http://www.irjse.in

ISSN: 2322-0015

## **ORIGINAL ARTICLE**

# Histopatholigical changes due to lead toxicity in gills of *P. ticto (hem)*.

Lalit Choudhary<sup>1</sup>, Tanay Vyas<sup>1</sup>, Naveent Raj Singh Chauhan<sup>2</sup>, Madhavi Bhoi<sup>2</sup>, Gopal Krishnan Yadav<sup>2</sup> and Seema Bharadwaj<sup>2\*</sup>

<sup>1</sup>Department of Zoology, Leo College, Banswara (Rajasthan) 327023 India.

## **Manuscript Details**

Received: 16.05.2019 Accepted: 20.07.2019 Published: 30.08.2019

ISSN: 2322-0015

Editor: Dr. Arvind Chavhan

#### Cite this article as:

Lalit Choudhary, Tanay Vyas, Naveent Raj Singh Chauhan, Madhavi Bhoi, Gopal Krishnan Yadavand Seema Bharadwaj. Histopatholigical changes due to lead toxicity in gills of *P. ticto (hem), Int. Res. Journal of Science & Engineering,* 2019, 7 (4): 92-95.

© The Author(s). 2019. Open Access
This article is distributed under the terms
of the Creative Commons Attribution
4.0 International License
(http://creativecommons.org/licenses/by/4.0/),
which permits unrestricted use, distribution, and
reproduction in any medium, provided you give
appropriate credit to the original author(s) and
the source, provide a link to the Creative
Commons license, and indicate if changes were
made.

#### **ABSTRACT**

Gill is a key organ for the direct action of pollutants in the aquatic environment. In the present study *P. ticto* was exposed with the sub lethal concentration of lead nitrate. The fishes were acclimatized and then exposed to the different concentration of lead nitrate. After the exposure, the histopathological changes in the gills were noted these changes showed the damages in tissue vacuole formation, lead deposition, shrinkage of lamella, fusion of tips of gill lamella and swelling of epithelial layer of gill.

**Key Words:** Histopathology, Toxicity, *P. ticto*, Lead nitrate.

## 1. INTRODUCTION

Heavy metals are generally water soluble, non-degradable, strongly bound to polypeptide and proteins. These metals are also very hard, strong, have very high boiling and melting points and are good conductors of heat and electricity. Elevated levels of lead in water arise principally from industrial discharges, high way run-off and weathering processes in area of natural lead mineralization. Lead is deposited from the atmosphere directly onto water surfaces. The pollution has entered in the environment through disposal of wastes and has entered in the organism through food chain [1]. Heavy metals pollute the water and disturb the whole aquatic ecosystem, hence they need exploration of different aspects of fish biology. Economical losses of fish farming are because of various water bound contaminants such as heavy metals, pesticides and sewage. These pollutants and eutrophication leading to adverse effects and cause of death of organisms in the aquatic system [2].

<sup>&</sup>lt;sup>2</sup>Department of Zoology, HDJ Govt. Girls College, Banswara (Rajasthan) 327023 India.

<sup>\*</sup>Corresponding author Email: <a href="mailto:seema377@gmail.com6">seema377@gmail.com6</a>

Choudhary et al., 2019 93

Heavy metals are being constantly discharged in the environment both from various human and natural activities viz. industrialization, vulcanization. Heavy metals are of greatest concern because after reaching the aquatic bodies they deteriorate the life sustaining quality of water and cause damage to both flora and fauna [3,4]. Lead, mercury and cadmium are of principal toxicological concern [5]. Pollution is an undesirable change in the physical, chemical or biological characteristics of land, air or water that show harmful effect on organisms [6]. Toxic impact brings about physiological, biochemical and pathological alterations in the organisms [7].

The LC50 values of zinc for the *Percocypris pingi* at 24 h, 48 h, 72 h and 96 h were 3.504 mg/L, 2.933 mg/L, 2.852 mg/L and 2.852 mg/L, respectively. But the LC50 values of copper at 24 h, 48 h, 72 h and 96 h were 1.730 mg/L, 1.389 mg/L, 1.340 mg/L and 1.340 mg/L, respectively [8]. The release of industrial, agricultural and domestic waste water into the aquatic environment cause pollution and fishes are often exposed with these metals. Especially in areas where the dilution effect is low and concentration is high [9,10]. Histological changes are providing an efficient and sensitive method to detect effects of contaminates in various tissues and organs of fish [11], Newton [12] worked on pollution of river of west Wales noticed that lead caused clogging and inflammation of gills and there was excess secretion of mucous due to lead toxicity. Bengeri and Patil [13] noticed the histopathological changes in the gill of *Puntis aurulius* due to lead toxicity.

## 2. MATERIAL AND METHODS

The living specimens of *P. ticto* were obtained from local ponds. Fishes were acclimatized to tube well water condition for a period of 15 days. No mortality occurred during this period. The fishes were washed with 0.1% KMnO<sub>4</sub> solution to obviate any normal infection, fishes were fed according to 1% body weight per day in the evening with dry shrimp powder. After acclimatization fishes were kept in different concentrations of lead nitrate. Methods of physicochemical analysis of water were followed as per description in APHA (1971). Paraffin sections of 5.0 to 8.0µ thickness were prepared for histopathological observations and were stained with double stain hematoxylin and eosin.

#### 3. RESULTS AND DISCUSSION

Histopathological observations in gills are marked. The gills show an outer layer of ciliated epithelium which exchange gasses, through circulation of water. Any hazardous impact on gill is noticed by histological changes in its epithelial lining. In control sections the gills show normal structure of its epithelial lining Figure-1. At 1.0 ppm concentration deterioration in the epithelial cells takes place. The layer of epithelium shrinks and gills appear to be thin long filaments Figure-2. At 5.0 ppm concentration the swelling of the epithelial layer takes place. The free gills filament at certain points in get joined together at their tips Figure-3. This obstruct free movement of gill filaments. This concentration deposits of lead are clearly observed within the epithelial lining Figure-4. This brings obstruction in the blood vessel for blood circulation in the gills. At the base of gill filament vacuole is created Figure-4. These histopathological changes are not observed in control section. These observations show similarities will earlier workers such as Fast et al., [15] found out the contents of Hg, Zn, Cd and Pb in fish Oncorhvnchus kisutch and O. tshwvtscha. Hodson et al., [16] conducted experiments on size and growth of rainbow trout with lead intoxication and noted that growth rate was affected with toxicity of lead. Giordano et al., [17] noticed the concentration of Hg, Cd and Pb in mussels of fish. Lu et al., [18] studied mean lead levels in fish flesh as 0.01-0.16 mg/kg. Carpenter [19,20] observed that Pb mucous film was formed over the gill surface of fishes due to toxicity of lead nitrate. He suggested that it was the main cause of death accompanied by respiratory distress. Ellis [21] pointed out that breathing distress in fishes is due to clogging of gills and direct damage is caused by heavy metal ions. Newton [12] suggested that lead caused clogging and inflammation of gills. Shukla [22] found that opercular beats of treated fishes were more in higher concentration of lead, while in recovery beats become normal. Oost et al., [9] noticed deleterious effects of heavy metal toxicity in different organs of fishes including gills, kidneys, liver and skin. Parashar and Banerjee [23] studied toxic impact of lethal concentration of lead nitrate on the gills of air-breathing catfish Heteropneustes fossilis (Bloch)<sup>23</sup>.

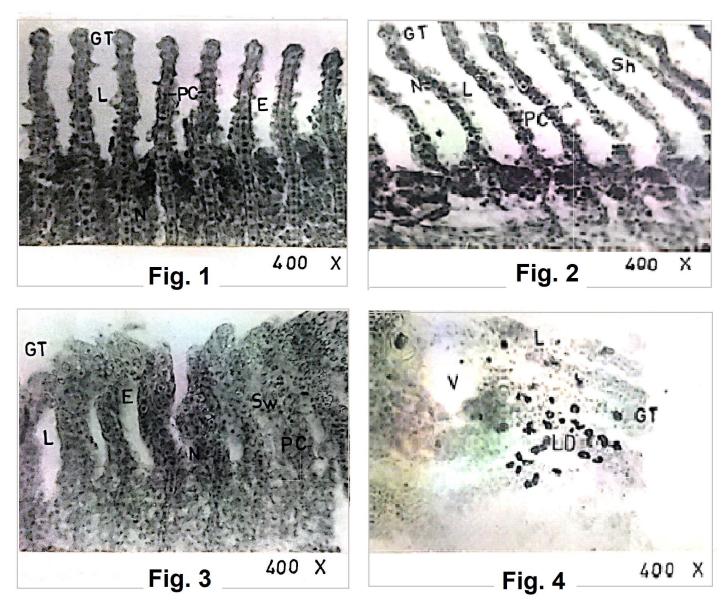


Fig. 1. L.S. of gill of control fish, showing normal structure of gill lamella.

Fig. 2. L.S. of gill of fish at 1.0 ppm concentration, showing shrinkage of lamella.

Fig. 3. L.S. of gill at 5. 0 ppm concentration, showing fused tips of gill lamella, and swelling of epithelial layer.

Fig. 4. L.S. of gill at 5. 0 ppm concentration, showing deposition of lead in gill lamella with formation of vacuole.

**Abbreviation** - E - Epithelium, GT - Gill tips, L - Gill lamella, LD - Lead deposits, N - Nucleus, PC - Pilaster cells, Sh - Shrinkage, Sw - Swelling, V - Vacuole)

## **CONCLUSION**

From the present study we can concluded that in control group the fish showed normal histoarchitecture of gills while treated fish is showed more notable changes in the gill architecture. The present study of histopathological effects of pollutants on the gills of *P. ticto* is an important

basic effort leading to our understanding of the true impact of pollutants on the ecosystem because the fishes show many histopathological changes in gills after exposed to different concentration of lead nitrate.

#### Conflict of interest

No conflict of interest influenced in this research.

Choudhary et al., 2019 95

#### REFERENCES

- 1. Bharadwaj S, Choudhary L. Histopathological changes in brain of *P. ticto* (HEM) due to lead toxicity. Inter. J. App. Envro. Sci., 2010; 5(2): 245-249.
- Dattatray GP. Clinicopathological Studies On Λ-Cyhalothrin and Lead Toxicities in Fish, Ph.D. Thesis, Department of Veterinary Pathology College of Veterinary Science, Guru Angad Dev Veterinary and Animal Sciences University Ludhiana, 2014.
- Semsettin A, Gey H, Suleyman G. Concentrations of heavy metals in water and chub, *Leuciscus cephalus* (*Linn*.) from the river Yildiz, Turkey. Journal of Environmental Biology, 2007; 28: 845-849.
- Yoon S, Sang-Seop H, Rana SVS. Molecular markers of heavy metal toxicity-A new paradigm for health risk assessment. Journal of Environmental Biology, 2008; 29: 1-14.
- Hammond PB, Clark CS. Faecal lead excretion in young children as related to sources of lead in their environment. Int. Arch. Environ., 1980; 46: 191-202. DOI: 10.1007/BF00380010.
- Bose MTJ, Ilavazhahan M, Tamilselvi R, Viswanathan M. Effect of Heavy Metals on The Histopathology of Gills and Brain of Fresh Water Fish Catla Catla. Biomedical & Pharmacology Journal, 2013; 6(1): 99-105.
- Shivakumar R, Mushigeri SB, Davie M. Effect of Endosulfan to freshwater fish *Ctenopharyngodonidellus*. J. Ecotoxl. Environ. Monit., 2005; 15: 113-116.
- 8. Zeng L, Huang L, Zhao M, Liu S, He Z, Feng J, Qin C, Yuan D. Acute Toxicity of Zinc Sulfate Heptahydrate (ZnSO4\*7H2O) and Copper (II) Sulfate Pentahydrate (CuSO4\*5H2O) on Freshwater Fish, *Percocypris pingi*. Fish Aqua. J., 2018; 9(1). DOI: 10.4172/2150-3508.1000240.
- 9. Oost RVD, Beyer J, Vermeule NPE. Fish bioaccumulation and biomarkers in environmental risk assessment: a review. Environ. Toxicol. Pharm., 2003; 13: 57–149.
- Bernet D, Schmidt H, Meier W. Burkhardt-Holm P, Wahli T. Histopathology in fish: proposal for a protocol to assess aquatic pollution. J. Fish Dis., 1999; 22: 25–34. DOI: 10.1046/j.1365-2761.1999.00134.x.
- 11. Rodrigues S, Antunes SC, Nunes B, Correia AT. Histological alterations in gills and liver of rainbow trout (*Oncorhynchus mykiss*) after exposure to the antibiotic oxytetracycline. Environmental Toxicology and Pharmacology, 2017; 53: 164–176. DOI: 10.1016/j.etap.2017.05.012.

12. Newton L. Pollution of rivers of waste water by lead and zinc mine effluents. Ann. APPL. Biol. Brit., 1944; 31: 1-11. DOI: 10.1111/j.1744-7348.1944.tb06200.x.

- 13. Bengeri KV, Patil HS. Histopathological changes in gills of *P. aurulius* induced by lead. J. Amm. Morphol. Physiol., 1987; 34(112): 113-116.
- 14. APHA. Standard methods for the analysis of water, sewage and industries water. American Public Health Association, Inc. New -York, 1971.
- 15. Fast RW, D'Itri FM, Barclay DK, Katase SA, Madenjian C. Heavy metal content of *Coho Oncorhynchus Kisutch* and Chinook Salmon *O. tishawytscha* reared in deep upwelled ocean waters in Hawaii (USA). J. World Aqua. Soc., 1990; 21(4): 271-276. DOI: 10.1111/j.1749-7345.1990.tb00539.x.
- Hodson, PV, Dixson DG, Spry DJ, Whittle DM, Sprague JB.
   Effect of growth rate and size of fish on rate of intoxication by waterborne lead. Canadian Fish Aquat. Sci., 1982; 39(9): 1243-1251.
- 17. Giordano R, Arata P, Ciaralli L, Rinaldi S, Giani M, Cicero AM, Costantini S. Heavy metal in mussels and fish from Italian Coastal waters. Mar. Pllut. Bull., 1991; 22(1): 10-14. DOI: 10.1016/0025-326X(91)90438-X.
- 18. Lu PY, Metcalf RL, Furman R, Vogel R, Hassett J. Model ecosystem studies of lead and cadmium and of urban sewage sludge containing these elements. Environ. Quality, 1975; 4: 505. DOI: 10.2134/jeq1975.00472425000400040017x.
- 19. Carpenter KE. The lethal action of soluble metallic salts on fishes. Brit. J. Expt. Biol., 1927; 4: 378.
- 20. Carpenter KE. Further researches on the action of metallic salts on fishes. J. Expt. Zool., 1930; 56(4): 407-422. DOI: 10.1002/jez.1400560403.
- 21. Ellis MM. Detection and measurement of pollution. U.S. Bur. Fish Bull., 1937; 48(22): 365-437.
- 22. Shukla S. Physico-chemical and histopathological observation on the effects of lead in certain organs of fresh water teleost *P. sophore* (Ham), M. Phill thesis, S.S. in zoology, Vikram university Ujjain (MP), India, 1989.
- 23. Parashar RS, Banerjee TK. Toxic impact of lethal concentration of lead nitrate on the gills of air-breathing catfish *Heteropneustes fossilis* (Bloch). Vet. Arhiv., 2002; 72(3): 167-183.

© 2013-2019 | Published by IRJSE