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ABSTRACT

Cytochrome P450 monooxygenases (CYPs) represent a large group and diverse family of enzymes involved in the myriad of biological processes in humans. In the present study, a total of 57 protein sequences of human CYPs retrieved from UniprotKB have been characterized for various physiochemical properties, homology search, motif and super family search and phylogenetic relationship. Physicochemical analysis showed that the isoelectric point values and GRAVY index ranged from 5.84 to 9.47 and 0.018 to -0.367, respectively. Many proteins (50 members, 87.8%) were in basic form, while few (7 members, 12.2%) were of acidic nature. Moreover, GRAVY index revealed that only CYP26C1 as hydrophobic while all others as hydrophilic. Phylogenetic analysis revealed that P450 proteins basically fall into two main clades and are divided into five subgroups. Motif analysis with MEME indicated presence, absence and even shuffling of motifs within clades. Clustering using the maximum likelihood analysis was also in accordance with P450s central roles in drug and xenobiotic metabolism as well as steroid hormone synthesis, fat-soluble vitamin metabolism, and the conversion of polyunsaturated fatty acids to biologically active molecules. Motif conservation within clusters showed the evolutionary pressure for maintenance of the structural and functional organization between different groups of protein. These results will help in the context of understanding the characteristics of the cytochrome P450 monooxygenase isoforms.

Key words: Cytochrome P450; in silico; maximum likelihood; motif analysis; phylogenetic analysis.

INTRODUCTION

Cytochrome P450 (CYP450) is a large family of heme-thiolate proteins that plays key role in xenobiotic metabolism, metabolizing most of the drugs and chemicals of toxicological importance. Besides that they also play major roles in diverse physiological processes including steroid and cholesterol biosynthesis, fatty acid metabolism (prostacyclin, thromboxane) and the maintenance of calcium homoeostasis.¹ These highly conserved genes are found distributed in different life forms, including prokaryotes (archaea, bacteria), unicellular eukaryotes (protists, fungi) and multicellular eukaryotes (plants and animals).² The human genome en-

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codes 57 P450 genes³ grouped into 18 mammalian families and 44 subfamilies. They are basically membrane-associated proteins⁴ located on the smooth endoplasmic reticulum of cells ubiquitously in the body, but predominantly expressed in the liver.⁵ Any imbalance in enzyme availability or its malfunction, e.g., due to a genetic mutation, may lead to a disease state in humans.^{6,7} In humans, CYP450 are known for their central role in phase I drug metabolism where they are of critical importance to two of the most significant problems in clinical pharmacology: drug interactions and inter-individual variability in drug metabolism.⁸ Besides detoxification, they also synthesize biologically active compounds such as steroids, prostaglandins, and arachidonate metabolites. CYP proteins have been identified in life forms like animals, plants, fungi, protists, bacteria, archaea, and even in viruses,⁹ but not in *Escherichia coli*.⁸

P450s are usually divided into gene families. Besides, they are also divided into four different classes depending on the redox partner required. They have a varied primary, secondary, and tertiary structures. Each P450, has a preferred set of substrates, must have a unique method of substrate recognition and an active site they all appear to have a similar structural core. Thus, the charge distribution and topography of the 'redox -partner binding region' of each class of P450 vary to accommodate the various types of redox partners there by providing a great variety of phylogenetically distributed isoform activities. Though much studied, still more remains poorly understood.

In the present study, we performed an *in silico* analysis on human P450 protein sequences by analyzing their biochemical features, homology, motif patterns, cluster and superfamily distribution to understand their functional evolution.

METHOD

The amino acid sequences of the human P450 in FASTA format were downloaded from the UniProt Knowledgebase (UniProtKB) (Table 1). Physicochemical analysis were per-

formed by methods explained by Brindha et al.¹⁰ Data on physiochemical properties were generated using tools like ProtParam, Protein calculator, Compute pI/Mw, ProtScale from Expert Protein Analysis System (EXPASY) proteomic server from the protein sequences.¹¹ The molecular weights (kilo dalton) were calculated by the addition of average isotopic masses of amino acid in the protein and deducting the average isotopic mass of one water molecule. The pI was calculated using pK values of amino acid according to Bjellqvist et al.¹² The atomic composition, extinction coefficients and aliphatic index was derived using the ProtParam tool, available at ExPASy. The Instability Index which predicts regional instability by calculating the weighted sum of dipeptides that occur more frequently in unstable proteins when compared to stable proteins was calculated using the approach of Guruprasad et al.¹³ The Grand average hydropathy (GRAVY) was calculated by adding the hydropathy value for each residue and dividing by the length of the sequence,¹⁴ respectively. Aliphatic index was calculated using the formula x* $(ALA) + a^{*}x (VAL) + b^{*}x (LEU) + b^{*}x (ILE)$ where a = 2.9 and b = 3.9 are constants.¹⁵ Mega 6.01¹⁶ was used for phylogenetic analysis by maximum likelihood method (ML). An optimal model of evolution for the aligned dataset was determined. Sequence divergences were then calculated using the best model which was identified by Mega 6.01. This model choice was guided by the need to avoid under parameterisation.¹⁷ Domain analysis was performed using Pfam 27.0.¹⁸ Motif analysis was done using ME-ME¹⁹ using the expectation maximization approach.

RESULTS AND DISCUSSION

Physicochemical characterization of P450 proteins

Humans have 57 CYP450 genes,³ which are subdivided to 18 families and 44 subfamilies. There are more than 9000 known CYP450 sequences.²⁰ Owing to its importance, many ani-

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Clade Gene/ Accession no. IWW	Gene/ Accession no. Mw	Accession no. MW	ΜM		d	EXTINCTIO	Instabili tv indev	Aliphati	GKAVY	Domain predicted by	Functio
amino acids	amino acids					efficient				predicted by	
Sub clade CYP3A4/503 P08684 57343.1 8.2	9 CYP3A4/503 P08684 57343.1 8.2	P08684 57343.1 8.2	57343.1 8.2	8.2	2	46215	41.19	95.47	-0.04	442(C).	
¹ СҮРЗА5/502 Р20815 57108.6 8.	CYP3A5/502 P20815 57108.6 8.	P20815 57108.6 8.	57108.6 8.	ω.	86	53080	32.31	96.08	-0.116	441(C).	-
CYP3A7/503 P24462 57525.5 5	CYP3A7/503 P24462 57525.5 9	P24462 57525.5 9	57525.5 9	0,	9.21	53205	29.92	97.59	-0.07	442(K).	Drug/xenobiotic metabolisr
CYP3A43/503 Q9HB55 57669.7	СҮРЗА43/503 Q9HB55 57669.7	Q9HB55 57669.7	57669.7		8.27	50225	33.92	98.03	-0.036	442(C).	
CYP4A11/519 Q02928 59347.8	CYP4A11/519 Q02928 59347.8	Q02928 59347.8	59347.8		8.96	85870	43.76	98.46	-0.186	457(C), 321(E).	
СҮР4А22/519 Q5TCH4 59245.8	CYP4A22/519 Q5TCH4 59245.8	Q5TCH4 59245.8	59245.8		9.21	74870	42.4	96.2	-0.208	457(C), 321(E).	
CYP4B1/511 P13584 58991.1	CYP4B1/511 P13584 58991.1	P13584 58991.1	58991.1		8.47	86330	44.19	85.11	-0.204	453(C), 315(E).	
CYP4F2/520 P78329 59853.4	CYP4F2/520 P78329 59853.4	P78329 59853.4	59853.4		6.6	99515	57.86	94.71	-0.118	468(C), 328(E).	
CYP4F3/520 Q08477 59846.7	CYP4F3/520 Q08477 59846.7	Q08477 59846.7	59846.7		7.59	102745	54.23	95.29	-0.074	468(C), 328(E).	
CYP4F8/520 P98187 59994.6	СҮР4F8/520 Р98187 59994.6	P98187 59994.6	59994.6		8.73	88640	52.23	94.88	-0.16	468(C).	Arachadonic acid and fatty
CYP4F11/524 Q9HBI6 60145.7	CYP4F11/524 Q9HBI6 60145.7	Q9HBI6 60145.7	60145.7		6.26	95630	55.43	94.18	-0.134	468(C), 328(E).	acid metabolism
CYP4F12/524 Q9HCS2 60269.8	CYP4F12/524 Q9HCS2 60269.8	Q9HCS2 60269.8	60269.8		7.02	92650	52.518	94.39	-0.126	468(C).	
CYP4F22/531 Q6NT55 61958.2 8	CYP4F22/531 Q6NT55 61958.2 8	Q6NT55 61958.2 8	61958.2 8	ω	3.95	83935	52.52	96.97	-0.201	475(C), 335(E).	
CYP4V2/525 Q6ZWL3 60724 7	CYP4V2/525 Q6ZWL3 60724 7	Q6ZWL3 60724	60724		7.19	102830	49.69	90.11	-0.284	467(C), 329(E).	
CYP4X1/509 Q8N118 58875.2	CYP4X1/509 Q8N118 58875.2	Q8N118 58875.2	58875.2		8.74	84060	43.32	88.55	-0.21	454(C).	
CYP4Z1/505 Q86W10 59085.9	CYP4Z1/505 Q86W10 59085.9	Q86W10 59085.9	59085.9		9.29	85995	52.4	90.16	-0.196	452(C).	
CYP5A1/533 P24557 60518.3	CYP5A1/533 P24557 60518.3	P24557 60518.3	60518.3		7.56	47620	44.89	90.75	-0.059	479(C).	Thromboxane A2 synthesis
Sub clade CYP11A1/521 P05108 60102	9 CYP11A1/521 P05108 60102	P05108 60102	60102		8.89	87445	34.54	89.06	-0.245	462(C).	
² CYP11B1/503 P15538 57572.9	CYP11B1/503 P15538 57572.9	P15538 57572.9	57572.9		9.4	69245	47.96	96.96	-0.1	450(C).	Key steps in steroid
CYP11B2/521 P19099 57560.1 9	CYP11B2/521 P19099 57560.1 9	P19099 57560.1	57560.1	0,	9.47	67755	42.31	99.11	-0.073	450(C).	sisalijilysolu
CYP24A1/514 Q07973 58875.4 E	CYP24A1/514 Q07973 58875.4 E	Q07973 58875.4 8	58875.4 8	ω	.94	80955	49.31	88.04	-0.367	428(C).	Vitamin D metaholism/inactivation
CYP27A1/531 Q02318 60234.7	CYP27A1/531 Q02318 60234.7	Q02318 60234.7	60234.7		9.05	78185	49.52	90.23	-0.185	476(C).	
CYP27B1/508 O15528 56504	CYP27B1/508 015528 56504	015528 56504	56504		9.34	66390	56.21	89.31	-0.179	455(C).	Bile acid biosynthesis, vitamin D activation

Sub clade	CYP26A1/497	043174	56198.6	8.96	51755	50.27	95.15	-0.081	442(C).	
Ω	CYP26B1/512	Q9NR63	57512.6	8.68	49765	35.09	98.44	-0.079	441(C).	Retinoic acid
	CYP26C1/522	Q6V0L0	57111	9.24	63535	47.86	97.43	0.018	459(C).	
	CYP46A1/500	Q9Y6A2	56821	9.15	51255	52.18	87.92	-0.212	437(C).	Cholesterol metabolism
	CYP51A1/503	Q16850	56803.7	8.72	64665	42.99	92.5	-0.151	449(C).	Cholesterol biosynthesis
Sub clade	CYP1A1/512	P04798	58167.3	8.61	63870	40.85	92.29	-0.084	224(F).	
4	CYP1A2/515	P05177	58294.3	9.18	64775	41.43	89.73	-0.161	226(F).	
	CYP1B1/543	Q16678	60845	9.18	52410	37.69	87.83	-0.134	470(C).	
	CYP2A6/494	P11509	56510.3	9.25	26025	40.66	80.85	-0.227	439(C), 107(F), 297(N).	
	CYP2A7/494	P20853	56425	7.69	24660	45.67	81.66	-0.18	439(R).	
	CYP2A13/494	Q16696	56687.5	9.31	27390	38.14	77.73	-0.275	297(N).	
	CYP2B6/491	P20813	56278.3	8.43	29590	37.42	93.14	-0.099	436(L).	
	CYP2C8/490	P10632	55824.7	8.8	33765	41.5	88.45	-0.167	435(C), 100(S), 204(N), 241(R).	
	СҮР2С9/490	P11712	55627.8	8.13	35130	45.12	94.65	-0.073	435(R)	:
	CYP2C18/490	P33260	55710.6	6.83	38235	45.76	89.49	-0.116	435(C).	Drug/xenobiotic metabolism
	CYP2C19/490	P33261	55931	7.11	33640	43.68	94.84	-0.098	435(K).	
	CYP2D6/497	P10635	55769.4	6.77	47815	44.63	95.15	-0.031	443(C), 301(D).	
	CYP2E1/493	P05181	56848.9	8.28	54820	33.65	91.14	-0.199	437(R).	
	CYP2F1/491	P24903	55501.2	6.93	31775	46.79	97.52	-0.047	436(C).	
	CYP2J2/502	P51589	57610.6	8.76	64650	33.98	86.1	-0.225	448(C).	
	CYP2R1/501	Q6VVX0	57359.3	7.25	67185	45.02	88.94	-0.094	448(G), 250(A).	
	CYP2S1/504	6DS96D	55816.6	8.8	42650	32.53	93.67	-0.024	440(G).	
	CYP2U1/544	Q7Z449	61987.1	8.63	76360	55.49	94.96	-0.078	490(C).	
	CYP2W1/490	Q8TAV3	53843.8	9.03	49430	45.43	104.12	-0.099	433(C).	
	CYP17A1/508	P05093	57370.5	8.72	63160	33.18	9.66	-0.154	442(C).	Testosterone and oestrogen biosvnthesis
	CYP19A1/503	P11511	57882.9	7.2	58830	38.23	98.81	-0.003	437(C), 309(D),	Oestrogen biosynthesis
	CYP21A2/494	P08686	55887.3	7.71	77265	49.27	106.19	-0.057	- 74 (M). 428(C).	(aronnatase) Steroid biosynthesis

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	Sub clade	CYP7A1/5	504 P22680	57660.7	8.48	60445	44.61	88.71	-0.21	444(C).	Rate-limiting step of bile
	Ŋ	CYP7B1/5	506 O75881	58255.9	8.2	56310	42.78	100	-0.041	449(C).	acid biosynthesis (cholesterol elimination)
Major droun 2		CYP8A1/5	500 Q16647	57104	6.8	80455	45.69	6.66	-0.13	440(C)	Prostacyclin and bile acid
9-04-0		CYP8B1/5	501 Q9UNU6	58068.4	8.78	06006	33.45	85.39	-0.165	440(C).	biosynthesis
		CYP39A1/-	469 Q9NYL5	54115.7	8.85	77975	38.29	102.45	-0.022	414(C).	Cholesterol metabolism
Table 2. l	Motifs obse	erved in th	ne human P450.								
Clade	Motif	Width	Regular expressi	uo							
Sub clade 1	e Motif 1	50	[gh]kalsd[el][df	e][il]ra[eQ][a\	/S][DI][TI]	F[MI]F[EA]	G[HY][DE]1	IT[AS]SG[L	I]S[WF][IV]LYEI	LA[TK]HP[ED][\	'HVJQ[EQ][RK][CL][RQ][EQ]E[I
	Motif 2	41	[HSD]P[YL]A[FY][II	-]PF[SG]AGPRN	NCIG[QM]RFA[ML]A	[EN][ML]K	VL][AV]LA	[LR][TV]L[LQ][F	kh NJF[RS][FL][I	-KJP[DC]
	Motif 3	50	[PG][WF]L[GK]D[G VAG][NDR][IV][MI	J[LHV][DAR]K.	JE[GD][D WL][EQR	e][ke]w[ks]	j[rq]hrr	irm]L[TS]P	(ATJF[HT][FS][N	ND][IK]LK[PE][Y	M][VIM][KP]I[FMI][AN][EQ]S[
	Motif 4	41	[PI][DN]GR[VFS][II	_]P[KA]GIV[VC]	LI[SP]IY[/	AGJLHH[ND	JP[KA][VY	JWP[DEN]F	EV[FY]DP[EF]F	kF[SD][PK][EK]N	-
	Motif 5	41	[den][pgk][ak][ei	PSJI[TE][WY]DI	DLAQ[LM][PE][YFJ[LT][TD]M[CV][IV][KN]E	STJLRL[YFH]PF	v[VA]PR[IV]SRx	[cl][tsk][kq][df][vi]
Sub clad∈ î	e Motif 1	41	I[KY][AG]N[VS][TN	JE[LM][LT][AL	[[AG][GS]	VDTT[SA][F	N][TP]L[LS	[WM][TA]	IL[YF]EL[AS]R[N	uh]pevqqal[f	(H]QE[ISV]
N	Motif 2	39	[HR][NP]F[AG][HS][LV]PFGFG[Vk	[M]R[QS]	c[LI]GRR[LI	JAE[LA]E[N	1][HLQ]L[LAJL[AHI][HQ]	irvjl[ka][hk][FYJE[VI][EQ]
	Motif 3	50	PLL[KR]A[AV]LKET S]R	'LRLYPV[VG][P	_][FGT][LI	u][ES]RVL[[ssj[ks]d[l	I]V[LV][GQ][DGN]Y[HL]IP	[KA][GN]T[LQ][vL]Q[LV][CF][HL]Y[ASV][LT][G
	Motif 4	50	[NRJF[IL][HDM]A[I W[KQ][DE]H[FTV]]	ILJ[EKY][VQT]N [EAL]AWD[CTV	IFM][KH: JIF[QKS][S][ST][TFS][YKS][GAV]]	VG][QPR][DK][NAI][LM][ML][FI CY][IT]	NV][MLT]P[RP	v][SDE]L[SFH][I	ĸĸj[wLs][FILT][SNR][PT]K[VT]
	Motif 5	50	[MS]V[CY][VL][AN EN]G[EP]EW	I][LS]PE[DL][VL	Jeklo[ro	J[QV][ED][GS][LP][H\	']PERMS[IL	JEPW[VK][AE][YHJR[QD][HY]F	R[GQ][HR]K[CY]G[VL][FL][LT]L[
Sub clade 3	e Motif 1	48	[YL][LIV][DG][CR][PIG[WHI[SNO]V[N	VC][IV]KE[TV][I ACI1[YV]SIIP][R	-MJRL[FL CTIIDVII7	NRJ[PT]P[I\ [NI[HO]][g][g][g][g][g][g][g][g][g][g][g][g][g]	[[GM][YFM]R[TMV][AV][L	r][kqrt][tp][f	Q][ET][LV][DAN]G[YF][QT]IP[K
	Motif 2	50	[ST]MG[FW]P[FL][MST]GA[ED]NV	FIJGET[LG][HQ][WM][LV][LV]Q[GR]	[SR][GKR]F	[hlq][sq]	[SM][RK]R[ER]	[KR]YG[FNT][VI	JJ[FY]KTHL[LF]GRP[LTV][IV]RV[
	Motif 3	39	[RK]F[HAST][YF][IF AS][EKT][LM][ALV]	FLV]PF[GS][GAI][/][/][/][/][/][/][/][/][/][/][/][/][/]	-]G[AHLR ACLSY][EF	V][RH][SRT tD][FW][ED	Jc(ILV)G[K QR]L	Qe][ehnQ][FL]A[KQY][AI	lmv][eflqv][l	.IV][KQ][VILT][LFIV][AMTW][V
	Motif 4	50	Geh[rh]lv[sr][s1 [filitams][rk][a]	NJ[EHQ]WP[A	ZRJS[ATV EICRSIY	'][RH][IMT]	[[]rd[sh]	GHN][TC][I	-V][SL][NG][AL	S][HIV][GD][DE	5][IPS]H[RK][NQR][RK][RK]KV
	Motif 5	45	[IV]	[SH][SA][KRW]	E[HLR]G[ehk][eR][LI	MP][DST]N	10[ea]lk[d	JEQ][SG][AST][iltvjel[li]f[ag][AG][FHY][AEF]TTASA[SA]TSL

Sub clade 4	Motif 1	50	R[YWJ][GA][LF]L[LY][LM][LM][KL][HY]P[ED][VJ][QET][AE][KR]V[QH]EE[LL][DE]RV[IV]G[RP]NR[QS]P[CS][LM][EQ]DR[AS][HK][ML]PY[TM][EDJAV[IVL]HE[VJ]Q
	Motif 2	32	F[ML]PF[SG][AI]GKRVCLGEGLARMELFLF[FL]TT[L]][LM]Q[NR]F
	Motif 3	50	G[LIV]P[HR][AR][VT]TRDT[KS][FL]RG[YF]L[IL]PKGT[ET][VI][FI][PT]NL[TW]SV[LH][HR]D[PE]KE[FW][PE]NPEEF[ND]PE[HR]FLD
	Motif 4	41	[SG][IL]E[ED]R[IV]QEEAR[CF]L[VI][ED][AE]LRKTK[GA][AS]P[FCI]DPTF[IL][LI][GS][CR]A[VP][SC]N[VI]I[CS]S
	Motif 5	29	GPR[PR]VVVLHGV[ED]A[VI][KR]EAL[VI]DQ[GA]E[ED]F[SA][GD]R[GP]
Sub clade 5	Motif 1	50	[PLT][GRW][ER]PP[LC][DIE][KLN]G[WLST][ILV]P[WY][L1]G[VCHY][AGV][LFM][ADENQ][FL][GR][KA][DNA][PAM][LAF][ERS]F[LIM][KERT][RAKT][MALN][QRK][EIKRT][KQ][HY]G[DHP][IVT]FT[VC1][LFKQ][LAV][GM]G[KNQR][YR][VFIM][TH][FV]
	Motif 2	41	[DN]G[KS][EKRT]K[TKV][DT]F[FY][KC][DNRT]G[KL][KR][L]][KH][CHNY]Y[LNTY]MP[FW]G[SAT]G[AHTV][SNT][IHK]C[PL]GR[FLS][FY]A[LIV][HMNS][ES][IV]K[QL]
	Motif 3	42	LWA[ST][QLV][AGS]N[TAM][GIV]P[ATV][AMST]FW[ALST][LM][LAFV][YFQ][LMV][LI][KRS][HN]P[ED][AI][MHIL][AKR]A[VAI][RMT][EDG][EG][IALV][DEKST][RSQ][VILT][LF][GEQW][EKNQS][AT][GER][QKL]
	Motif 4	15	[DN]P[EDK][IY][FYH][PET][DEA]P[ELT][VELT]F[KR][YP][DEN]R
	Motif 5	41	L[DNQ][SDH][LT][P1][VC]L[DE]S[VIS][ILV][EFKS]E[SAT]LRL[SRT][AS][AY][PS][TFL][ILNT][ILT]R[EFLT][VA][EHKV][EV]D[LFV][TA][LM][HKP S][MLS][EAS][DST]G[DQRS][EY][CFNV]

mals posses more CYP genes than humans like mice have genes for 101 CYPs, and sea urchins have even more (perhaps as many as 120 genes).²¹ Apart from humans, CYPs from insects, including CYP6G1 from Drosophila melanogaster²² and CYP6Z1 from Anopheles gambiae²³ have also been heavily studied so as to understand pesticide resistance. The amino acids composition and physicochemical features of protein is essential for understanding the evolutionarily conserved structure and function.²⁴ In the present study these 57 p450 protein sequences from human were analyzed by using bioinformatics tools. Physicochemical analysis showed that pIvalues 5.84 to 9.47. Isoelectric point (pI) is a pH value where net charge of protein is zero and shows whether protein character is acidic or basic. It was observed that most of proteins (50 members, 87.8%) were considered as basic character (pI \geq 7), while 7 protein sequences (12.2%) were considered as acidic (pI \leq 7). The most basic protein was found as CYP11B2 while the most acidic protein was found as CYP20A1. The GRAVY index ranged from 0.018 to -0.367. Positive GRAVY value corresponds to hydrophobic protein structure while negative GRAVY value means hydrophilic protein structure. GRAVY index revealed that except CYP26C1, all others as hydrophilic in nature. P450s are designed to break down environmentally toxic compounds such as polycyclic aromatic hydrocarbons and fluorocarbons, but the substrates of these proteins are predominately hydrophobic compounds and made more water soluble by monooxygenation.²⁵ The relative volume of valine and leucine/isoleucine side chains in comparison to the side chains of alanine is demonstrated by Aliphatic index.¹⁵ The dipeptides and tripeptides composition and their frequency of occurrence have been associated with solubility and folding of over-expressed proteins.²⁶ In general, proteins are mostly in the range of 42 to 62 kDa. The smallest protein was CYP27C1 containing 372 amino acids, had a molecular weight of 42.6 kDa. Proteins CYP4F22 with 531 amino acids and CYP2U1 with 544 amino acids had approximately were found to be the heaviest,

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Figure 1. Radial representation of the Maximum Likelihood tree of the human P450 proteins.

sharing a molecular weight of 61.9 kDa. Total amino acid composition analysis revealed that leucine is the major composition in all P450 proteins, and up to five motifs were determined by using MEME tool from proteins of every clade. The intron-exon organization of P450 genes exhibits a diversity of gene structure clearly indicating that multiple gains and losses of introns have occurred during the evolution of P450 genes in diverse species, with little conservation of intron positions among divergent P450 families.^{27,28}

Phylogenetic relationship of P450 proteins

The phylogenetic tree was constructed using 57 P450 protein sequences maximum likelihood method (ML). LG+G+I was used as the model. Two major groups were observed on the phylogenetic tree, the smaller one had CYP7, CYP8 and CYP39 group of proteins while the the larger clade, contained the rest of the proteins. Apart from the major clades, CYP20A1 was found to be alone. Clustering of the proteins was mainly on the identity. If an isoform has an identity of

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a)



b)



c)



Figure 2. Combined block diagrams of the conserved protein motifs in P450 proteins using MEME server. (a) Sub clade 1 consisting families 3, 4 and 5. (b) Sub clade 2 consisting families 11, 24 and 27. (c) Sub clade 3 consisting families 26, 46 and 51.



Figure 2. (d) Sub clade 4 consisting families 1, 2, 17, 19 and 21. (e) 7, 8 and 39.

more than 40%, it belongs to the same CYP gene family and if approximately 55-60%, then they belong to the same subfamily such as CYP1A, CYP2B, etc.^{20,29}

The bigger group had four sub-clades (Fig. 1). Each cluster of proteins was subjected to motif analysis. The motif and their regular expression are represented in Table 1. Variations among various P450enzymes in terms of physiochemical parameters are given in Table 2.

Major clade 1 comprises the first biggest group of proteins containing about 24 protein sequences with two sub groups. One represents families 3, 4 and 5 with functions related to drug/xenobiotic and fatty acid metabolism, while the other with families 11, 24 and 27 with functions related to bile acid biosynthesis and vitamin D activation. The pI of the proteins ranges from 6.26 to 9.47. Motif analysis showed absence of motif 3 in CYP11B2 and CYP27C1 (Fig. 2a).

Sub clade 1 consists of families 3, 4 and 5 with functions related to drug/xenobiotic and fatty acid metabolism. The pI of the proteins ranges from 6.26 to 9.29.

Sub clade 2 comprised of families 11, 24 and 27 with functions related to bile acid biosynthesis and vitamin D activation. The pI of the proteins ranges from 8.89 to 9.47. Motif analysis showed absence of motif 4 in CYP27B and motif 5 in CYP27C1 (Fig. 2b). All the proteins in this clade are of mitochondrial.

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Sub clade 3 represent 5 protein sequences belonging to families 26, 46 and 51. Cholesterol metabolism and retinoic acid metabolism were observed as the main functions of this group. The pI ranged from 8.68 to 9.24. The only protein (CYP26C1) that is hydrophobic in nature falls in this cluster. Domain analysis showed that as cystein as the key residue of all the proteins in this segment. Motif analysis showed absence of motif 2 in CYP46A1and motif 4 in CYP51A1 (Fig. 2c).

Sub clade 4, represent 22 protein sequences belonging to families 1, 2, 17, 19 and 21. These had functions related to drug/xenobiotic and steroid metabolism. The pI ranged from 6.77 to 9.31. Motif analysis showed absence of motif 4 in CYP1B1and motif 5 in CYP19A1 (Fig. 2d). CYP1A5 and CYP3A37 in turkeys were found to be very similar to the human CYP1A2 and CYP3A4 respectively, in terms of their kinetic properties as well as in the metabolism of aflatoxin B1.³⁰

Clade 5 represent 5 protein sequences belonging to families 7, 8 and 39. These had functions related to cholesterol metabolism, prostacyclin and bile acid biosynthesis. The pI ranged from 6.8 to 8.85. Domain analysis showed that cystein as the key residue. Motif analysis showed absence of motif 2 in CYP39A1 (Fig. 2e).

The protein CYP20A1 function of this protein is still not attributed. This was separated from the other two major clades. A similar protein was found in *Rattus norvegicus* (Q6P7D4) with slightly different physiochemical properties.

Computational analyses indicate that the human P450 proteins have wide sequence diversity, physicochemical properties and distinct phylogenetic tree topology. Cluster analysis of all retrieved sequences indicates the evolutionary history of P450 proteins. Conserved motifs within clusters showed the evolutionary pressure to maintain important residues for the structural and functional organization of different groups of protein. The data represented here can also be important for better understanding of physiological properties and roles of P450 family proteins in humans.

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