

# Platelet Concentrates

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## Introduction

**P**latelets play an important role in hemostasis and wound healing. Platelet derived growth factors are well known source of healing cytokines. Various techniques of autologous platelet concentrates have been developed and applied in oral and maxillofacial surgery. Platelets are non-nucleated cytoplasmic fragments derived from bone marrow megakaryocytes and measure 2–3  $\mu$ m in diameter. They contain many granules, lesser mitochondria and two prominent membrane structures, one the surface connected canalicular system and the other is dense tubular system.

$\alpha$ -granules are intracellular storage pool of proteins important for wound healing which including platelet-derived growth factor (PDGF), transforming growth factor (TGF- $\beta$ ), and insulin-like growth factor (IGF-I). After activation the  $\alpha$  granules fuse with the platelet cell membrane. Few secretory proteins are transformed to a bioactive state. The active proteins are then secreted, allowing them to bind to transmembrane receptors of the target cells.

Bound, intracellular signal proteins are activated resulting in the expression of a gene sequence that directs cellular proliferation, collagen synthesis, osteoid production etc. Platelet concentrates were originally used in transfusion medicine, for the treatment and prevention of haemorrhage due to severe thrombopenia, which is often caused by medullary aplasia, acute leukaemia or significant blood loss during long-lasting surgery. The standard platelet concentrate for transfusion has been named platelet-rich plasma (PRP) and classically contains  $0.5 \times 10^{11}$  platelets per unit.

The use of blood-derived products to seal wounds and stimulate healing started with the use of fibrin glues, which were first described in 1970's and are constituted of concentrated fibrinogen (polymerization induced by thrombin and calcium). They are used for topical hemostasis and tissue sealing

and as melting agents for particulate bone substitutes. Risk of cross infection for commercial adhesives (Tisseel, Baxter healthcare) led to the development of autologous fibrin sealants from the patient's own plasma. However, their fabrication resulted in less reproducible or less satisfactory rheologic properties. Autologous fibrin glues were considered the best choice to avoid contamination risk, but their use remains very limited owing to the complexity and the cost of their production protocols.

Consequently, the use of platelet concentrates to improve healing and to replace fibrin glues, as first described by Whitman et al in 1997.

## Aim:

Aim of this paper/poster is to review the history and various types of platelet concentrates and their uses in dentistry.

## Classification of Platelet Concentrates

Platelet concentrates now can be broadly classified as first generation platelet concentrates and second generation platelet concentrates.

## First Generation Platelet Concentrate

**Leucocyte-poor or pure platelet-rich plasma (P-PRP):** Pure platelet concentrates for topical use were first developed as an additional application of the classical transfusion platelet units and were first reported for maxillofacial surgery by Whitman et al in 1997.

two type of methods were employed for its production

- Automated protocols for P-PRP: plasmapheresis with a laboratory cell separator and Vivostat PRF: either in a discontinuous flow set up (in which the patient stays connected to the machine and the blood filtering continues until the desired quantity of platelets has been collected) or starting from a bag of harvested blood with anticoagulant.
- The Vivostat PRF centrifuge (Vivolution, Denmark can be considered as an advanced cell separator, and it was originally designed to produce the Vivostat Fibrin sealant. The use of a

specific preparation kit with this centrifuge allows the production of a leucocyte-poor platelet concentrate for surgical use. However, Vivostat PRF has been used in only a few published studies, and this system is cumbersome and very expensive for daily practice. Moreover, its platelet collection efficiency is rather low and platelets are damaged during the process.

- Manual protocols for modified P-PRP: Anitua's PRGF: One of the first platelet concentrate protocols (PRGF, which stands for either plasma rich in growth factors or preparation rich in growth factors) was described in 1999 by Anitua and has been commercialized by BTI (BioTechnology Institute, Vitoria, Spain). In this protocol, venous blood is collected and centrifuged in several small tubes to obtain the three typical layers: RBCs, 'buffy coat' and acellular plasma. The upper part of the acellular plasma is called plasma poor in growth factors (PPGF) and is discarded from each tube by careful pipetting to avoid creating turbulences. The remaining plasma is termed PRGF and is collected with a pipette, using only 'eyeballing' as a measuring tool.

**Leucocyte- and platelet-rich plasma (L-PRP):** The initial objective of developing alternative easy-to-handle methods was to make it possible to use platelet concentrates in daily practice without having the support of a transfusion laboratory. Without a cell separator, elimination of leucocytes becomes more difficult, and the resulting platelet concentrates therefore contain a high quantity of leucocytes, which were not initially desired.

- Automated protocols for L-PRP: SmartPREP, PCCS, GPS and Magellan: Automated systems for L-PRP have been developed in the form of PCCS (Platelet Concentrate Collection System) by 3I (Palm Beach Gardens, USA) in 2006 and SmartPREP by Harvest Corp (Plymouth, USA). The centrifuges used have been

designed to take a customized collection and centrifugation device, which consists of two connected compartments. In the PCCS method, citrated whole blood is transferred into the first compartment and centrifuged briefly to obtain the three layers (RBC, buffy coat, PPP). Then, by opening of a tubule and using air pressure, the superficial layers (i.e. PPP and buffy coat) are transferred to the second chamber and centrifuged again but for a longer period. Finally, using the same air pressure system, most of the PPP layer is transferred back into the first compartment and thus discarded.

- Manual protocols for L-PRP: Curasan, Friadent-Schüttze, Regen and Plateltex: Two similar protocols, each using a two-step centrifugation procedure, were marketed by Curasan (Kleinostheim, Germany) and Friadent-Schüttze (Vienna, Austria) respectively. Each method follows the first principal step described above and shown in Figure, in which a first centrifugation step separates the blood components into three layers of RBCs, 'buffy coat' and PPP. The PPP and buffy coat layers are then carefully collected, avoiding RBC contamination, and transferred to another tube, where they are subjected to a second centrifugation step at high speed, which separates the sample again into its components. After the second centrifugation step, most of the PPP layer is discarded using the 'eyeballing' method. The PRP concentrate obtained with this method is composed of a high quantity of platelets, leucocytes and circulating fibrinogen, but it also contains residual RBCs. The concentrate is applied with bovine thrombin and calcium chloride.

**Leucocyte-poor or pure platelet-rich fibrin (P-PRF) concentrates:** In this category, there is only one method available. The Fibrinet PRFM kit by Cascade Medical (New Jersey, USA) contains two tubes, one for blood collection and another for PRFM clotting, together with a transfer device. A small amount of blood (typically 9 mL) is drawn into a collection tube, which contains tri-sodium citrate as an anticoagulant and a proprietary separator gel, and centrifuged for six minutes at high speed. The three typical layers of RBCs, buffy coat and PPP are obtained. Buffy coat and PPP are easily transferred to a second tube containing  $\text{CaCl}_2$  with the help of a specifically designed tube connection system. The clotting process is triggered by the presence of  $\text{CaCl}_2$  and the tube is immediately centrifuged for 15 min, after which a stable PRFM clot can be collected. The company claims that the system produces a 'natural' platelet concentrate owing to the absence of bovine thrombin. However, this claim is doubtful because the blood is mixed with anticoagulant and separation gel, leading to what could be considered unnatural

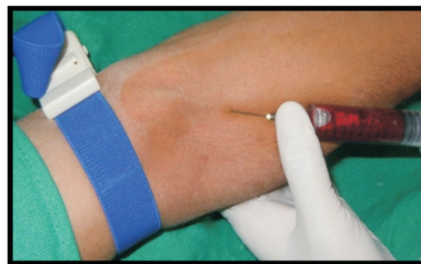
conditions.

Advantages of Platelet Gel and PRP Over Fibrin Sealants Safe autogenous preparation, free from concerns over transmissible diseases such as HIV, hepatitis, West Nile fever and Creutzfeld-Jacob disease (mad cow disease).

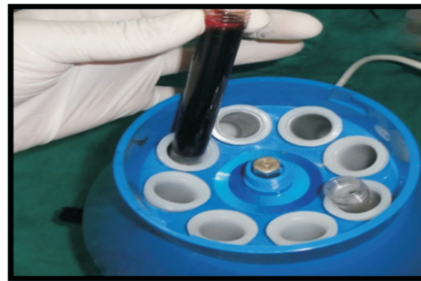
Convenient for patient since blood is collected in the immediate preoperative period.

More patients are eligible for this procedure because the criteria of blood bank donation do not have to be met, this would include children up to age 6, weights up to 25 kg, the elderly, those whose medical condition would preclude the blood bank from drawing a unit of whole blood.

Presence of platelets brings cytokines and growth factors to the site of surgery in a manner that would not occur with fibrin glue.



Withdrawal of blood



Centrifugation of blood

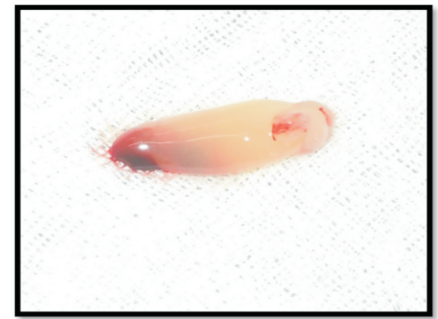


Centrifuged blood clot

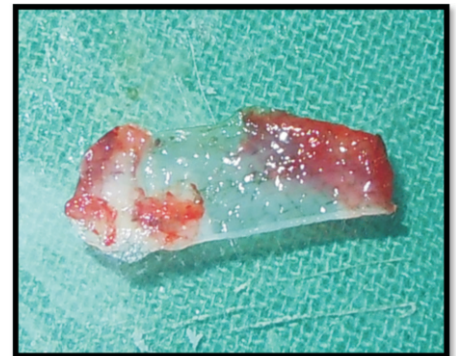


PRF clot

**Second generation Platelet concentrate: Leucocyte- and platelet-rich fibrin (L-PRF) concentrates:** Choukroun's PRF: Choukroun's PRF protocol is a simple and free technique developed in France by Choukroun et al.. It can be considered as a second-generation platelet



PRF clot is compressed between two glass slabs



PRF membrane

Figure : Preparation of Choukroun's PRF

concentrate because the natural concentrate is produced without any anticoagulants or gelling agents. Venous blood is collected in dry glass tubes and centrifuged at low speed (Process protocol, Nice, France). In the absence of anticoagulants, platelet activation and fibrin polymerization are triggered immediately. Therefore, after centrifugation, three layers are formed: the RBC base layer, acellular plasma top layer and a PRF clot in the middle (Figure 3). The PRF clot forms a strong fibrin matrix with a complex three-dimensional architecture, in which most of the platelets and leucocytes from the harvested blood are concentrated. When pressed between two gauzes, the PRF clot becomes a strong membrane, and some applications of this autologous biomaterial have been described in oral, maxillofacial, ENT (ear, nose, throat) and plastic surgery.

## Properties of PRF

- The biochemical analysis of the PRF composition indicates that this biomaterial consists of an intimate assembly of cytokines, glycanic chains, structural glycoproteins enmeshed within a slowly polymerized fibrin network. These biochemical components have well known synergistic effects on healing processes.
- PRF is not only a platelet concentrate but also an immune node able to stimulate defense mechanisms. It is likely that the significant inflammatory regulation noted on surgical sites treated with PRF is the outcome of retro control effects from cytokines trapped in the fibrin network and released during the remodeling of this

initial matrix.

(3) Role of fibrin matrix of PRF:

Fibrin is the natural guide of angiogenesis. Fibrin constitutes a natural support to immunity.

Fibrin and wound coverage: Fibrin matrix guides the coverage of injured tissues, affecting the metabolism of epithelial cells and fibroblasts

**Clinical Implications of PRF**

- In sinus lift procedures.
- Socket preservation.
- PRF membrane has been used for gingival recession coverage with coronally advanced or lateral pedicle flap for multiple and single recession respectively. PRF acts both as healing and interpositional biomaterial.
- Filling of cystic cavity.
- In the treatment of combined periodontic endodontic lesion/furcation defect.

**Advantages of PRF over PRP**

No need of addition of bovine thrombin or other anticoagulants so it is completely safe. Standard production protocol.

**Limitations of PRF Technology**

- (1) Only a limited volume of PRF can be used. Because it is obtained from an autologous blood sample, the quantities produced are low. This fact limits the systematic utilization of PRF for general surgery.
- (2) PRF tissue banks are unfeasible. The fibrin matrix contains all the circulating immune cells and all the highly antigenic plasmatic molecules. That is why PRF membranes are totally specific to the donor and cannot constitute an allogenic graft tissue

**Conclusion**

Platelet concentrates are rich in growth factors. The role of growth factors in regeneration is undisputable, several in vitro and in vivo studies have provided evidence that the application of growth factors may increase tissue regeneration. Platelet rich and platelet-poor plasma have demonstrated results in the soft tissue regeneration, they might promote myofibroblastic differentiation, cell migration and extracellular matrix remodeling. but there are concerns whether these growth factors are

available at the site for enough period of time? It is still uncertain which concentrations of platelet rich plasma are optimal for promoting wound healing also it was observed that the growth factors released by platelet concentrates gets washed away by GCF flow leaving very less or no available growth factors for regeneration. Hence further studies are required to assess the role of platelet concentrates in periodontal regeneration.

**Difference Between first & second Generation Platelet Concentrates.<sup>1</sup>**

First Generation-cPRP	Second Generation-PRP
Use of Bovine Thrombin & Calcium Chloride (Anticoagulants)	No Anticoagulants Used
Sudden Fibrin polymerization depending on the amount of surgical additives (Thrombin & Calcium Chloride)	Slow Natural polymerization on contact with glass particles of the test tube results in physiologic thrombin concentration
3-D organization of a fibrin network-condensed tetra Molecular or bilateral Junctions constituted with strong thrombin concentrations. Allows the thickening of fibrin polymers; this leads to a rigid network, not very favorable to cytokine enmeshment & cellular Migration.	3-D network-connected trimolecular or equilateral junctions-allows the establishment of a fine & flexible fibrin network able to support cytokines enmeshment & cellular migration.
The 3-D Structure provides great resistance of such a gel, appropriate to firmly seal biologic tissues.	The 3-D Structure gives elasticity & flexibility to the PRF membrane.