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Diuretic and antioxidant activities of the aqueous extract of leaves of *Cassia occidentalis* (Linn.) in rats

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ABSTRACT

Objective: To assess the putative diuretic and antioxidant properties of *Cassia occidentalis* (*C. occidentalis*) leaves' aqueous extract.

Methods: Adult rats were administered with *C. occidentalis* leaves aqueous extract acutely (24 h) and subchronically (7 d), at doses 80, 160, 240, 320, and 400 mg/kg (*per os*). Negative control group received only an equivalent volume of distilled water, while the two positive control groups received the diuretic drugs furosemide (20 mg/kg, *ip.*) and hydrochlorothiazide (HCTZ) (20 mg/kg, *ip.*). Urinary elimination of electrolytes in response to treatments was evaluated, together with changes in concentrations of creatinine, urea, aldosterone, glucose, and albumin in urine and plasma. Various urinary indicators of kidney function and plasmatic markers of oxidative stress were also assessed. **Results:** The acute administration of *C. occidentalis* increased the urinary excretion of 107.58% at the higher dose tested, compared to negative control. The reference drugs furosemide and HCTZ induced increases of 84.27% and 48.05%, respectively. Acutely, the extract induced Na⁺ and Cl⁻ elimination, whereas subchronically an increase in K⁺ elimination was also observed. The extract also improved the kidney function indexes and oxidative stress markers. These effects were dose-dependent and comparable with positive control observations.

Conclusions: Our findings strongly suggest that *C. occidentalis* aqueous extract has diuretic and antioxidant activities, and deserves further studies considering the potential for the treatment of hypertension.

1. Introduction

Arterial hypertension is among the most frequent pathologies in elderly worldwide, with an incidence ranging from 40% (about 65 years patients) to 90% (patients older than 85) in developed countries [1,2]. This pathology raises more concerns as it constitutes a major risk of cardiovascular accident. In developing countries, partly due to the relatively inexpensive

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costs several hypertensive patients have been relying on medicinal plants for their treatment. Furthermore, various reports suggesting that conventional antihypertensive drugs may increase the risk for cardiovascular accidents have resulted in an increased interest of the research community for medicinal plants [3,4]. Not surprisingly, WHO reports (particularly in the last decade) have been encouraging studies on medicinal plants and alternative medicine for priority diseases like hypertension and its cardiovascular complications.

Cassia occidentalis (*C. occidentalis*) is a tropical plant used in African and Asian traditional medicines to treat or improve a number of diseases and conditions, in particular cardiovascular disorders [5,6]. As in various other countries, in Cameroun roasted seeds are used as coffee substitutes, while other parts of the plants are used by traditional healers to treat metabolic and cardiovascular diseases. Interestingly, experimental

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evidence supports the applications in traditional medicine. Phytochemical studies of C. occidentalis leaves revealed the presence of a number of pharmacologically active families of molecule, including tanins, saponins, cardiac glycosides, terpenoides and anthroquinones, terpenes, and inorganic elements [7,8]. Extracts of this plant were reported antifungal, antiviral [9,10], antibacterial, anthelmintic [11,12], antispasmodic, anti-inflammatory analgesic, antipyretic, [13,14] and hepatoprotective properties in humans and experimental models [15,16]. Verma and colleagues (2010) showed the effect of ethanolic extract of C. occidentalis for the management of alloxan-induced diabetic rats [17,18], and Sreejith and colleagues reported anti-allergic, anti-inflammatory and antilipid peroxiding effects [19].

The present work aim was to measure the diuretic and antioxidant activities of the aqueous extract of *C. occidentalis* leaves in rats.

2. Materials and methods

2.1. Animals and procedures

Rattus norvegicus $[(172.3 \pm 4.3) \text{ g}]$ of both sexes obtained from Yaoundé's Pasteur Institute (Cameroon) were reared in the Department of Biological Sciences, Faculty of Sciences (University of Ngaoundéré, Cameroon). Animals were housed under controlled temperature $[(24 \pm 2) \ ^{\circ}C]$ and relative humidity $[(45 \pm 10)\%]$, and had *ad libitum* access to food (pellets from Cameroonian National Veterinary laboratory) and tap water. Animal health status and housing conditions were monitored by a veterinary physician.

Preliminary tests were performed as previously described [20]. Briefly, rats received distilled water per os (10 mL/kg body weight), and placed individually in metabolic cages. After 6 h, urine was collected and the volume measured. Animals excreting at least 40% of the volume of distilled water received were selected for the study, and conversely, those excreting less than 40% were excluded. Then, selected animals were placed individually in metabolic cages, and allowed 7 d for acclimation. Eight experimental groups were obtained by treating n = 5 rats (per group) with a specific solution, *i.e.*: vehicle solution (distilled water, per os) for the negative control group, one of the 5 different doses of extract investigated for the 5 test groups (per os), and the diuretic drugs furosemide (20 mg/kg, ip.) or hydrochlorothiazide (HCTZ) (20 mg/kg, ip.) for the positive control groups. Animals were sacrificed by decapitation at the end of the experiment. Arteriovenous blood was collected in heparinized tubes and centrifuged (3000 rev/min for 10 min). The plasma collected was stored at -20 °C for biochemical analyses. The liver and kidneys were dissected out, cleaned of fat material, weighed and stored at -20 °C for biochemical analyses.

Experimental procedures were approved by the institutional Animal Care and Use Committee and the research was approved by the Ethics Committee of the University of Ngaoundéré.

2.2. Plant extract preparation

2.2.1. Plant material collection

Leaves of *C. occidentalis* Linn were harvested in Mora (60 km from Maroua, the largest city of the Far North region of

Cameroon) during rainy season. They were identified by experts of the National Herbarium of Cameroon and a sample was deposited (specimen N0 21057/SFR/CAM).

2.2.2. Aqueous extract preparation

Fresh leaves of *C. occidentalis* were soaked in distilled water (1 000 g for 1 L at room temperature) for 12 h. The macerate was filtered through Whatman filter paper No 3, and the filtrate concentrated in a rotary evaporator at 40 °C for 24 h. This process was repeated until an oily paste extract was obtained (130 g), which represented the concentrated crude extract of *C. occidentalis* leaves. The extract was stored at -20 °C until use.

2.2.3. Aqueous extract doses

The solution of *C. occidentalis* extract with the highest concentration tested was prepared by dissolving 800 mg of the concentrated crude extract obtained previously in 10 mL of distilled water (80 mg/mL concentration). The other solutions used in the study were 4:5, 3:5, 2:5, and 1:5 dilutions of this solution in distilled water. Solutions were given *per os* in a volume of 5 mL/kg body weight, thus, the increasing doses of aqueous extract of *C. occidentalis* tested were 80, 160, 240, 320, and 400 mg/kg.

2.3. Diuretic effect evaluation

2.3.1. Acute diuretic effect evaluation

Urine was collected and the volume determined each hour from the treatment for 6 h (*i.e.* 1, 2, 3, 4, 5, and 6 h after treatment) and 24 h after in all experimental groups. Electrolyte concentrations (Na⁺, K⁺, and Cl⁻) were measured in 24-h urine and in blood plasma obtained from animals sacrificed 24 h after treatment.

2.3.2. Subchronic diuretic effect evaluation

Based on preliminary observations from acute diuretic effect evaluation, the experimental group receiving the highest dose of extract tested (400 mg/kg), and the positive and negative control groups were treated daily for 7 d at the same time each day. Urine was collected daily, its volume measured, and electrolyte concentrations (Na⁺, K⁺, Cl⁻) determined.

2.4. Determination of urinary and/or plasma concentrations

2.4.1. Osmolarity and electrolytes

Osmolarity of plasma and urine samples were measured by cytometry using an osmometer (Knauer). Urinary and plasma concentrations of Na⁺, K⁺ and Cl⁻ ions were evaluated using flame photometry (Jenway PFP 7, Bibby Scientific, USA), following standard protocols. Urinary natriuresis was measured during the diuretic response, particularly at the maximum excretion rate. Doses of Na⁺ and K⁺ were calculated as indicators of saluretic activity and the ratio Na⁺/K⁺ was calculated for the natriuretic activity. And to estimate the carbonic anhydrase inhibition activity, the ratio of Cl⁻ ions to Na⁺ and K⁺ ions was calculated [21].

Osmolar clearance was calculated using plasma osmolality, urinary osmolarity and urine flow (V) according to the following formula:

Osmolar clearance = urinary osmolarity × V/plasma osmolality

2.4.2. Concentrations of other blood molecules

A two-way digital spectrophotometer (Secomam RS232C, Secomam SAS, France) was used to determine the concentrations of urea, glucose, albumin, and creatinine in plasma and urine samples. Similarly malondialdehyde concentration was determined in plasma, and catalase, hydroperoxide and protein concentrations were determined in hemolysates of blood pellets and in liver homogenates. Aldosterone concentration in the plasma was measured using radioimmunoassay (assay kit Aldo RIACT, ALPCO Diagnostics, USA).

2.5. Phytochemical studies

In order to identify the chemical structure of the compounds responsible for the diuretic activity, preliminary tests of the phytochemical study were conducted following the procedures described by Trease and Evans (1983) ^[22]. Briefly, Essential oils from the aqueous extract of *C. occidentalis* and urine were extracted with hexane. These extracts were then stitched onto plates of thin layer chromatography on silica, the first disclosure was obtained by ultraviolet radiation (254 nm and 365 nm) and then with vanillin. Analytical tests for the identification of different families of metabolites in crude extracts of the leaves were performed at the national Institute of Medicinal Plants for Medicinal research (Cameroon).

2.6. Statistical analyses

Data from test groups and positive control groups were compared to negative control group using one-way ANOVA followed by LSD test for post hoc analysis, using Origin software (OriginLab, Northampton, MA, USA). Changes with P values lower than 0.05 were considered significant. Data are presented as mean \pm SEM.

3. Results

3.1. Effect of the acute administration of C. occidentalis extract on urination and associated urinary and serum parameters

3.1.1. Overload eliminated after 1 h

The overload eliminated after 1 h by rats acutely treated with the three highest doses of *C. occidentalis* aqueous extract tested (240, 320, and 400 mg/kg), administered with an equivalent volume of distilled water (negative control), or rats treated with one of the two diuretic drugs (positive control) is shown in Figure 1A. All treatments induced at least a 3-fold increase in the overload elimination compared with negative control group. The response of the extract was dose-dependent, and eliminated 78.79% of overload at 240 mg/kg (P < 0.001 against negative control group), 90.87% at 320 mg/kg (P < 0.01) and 126.87% at 400 mg/kg (P < 0.001). Furosemide eliminated 100.00% (P < 0.001), HCTZ 88.43% (P < 0.001), whereas only 35.35% overload was eliminated in the negative control group.

3.1.2. Latency to first urination, urine volume and pH

The bar graph of Figure 1B represents the latency to first urination of rats acutely treated with the three highest doses of

10 2 5 1 0 0 dH2O нстг 240 320 400 Furos. C. occidentalis extract (mg/kg) Figure 1. Overload eliminated and urination latency. A. Overload eliminated after 1 h by rats acutely treated with the three highest doses of C. occidentalis aqueous extract tested, administered with an equivalent volume of distilled water (negative controls), or rats treated with one of the two diuretic drugs used as positive controls. Note that all treatments induced at least a 3-fold increase in the overload elimination compared with negative controls. Note also that the response of the extract was dose-dependent. B. Latency to first urination (bars) and urine pH of the same rats. Note the 2-fold decrease in all groups compared with negative controls. Note also the dose-dependent response of the extract, and the absence of significant change in the pH of the urine. dH₂O, distilled water; Furos, furosemide. ANOVA + LSD test against negative control group: *P < 0.05, **P < 0.01, ***P < 0.001.

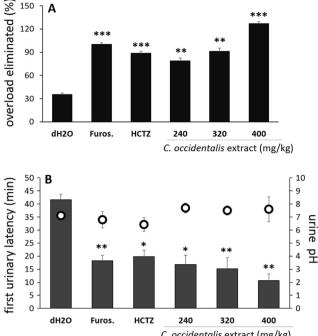
C. occidentalis aqueous extract tested, and of negative and positive control groups. Negative control group average first urination latency was 42 min, furosemide 18.27 min (P < 0.01), and HCTZ 19.82 min (P < 0.05). The extract displayed a dose-dependent decrease as the first urination latency with 17 min (P < 0.05), 15 min (P < 0.01), and 11 min (P < 0.01) at doses 240, 320 and 400 mg/kg.

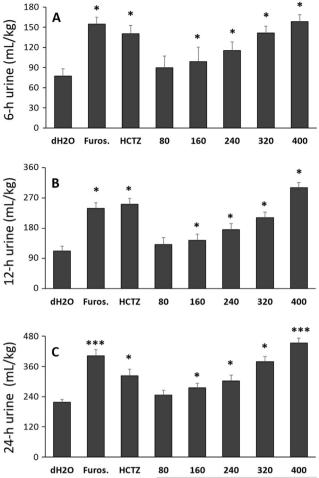
The urinary pH (7.11 in negative control group) was slightly decreased in groups treated with furosemide (6.79) or HCTZ (6.42), and slightly increased in groups receiving the extract (7.7, 7.5, 7.6 at doses 240, 320 and 400 mg/kg). However, these changes were not statistically significant (Figure 1B).

Figure 2 shows the changes in the total volume of urine excreted by rats following acute treatment with various doses of *C. occidentalis* aqueous extract (80, 160, 240, 320, and 400 mg/ kg). On the overall, the extract and the diuretic drugs increased the volume of urine excreted. After 24 h, an average volume of urine of 218.34 mL/kg body weight was excreted in the control group, whereas 402.35 mL/kg of urine were excreted in furosemide-treated (P < 0.001), 323.27 mL/kg in HCTZ-treated (P < 0.05), and 453.24 mL/kg (P < 0.001), in groups receiving the extract at 400 mg/kg, respectively (Figure 2C).

3.1.3. Electrolyte excretion

The effect of acute treatment with *C. occidentalis* aqueous extract or diuretic drugs (compared with negative control





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control group), respectively, to 167.39 and 168.36 mEq/kg at the dose 400 mg/kg. The urinary excretion of Na⁺ and Cl⁻ induced by furosemide were 93.38 and 94.67 mEq/kg (P < 0.05), respectively, and HCTZ 72.43 and 75.41 mEq/kg (P < 0.05) (Figure 3C, F). No statistically significant change was observed in the mount of K⁺ excreted in the urine after 6 h (Figure 3G), 12 h (Figure 3H) and 24 h (Figure 3I).

3.1.4. Urinary indexes of kidney function

The effect of C. occidentalis aqueous extract and diuretic drug treatment (compared to negative control group) on indexes of kidney function in urine produced in the 24 h following the treatment are shown in Table 3. Treatments with the various doses of extract and the diuretic drugs decreased the glomerular filtration rate from 1.66 mL/min in the negative control group to 1.37 mL/min (furosemide, P < 0.001), 1.38 mL/min (HCTZ, P < 0.001), and 1.43 mL/min (extract at 400 mg/kg, P < 0.01). Creatinine clearance and urea concentration in the urine were significantly decreased as well, but no significant change was observed in urinary creatinine rate (Table 3). Osmolar clearance and free water clearance were increased, while urinary osmolarity decreased. Glucose and albumin were not detected in the urine.

3.1.5. Serum parameters

Concentrations of Na⁺ and K⁺ ions were significantly increased (596.44% and 311.17% respectively in animals receiving the extract at 400 mg/kg, P < 0.05) (Table 4). Furosemide and HCTZ and the extract (at 400 mg/kg) induced significant increases (4.56%, 2.31%, and 4.57% respectively, P < 0.05) in albumin level (Table 4). Albumin level increased from 43.17 g/L in the control group to 45.18 g/L in animals treated with the extract at 400 mg/kg.

Increases in plasma osmolality and aldosterone levels were observed (P < 0.001) (Table 4). Similarly, the extract and diuretic drugs induced significant increases (P < 0.05) in serum creatinine and urea levels. Despite a slight increase in rat receiving the extract and diuretic drugs, glycemia ranged between (91.12 \pm 6.34) mg/dL and (97.65 \pm 7.15) mg/dL in all experimental groups.

3.2. Effect of the sub-chronic administration of C. occidentalis extract on urination and urine parameters

3.2.1. Urine volume

The total volumes of urine excreted daily by rats following subchronic treatment with the highest dose of C. occidentalis aqueous extract tested (400 mg/kg), administration of an equivalent volume of distilled water (negative controls), or treatment with one of the two diuretic drugs used as positive controls are shown in Figure 4A. C. occidentalis and diuretic treatments significantly increased the volume of urine excreted, compared to the negative control group. Compared to the negative control group (y = 1.91x + 208.4, $R^2 = 0.77$), furosemide (y = 21.3x + 389.9, $R^2 = 0.90$), HCTZ (y = 14.7x + 292.3, $R^2 = 0.99$), and extract-treated groups (y = 23.5x + 425.6, $R^2 = 0.99$) had a significantly higher and positive slope (P > 0.01), *i.e.* the effects were more marked with time. Notably, from the first day of treatment the volume of urine excreted daily was significantly increased by all these treatments, compared to the negative control group.

The total volume of urine excreted after 6 h (A), 12 h (B), and 24 h (C) by rats following acute treatment with various doses of C. occidentalis aqueous extract, administration of an equivalent volume of distilled water (negative controls), or treatment with one of the two diuretic drugs used as positive controls. Note that the highest doses of extract and the diuretic drugs increased the volume of urine excreted. Note also that the response of the extract was dose-dependent. dH2O, distilled water; Furos, furosemide. ANOVA + LSD test against negative control group: *P < 0.05, **P < 0.01, ***P < 0.001.

Figure 2. Extract acute administration and urine volume.

C. occidentalis extract (mg/kg)

groups) on various electrolyte excretion indexes and parameters in the urine produced in the 24 h following treatment are shown in Figure 3, and Tables 1 and 2.

As shown in Table 1, saluretic and natriuretic activities were significantly increased by diuretic drugs (P < 0.001) and by the extract (P < 0.001 at the highest dose tested). Similarly, carbonic anhydrase inhibition was also increased by diuretic drugs (P < 0.01) and by the extract (P < 0.01) at the highest dose tested). On the same hand, carbonic anhydrase inhibition, saluretic, natriuretic, diuretic, as well as Na⁺ and Cl⁻ indexes were high for in all groups, but the extract effect at 400 mg/kg (higher dose used) was 2-fold higher than the diuretic drugs at the dose used (Table 2). Notably, K⁺ index was only slightly increased by the diuretic drugs and the extract (Table 2).

All treatments increased the urinary excretion of Na⁺ and Cl⁻ compared to negative control group; the extract response was dose-dependent (Figure 3). After 24 h, the extract significantly (P < 0.05) increased the urinary excretion of Na⁺ (Figure 3C) and Cl⁻ (Figure 3F) from 16.58 to 16.67 mEq/kg (negative

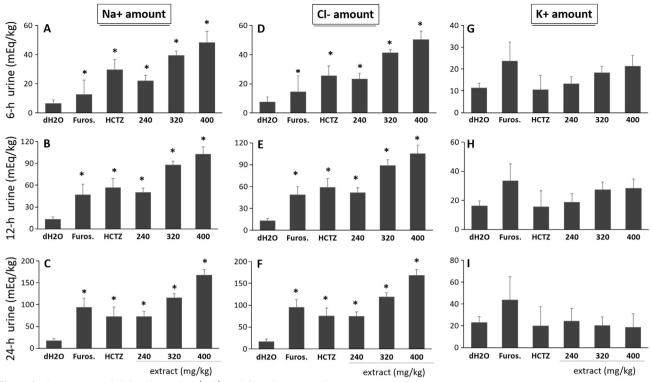


Figure 3. Extract acute administration and Na⁺, K⁺, and Cl⁻ urinary excretion.

A–C. Estimated amount of Na⁺ excreted in the urine after 6 h (A), 12 h (B), and 24 h (C) by rats following acute treatment with various doses of *C. occidentalis* aqueous extract, administration of an equivalent volume of distilled water (negative controls), or treatment with one of the two diuretic drugs used as positive controls. Note that all treatments increased the urinary excretion of Na⁺ compared to negative control group, and the dose-dependent response of the extract. Note also that the highest dose of extract tested induced a 10-fold increase in the amount of Na⁺ excreted, whereas the diuretic drugs induced only a 5-fold increase. D–F. Estimated amount of Cl⁻ excreted in the urine after 6 h (D), 12 h (E), and 24 h (F) by the same rats. Note the similar observations as in A–C. G-I. Estimated amount of K⁺ excreted in the urine after 6 h (G), 12 h (H), and 24 h (I) by the same rats. Note that no statistically significant change was observed. dH₂O, distilled water; Furos, furosemide. ANOVA + LSD test against negative control group: **P* < 0.05, ***P* < 0.01, ****P* < 0.001.

Table 1

Effect of acute treatment with *C. occidentalis* aqueous extract on carbonic anhydrase inhibition, and on saluretic and natriuretic activities, observed in urine produced in the 24 h following the treatment.

Experimental gr	oups	Saluretic (Na ⁺ + Cl ⁻)	Natriuretic (Na ⁺ /K ⁺)	CAI $[Cl/(Na^+ + K^+)]$
dH ₂ O		33.3 ± 3.3	0.7 ± 1.0	0.4 ± 0.5
Extract	240 mg/kg	$147.0 \pm 4.5^{***}$	$1.4 \pm 1.1^*$	$0.8 \pm 0.4^*$
	320 mg/kg	234.1 ± 3.8***	$5.7 \pm 1.2^{**}$	$0.9 \pm 0.6^{*}$
	400 mg/kg	$335.8 \pm 5.2^{***}$	$9.1 \pm 1.0^{***}$	$0.9 \pm 0.5^{**}$
Furosemide		$188.1 \pm 7.9^{***}$	$2.1 \pm 0.9^{***}$	$0.7 \pm 0.4^{**}$
HCTZ		$147.8 \pm 7.8^{***}$	$3.6 \pm 1.2^{***}$	$0.8 \pm 0.5^{**}$

CAI: carbonic anhydrase inhibition. Values are mean \pm SEM, n = 5. ANOVA + LSD test vs. negative control: *P < 0.05, **P < 0.01, ***P < 0.001.

Table 2

Effect of acute treatment with C. occidentalis aqueous extract on electrolyte excretion indexes in urine produced in the 24-h following the treatment.

Experimental groups		Electrolyte indexes		K^+ index	Other indexes		Natriuretic (Na ⁺ /k ⁺)	CAI [Cl/(Na + K)]
		Na ⁺ index	Cl ⁻ index		Diuretic index	Saluretic (Na ⁺ + Cl ⁻)		
Extract	240 mg/kg	4.4	4.5	1.1	1.4	4.4	2.0	1.8
	320 mg/kg	7.0	7.1	0.9	1.7	7.0	7.9	2.1
	400 mg/kg	10.1	10.1	0.8	2.1	10.1	12.6	2.1
Furosen	nide	5.6	5.7	1.9	1.8	5.7	3.0	1.6
HCTZ		4.4	4.5	0.9	1.5	4.4	3.5	2.0

CAI index: CAI activity in test group/CAI activity in control group. Cl⁻ index: chloride excretion in test group/chloride excretion in control group. Diuretic index: urine volume of test group/urine volume of control group. K⁺ index: potassium excretion in test group/potassium excretion in control group. Na⁺ index: sodium excretion in test group/sodium excretion in control group. Natriuretic index: natriuretic activity in test group/natriuretic activity in control group. Saluretic index: saluretic activity in test group/saluretic activity in control group. Values are mean \pm SEM, n = 5.

Table 3

Effect of acute treatment with C. occidentalis aqueous extract on indexes of kidney function in urine produced in the 24-h following the treatment.

Experimental groups	Glomerular filtration rate	Urinary osmolarity	Osmolar clearance	CH ₂ O	Urea	Creatinine	Creat C
	(mL/min)	(mosmol/kg)	(µL/min)	(µL/min)	(mg/24 h)	(mg/24 h)	(µL/min)
dH ₂ O Extract 240 mg/kg 320 mg/kg 400 mg/kg Furosemide HCTZ	$1.52 \pm 0.14*$	$\begin{array}{l} 200.20 \pm 17.10 \\ 111.10 \pm 18.20^{***} \\ 113.10 \pm 13.20^{***} \\ 124.40 \pm 17.30^{***} \\ 170.20 \pm 28.10^{***} \\ 166.30 \pm 12.10^{***} \end{array}$		$56.0 \pm 1.2 54.0 \pm 1.3 56.0 \pm 1.1 66.0 \pm 1.6^{***} 68.0 \pm 1.5^{***} 66.0 \pm 1.8^{***}$		$23.2 \pm 4.5 24.2 \pm 3.8 26.2 \pm 2.6 27.1 \pm 3.4$	28.0 ± 1.2 $25.0 \pm 1.6*$ $22.0 \pm 1.4**$ $20.0 \pm 1.3***$ $21.0 \pm 1.2**$ $19.0 \pm 1.4***$

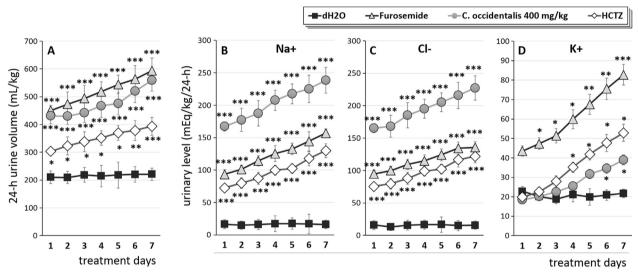
CH₂O: free water clearance. Creat C: creatinine clearance. Values are mean \pm SEM, n = 5. ANOVA + LSD test vs. negative control: *P < 0.05, **P < 0.01.

Table 4

Effect of acute treatment with C. occident	<i>lis</i> aqueous extract on serum parameters.
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Groups	Na ⁺ (mEq/L)	K ⁺ (mEq/L)	Glucose (mg/dL)	Creatinine (mg/dL)	Urea (mg/dL)	Albumin (g/L)	Aldosterone (pg/mL)	Plasma osmolality (mosmol/kg)
dH ₂ O	1.7 ± 0.6	1.8 ± 0.9	91.1 ± 6.3	0.6 ± 0.3	22.2 ± 2.4	43.2 ± 4.1	283.2 ± 22.2	259.2 ± 21.1
Extract 240 mg/kg	7.6 ± 1.7***	$5.0 \pm 2.8^{***}$	92.2 ± 7.6	$0.7 \pm 0.6^{*}$	23.5 ± 1.3	44.1 ± 3.1	285.2 ± 18.6	266.3 ± 22.1**
320 mg/kg	9.9 ± 1.4***	$5.7 \pm 3.6^{***}$	94.2 ± 8.3	$0.7 \pm 0.3^{**}$	$24.4 \pm 3.2^{**}$	$45.2 \pm 4.1^{**}$	291.2 ± 19.3**	268.5 ± 31.2***
400 mg/kg	$11.8 \pm 1.7^{***}$	$6.5 \pm 3.7^{***}$	97.7 ± 7.2	$0.9 \pm 0.2^{***}$	$26.9 \pm 4.3^{***}$	$45.2 \pm 5.2^{***}$	$303.4 \pm 87.7^{***}$	272.1 ± 42.1***
Furosemide	$12.4 \pm 1.2^{***}$	$7.4 \pm 2.3^{***}$	97.3 ± 6.4	$0.7 \pm 0.1^{***}$	$24.6 \pm 4.5^{**}$	45.1 ± 3.3**	$301.2 \pm 84.9^{***}$	273.5 ± 32.1***
HCTZ	7.8 ± 1.6***	$10.8 \pm 3.5^{***}$	95.4 ± 5.2	$0.7 \pm 0.2^{***}$	27.3 ± 3.1***	44.2 ± 2.3***	$295.3 \pm 77.7 ***$	$272.3 \pm 44.2^{***}$

Values are mean \pm SEM, n = 5. ANOVA + LSD test vs. negative control: *P < 0.05, **P < 0.01, ***P < 0.001.





A. Total volume of urine excreted daily by rats following subchronic treatment with the highest dose of *C. occidentalis* aqueous extract tested, administration of an equivalent volume of distilled water (negative controls), or treatment with one of the two diuretic drugs used as positive controls. Note that from the first day of treatment to the last all treatments significantly increased the volume of urine excreted daily, compared to the negative control group. B–D. Estimated amounts of Na⁺ (B), K⁺ (C), and Cl⁻ (D) excreted in the urine by the same rats daily. Note that all treatments increased the urinary excretion of Na⁺ and Cl⁻ from the first day of treatment to the last one, compared to the negative control group, unlike K⁺ excretion, which started to be marked from the fifth day of treatment with the extract, and on the second or fourth day of treatment with the diuretic drugs used in positive control groups. dH₂O, distilled water. ANOVA + LSD test against negative control group: **P* < 0.05, ***P* < 0.01, ****P* < 0.001.

3.2.2. Urinary electrolyte excretion

The estimated amounts of Na⁺ (Figure 4B), Cl⁻ (Figure 4C), and K⁺ (Figure 4D) excreted daily in the urine by rats following subchronic treatment with *C. occidentalis* extract at 400 mg/kg, administration of an equivalent volume of distilled water, or treatment with one of the two diuretic drugs used as positive controls are shown in Figure 4. Na⁺ and Cl⁻ amounts were significantly increased by all these treatments, compared to the negative control group from the first day of treatment. All experimental groups had a significantly higher slope (P > 0.01) than the negative control group (y = 0.14x + 15.9, $R^2 = 0.56$): furosemide (y = 10.6x + 81.9, $R^2 = 0.99$), HCTZ (y = 9.4x + 60.9, $R^2 = 0.98$), and extract-treated groups (y = 12.1x + 154.6, $R^2 = 0.98$). The effects of treatment with extract or diuretic drugs on K⁺ amount excreted was also more marked with time. However, the effect were more marked from two to six days of treatment (Figure 4D): furosemide (y = 6.8x + 33.9, $R^2 = 0.98$), HCTZ (y = 5.8x + 12.1, $R^2 = 0.99$), and extract-treated groups (y = 3.6x + 13.2, $R^2 = 0.97$).

Table 5

Effect of the subchronic administration of C occidentalis aqueous extract on markers of oxidative stress.

	Experimental groups	MDA (µM/100 g of tissue)	ROOH (µM/100 g of tissue)	CAT (mMH ₂ O ₂ /min per g of protein)	Protein (g/100 g of tissue)	Glutathione (mmol/L)
Liver homogenates	dH ₂ O	9.96 ± 1.32 $7.80 \pm 0.86^{*}$	1.37 ± 0.09 $0.76 \pm 0.25^{***}$	0.02 ± 0.02 $0.17 \pm 0.06^{*}$	56.04 ± 0.52 $30.29 \pm 5.18^{**}$	-
Blood plasma	400 mg/kg extract dH ₂ O	20.96 ± 1.50	$0.76 \pm 0.25^{++++}$ 0.03 ± 0.01	-	- 50.29 ± 5.18**	- 0.43 ± 0.04
	400 mg/kg extract	8.10 ± 3.12***	$0.06 \pm 0.01^*$	_	-	$0.24 \pm 0.03^{**}$
Blood pellet	dH ₂ O	-	-	0.01 ± 0.00	46.81 ± 8.01	_
hemolysates	400 mg/kg extract	-	-	$0.13 \pm 0.03^{***}$	$54.81 \pm 0.29*$	-

CAT: catalase. MDA: malondialdehyde. ROOH: hydroperoxide. Values are mean \pm SEM, n = 5. ANOVA + LSD test vs. negative control: *P < 0.05, **P < 0.01, ***P < 0.001.

3.2.3. Oxidative stress markers

Table 5 showed the effects of the subchronic administration of *C. occidentalis* aqueous extract (400 mg/kg) on markers of oxidative stress. Catalase activities in liver homogenates and in hemolysates were significantly decreased (P < 0.05). The extract also induced a significant decrease in hydroperoxide amount in liver homogenates (P < 0.001), and an increase in blood plasma (P < 0.05). Plasma and liver malondialdehyde amounts were significantly decreased (P < 0.05). Glutathione concentration in plasma were decreased (P < 0.01). Protein amounts were decreased in liver homogenates (P < 0.01) and increased in hemolysates (P < 0.05).

3.3. Phytochemical study

Phytochemical screening performed on crude extracts revealed the presence of several primary and secondary metabolites such as fatty acids, anthraquinones, glycosides, anthracenes, saponins, tannins, and coumarins. Phenolic compounds, triterpenes, volatile oils and sterols, but also flavonoids and alkaloids were also present in the extract. Thin layer chromatography showed that the hexane extract and urine of treated rats contained four chemical fractions. These initial observations and findings suggest that the aqueous extract of leaves of *C. occidentalis* contains several chemical compounds which biological potential activity deserves further investigation.

4. Discussion

Results of the present study suggest that the aqueous extract of leaves of C. occidentalis administrated per os had stronger diuretic effects than reference drugs at doses used, both in acute (24 h) and subchronic (7 d) studies. The acute administration of the extract at the dose with the more marked response (400 mg/ kg) induced an increase of 107.58% in urinary excretion (compared to negative control group), against 84.2% and 48.05% with furosemide and HCTZ, respectively. Still in the acute study, the extract also accelerated the elimination of fluid overload and decreased the latency of the first urination (152.43%, at 400 mg/kg, against 114.08% and 42.34% furosemide and HCTZ, respectively) and the diuretic index of groups treated with the extract was higher (2.07 at 400 mg/kg) than furosemide-treated (1.84) and HCTZ -treated (1.48). The pharmacological response of the extract was dose-dependent and comparable results were also obtained in the subchronic study. Notably, the volume of urine excreted increased with the dose of extract, particularly in the first hour after administration. Similar

observations were reported in studies assessing the other plants with diuretic activity such as *Randia echinocarpa* ^[23], and *Ficus glumosa* ^[20]. Such rapid diuretic activity may be due to very high concentration of active molecules of the saponin, flavonoïd and anthraquinone families ^[24], which presence in extracts of *C. occidentalis* was detected by phytochemical analysis in our study and previously reported ^[25].

C. occidentalis extract caused a more marked increase in natriuresis than furosemide and HCTZ compared to the negative control group, more specifically 909.59% at 400 mg/kg against 463.20% and 336.85%, respectively. Furosemide increases urinary excretion of sodium by inhibiting Na⁺/K⁺/2Cl⁻ symporter (co-transporter system) in the thick ascending limb of the Henley loop [20], while HCTZ inhibits the Na⁺/Cl⁻ symporter (cotransporter system) in the distal convoluted tubule, by competing for the Cl⁻ binding site, and increasing the excretion of Na+ and Cl⁻ [20]. Whether the extract induces the suppression of renal tubular reabsorption of water and electrolytes by one of these processes or by another mechanism is still to be determined. However, although during acute testing the extract only induced a strong elimination of Na⁺, as both diuretic reference drugs, K⁺ elimination by extract subchronic treatment became marked only from 6 d of treatment onward, against 2 d for furosemide and 4 d for HCTZ-treated. These observations suggest that C. occidentalis acted as a K⁺ -sparing diuretic [26,27].

C. occidentalis was well tolerated with an encouraging safety profile in subchronic administration. Glucose and albumin were not present in treated rats' urine, and no significant change was observed in the urinary creatinine levels. Instead, a marked reduction was observed in the concentration of urea in the urine compared to negative control group, the K⁺ plasmatic concentration was increased, and Na⁺ and Cl⁻ concentrations in the plasma were significantly decreased. Taken together, these results indicate that *C. occidentalis* may act as a loop diuretic which inhibit the Na⁺/K⁺/Cl⁻ co-transporter system in the thick ascending loop of the nephron, thus increasing natriuresis and kaliuresis [28].

C. occidentalis also caused the acidification of urine. There was a significant reduction in the osmolarity of urine in rats treated with the extract. *C. occidentalis* may impair the basal secretion of ADH and reduce the responsiveness of uriniferous tubules to the action of ADH. Inhibition of ADH causes polyurea with low osmolarity ^[29]. Furthermore, in our study radioimmunoassay revealed a decrease in aldosterone in animals treated with the extract, further suggesting that the stimulation of diuresis by the aqueous extract of the leaves of

C. occidentalis may be comparable to furosemide mechanism. Notably, glomerular filtration measured by creatinine clearance was reduced, together with glomerular filtration rate, probably due to increases in the Na⁺ load available for Na⁺/K⁺ exchange [30,31], which indicates that the increase in diuresis may also originate from changes in glomerular filtration, besides changes at tubular level.

Findings of the study also indicated that *C. occidentalis* may have antioxidant effects. Comparable observations were reported in a number of other plants with diuretic properties [32]. In addition, the extract also decreased hydroperoxide levels in homogenates, malondialdehyde levels in plasma, and the activity of catalase in homogenates and hemolysates, which are markers of oxidative stress [33].

The results of the present study strongly suggest that *C. occidentalis* leaves' aqueous extract have potent and doseresponse diuretic and antioxidant properties, both in acute and subchronic use in rats. These findings justify at least partly the use of this extract in folk medicine for the treatment of hypertension. Future studies aimed at identifying the active principles accounting for these effects of *C. occidentalis* leaves' aqueous extract may lead to the discovery of a potent diuretic, potentially with antioxidant properties.

Conflict of interest statement

We declare that we have no conflict of interest.

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References

- [1] Fonarow GC, Smith EE, Reeves MJ, Pan W, Olson D, Hernandez AF, et al. Hospital-level variation in mortality and rehospitalization for medicare beneficiaries with acute ischemic stroke. *Stroke* 2011; **42**: 159-166.
- [2] Barakat LA, Mahmoud RH. The antiatherogenic, renal protective and immunomodulatory effects of purslane, pumpkin and flax seeds on hypercholesterolemic rats. *N Am J Med Sci* 2011; 3: 411-417.
- [3] Brandi L. 1alpha(OH)D3 One-alpha-hydroxy-cholecalciferol-an active vitamin D analog. Clinical studies on prophylaxis and treatment of secondary hyperparathyroidism in uremic patients on chronic dialysis. *Dan Med Bull* 2008; 55: 186-210.
- [4] Strand V. Are COX-2 inhibitors preferable to non-selective nonsteroidal anti-inflammatory drugs in patients with risk of cardiovascular events taking low-dose aspirin? *Lancet* 2007; 370: 2138-2151.
- [5] Seethapathy GS, Ganesh D, Santhosh Kumar JU, Senthilkumar U, Newmaster SG, Ragupathy S, et al. Assessing product adulteration in natural health products for laxative yielding plants, Cassia, Senna, and Chamaecrista, in Southern India using DNA barcoding. *Int J Leg Med* 2014; http://dx.doi.org/10.1007/s00414-014-1120-z.
- [6] Yadav JP, Arya V, Yadav S, Panghal M, Kumar S, Dhankhar S. *Cassia occidentalis* L.: a review on its ethnobotany, phytochemical and pharmacological profile. *Fitoterapia* 2010; 81: 223-230.

- [7] Bukhari NA, Siddique I, Perveen K, Siddiqui I, Alwahibi MS. Synthetic seed production and physio-biochemical studies in *Cassia angustifolia* Vahl. – a medicinal plant. *Acta Biol Hung* 2014; 65: 355-367.
- [8] Epifano F, Fiorito S, Locatelli M, Taddeo VA, Genovese S. Screening for novel plant sources of prenyloxyanthraquinones: *Senna alexandrina* Mill. and *Aloe vera* (L.) Burm. F. *Nat Prod Res* 2015; 29: 180-184.
- [9] Chen L, Yang Y, Yuan P, Yang Y, Chen K, Jia Q, et al. Immunosuppressive effects of A-type procyanidin oligomers from *Cin*namonum tamala. Evid Based Complement Alternat Med 2014; 2014: 365258.
- [10] Cong Q, Shang M, Dong Q, Liao W, Xiao F, Ding K. Structure and activities of a novel heteroxylan from *Cassia obtusifolia* seeds and its sulfated derivative. *Carbohydr Res* 2014; **393**: 43-50.
- [11] Shao F, Chen HJ, Liu RH, Hou YC, Ren G, Huang HL, et al. Effects of heishunpian total alkaloids on *Cassia acutifolia* induced mice diarrhea and contraction of isolated intestinal smooth muscle in rats. *Zhong Yao Cai* 2013; **36**: 1805-1809.
- [12] Somova LO, Nadar A, Rammanan P, Shode FO. Cardiovascular, antihyperlipidemic and antioxidant effects of oleanolic and ursolic acids in experimental hypertension. *Phytomedicine* 2003; 10: 115-121.
- [13] Nakamura S, Xu F, Ninomiya K, Nakashima S, Oda Y, Morikawa T, et al. Chemical structures and hepatoprotective effects of constituents from *Cassia auriculata* leaves. *Chem Pharm Bull* (*Tokyo*) 2014; 62: 1026-1031.
- [14] Purushotham KN, Annegowda HV, Sathish NK, Ramesh B, Mansor SM. Evaluation of phenolic content and antioxidant potency in various parts of *Cassia auriculata* L.: a traditionally valued plant. *Pak J Biol Sci* 2014; **17**: 41-48.
- [15] Silva CR, Monteiro MR, Rocha HM, Ribeiro AF, Caldeira-de-Araujo A, Leitao AC, et al. Assessment of antimutagenic and genotoxic potential of senna (*Cassia angustifolia* Vahl.) aqueous extract using *in vitro* assays. *Toxicol Vitro* 2008; 22: 212-218.
- [16] Matsuzawa N, Takamura T, Kurita S, Misu H, Ota T, Ando H, et al. Lipid-induced oxidative stress causes steatohepatitis in mice fed an atherogenic diet. *Hepatology* 2007; 46: 1392-1403.
- [17] Wang X, Li Q, Shen L, Yang J, Cheng H, Jiang S, et al. Fumigant, contact, and repellent activities of essential oils against the darkling beetle, Alphitobius diaperinus. *J Insect Sci* 2014; 14: 75.
- [18] Verma L, Singour PK, Chaurasiya PK, Rajak H, Pawar RS, Patil UK. Effect of ethanolic extract of *Cassia occidentalis* Linn. for the management of alloxan-induced diabetic rats. *Pharmacogn Res* 2010; 2: 132-137.
- [19] Sreejith G, Latha PG, Shine VJ, Anuja GI, Suja SR, Sini S, et al. Anti-allergic, anti-inflammatory and anti-lipidperoxidant effects of *Cassia occidentalis* Linn. *Indian J Exp Biol* 2010; 48: 494-498.
- [20] Ntchapda F, Djedouboum A, Kom B, Nana P, Bonabe C, Maguirgue K, et al. Diuretic activity of the aqueous extract leaves of *Ficus glumosa* Del. (Moraceae) in rats. *Sci World J* 2014; 2014: 693803; http://dx.doi.org/10.1155/2014/693803.
- [21] Vogel GH. Drug discovery and evaluation: pharmacological assays. Germany: Springer- Verlag; 2002.
- [22] Trease GE, Evans MC. *Textbook of pharmacognosy*. Tindall, London: Bailliere; 1983.
- [23] Vargas Solis R, Perez Gutierrez RM. Diuretic and urolithiatic activities of the aqueous extract of the fruit of *Randia echinocarpa* on rats. *J Ethnopharmacol* 2002; 83: 145-147.
- [24] Maghrani M, Zeggwagh NA, Haloui M, Eddouks M. Acute diuretic effect of aqueous extract of *Retama raetam* in normal rats. *J Ethnopharmacol* 2005; **99**: 31-35.
- [25] Takahashi M, Sakurai K, Fujii H, Saito K. Identification of indicator components for the discrimination of Cassia plants in health teas and development of analytical method for the components. *J AOAC Int* 2014; **97**: 1195-1201.
- [26] Kreydiyyeh SI, Usta J. Diuretic effect and mechanism of action of parsley. J Ethnopharmacol 2002; 79: 353-357.
- [27] Rang HP, Dale MM, Ritter JM. *Pharmacology*. London: Churchill Livingstone; 1995.

- [28] Ratnasooriya WD, Pieris KP, Samaratunga U, Jayakody JR. Diuretic activity of *Spilanthes acmella* flowers in rats. *J Ethnopharmacol* 2004; **91**: 317-320.
- [29] Osorio FV, Teitelbaum I. Mechanisms of defective hydroosmotic response in chronic renal failure. *J Nephrol* 1997; **10**: 232-237.
- [30] Jouad H, Lacaille-Dubois MA, Eddouks M. Chronic diuretic effect of the water extract of *Spergularia purpurea* in normal rats. *J Ethnopharmacol* 2001; 75: 219-223.
- [31] Jackson EK. Drugs affecting renal and cardiovascular function. In: Hardman JC, Gilman AG, Limbird LE, editors. *Goodman and*

Gilman's the pharmacological basis of therapeutics. New York: Pergamon press; 1996, p. 685-713.

- [32] Gupta RK, Kesari AN, Diwakar S, Tyagi A, Tandon V, Chandra R, et al. *In vivo* evaluation of anti-oxidant and anti-lipidimic potential of *Annona squamosa* aqueous extract in type 2 diabetic models. *J Ethnopharmacol* 2008; **118**: 21-25.
- [33] Ly J, Maquet P. Stroke and aging. *Rev Med Liege* 2014; 69: 315-317.