

Asian Pacific Journal of Tropical Biomedicine



Journal homepage: www.apjtb.org

doi: 10.4103/2221-1691.244160

©2018 by the Asian Pacific Journal of Tropical Biomedicine.

Endophytic actinobacteria of medicinal plant Aloe vera: Isolation, antimicrobial, antioxidant, cytotoxicity assays and taxonomic study

Ahmed Nafis¹², Ayoub Kasrati², Asma Azmani¹, Yedir Ouhdouch¹, Lahcen Hassani¹

¹Laboratory of Biology and Biotechnology of Microorganisms, Faculty of Sciences Semlalia, Cadi Ayyad University, PO Box 2390, Marrakech, Morocco

²Department of Biology, Faculty of Sciences Semlalia, Cadi Ayyad University, Marrakech, Morocco

ARTICLE INFO

Article history: Received 16 August 2018 Revision 7 September 2018 Accepted 3 October 2018 Available online 29 October 2018

Keywords: Actinobacteria Medicinal plant Aloe vera Antimicrobial activity Cytotoxicity Antioxidant assays Molecular identification

ABSTRACT

Objective: To explore the new sources of novel bioactive compounds having pharmaceutical and agricultural interest and to search the endophytic actinobacteria from medicinal plants. Methods: NAF-1 an endophyte actinobacteria was isolated from leaves of medicinal plant Aloe vera collected in Marrakesh, Morocco using Bennett agar as selective medium. NAF-1 was tested for its antimicrobial activity against five pathogenic bacteria such as Staphylococcus aureus PIC 53156, Micrococcus luteus ATCC381, Bacillus subtilis ATCC 14579, Pseudomonas aeruginosa DSM 50090 and Escherichia coli ATCC 8739 and four human clinic fungi belonging to the Candida, Aspergillus and Microsporum genera. Several antioxidant activities were studied such as DPPH free radical scavenging, β -carotene and linoleic acid and reducing power assays. The total of phenol and flavonoid was also calculated. Using Artemia salina shrimp assay, the cytotoxicity of NAF-1 crude extract was determined. Results: The results revealed that the actinobacteria showed a high activity (≥20 mm) against only Gram positive bacteria but it had a moderate activity (between 13 and 15 mm) against Human clinic fungi. The isolate also exhibited a LD₅₀ of 14.20 μ g/mL in the cytotoxicity assay. The result showed that the crude extract presented an interesting free radical-scavenging activity with IC_{50} value of (5.58 \pm 0.26) µg/mL and a high value of phenolic and flavonoid compounds with (15.41 \pm 0.18) µg GAE/mg extract and $(11.41 \pm 0.06) \mu g$ QE/mg extract respectively. Moreover, the taxonomic position of our endophyte actinobacteria using the morphological and physiological criteria and using 16S rRNA gene sequence (polyphasic approach) showed that the NAF-1 isolate was similar to Streptomyces hydrogenans which was never described as an endophyte actinobacteria. Conclusions: This isolated strain appears promising resources of bioactive agents and can be exploited to produce therapeutic agents active against pathogenic disease.

1. Introduction

Pathogens multi-drug resistance capacity becomes more and more severe due to abusive use of antibiotics. One way to solve this problem is to look permanently for new bioactive compounds especially from natural resources[1].

E-mail: ahmed.nafis@edu.uca.ac.ma

Endophytic actinobacteria are antagonists microorganisms that act inside the internal tissues of plants without having any negative

©2018 Asian Pacific Journal of Tropical Biomedicine Produced by Wolters Kluwer-Medknow

How to cite this article: Nafis A, Kasrati A, Azmani A, Ouhdouch Y, Hassani L. Endophytic actinobacteria of medicinal plant Aloe vera: Isolation, antimicrobial, antioxidant, cytotoxicity assays and taxonomic study. Asian Pac J Trop Biomed 2018; 8(10): 513-518.

[™]First and corresponding author: Ahmed Nafis, Laboratory of Biology and Biotechnology of Microorganisms, Faculty of Sciences Semlalia, Cadi Ayyad University, PO Box 2390, Marrakech, Morocco. Tel: +212610170760

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-Non Commercial-Share Alike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms

For reprints contact: reprints@medknow.com

influence on the plant host[2]. Medicinal plants constitute huge diversity of endophytic actinobacteria, which are filamentous Grampositive bacteria, with important economic value because of their ability to produce high potential bioactive substances including antimicrobial, anti-cancer and other pharmaceutical compounds[3,4]. Among members of actinobacteria, *Streptomyces* genus remains the richest sources of valuable natural product with 7 600 (76%) bioactive metabolites[5,6].

To date, few are known about the antibiotics produced by endophytic actinobacteria. Thus, the present study is focused on isolating actinobacteria from the leave tissues of medicinal plant *Aloe vera* (*A. vera*), on evaluating the antimicrobial, antioxidant and cytotoxic activities of its secondary metabolites and on identifing it on the basis of the morphological, physiological and 16S rRNA gene analysis.

2. Materials and methods

2.1. Sample collection and isolation of endophytic actinobacteria

Strain NAF-1 was isolated from the leaves of a medicinal plant *A. vera* collected in Marrakech, located in the north of the foothills of the Atlas Mountains, Morocco. Firstly, the samples of leaves were washed by distillated water for 1-2 min to remove soil particles and then surface sterilized with 70% ethanol for 10 min and 1% sodium hypochlorite for 15 min. In order to reduce the opportunity of endophytic fungi emergence, the leaves were soaked 10 min in 10% NaHCO₃ solution to inhibit the growth of fungi. Subsequently, the plant materials were rinsed three times with sterilized distillated water[7–9] and the leaves of plant were cut into small fragments. Finally, the pieces were transferred to Petri dishes of Bennett agar medium containing 50 µg/mL of cycloheximide and 100 µg/mL of nalidixic acid, and the endophytic actinobacteria were observed after incubation at 28 °C for 1-4 weeks.

2.2. Screening for antimicrobial activity of endophytic actinobacteria

Strain NAF-1 was grown on Bennett agar medium during 14 days. Agar cylinders were cut and placed on Muller-Hinton medium for bacteria and Sabouraud medium for fungi (yeasts and molds), previously inoculated by the test microorganisms. Plates were kept during 4 h at 4 $^{\circ}$ C for a good diffusion of secondary metabolites produced, then incubated at 37 $^{\circ}$ C for bacteria and 30 $^{\circ}$ C for fungi.

Antibacterial activity was evaluated *in vitro* using three Gram positive bacteria, *Staphylococcus aureus* PIC 53156, *Micrococcus luteus* ATCC381 and *Bacillus subtilis* ATCC 14579 and two Gran negative bacteria, *Pseudomonas aeruginosa* DSM 50090, *Escherichia coli* ATCC 8739. While the antifungal activity was achieved against four clinic fungi species: *Candida albicans* CCMM-L4, *Candida tropicalis* DSM11953, *Aspergillus niger* CCMM-M100 and *Microsporum canis* CCMM-M103.

2.3. Cytotoxicity assay of NAF-1 crude extract

Artemia salina brine shrimp was used as a model to study cytotoxicity of crude extract of strain NAF-1. After hatching of 5 mg of brine shrimp eggs in 500 mL of sterilized seawater under constant aeration at 37 °C for 24 to 48 h, the active nauplii were collected and used for the test. Several concentrations were prepared by dilution from 100 µg/mL to 0.753 µg/mL of crude extract obtained by extraction from fermentation culture using ethyl acetate and concentrated under vacuum. A total of 0.5 mL of each dilution were added to 10 to 20 larvae (0.5 mL) of mature nauplii into separate well of plastic 24-well tissue culture plate. After 24 h, the LD₅₀ was calculated on the basis of the mortality rate[10].

2.4. Antioxidant activity of NAF-1 crude extract

2.4.1. Determination of 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging and reducing power activities

To determine the antioxidant activity (IC₅₀) of the crude extract obtained from the culture fermentation of strain NAF-1, we used the stable radical DPPH according to Şahin *et al.*[11]. Concerning the reductive potential, it was determined by the Fe³⁺ to Fe²⁺ transformations in the presence of the crude extract, using the method of Gülçin *et al.*[12]. The BHT and quercetin were used as positive control.

2.4.2. β -carotene/linoleic acid bleaching assay

The crude extract's ability to prevent beaching of β -carotene, by its oxidation in the presence of O₂ molecule, was performed by Miraliakbari & Shahidi[13] method with small modifications. The absorbance values were measured at 470 nm to calculate the inhibitions percentages (I %) of the sample.

2.4.3. Determination of total phenols and flavonoids content

Total phenolic content of the crude extract of our strain which was expressed in µg of gallic acid equivalent/mg extract (µg GAE/ mg extract) was determined by the Folin-Ciocalteu micro-method as described by Ali *et al.*[14]. On the other hand, the total flavonoid content was determined by the method of Zengin *et al.*[15] and expressed in µg of quercetin equivalent/mg extract (µg QE/mg extract).

2.5. Polyphasic taxonomy of endophytic actinobacteria

2.5.1. Morphological and physiological characterization

The cultural features of the strains NAF-1 were characterized following the directions given by the International Streptomyces Project (ISP) media[16,17] and the Bergey's Manual of Systematic Bacteriology^[18]. Our promising isolate was tested for their ability to grow at pH 5 to 9 and at a temperature range from 27 $^{\circ}$ C to 37 $^{\circ}$ C^[19]. The production of melanoide pigments, considered a useful criterion for taxonomic studies, was carried out on ISP6 and ISP7 agar[20].

The study of the physiological criteria such us using different carbon and nitrogen sources has been carried out on the ISP9 base medium to which carbon and nitrogen sources were added to a final concentration of 1%[21]. The plates were kept at 28 $^{\circ}$ C and the growth was evaluated visually.

2.5.2. DNA extraction, polymerase chain reaction (PCR) amplification and sequencing of 16S rRNA

Strain NAF-1 was grown under orbital shaking during 4 days at 28 $^{\circ}$ C in Erlenmeyer flasks containing 50 mL of Bennett medium liquid. Biomass was separated using centrifugation at 8000 rpm for 10 min and washed several times with sterile distillated water. The genomic DNA of the pellet was extracted using Bacterial DNA kit according to the manufacturer instructions.

A PCR using primers 27 Fbac (5'AGAGTTTGATCMTGGCTCAG3') and 1492 Runi (5' TACGGYTACCTTGTTACGACTT 3') was used to amplify the 16S rDNA gene. The conditions used for thermal cycling were as follows: denaturation at 95 $^{\circ}$ C for 5 min, followed by 30 cycles of amplification at 95 $^{\circ}$ C for 1 min, annealing at 54-55 $^{\circ}$ C for 1 min, extension at 72 $^{\circ}$ C for 2 min and extra extension at 72 $^{\circ}$ C for 10 min and then cooling to 4 $^{\circ}$ C[22].

The purified 16S rRNA was sequenced directly using the same primers and was compared for similarity with those contained in genomic database banks, using NCBI BLAST. The phylogenetic tree was constructed using the neighbor-joining method with the Mega 6 software[23].

3. Results

3.1. Antimicrobial activity of the isolated NAF-1 strain

In this study, the antimicrobial activity was screened on agar medium and the results revealed that the maximum zone of inhibition was recorded against *Staphylococcus aureus* (25 mm) and *Micrococcus luteus* (25 mm) followed by *Bacillus subtilis* (20 mm) but no activity against the Gram negative bacteria. Furthermore, our actinobacteria was active against all yeasts and filamentous fungi used in our test. The maximum of inhibition was observed against *Aspergillus niger* (15 mm) followed by *Candida tropicalis* (14 mm), *Candida albicans* (13 mm) and *Microsporum canis* (13 mm).

3.2. Cytotoxicity assay

The crude extract of strain NAF-1 was subjected to Brine shrimp lethality assay to find the cytotoxicity expressed in LD_{50} . The extract studied showed significant lethality against mature nauplii with 14.20 µg/mL as LD₅₀ (Figure 1).

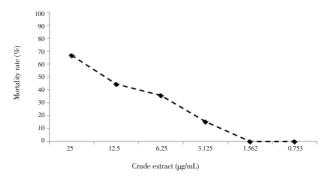


Figure 1. Effect of various concentrations of crude extract (strain NAF-1) against *Artemia salina* shrimp larvae.

3.3. Antioxidant activity

To evaluate the antioxidant activity of NAF-1 crude extract, several tests were performed, and the IC₅₀ of crude extract and those of BHT and quercetin were given in Table 1. The result showed that our crude extract exhibited an interesting free radical-scavenging activity with IC₅₀ value of $(5.58 \pm 0.26) \mu g/mL$ better than those in BHT and quercetin. Concerning β -carotene acid bleaching test, IC₅₀ value of the crude extract was less effective than BHT and more effective than quercetin. Furthermore, the reducing power IC₅₀ values were significantly lower than those obtained for synthetic antioxidant agents. Our crude extract showed a high value of phenolic and flavonoid compounds with (15.41 ± 0.18) μ g GAE/mg extract and (11.41 ± 0.06) μ g QE/mg extract respectively.

Table 1

IC ₅₀ values of cruc	e extract, BHT	and quercetin.
---------------------------------	----------------	----------------

Antioxidant tests	NAF-1 extract	BHT	Quercetin
	(µg/mL)	(µg/mL)	(µg/mL)
DPPH assay	5.58 ± 0.26	4.21 ± 0.08	1.07 ± 0.01
Reducing power assay	6.25 ± 0.19	7.09 ± 0.10	2.29 ± 0.10
β -Carotene/linoleic acid assay	5.15 ± 0.53	4.30 ± 0.33	0.95 ± 0.02

Values represent mean ± standard deviations for triplicate experiments.

3.4. Taxonomic study of Endophytic actinobacteria NAF-1

The promising strain NAF-1 was the subject of a taxonomic study based on the determination of its morphological, biochemical and molecular criteria. The phenotypic appearance and cultural characteristics were determined on ISP culture media. The importance of growth, the development of aerial mycelium on each medium, the color of the aerial and substrate mycelium as well as the presence of diffusible or non-diffusible pigments in the agar were observed and the all results were noted in the Table 2.

Strain NAF-1 showed a good growth and sporulation and abundant mycelium in all media used. The color of the aerial mycelium was white but the coloration of substrate mycelium was variable, it was brownish yellow in ISP1, brown in ISP2, ISP5 and Bennett and gray in ISP3 and ISP4 (Table 2). The diffusible pigments and melanoids were produced on ISP6 and ISP7 respectively.

In the case of the use of carbon sources, the NAF-1 isolate was capable to degrade the glucose, fructose, arabinose, galactose, rhamnose and mannitol but concerning the use of amino acids, the NAF-1 strain was able to use asparagine and arginine. Regarding the pH, we found that the strain NAF-1 was able to develop at different pH values (5, 6, 7, 8 and 9) so it tolerated neutral, slightly acidic and basic pH. In addition, the results showed that our strain grew at temperatures ranging from 27 $^{\circ}$ C to 37 $^{\circ}$ C with an optimum at 30 $^{\circ}$ C so it preferred mesophilic growth temperatures.

According to these morphological, physiological and biochemical studies, it is impossible to determine exactly the species to which our isolate belongs, so it is necessary to make recourse to the molecular identification of 16S RNA.

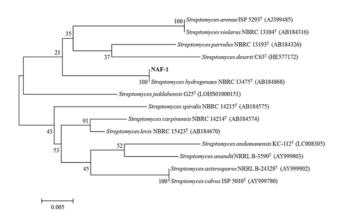
Indeed, the sequencing of the NAF-1 16S RNA yielded a nucleotide sequence of 1 395 bp having 100% of similarity with *Streptomyces hydrogenans* (*S. hydrogenans*) NBRC 13475^T (Figure 2). In addition, the comparison of the profiles using some carbon sources and nitrogen substrates showed that NAF-1 had a great resemblance to *S. hydrogenans* except that this latter is unable to use fructose and mannitol.

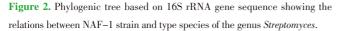
Table 2

Phenotypic characteristics of NAF-1 in ISP media after 2 weeks of incubation at 28 $^\circ\!\!\mathbb{C}.$

Medium	Growth*	Sporulation [*]	Aerial	Substrate
			mycelium	mycelium
Tryptone-yeast extract	+++	+++	White	Brownish
agar (ISP1)				yellow
Yeast extract-malt extract	+++	+++	White	Brown
agar (ISP2)				
Oatmeal agar (ISP3)	+++	+++	White	Gray
Inorganic salt-starch agar	+++	+++	White	Gray
(ISP4)				
Glycerol Asparagine	+++	+++	White	Brown
Agar (ISP5)				
Bennett agar	+++	+++	White	Brown

*: +++ Abundant, ++ moderate and + less.





4. Discussion

Actinobacteria from medicinal plants are the new source of several bioactive substances with high therapeutic potential^[3], and their molecules produced may be associated with the properties of the host medicinal plant^[24]. We investigate the application of endophytic actinobacteria, especially from *A. vera*, in order to evaluate the species richness and biological properties to decrease the problem of antibiotic resistance emergence.

According to the results of the antimicrobial activity, we noticed that our NAF-1 strain showed a high activity against the Gram positive bacteria and was not active against the negative ones. This different sensitivity could be attributed to the high level of lipopolysaccharides that contain the Gram positive bacteria membrane, which could make the cell wall impermeable to bioactive compounds[25].

On the other hand, the isolated actinobacteria showed a very interesting activity against the clinic fungi. These data suggest that NAF-1 strain produces metabolites with a broad spectrum activity which are required to treat multidrug resistant pathogens, such as MRSA, *Pseudomonas aeruginosa*, and *Acinetobacter baumannii* that are considered one of the most urgent issues in modern healthcare[26–28]. This result also confirms that the endophytic actinobacteria are often associated with a high antimicrobial properties[29,30].

The brine shrimp lethality assay was considered one of the most useful tests used to study the toxicity of bioactive substances[31]. In our bioassay, the cytotoxicity was similar to those obtained from actinobacteria by many research works using *Artemia salina* as model[32,33]. Several studies reported that, if the toxicity using brine shrimp larvae displayed LD₅₀ lower than 1 000 µg/mL of natural extract, it implies that it contains physiologically active principles[34]. That idea was confirmed by the highest antioxidant activity and the values of total phenolic and flavonoid calculated. Those activities can be explained by the presence of some class of compounds, for example the fatty acids. Moreover, the benzyl alcohol and esters were previously reported to have an interesting antioxidant activity[35].

To confirm the reliability of the preliminary taxonomic characterization, the bioactive endophytic actinobacteria NAF-1 was subjected to 16S rRNA gene sequence analysis. The sequences BLAST suggest that our isolate is *S. hydrogenans* with 100% of similarity. This result demonstrates that the most abundant isolates of the endophytic community belong to the genus *Streptomyces* which was consistent with various reports on other plant hosts[7,36,37]. We reported for the first time the isolation of strain closely related to the species *S. hydrogenans* as endophyte. *S. hydrogenans* was originally isolated from soil in several works[29,38,39].

Several strains of endophytic actinobacteria show a different range of bioactivities which can be exploited therapeutically. For example, 43.4% of endophytic actinobacteria recovered from leaves and roots of maize showed an antibacterial and antifungal activity against at least one of bacteria and yeast tests[40]. Similarly, another study revealed that 55% of endophytic actinobacteria isolated from leaves of *Paeonia lactiflora* and *Trifalium repens* inhibited the growth of *Rhizoctonia solani* a significant pathogen of plants[41]. *Streptomyces longisporoflavus* an endophytic actinobacteria was isolated from *Lysimachia ciliate* in India which has anti-diabetic activity[42].

The study conducted by Zhang *et al.*[43], allows to obtain 12 actinobacteria stains which are able to inhibit the growth of penicillin resistant *Staphylococcus aureus*, the majority of them were belonging to the genus *Streptomyces* and they isolated from *Artemisia argyi*, *Radix platycodi*, *Achyranthes bidentata* and *Paeonia lactiflora*. Searching for novel bioactive substances with interesting biological activity in a diverse environment has attracted more attention in recent decade due to the increased incidence of multiple resistances to the used drugs[44].

A. vera provides a new source for potential endophytic actinobacteria with promising and new biologically secondary metabolites. The study shows that the isolation from a unique niche (medicinal plants) to explore endophytic actinobacteria will facilitate the discovery of new antimicrobial agents with significant biological activity.

Conflict of interest statement

The authors declared no conflict of interest.

Acknowledgements

The corresponding author thanks a lot all the co-authors for help.

References

- [1] Golinska P, Wypij M, Agarkar G, Rathod D, Dahm H, Rai M. Endophytic actinobacteria of medicinal plants: Diversity and bioactivity. *Antonie Van Leeuwenhoek* 2015; **108**(2): 267-289.
- [2] Kumari Kadiri S, Yarla NS, Vidavalur S. Screening and isolation of antagonistic actinobacteria associated with marine sponges from Indian coast. J Microb Biochem Technol 2014; s8(01): 1-4.
- [3] Savi DC, Shaaban KA, Vargas N, Ponomareva LV, Possiede YM, Thorson JS, et al. *Microbispora* sp. LGMB259 endophytic actinomycete isolated from *Vochysia divergens* (Pantanal, Brazil) producing β-carbolines and indoles with biological activity. *Curr Microbiol* 2015; **70**(3): 345-354.
- [4] Gos FMWR, Savi DC, Shaaban KA, Thorson JS, Aluizio R, Possiede YM, et al. Antibacterial activity of endophytic actinomycetes isolated from the medicinal plant *Vochysia divergens* (Pantanal, Brazil). *Front Microbiol* 2017; 8: 1642.
- [5] Olano C, Méndez C, Salas JA. Antitumor compounds from actinomycetes: From gene clusters to new derivatives by combinatorial biosynthesis. *Nat Prod Rep* 2009; 26(5): 628.

- [6] Bérdy J. Thoughts and facts about antibiotics: Where we are now and where we are heading. J Antibiot (Tokyo) 2012; 65(8): 385-395.
- [7] Qin S, Li J, Chen HH, Zhao GZ, Zhu WY, Jiang CL, et al. Isolation, diversity, and antimicrobial activity of rare actinobacteria from medicinal plants of tropical rain forests in Xishuangbanna, China. *Appl Environ Microbiol* 2009; **75**(19): 6176-6186.
- [8] Sunaryanto R, Mahsunah AH. Isolation, purification, and characterization of antimicrobial substances from endophytic actinomycetes. *Makara J Sci* 2013; **173**(3). Doi:10.7454/mss.v17i3.
- [9] Prashith Kekuda TR. Isolation, characterization and antimicrobial potential of endophytic actinomycetes. *Int J Curr Microbiol Appl Sci* 2016; 5(57): 100-116.
- [10]Samri SE, Mohamed B, Abdelmounaim J, Houda A, Said EM, Abdellatif EM, et al. Preliminary assessment of insecticidal activity of Moroccan actinobacteria isolates against mediterranean fruit fly (*Ceratitis capitata*). *African J Biotechnol* 2015; 14(10): 859-866.
- [11]Şahin F, Güllüce M, Daferera D, Sökmen A, Sökmen M, Polissiou M, et al. Biological activities of the essential oils and methanol extract of *Origanum vulgare* ssp. vulgare in the Eastern Anatolia region of Turkey. *Food Control* 2004; **15**(7): 549-557.
- [12]Gülçin İ, Huyut Z, Elmastaş M, Aboul-Enein HY. Radical scavenging and antioxidant activity of tannic acid. Arab J Chem 2010; 3(1): 43-53.
- [13]Miraliakbari H, Shahidi F. Antioxidant activity of minor components of tree nut oils. *Food Chem* 2008; **111**(2): 421-427.
- [14]Ali BEHI, Bahri R, Chaouachi M, Boussaïd M, Harzallah-Skhiri F. Phenolic content, antioxidant and allelopathic activities of various extracts of *Thymus numidicus* Poir. organs. *Ind Crops Prod* 2014; 62: 188-195.
- [15]Zengin G, Sarikurkcu C, Aktumsek A, Ceylan R. Sideritis galatica Bornm.: A source of multifunctional agents for the management of oxidative damage, Alzheimer's's and diabetes mellitus. J Funct Foods 2014; 11(2): 538-547.
- [16]Shirling EB, Gottlieb D. Methods for characterization of *Streptomyces* species. *Int J Syst Bacteriol* 1966; 16(3): 313-340.
- [17]Mohamed H, Miloud B, Zohra F, García-Arenzana JM, Veloso A, Rodríguez-Couto S. Isolation and characterization of actinobacteria from Algerian Sahara soils with antimicrobial activities. *Int J Mol Cell Med* 2017; 6(2): 109-120.
- [18]Bergey DH, Whitman WB, Goodfellow M, Kämpfer P, Busse HJ. Bergey's manual of systematic bacteriology: Volume 5: The actinobacteria. New York: Springer; 2012.
- [19]Boudemagh A. Isolation, from Saharan soils, of actinomycetal bacteria producing antifungal molecules, molecular identification of active strains. PhD Thesis. University of Mentouri Constantine Faculty of Science Algeria; 2007.
- [20]Boudemagh A, Kitouni M, Boughachiche F, Hamdiken H, Oulmi L, Reghioua S, et al. Isolation and molecular identification of actinomycete microflora, of some saharian soils of south east Algeria (Biskra, EL-Oued and Ourgla) study of antifungal activity of isolated strains. *J Mycol Med* 2005; **15**(1): 39-44.
- [21]Ayari A, Morakchi H, Djamila KG. Identification and antifungal activity

of *Streptomyces* sp. S72 isolated from Lake Oubeira sediments in North-East of Algeria. *African J Biotechnol* 2012; **11**(2): 305-311.

- [22]Li WJ, Xu P, Schumann P, Zhang YQ, Pukall R, Xu LH, et al. Georgenia ruanii sp. nov., a novel actinobacterium isolated from forest soil in Yunnan (China), and emended description of the genus Georgenia. Int J Syst Evol Microbiol 2007; 57(7): 1424-1428.
- [23]Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. MEGA5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Mol Biol Evol* 2011; 28(10): 2731-2739.
- [24]Santos IP dos, Silva LCN da, Silva MV da, Araújo JM de, Cavalcanti M da S, Lima VL de M. Antibacterial activity of endophytic fungi from leaves of *Indigofera suffruticosa* Miller (Fabaceae). *Front Microbiol* 2015; 6: 350.
- [25]Gebreyohannes G, Moges F, Sahile S, Raja N. Isolation and characterization of potential antibiotic producing actinomycetes from water and sediments of Lake Tana, Ethiopia. *Asian Pac J Trop Biomed* 2013; 3(6): 426-435.
- [26]Smith PA, Roberts TC, Romesberg FE. Broad-spectrum antibiotic activity of the arylomycin natural products is masked by natural target mutations. *Chem Biol* 2010; **17**(11): 1223-1231.
- [27]Çıkman A, Parlak M, Bayram Y, Güdücüoğlu H, Berktaş M. Antibiotics resistance of *Stenotrophomonas maltophilia* strains isolated from various clinical specimens. *Afr Health Sci* 2016; **16**(1): 149.
- [28]Paulus C, Rebets Y, Tokovenko B, Nadmid S, Terekhova LP, Myronovskyi M, et al. New natural products identified by combined genomics-metabolomics profiling of marine *Streptomyces* sp. MP131-18. *Sci Rep* 2017; 7: 42382.
- [29]Manhas RK, Kaur T. Endophytic actinobacteria associated with *Dracaena cochinchinensis* Lour.: Isolation, diversity, and their cytotoxic activities. *Biomed Res Int* 2017; **2017**: 11.
- [30]Salam N, Khieu TN, Liu MJ, Vu TT, Chu-Ky S, Quach NT, et al. Endophytic actinobacteria associated with *Dracaena cochinchinensis* Lour.: Isolation, diversity, and their cytotoxic activities. *Biomed Res Int* 2017; 2017: 1308563.
- [31]Sudhakesavan S, Vijayalakshmi S, Nandhini SU, Latha MB, Selvam MM. Application of brine shrimp bioassay for screening cytotoxic actinomycetes. *Int J Pharm Pharm Sci Res* 2011; 1(3): 104-107.
- [32]Prashith Kekuda TR, Shobha KS, Onkarappa R, Goutham SA, Raghavendra HL. Screening biological activities of a *Streptomyces* species isolated from soil of Agumbe, Karnataka, India. *Int J Drug Develop Res*

2012; **4**(3): 104-114.

- [33]Tanvir R, Sajid I, Hasnain S. Biotechnological potential of endophytic actinomycetes associated with Asteraceae plants: Isolation, biodiversity and bioactivities. *Biotechnol Lett* 2014; 36(4): 767-773.
- [34]Sudha S, Selvam M. In vitro cytotoxic activity of bioactive metabolite and crude extract from a new Streptomyces sp. SU. J Pure Appl Microbiol 2013; 7(3): 2331-2336.
- [35]Keawsa-Ard S, Kongtaweelert S. Antioxidant, antibacterial, anticancer activities and chemical constituents of the essential oil from *Mesua ferrea* leaves. *Chiang Mai J Sci* 2012; **39**(3): 455-463.
- [36]Li J, Zhao GZ, Huang HY, Qin S, Zhu WY, Zhao LX, et al. Isolation and characterization of culturable endophytic actinobacteria associated with *Artemisia annua L. Antonie Van Leeuwenhoek* 2012; **101**(3): 515-527.
- [37]Shutsrirung A, Chromkaew Y, Pathom-Aree W, Choonluchanon S, Boonkerd N. Diversity of endophytic actinomycetes in mandarin grown in northern Thailand, their phytohormone production potential and plant growth promoting activity. *Soil Sci Plant Nutr* 2013; **59**(3): 322-330.
- [38]Kaur T, Manhas RK. Antifungal, insecticidal, and plant growth promoting potential of *Streptomyces hydrogenans* DH16. *J Basic Microbiol* 2014; **54**(11): 1175-1185.
- [39]Kulkarni M, Gorthi S, Banerjee G, Chattopadhyay P. Production, characterization and optimization of actinomycin D from *Streptomyces hydrogenans* IB310, a(n antagonistic bacterium against phytopathogens. *Biocatal Agric Biotechnol* 2017; **10**: 69-74.
- [40]Araújo JM de, Silva AC da, Azevedo JL. Isolation of endophytic actinomycetes from roots and leaves of maize (*Zea mays L.*). *Brazilian Arch Biol Technol* 2000; 43(4): 447-451.
- [41]Gu Q, Liu N, Qiu DH, Liu ZH, Huang Y. Isolation, classification and antimicrobial activity of endophytic actinomycetes from plant leaves. *Wei Sheng Wu Xue Bao* 2006; **46**(5): 778-782.
- [42]Akshatha VJ, Nalini MS, D'Souza C, Prakash HS. Streptomycete endophytes from anti-diabetic medicinal plants of the Western Ghats inhibit alpha-amylase and promote glucose uptake. Lett Appl Microbiol 2014; 58(5): 433-439.
- [43]Zhang X, Ren K, Zhang L. Screening and preliminary identification of medicinal plants endophytic actinomycetes used for inhibiting penicillinresistant *Staphylococcus aureus*. *Int J Biol* 2012; **4**(2):119.
- [44]Rao HCY, Rakshith D, Satish S. Antimicrobial properties of endophytic actinomycetes isolated from *Combretum latifolium* Blume, a medicinal shrub from Western Ghats of India. *Front Biol (Beijing)* 2015; 10(6): 528-536.