

CODEN (USA): IAJPBB

ISSN: 2349-7750

INDO AMERICAN JOURNAL OF PHARMACEUTICAL SCIENCES

Available online at: <u>http://www.iajps.com</u>

Research Article

DEVELOPMENT AND VALIDATION OF RP-HPLC METHOD FOR QUANTIFICATION OF ACYCLOVIR IN TABLETS

Shital S.Patil^{*}, P. A. Salunke, R. S. Wagh, Dr. S. D. Barhate

Shree Sureshdada Jain Institute Of Pharmaceutical Education And Research, Jamner, (M.S.) India.

Abstract:

A reverse phase method has been developed for the quantitative estimation of Acyclovir in tablet. The quantification was carried out using RP stainless steel column Water HEMA C18 (250 x 4.6) mm 5 μ column packing in isocratic mode with mobile phase containing methanol, acetonitrile and water in the ratio of 45:45:10, Flow rate of 1.0 ml/min and the detection wavelength were set at 264 nm and the linearity was found to be in the range of 2-10 μ g/ml for acyclovir. The proposed method was found to be simple, precise, accurate, and reproducible for the estimation of acyclovir.

Keywords: Acyclovir, method development, validation, high performance liquid chromatography.

Corresponding author: Shital S. Patil, Shree Sureshdada Jain Institute Of Pharmaceutical Education and Research, Jamner, (M.S.) India. sspatil26888@gmail.com



Please cite this article in press as Shital et al, **Development and Validation of RP-HPLC Method for** *Quantification of Acyclovir in Tablets*, Indo Am. J. Pharm. Sci, 2016; 3(1).

INTRODUCTION:

Acyclovir is Guanosine derivative and has potent antiviral activity essentially against herpes simplex type I virus and HSV-2, EBV, VZV [1]. The chemical name for acyclovir is 2 amino-1,9-dihydro-9- [(2-hydroxyethoxy) methyl]-6Hpurine- 6-one, or 9- [(2-hydroxyethoxy) methyl]- guanine. Its molecular formula is C8H11N5O3, and molecular weight 225.21 g/mol [4]. Acyclovir (Fig-1) is commonly used as the free acid form in solid oral dosage forms, whereas the sodium salt is used in parenteral dosage forms. [5]. Acyclovir is normally present in a hydrated form consisting of three acyclovir molecules to two molecules of water, [6] corresponding to a theoretical water content of about 5%, but dose and solubility are normally expressed in units of anhydrous acyclovir. Acyclovir is described as "slightly soluble in water" in different Pharmacopoeias. The solubility of acyclovir in most of the literature are range from 1.2 to 1.6 mg/mL at room temperature (22 to 250C). [7].

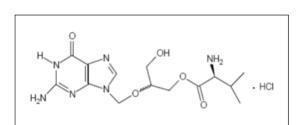


Fig 1: Chemical Structure of Acyclovir MATERIAL AND METHODS:

Materials

Acyclovir was obtained from AurochemPharmceuticals (I) Pvt. Ltd, Thane, India, HPLCgrade acetonitrile, AR grade Methanol was procured from Merck, India and Bransted HPLC water was used as received.

HPLC analysis

The analysis was performed on a chromatographic system of Waters -UV Detector, equipped with manual sampler and DATA ACE software. The chromatographic column was RP stainless steel column Water HEMA C18 (250 x 4.6) mm, 5 μ column packing in isocratic mode. HPLC instrument was operated at ambient temperature. The flow rate of the mobile phase was maintained at 1.0 ml/min. Detection was carried out at 264 nm Retention time of Acyclovir was about 1.757 min. Run time was set for 10 min.

Acyclovir standard solution

Accurately weighed 10 mg of Acyclovir tablet was taken into a 10 ml volumetric flask. 5 ml of mobile phase was added to it, sonicated to dissolve and diluted to volume with mobile phase and mixed thoroughly.

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Acyclovir standard stock solution

One milliliter of Solution A was diluted to 10 ml with the mobile phase.

Study of Spectra and Selection of Wavelength:

The aliquot portions of standard stock solutions of Acyclovir were diluted appropriately with distilled water to obtain concentration 10 μ g/mL of drug. The solutions of drug was scanned in the range of 400 – 200 nm. The UV absorbance spectrum of Acyclovir is shown in **Figure 2**.

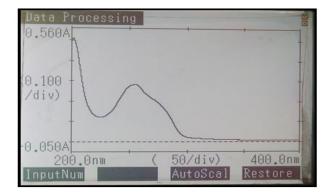


Fig 2: UV- Spectrum of Acyclovir

From the spectrum wavelengths selected for estimation of drug was 264 nm as λ max of Acyclovir.

RESULT AND DISCUSSION:

Linearity

The developed method has been validated as per ICH guidelines Each 10 ml of the standard solution of Acyclovir in the concentration range of 2-10 μ g/ml each were injected into the chromatographic system. The chromatograms were developed and the peak area was determined for each concentration of the drug solution. Calibration curves of Acyclovir were obtained by plotting the peak area ratio versus the applied concentrations of acyclovir (fig. 3).

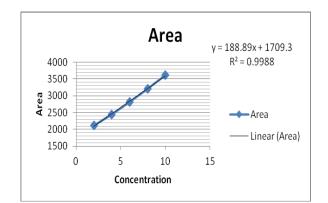


Fig.3: Linearity of Acyclovir

The resultant correlation coefficient (R) should be ≥ 0.99 for Acyclovir. As per linearity graph of Acyclovir. It was linear with co-efficient of correlation (R) of more than 0.99. The linearity within the range of (80 % to 120%), the standard limits concentration was established.

Repeatability of the method was checked by injecting six replicate injections of the solution 20μ l each of Acyclovir, the RSD was found to be 0.74. The relative standard deviation of reproducibility and repeatability with respect to peak area and retention time are well within the acceptance criteria. Hence, the method was suitable.

Precision

1		Table 1: In	ter day Precis	ion study		
Sr No.	Conc.	Area	ĨI	Mean	SD	RSD
1	2	2111.61	2151.3	2131.46	28.07	1.32
2	4	2458.12	2483.87	2471.00	18.21	0.74
3	6	2871.41	2916.05	2893.73	31.57	1.09
		Table 2: In	tra day Precis	ion study		
Sr No.	Conc.	Area	II	Mean	SD	RSD
1	2	2111.61	2151.3	2131.46	28.07	1.32
2	4	2458.12	2483.87	2471.00	18.21	0.74
3	6	2871.41	2916.05	2893.73	31.57	1.09
		Table 3: Reso	olution Study o	of Acyclovir		
Sr. no	RT		TP		TF	
1	1.757		5527		1.75	
2	1.757		5643		1.74	
3	1.757		5612		1.76	
4	1.757		5631		1.72	
5	1.757	5515			1.71	
6	1.757		5495		1.72	

RT- Retention Time, TP- Theoritical Plate, TF- Tailing Factor

Table 4: Accuracy study

Conc.	Area	Mean	SD	% RSD	% Accuracy	% Recovery
	3857.44					
80	3787.23	3797.63	2.97	1.82	74.2	92.53
	3748.65					
	1519.22					
100	1528.21	1530.29	0.65	1.73	97.50	97.50
	1543.46					
	5732.47					
120	5834.21	5820.97	4.37	1.65	122.60	102.17
	5896.23					

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Accuracy

The accuracy of the method was tested by carrying out recovery studies at different spiked levels. The estimation was carried out as described earlier. At each level, three determinations were performed and results obtained. The amounts recovered and the values of percent recovery were calculated. The results for Acyclovir have been displayed in Table 4. The accuracy and recovery results obtained with all the three different concentration levels applied (80%,100% & 120%) are well within the acceptance criteria which shows that the method is accurate. *Repeatability*

Repeatability was carried out by using a minimum of 6 determinations at 80 percent of the test concentration.

Sr No.	Conc.	Peak Area	Amt Found	%Amt Found
1	2	3406.7	136.77	45.59
2	2	3446.3	138.87	46.29
3	2	3445.3	138.82	46.27
4	2	3505.6	142.01	47.34
5	2	3383.4	135.54	45.18
6	2	3396.3	136.22	45.41
		Mean	138.04	46.01
		SD	2.37	0.79
		%RSD	1.72	1.72

Table 5: Repeatability study

Robustness

The evaluation of robustness should be considered during the development phase and depends on the type of procedure under study. It should show the reliability of an analysis with respect to deliberate variations in method parameters. For eg. Flow rate, Mobile phase composition results were shown in tables 6 and 7.

Specificity

The specificity of the method was checked for the interference of retention time of a blank solution

(without any sample) and then a drug solution of 20 μ l was injected into the column, under optimized chromatographic conditions, to demonstrate the separation of Acyclovir. There was no interference of blank on retention time of Acyclovir.

Limit of Quantization (LOQ)

It is the smallest level of analyte that gives a measurable response and standard deviation and slope for the peak area responses was calculated shown in Table 8.

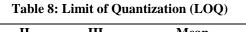
Table 6: Robi	stness study:	Flow change
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Sr No.	Conc.	Area	Sr No.	Conc	ng/Band	Area
1	10	3283.06	1		10	3209.65
2	10	3247.36	2		10	3259.15
	Mean	3265.21			Mean	3234.40
	SD	25.24			SD	35.00
	%RSD	0.77			%RSD	1.08

Sr. No.	Conc.	Area	Sr. No.	Conc	Area
1	2	14966.4	1	2	14963.2
2	2	14593.9	2	2	14669.6
	Mean	14780.15		Mean	14816.40
	SD	263.40		SD	207.61
	%RSD	1.78		%RSD	1.40

Table 7: Robustness study: Mobile phase

Sr No.	Conc.	Area I	II	III	Mean	SD	%RSD
1	2	2649.69	2672.3	2692.6	2671.53	21.47	0.80
2	4	3220.51	3129.9	3135.9	3162.10	50.67	1.60
3	6	3477.69	3488.4	3498.7	3488.26	10.51	0.30



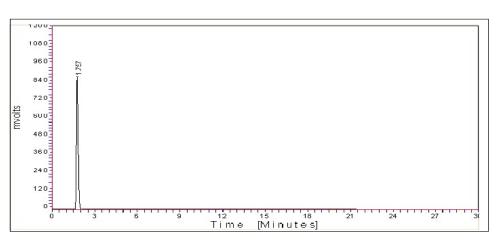


Fig 4: Typical chromatograph of Acyclovir

Table 9: System Suitability

System Suitability Component Parameter	Acyclovir
Retention times (RT) min	1.757
Tailing factor (AS)	1.75
Slope	8939.2
Intercept	4900.2
Coefficient of variance	0.9994
Linear range	1-3 mg/ml

System Suitability

System suitability testing is an integral part of many analytical procedures. The tests are based on the concept that the equipment, electronics, analytical operations, and samples to be analyzed constitute an integral system that can be evaluated as such. The system suitability is given in table no.9

CONCLUSION:

The developed method was validated in terms of accuracy, linearity and precision. A good linear relationship was observed. Acyclovir in the concentration ranges of 2-10 μ g/ml /ml. The correlation coefficient for Acyclovir was found to be 0.9994. Selectivity experiment showed that there is no interference or overlapping of the peaks either due to diluents with the main peak of Acyclovir. The percentage RSD for precision is <2 which confirms that method is sufficiently precise and the total runtime required for the method is only 5 min for eluting Acyclovir. The proposed method is simple, fast, accurate, and precise and can be used for routine analysis in quality control for Acyclovir.

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