Nigeria, and Panama [46–50], and grows in swampy forests along streams. It produces egg-sized fruits of about 5–7 cm long and 3 cm in diameter, which are covered by imbricate glossy reddish–brown scales; each fruit contains one seed. A well-dried seed is hard and has a brown outer part and white shiny inner part.

In literature, no study has clearly evaluated the possibility of using *R taedigera* seed as an adsorbent. With recent developments in the need to source locally for available materials to prepare adsorbent, since metal ion and dye pollutants pose a serious threat to biological and environmental sources, given their harmful effects on human health and the environment [51, 52 hence; a need to check the potential of *R. taedigera* seed as a useful adsorbent This present study is aimed at preparation and characterization of adsorbent from *Raphia taedigera* seed considered as waste, for the recovery of methylene blue dye from aqueous solution.

2. Materials and Methods

2.1. Sample collection and treatment

R. taedigera seeds were obtained locally from a farmland in Ise-Ekiti, washed and sundried for several days until moisture evaporation was completed. The dried samples were crushed using 911MPE-24 laboratory crusher. The crushed sample was kept in an airtight plastic container for further analysis.

2.2. Preparation of Raphia taedigera adsorbent

2.2.1. Carbonization of samples

The carbonization was done as reported by [53]. The dried and crushed *R. taedigera* seeds were carbonized by placing 100 g of the seed in a muffle furnace (Carbolite, Sheffield, England, LMF4) at 350°C for about 2 hours. During carbonization, an exhaust pipe was used to remove steam to set the process oxygen-deficient. This made the material to thermally decompose to porous carbonaceous materials and compounds of the hydrocarbon. After the carbonization process, the carbonized biochars were

allowed to cool to room temperature, washed with double distilled water to a pH of about 7 and dried to constant weight in an oven at 105° C. The carbonized biochars were sieved with 100-mm mesh Tyler sieve and kept in an airtight polythene bag.

2.2.2. Activation of the carbonized samples

The activation was done as reported by [53]. Both acidic and basic activations were done on the material. 20 g carbonized material was carefully weighed and quantitatively transferred into a clean and dry 500 mL beaker containing 200 mL 0.1 M sulphuric acid for acidic activation and 200 mL of 0.1 M sodium hydroxide was used for alkaline activation of another 20 g of the carbonized material. The contents of the beakers were carefully homogenized and left to stand for about 24 hours. The impregnated materials were diluted with 200 mL distilled water to rinse off the sulphuric acid and sodium hydroxide used as activating agents respectively. The process of washing continued until the pH of 7.0 was attained and transferred into an oven and dried to constant weight at $105\,^{\circ}\text{C}$ for about 4 hours. It was sieved with 100-mm mesh to obtain a fine powder of *Raphia taedigera* activated biochars for both the acid (RTA) and the base (RTB). The RTA, RTB, and RTR (raw *Raphia taedigera* seed powder) were stored in a separate airtight plastic container for further use.

2.3. Material Characterization

The morphology of the samples was carried out using scanning electron microscopy (SEM) to evaluate the microstructure characteristics. TGA/DTA was used to determine the thermal behaviour of the samples using thermogravimetric Analyzer Perkin-Elmer TGA 4000 at the temperature range of 30 - 900°C while Fourier-transform infrared spectrophotometer (FTIR) was engaged to evaluate the molecular bonding present in the materials.

2.4. Preparation of Methylene Blue (MB)

 $100\,$ mg/L of methylene blue (MB) stock solution was prepared by quantitatively dissolving $0.1\,$ g MB in distilled water in $1,000\,$ mL volumetric flask and made up to the mark with distilled water. The maximum absorption wavelength was determined by UV spectrophotometer. $0.1\,$ M of HCl and $0.1\,$ M NaOH solutions were also prepared to adjust the pH. All reagents used were of analytical grade.

2.5. Evaluation of Effect some Parameters on the Adsorption

2.5.1. Effect of Adsorbent Dose

The effect of the adsorbent dose was experimented using different adsorbent dosages of 0.1 g, 0.3 g, 0.5 g, 0.5 g, 0.7 g and 0.9 g. 10 mL of 100 mg/L MB was introduced into a beaker containing each of the adsorbent doses for 15 minutes at room temperature. The solution was filtered, and the concentrations of the filtrates were determined using a UV spectrophotometer and the amount of MB adsorbed was calculated using Eq 1.

$$q_e = \frac{(C_o - C_e)V}{W} \tag{1}$$

Where, q_e is the amount of MB adsorbed per unit mass in mg/g, C_o is the initial MB concentration in mg/L, C_e is the MB concentration at equilibrium in mg/L, V is the volume of MB solution in liter and W is the mass of the adsorbent in grams.

The adsorption efficiency was calculated using the equation below:

Adsorption efficiency =
$$\frac{(C_o - C_e)}{C_o} \times 100$$
 (2)

2.5.2. Effect of Initial Solution Concentration

The effect of initial concentrations of MB was evaluated using different concentrations of 20 mg/L, 40 mg/L, 60 mg/L. 80 mg/L and 100mg/L. 0.5 g of the adsorbent was introduced into a conical flask containing 10 mL of each of the solution and allowed to stand for 15 minutes at room temperature. The solution was filtered, and the concentrations of the filtrates were determined. The amount of MB adsorbed and percentage efficiency of the adsorbents were calculated using equations 1 and 2 respectively.

3. Results and Discussion

3.1. Surface morphology studies.

The surface morphology studies of RTR, RTA and RTB were done using Scanning electron microscopy and presented in Figures 1, 2 and 3 respectively. The photomicrographs at a magnification of 300x, 500x and 1000x are given as A, B and C respectively. As expected, pyrolysis and surface activation, altered to a noticeable level, the surface morphologies of the biochars (RTA and RTB) compared to raw *Raphia taedigera* (RTR). Figure 1 shows scattered aggregated surface, while figures 2 and 3 show aggregated and rough surface morphology with links between the surface particles creating many more pores at the surfaces. The roughness and uneven nature of the surfaces observed in figures 2 and 3 are due to the activation of the porous carbon using H₂SO₄ and NaOH solutions respectively.

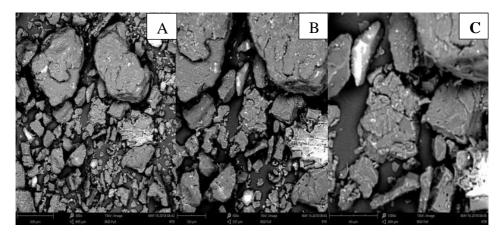


Fig. 1. SEM images of RTR at 300x (A), 500x (B) and 1000x (C)

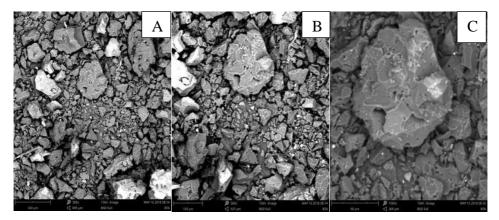


Fig. 2. SEM images of RTA at 300x (A), 500x (B) and 1000x (C).

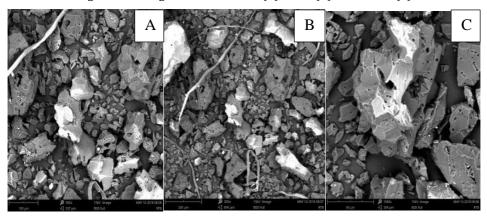


Fig. 3. SEM images of RTB at 300x (A), 500x (B) and 1000x (C)

3.2. FTIR Spectral Analysis

Figures 4, 5 and 6 show the FTIR spectra of raw R. taedigera seed powder, acid activated R. taedigera seed and base activated R. taedigera seed respectively. Again significant changes were observed in the materials in term of band shift, band disappearances and new band formations due to pyrolysis and activation of RTA and RTB. As shown in the spectra, a strong and sharp band at about 3700 - 3500 cm-1 due to free OH group, a broad band at 3600 - 3200 cm⁻¹ representing bonded OH group, a band at about 3000 -2850 cm⁻¹ corresponding to aliphatic C-H group, a band at about 1820 - 1670 cm⁻¹ indicating C=O stretch, a band in about 1385 – 1345 cm⁻¹ indicating N-O stretch of nitro molecule, a band at about 1300 - 1000 cm⁻¹ corresponds to C-O stretch of ester and a strong band at about 800 - 600 cm⁻¹ indicating C-Cl of alkyl halide were observed in the three materials. Comparing the spectra of RTA and RTB (Figures 5 and 6) with RTR (figure 4), two new bands corresponding to free OH group at about 3700 - 3500 cm⁻¹, new band at about 1600 - 1400 cm⁻¹ corresponding to C=C of aromatic and a band at about 1354 cm⁻¹ corresponding to C-N stretching of amine were observed. However, the extra band at 2374 cm⁻¹ indicating CN stretch of nitrile and a band at 1241 cm⁻¹ corresponding to CN stretch of amine observed in RTA spectral were not found in RTB spectral. The changes in the FTIR spectra of the materials may be due to the introduction of activating agents for surface modifications [54, 53] of RTA and RTB.

Undoubtedly, the shifts in the spectra indicated the prepared materials would be of great use as adsorbents for heavy metals and dye removal. This is in line with reports of previous studies as reported by [55, 56, 57].

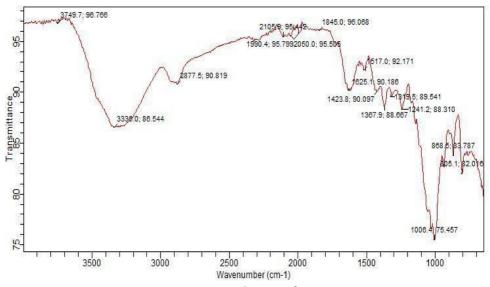


Fig. 4. FTIR Spectra of RTR

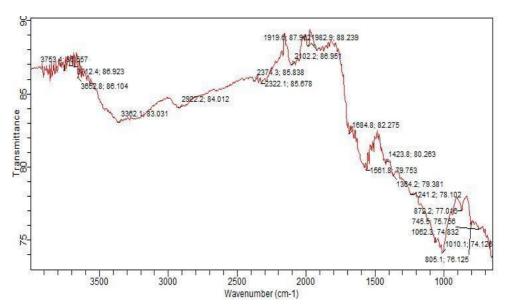


Fig. 5. FTIR Spectra of RTA

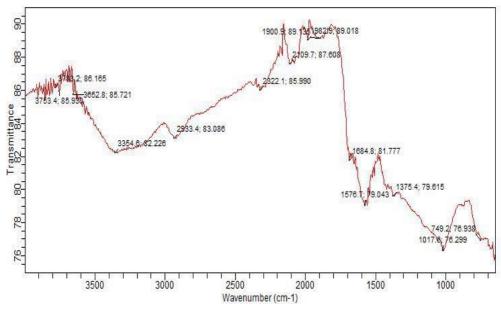


Fig. 6. FTIR Spectra of RTB

3.3. Thermal Behaviour

Thermogravimetric analysis (TGA) reveals the thermal stability of the materials while Differential thermogravimetric analysis (DTA) gives the temperature of the maximum decomposition rate. The TGA and DTA curves of raw R taedigera, acid and base biochars are presented as Figure 7, 8 and 9 respectively. According to the results of TGA presented in Figures 7, 8, and 9, there was slight loss of weight in the materials within a temperature range of 50 - 130°C. The loss of weight can be attributed to the removal of week-bonded water molecules within the materials. The second stage of weight loss which was a major weight loss in the materials was observed at around 350 - 500°C for RTR and at about 350 - 550°C for RTA and RTB. This second stage loss of weight was due to decomposition of glycosyl units of cellulose or lignin followed by the formation of char (Vilar et al., 2009). Also, as shown in the DTA Figures, the peaks corresponding to maximum degradation temperatures of RTR, RTA and RTB were observed at 380°C, 425°C and 410°C respectively. This indicates the activated materials exhibited greater thermal stability. This may be as a result of pyrolysis and the ability of acid and base used as activating agents to further burn out more of organic compounds from the carbonized material.

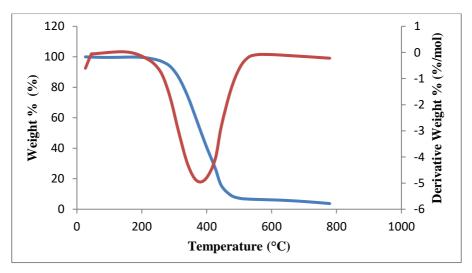


Fig. 7. TGA/DTG curves for RTR

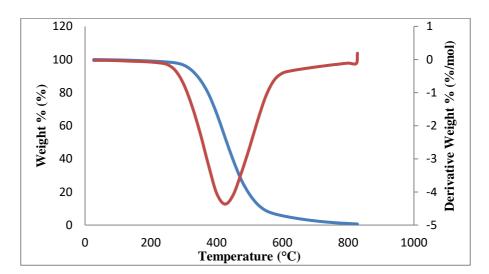


Fig. 8. TGA/DTG curves for RTA

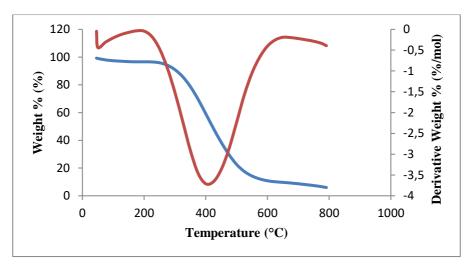


Fig. 9. TGA/DTG curves for RTB

3.4. Results of Parameters Evaluation

3.4.1. Determination of Maximum Absorption Wavelength.

Maximum absorption wavelength of the stock solution was determined using UV-1800 series spectrophotometer between the wavelength 200 and 900 nm. The scanning revealed the maximum absorption wavelength of the methylene blue to be 659 nm as obtained from the curve of absorbance against wavelength as presented as Figure 10. The maximum absorption wavelength was used for subsequent determinations.

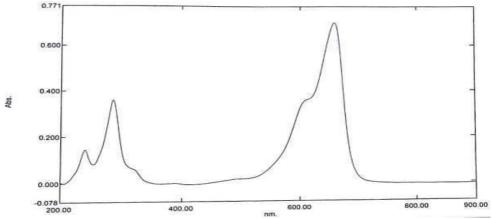


Fig. 10. Absorption Curve of Methylene Blue as a function of Wavelength

3.4.2. Effect of Biochar Dosage

The effect of adsorbent dosage on Methylene blue was determined at room temperature (298 K), adsorbent concentration (100 mg/L) and time (15 minutes). The result, as presented in Figure 11 showed an increase in adsorption as the dosage increases [58, 59, 60] up to 0.7 g. The increase may be due to the availability of a larger active surface area [61] and pore size. Maximum adsorptions of 99.207% and 89.997% were observed at 0.7 g for RTA and RTB respectively. Reduction in the adsorption was observed with a

dosage above 0.7 g for both adsorbents. The reduction can be attributed to the initial vacant sites on the adsorbents being quickly and completely filled with molecules of MB.

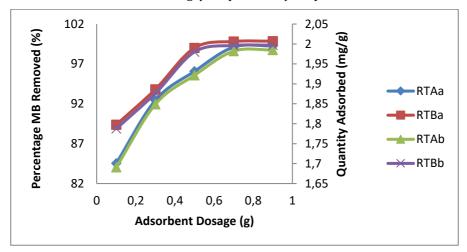


Fig. 11. Effect of dosage on adsorption capacity of RTA and RTB

RTAa and RTBa are percentage of MB removed by RTA and RTB respectively, while RTAb and RTBb are quantity of MB adsorbed per unit mass of RTA and RTB respectively

3.4.3. Effect of Initial Solution Concentration

The effect of initial methylene blue concentration was evaluated with 20.00 mg/L, 40.00 mg/L, 60.00 mg/L, 80.00 mg/L and 100.00 mg/L of methylene blue at constant temperature (298K), time (15 min) and adsorbent dose (0.5 g). Figure 12 revealed that adsorption increases with concentration. This implies that the diffusion of methylene blue is accelerated with increasing concentration due to the increase in forces of attraction of the concentration gradients [62] leading to effective interactions between dye particles and adsorbent surface [63]. On the other hand, the continuous increase of the initial concentration of MB reduces the adsorption capacity of the adsorption; this may be due to the equilibrium between the MB particles and the surface of the adsorbent.

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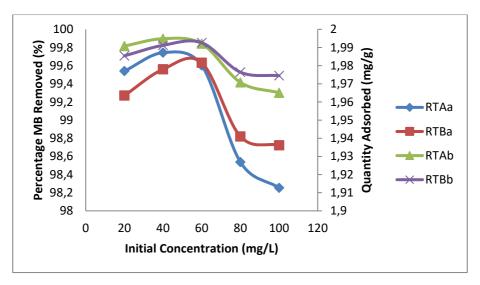


Fig. 12. Effect of initial concentration on adsorption capacity of RTA and RTB

RTAa and RTBa are percentage of MB removed by RTA and RTB respectively, while RTAb and RTBb are quantity of MB adsorbed per unit mass of RTA and RTB respectively

4. Conclusion

In the present study, new potential acidic activated *Raphia taedigera* seed and alkaline activated *Raphia taedigera* seed biochars were prepared from *Raphia taedigera* seed. The biochars were prepared by carbonization and chemical activation using 0.1 M sulphuric acid and 0.1 M sodium hydroxide to produce RTA and RTB respectively. Scanning electron microscopy (SEM), Fourier-transform infrared (FTIR) spectrophotometry, and thermogravimetric/Differential thermal analyses (TG/DTA) were carried out on the prepared biochars and raw *R. taedigera* seed.

- The results of the analyses revealed that acid and base activated materials could be excellent adsorbents for removal of dyes from industrial and agricultural wastewater.
- Considering the results from scanning electron microscope, the activated materials have aggregated and rough surface morphologies with links between surface particles creating porous structure with greater homogeneity.
- Results of thermal analyses, showed higher thermal stability of the biochars with 380°C, 425°C and 418°C maximum decomposition temperatures for RTR, RTA and RTB respectively.
- Fourier-transform infrared spectrophotometry result revealved a large number of peaks corresponding to different functional groups such as hydroxyl group, ketonic group, carboxylic group, alkene group among others, which are all capable of aiding adsorption process.
- The adsorbents performance evaluations using different initial concentrations of methylene blue and adsorbent dosage were evaluated. The results revealved the percentage efficiencies of RTA and RTB to be within 98.245 % to 99.743 % and 98.723 % to 99.632 % respectively for the methylene blue initial concentrations range of 20 to 100 mg/L. Also, 84.542 % to 99.256 % and

89.375 % to 99.851 % efficiencies were observed for RTA and RTB respectively, for the effect of adsorbent dosages range of 0.1 g to 0.9 g.

The results of the preliminary investigations showed great potentials of the materials being used as adsorbents. Further studies are required to investigate the mechanism involved and the efficiencies of the prepared materials.

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