IJAPC

VOLUME 9 ISSUE 2 2018

www.ijapc.com E ISSN 2350-0204

GREENTREE GROUP PUBLISHERS



RESEARCH ARTICLE

www.ijapc.com e-ISSN 2350-0204

Pharmacological Evaluation, Phytochemical Screening and Analytical Profiling of Various Extracts of Clerodendron serratum: An Medicinal Herb

Sapna Malviya^{1*} and P. Dwivedi²

^{1,2}Govt. Madhav Science P. G. College, Ujjain (M.P.), India

ABSTRACT

Clerodendron serratum belongs to family *verbenaceae* known as *Bharangi* commonly found in the India. The *Clerodendron serratum* showed anti-fungal, anti-oxidant, tuberculosis, antiasthmatic, anti-bacterial and anti-inflammatory properties. The *Clerodendron serratum* contains D-mannitol, cleroflavone, apigenin, scutellarein, serratagenic acid, queretaroic acid and γ -sitosterol. The phytochemical screening was performed on petroleum ether, chloroform and methanolicextractsof the plant. Analytical techniques (Thin Layer Chromatography, Fourier Transmittance Infrared and High Performance Thin Layer Chromatography) and chemical test confirmed the presence of alkaloids, flavanoids, phenols, terpenoids, steroids and saponins in plant extracts. Most of the phytochemicals were present in chloroform and methanol extracts. The alkaloid found in chloroform and methanol extract respectively at R_f 0.42 and 0.45. Flavanoids and phenol were present in chloroform and methanol extract. Terpenoids and saponins were present in methanol extract while steroids were observed in chloroform extract only. Further antibacterial and antifungal activities were also performed which showed positive result for plant extracts.

KEYWORDS

Clerodendronserratum, Antibacterial, Antifungal, Phytochemical Screening, TLC, HPTLC



INTRODUCTION

In a present scinerio drugs have been derived directly or indirectly from traditional medicinal plants. Microscopic and macroscopic descriptions of medicinal plants is the first step towards establishing the identity and purity and such work carried before should be out performingother tests. Anatomical characters are also helpful for the identification of drug when morphological features are indistinct¹⁻⁵.

In India there are about 7500 species of the flowering plants known to have medicinal properties. Various parts of the plant may be used for medicinal purposes uch as; root, rhizome, wood, bark, flower, fruit and seed. Bark is outermost covering of old stem and trunk. The term bark is used most often in a non-technical context and refers to all tissue outside the vascular cambium of the axis, in either a primary or secondary state of growth⁶⁻⁸. The outer layer protects the tree from hot or chilly winds and it may be impregnated with certain chemical substances. The secondary metabolites like tannins, phenolic, steroids, alkaloids, crystals of calcium carbonate, calcium magnesium oxalate and oxide etc. embedded in the barks. Therefore bark is used as medicine to treat several diseases includingstem bark as well as root barks. In

Ayurvedicsystem the barks are used in the form of fine powder (*Churnas*), infusion (*Fanta*), decoction (*Kadha*) or fermented decoction (*Arishtha*) or can be made into pills (*Vati* or *Guti*)⁹⁻¹².

Considering importance of bark present investigation were planned to perform phyto-chemical and pharmacological evaluation of bark extracts of *Clerodendronserratum*. The study was aimed to explore folklore use of plant bark.

MATERIALS AND METHODS Sample collection

The *Clerodendron serratum* was collected from the Indore (*Malwa*) region India. The authentication of plant was done in the Ayurveda department Indore. The bark of plant was scraped with the help of knife, in the month of November 2016 and dried in shade. After drying the bark was grinded into course powder and stored in plastic vessel.

Extract preparation

The course powder of bark of *Clerodendron serratum* was divided into three parts. Each part having 25 g of bark powder extracted with petroleum ether, chloroform and methanol. The first 25g bark powder was wrapped into thimble and kept in upper chamber of Soxhlet apparatus (Fig.1) and in lower portion solvent was

present, heated to evaporation and solvent reaches to thimble (upper portion) and passes through the sample powder as a result extraction was starts. same procedure was adopted for chloroform and methanol extracts.

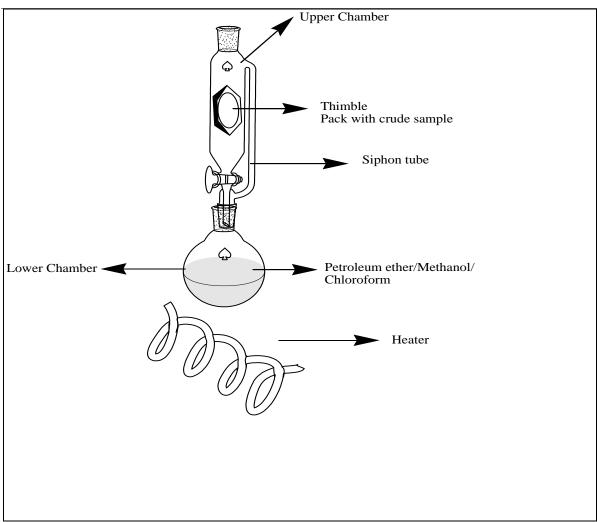


Fig. 1 The Soxhlet apparatus used in extraction The petroleum ether, chloroform and methanolic extracts were collected, filtered with Whatmann No. 1 filter paper, evaporated till drying, stored in air tight container and analyzed further¹³.

Thin layer chromatography

The thin layer chromatography profiling of dried extracts of petroleum ether, chloroform andmethanol was performed using silicagel plate. The three TLC plates were taken and 50 μ l crude extracts applied on 1 centimeter above the TLC plate with the help of micro-pipette. After sample application the plates were dried and kept in the chamber equipped with solvent system; Ethyl Acetate: Chloroform: Water: methanol 5:3:1:1. The solvent mixture was allowed to travel ³/₄th of plate height, after that plate was removed and dried. The dried plate kept in UV chamber and then in iodine chamber to detect spots. The spots were scraped and analyzed further for chemical evaluation¹⁴.

The scraped bands of TLC plate were dissolved in10 mL respective solvent, filtered and then subjected to following tests:

Chemical Test

Discussed in Table 1.

| | | | Extracts of Clerodendron serratum | | | | | |
|------------------------------------|------------|---------------------------|-----------------------------------|----------------------------|------------|----------------------|------------------|----------------------------|
| S. No. Constituents Test performed | | | Petroleum Ether Extract | | Chloroform | | Methanol extract | |
| | | | Results | <i>R_f</i> Value | Results | R _f Value | Results | <i>R_f</i> Value |
| 1 | Alkaloids | Mayer's Test | -ve | - | +ve | 0.42 | +ve | 0.45 |
| 1 | Aikaiolus | Wagner's Test | -ve | | +ve | 0.42 | +ve | |
| 2 | Flavanoids | Ammonia Reduction Test | -ve | - | +ve | 0.30 | +ve | 0.32 |
| 3 | Phenol | Ferric Chloride Test | -ve | - | +ve | 0.70 | +ve | 0.71 |
| 4 | Terpenoids | Salkowski's Test | +ve | 0.54 | -ve | - | +ve | 0.53 |
| 5 | Steroids | Salkowski's Test | -ve | - | +ve | 0.60 | -ve | - |
| 6 | Saponins | Froth Test | +ve | 0.80 | -ve | - | +ve | 0.80 |

| Table 1 Phytochemical screening chemical test and TLC results (R_f Value) of extracts of Clerodendron serratum | |
|---|--|
| Extracts of Clerodendron serratum | |

IR Spectroscopy

The dried petroleum ether, chloroform and methanol extract of plant Clerodendron serratum kept in oven for drying then triturated in mortal pestle along with dried KBr. The blank reading of KBr analyzed in FTIR (Fourier transform infrared) to avoid the interference of it in sample. The triturated extract kept in sample cell of FTIR and instrument allowed to run spectra which generated peaks of functional group present in sample¹⁰.

High performance thin layer chromatography (HPTLC)

The silica gel GF HPTLC plate were used for analysis and activated in oven prior to spotting. The sample of dried petroleum ether, chloroform and methanol extract of

plant was applied with an automatic applicator. Thesolvents used asmobilephase were of composed of Ethyl Acetate: Chloroform: Water: methanol 5:3:1:1. The solvent mixture was filtered and kept in chamber for saturation. The silica gel GF HPTLC plate kept in HPTLC chamber and mobile phase was allowed to run for few hours. After development plate was removed from chamber dried to avoid contamination. The plate was kept in UV chamber for confirmation of spot. The TLC scanner used to detect spot at 200 and 800 nm^{15} .

Pharmacological evaluation

The pharmacological evaluation of extracts of *Clerodendron serratum* was performed as follows:

a. Anti-bacterial activity: Cup-plate method

The culture medium was made with nutrient agar in Ultra-Violet laminar air flow and sterilization was done at 60°C in an autoclave. The medium was poured on glass plate and mixed with bacterial suspension and after drying cups were prepared over it. The samples were then poured over bacterial cup and plate and activity was measured by visual inspection.¹⁶⁻¹⁸

b. Antifungal activity by inhibitory zone estimation:

Disk diffusion method was used to determine antifungal activity. Sample was prepared and disk was dipped in it with solid culture medium. All plates of different extract were kept for incubation at 37 °C in UV chamber for 48 h. after incubation inhibition was measured by the round scale¹⁶⁻¹⁸.

RESULTS AND DISCUSSION

The plant sources were widely used for medicinal purpose since ancient times due to active chemical constituents present in it. The results of phytochemical screening are compiled in table-1 which confirms presence of secondary metabolite which contributed towards antimicrobial activity. The TLC was also performed using mixture of ethyl acetate: chloroform: water: methanol as mobile phase. The alkaloid was present in the chloroform and methanolic extract of plant respectively at R_f value 0.42 and 0.45. The alkaloid was confirmed in scrap part of TLC by Mayer's Test and Wagner's Test. The flavonoid was confirmed by ammonia reduction test at R_f value 0.30 and 0.2 in chloroform and extract, respectively. The methanol terpenoids was confirmed by Salkowski's Test at R_f value 0.54 and 0.53 in petroleum ether and methanol extract of plant respectively. The steroid was present in chloroform extract of plant at R_f value 0.60. The saponin was confirmed by Froth Test at R_f value 0.80 and 0.80 in petroleum ether and methanol extract respectively. The investigation confirmed of presence important phytochemical in plant extracts which are known for therapeutic value.

The FTIR technique was used to identify functional group present in the extract. The narrow -NH group peak obtained at 3502 cm⁻¹, broad -OH peak at 3400 cm^{-1} , =C-H peak at 2200 cm⁻¹, -C-H peak at 2000 cm⁻¹, C-O peak at 1800 and -CHO peak at 1700 cm⁻¹. The results of IR confirmed study presence of characteristics peaks of Alkaloids, Flavanoids, Terpenoids, Steroids and Saponins.

The HPTLC also performed which

confirmed the presence of Alkaloids, Flavanoids, Phenol, Terpenoids, Steroids and Saponinsin the extract. The spots for Alkaloids, Flavanoids, Phenol, Terpenoids, Steroids and Saponins found in TLC respectively at R_f value 0.45, 0.32, 0.71, 0.53, 0.60 and 0.80. The TLC and HPTLC results were calculated and found similarity between both. The antibacterial activity of extract chloroform (petroleum ether. and methanolic) of Clerodendron serratum performed by a cup-plate technique. The petroleum chloroform ether, and methanolic extract showed positive result *Staphylococcus* against aureus and Salomonella typhimurium (Table 2).

Table 2 Anti-bacterial activity of petroleum ether, chloroform and methanol extract of *Clerodendron serratum*

| S.No. | Bacteria | Clerodendron serratum | | | |
|-------|------------------------|-----------------------|------------|------------------|--|
| | | Petroleum | chloroform | Methanol extract | |
| | | Etherextract | Extract | | |
| 1 | Staphylococcus aureus | +ve | +ve | +ve | |
| 2 | Salomonellatyphimurium | +ve | +ve | +ve | |

The Anti-fungal activity of extract (petroleum ether, chloroform and methanolic) of *Clerodendron serratum* performed through a zone inhibition technique. The petroleum ether extract showed negative result against *A.flavus* while showed positive result against *P*.

notatum, A.niger and *A.fumigatus.* The chloroform extract showed positive result against *A.flavus* and *P. notatum*, while showed negative result against *A.niger* and *A.fumigatus.* The methanolic extract showed positive result against *A.flavus* and *P. notatum* (Table 3).

Table 3 Anti-fungal activity of petroleum ether, chloroform and methanol extract of Clerodendron serratum

| | | Clerodendronserratum | | | |
|-------|-------------|-------------------------|-----------------------|------------------|--|
| S.No. | Fungus | Petroleum ether extract | Chloroform extract | Methanol extract | |
| 1 | A.flavus | -ve | +ve | +ve | |
| 2 | P. notatum | +ve | +ve | +ve | |
| 3 | A.niger | +ve | -ve | -ve | |
| 4 | A.fumigatus | +ve | -ve | -ve | |

CONCLUSION

The Clerodendron serratum is an important medicinal used plant from ancient times. The results of phytochemical analysis suggested that Clerodendron serratum bioactive phytochemicals possess of medicinal importance. The phytochemical screening initiated by TLC profiling and chemical test confirmed presence of Alkaloids, Flavanoids, Phenol, Terpenoids, Steroids and Saponins. The pharmacological evaluation of extracts was also performed. The petroleum ether, chloroform and methanolic extract of plant showed potential therapeutic response against bacteria and fungus. The study concluded that the bark of plant Clerodendron serratum may be further recommended as potent antimicrobial agents.

Acknowledgement

We are thankful to the Govt. Madhav Science P. G. College, Ujjain (M.P.) for providing the facilities.

REFERENCES

1. John, D. B. A. Petchimuthu, K. Nirmal K. N. el at., (2007). Phytochemical study on *Indigoferaspp Linn. (Fabaceae)*. Journal of economic and taxonomic botany vol. 31 No. 4, 948.

2. John D. B. A., Petchimuthu K., Nirmal K. N. and Rekha, G.S. (2008). Preliminary phytochemical study on a medical plant *Mimosa pudica* L, (*Mimosaceae*). Journal of economic and taxonomic botany. 32 (1), 86-89.

3. Anonymous, 1996. Indian Pharmacopoeia, Government of India. Ministry of Health and family Welfare, The controller of publications civil lines, New Delhi- Vol. I &II.

 Alexander J.D., 1940. Plant microtechnique, McGraw Hill, London, England.
 Babu, L.B. Nambigan and Sudarshan, S. (1990). Comparative valuation of biochemical constituent of selected tuber crops, Journal of root crops. 270-273.

6. Esau, K. (1977). Anatomy of seed plants. 2nd edition, New York, USA.

7. Gamble, J.S. (1915). Flora of Presidency Madras, (BSI reprint, 1957), BSI, Calcutta, India, 1-3.

8. Ghosal, S. (2002). Herbal health products in global perspective. Science and Culture; 68(1-4): 33-40.

9. Haines, H.H. (1921). The Botany of Bihar and Orissa (BSI reprint, 1961), Calcutta, India.

10. Harborne J.B., 1988. Phytochemical methods, Gouda to modern technique of Plants Analysis. Chapman and Hall II Ed., New York.

11. Jayshree, D. P. and Kumar, V. (2008). *Annonasquamosa* L., Phytochemical analysis and antimicrobial screening, Journal of pharmacy research.1(1), 34-38.

12. Demirdöven, N. Cheatum, C. M. Chung, H. S. Khalil, M. Knoester, J. Tokmakoff A. (2004). "Two-dimensional infrared spectroscopy of antiparallel betasheet secondary structure". Journal of the American Chemical Society. 126 (25): 7981–90.

 Crombie D.S. and Chmurzynski L. (1997). High-performance liquid chromatographic determination of quinine in rat biological fluids. J Chromatogra. B: Biomed Sci Appl. 693, 423-429.

14. Ravishankara, M.N. Shrivastava, N. Padh H. and Rajani M. (2001). HPTLC Method for the Estimation of Alkaloids of Cinchona officinalis Stem Bark and its Marketed Formulations. Planta Med. 67, 294-296.

15. Mroczek T. and Glowniak K. (2000). TLC and HPTLC assay of quinoline and quinuclidine alkaloids in Cinchona cortex and pharmaceutical preparations. J Planar Chromatogr. 13, 457-462.

16. Sutar N, Garai R, Sharma US, Sharma UK. Anthelmintic activity of Platycladusorientalis leaves extract.
International Journal of Parasitology Research 2010; 2(2):1-3.

17. Shete, S. A. Shah, G. N. Walke, S. S. Patil, V. S. Patil K. D. and Killedar, S. G. (2013). Standardization and Antibacterial Activity of Couroupita Guianensis Fruit Shell Extract, Int. J. Bioassays, 2(1), 360-364.

Manikandan, A. Rajendran, R. Balachandar, S. Sanumol M. S. and Mary
 S. M. 2015. Antimicrobial activity of ailanthus excels roxb. collected from coimbatore district, Tamilnadu, India. 4,(03), 697-704.