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## Antidiarrhoeal efficacy of *Mangifera indica* seed kernel on Swiss albino mice

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### ABSTRACT

**Objective:** To examine the antidiarrhoeal activity of alcoholic and aqueous seed kernel extract of *Mangifera indica* (*M. indica*) on castor oil-induced diarrhoeal activity in Swiss albino mice. **Methods:** Mango seed kernels were processed and extracted using alcohol and water. Antidiarrhoeal activity of the extracts were assessed using intestinal motility and faecal score methods. **Results:** Aqueous and alcoholic extracts of *M. indica* significantly reduced intestinal motility and faecal score in Swiss albino mice. **Conclusions:** The present study shows the traditional claim on the use of *M. indica* seed kernel for treating diarrhoea in Southern parts of India.

## 1. Introduction

Gastroenteritis is a clinicopathological term that refers to inflammation of the intestines and results in diarrhoea. Diarrhoeal diseases contribute in a big way to infant and childhood mortality and morbidity in India<sup>[1,2]</sup>. Diarrhoea is an intestinal disorder characterized by abnormal fluidity and frequency of faecal evaluation, generally the result of increased motility in the colon. The diarrhoea is caused by a variety of enteric pathogenic bacteria such as enterotoxigenic *Escherichia coli*, enteropathogenic *Escherichia coli*, enteroinvasive *Escherichia coli*, *Salmonella* sp., *Shigella flexneri*, *Vibrio cholerae*, *Campylobacter jejuni* etc., viruses, protozoans and helminthes can also cause diarrhoea in human<sup>[3–6]</sup>. The incidence of diarrhoeal

still remains high despite the efforts of many government and international organizations to curb it. It is therefore important to identify and evaluate available natural drugs as alternatives to currently used anti-diarrhoeal drugs, which are not always free from adverse effects. A range of medicinal plants with antidiarrhoeal properties is widely used for traditional healers<sup>[7–11]</sup>.

*Mangifera indica* Linn. (family Anacardiaceae) (*M. indica*) is a large evergreen tree reaching a height of 15 m, indigenous to India. Both unripe and ripe fruits are widely used by the local population. There are few studies on the medicinal value of seeds of *M. indica*. *M. indica* along with *Artocarpus inegrefolia* was given to treat dysentery. *M. indica* seed kernel is a promising source of food additive and enhance oxidative stability of food and used as food preservative and feed additive<sup>[12–16]</sup>. Various extracts from the seed kernel, leaves and barks of *M. indica* is active against human pathogens<sup>[17–21]</sup>, antioxidant<sup>[22–26]</sup>, anti-allergic<sup>[27]</sup>, anti-tyrosinase<sup>[28, 29]</sup> anticarcinogenic<sup>[30]</sup> and promote endothelial cell migration<sup>[31]</sup>. The seed of *M. indica* is reported in

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traditional medicine to cure vomiting, dysentery and burning. Decoction of seed kernel is generally prescribed for diarrhea among the rurals of Kanyakumari district, Tamilnadu, India. As *M. indica* is widely available, cheap and commonly used, it is worthwhile to study its potential antidiarrhoeal activity, which if proved would be an easily accessible home remedy for diarrhoea. In the present study, we have analysed the antidiarrhoeal effects of an alcoholic and aqueous extract of seed kernel of *M. indica* on castor oil-induced diarrhoeal activity in mice.

## 2. Materials and methods

### 2.1. Preparation of extracts

The fruits of *M. indica* were purchased from the local market and seeds were separated from the fruit. The hard seed coat (kernel) was removed and the seeds were dried. These dried seeds were coarsely powdered and stored in closed container for further use. Known quantity of powdered seed kernel of *M. indica* was taken and the crude extract was prepared by adding water and 100% of ethyl alcohol. The collected extracts were dried in a vacuum desiccator and stored in a sterile container for further use.

### 2.2. Antidiarrhoeal study

Swiss albino mice (20–35 g) of either sex obtained from King Institute of Preventive Medicine, Guindy, Chennai were used. They were housed in polypropylene cages in the departmental animal house at (26±2)°C for one week before and during the experiments. Fresh dry husks were used as bed material. They were fed commercially with standard pellet diet and distilled water. Food was withdrawn 18–24 h before the experiment though water was allowed *ad libitum*. Mice were divided into seven groups of 6 animals each. Group I: Normal control; Group II: Castor oil control (Disease control); Group III: Castor oil + Lopramide; Group IV: Castor oil + 200 mg/mL Alcoholic extract; Group V: Castor oil + 400 mg/mL Alcoholic extract; Group VI: Castor oil + 200 mg/mL Aqueous extract; Group VII: Castor oil + 400 mg/mL Aqueous extract.

### 2.3. Effect of extract on castor oil induced diarrhoea

First group served as the control and received distilled water. All other 6 groups received castor oil at a dose of 0.1 mL per animal orally. The second group served as castor oil control. Thirty minutes after castor oil administration, the third group received lopramide. The fourth and fifth groups received alcoholic extract of *M. indica* seed kernel (200 mg/kg b.w. & 400 mg/kg b.w. respectively), sixth and seventh

group received aqueous extract of *M. indica* seed kernel (200 mg/kg b.w. & 400 mg/kg b.w. respectively). Following administration, the animals were placed separately in cages with filter paper, which was changed every hour. The total number of faeces and diarrhoeal faeces excreted was recorded for a period of 76 h. The total score of diarrhoeal faeces of diseased control group was considered as 100%.

### 2.4. Effect of extract on intestinal propulsion

Extraction on intestinal propulsion of Swiss albino mice was done by using charcoal meal as a diet marker. First group served as the control and received distilled water. All other 6 groups received castor oil at a dose of 0.1 mL per animal orally. The second group served as castor oil control. 30 min after castor oil administration, the third group received saline. The fourth and fifth groups received alcoholic extract of *M. indica* seed kernel (200 mg/kg b.w. & 400 mg/kg b.w. respectively), sixth and seventh group received aqueous extract of *M. indica* seed kernel (200 mg/kg bw & 400 mg/kg b.w. respectively). Each animal was given 1 mL of charcoal meal orally (3% deactivated charcoal in 10% aqueous feed) after 30 minutes of castor oil administration. All animals were sacrificed after 30 minutes of charcoal meal administration and the distance covered by the charcoal meal in the intestine, from the pylorus to the caecum was measured and expressed as a percentage of distance moved.

## 3. Results

The effect of the extract on castor oil induced diarrhoea in mice showed decrease in the weight of faecal matter passed by the animals. Faecal passage was significantly reduced in aqueous and alcoholic extract of *M. indica* seed kernel treated animals from 74% to 88% when compared to castor oil control on day one and 132%–138% on day three. On day 3 plant extract treated animals reduced faecal score drastically, which is similar to the lopramide treated group (Table 1).

Table 2 revealed the charcoal mobility pattern. Charcoal mobility directly demonstrates the intestinal movement pattern in different groups of animals. Normal animal stomach mobility is slower than other groups (23.5 cm) whereas castor oil induced animals have increased intestinal mobility upto 147%. Lopramide treated animals showed only 7% increase in mobility when compared to normal animals. Animals treated with aqueous and alcoholic extracts from the seed kernel of *M. indica* also showed reduced intestinal mobility. This effect is slightly lower than the effect of lopramide.

**Table 1**Antidiarrhoeal activity of alcoholic and aqueous extract of *M. indica* seed kernel on castor-oil induced diarrhoea.

Groups	Faecal score		
	Day 1	Day 2	Day 3
Group I	3.50±0.96	3.50±1.12	3.80±1.07
Group II	8.30±1.10*	8.20±1.07*	8.50±0.96*
Group III	5.00±1.18*	4.50±0.96*	3.00±1.07*
Group IV	5.70±1.54*	5.00±0.82*	3.50±0.50*
Group V	5.60±0.81*	4.80±0.81*	3.30±0.47*
Group VI	5.40±0.11*	5.40±0.14*	3.85±0.10*
Group VII	5.60±0.12*	4.80±0.21*	3.40±0.48*

Data were expressed as Mean±SD, \*  $P < 0.01$  comparing with the group I.**Table 2**Effect of alcoholic and aqueous extract of *M. indica* seed kernel on small intestinal transit.

Groups	Charcoal mobility(cm)
Group I	23.50±2.07
Group II	58.00±1.75*
Group III	25.20±3.58*
Group IV	29.20±3.18*
Group V	28.00±2.65*
Group VI	29.00±1.78*
Group VII	26.00±1.86*

Data were expressed as Mean±SD, \*  $P < 0.01$  comparing with the group I.

#### 4. Discussion

The seed kernel extract of *M. indica* inhibited the electrolyte permeability in the intestine due to castor oil through the inhibition of prostaglandin release. Castor oil induces intestinal permeability and also stimulates prostaglandins release. Secondary metabolites such as saponin, flavonoid, glycosides, tannins and alkaloids present in the seed kernels of *M. indica* have been implicated as having antidiarrhoeal activity and inhibit the intestinal mobility. Reduction of intestinal mobility may be due to the presence of tannins and tannic acid in the seed kernel extract of *M. indica*. Tannins have been implicated as the bitter principle present in the seed kernel of *M. indica*[32–34]. Dried mango seed contain 15% tannin served as astringent in cases of diarrhea, dysentery, urethritis etc[20].

Plants are tremendous source for discovering new products with medicinal value for drug development. Traditionally tribal people of India consume mango seed kernel in roasted form during starvation as it is rich in starch. Hence it is assumed to be suitable for human consumption. It enhances oxidative stability of food and it is used as a food preservative. Past studies revealed that the mango seed kernels are free from toxic substances and seems to be a safe source of antioxidant[20]. These reports establish the use of *M. indica* as an antidiarrhoeal medicine as claimed

by the traditional medicine.

#### Conflict of interest statement

The author does not have conflict of interest from the present study.

#### References

- [1] Pal BB, Khuntia HK, Samal SK, Kar SK, Patnaik B. Epidemics of severe cholera caused by El Tor *Vibrio cholerae* O1 Ogawa possessing the ctxB gene of the classical biotype in Orissa, India. *Inter J Infect Dis* 2010; **14**(5): e384–e389.
- [2] Krishna BVS, Patil AB, Chandrasekhar MR. Fluoroquinolone-resistant *Vibrio cholerae* isolated during a cholera outbreak in India. *Trans Royal Soc Trop Med Hyg* 2006; **100** (3): 224–226.
- [3] Kitaoka M. Antibiotic resistance mechanisms of *Vibrio cholerae*. *Med Microbiol* 2011; **60**: 397–407.
- [4] Pazhani GP, Ramamurthy T, Mitra U, Bhattacharya SK, Niyogi SK. Species diversity and antimicrobial resistance of *Shigella* spp. isolated between 2001 and 2004 from hospitalized children with diarrhoea in Kolkata (Calcutta), India. *Epidem Infect* 2005; **133**: 1089–1095.
- [5] Rose A, Roy S, Abraham V, Holmgren G, George K, Balraj V, et al. Solar disinfection of water for diarrhoeal prevention in southern India. *Arch Dis Child* 2006; **91**: 139–141.
- [6] Baldi F, Bianco MA, Nardone G, Pilotto A, Zamparo E. Focus on acute diarrhoeal disease. *World J Gastroenterol* 2009; **15**(27): 3341–3348.
- [7] Dash SK, Padhy S. Review on ethnomedicines for diarrhoea diseases from Orissa: prevalence versus culture. *J Human Eco* 2006; **20**(1): 59–64.
- [8] Kiruba S, Jeeva S, Das SSM. Enumeration of ethnoveterinary plants of Cape Comorin, Tamil Nadu. *Indian J Trad Know* 2006; **5**(4): 576–578.
- [9] Jasmine TS, Jeeva S, Febreena GL, Mishra BP, Laloo RC. Wild edible plants of Meghalaya, North-east India. *Nat Prod Rad* 2007;

- 6(5): 410–426.
- [10]Jeeva S. Horticultural potential of wild edible fruits used by the Khasi tribes of Meghalaya. *J Hort For* 2009; **1**(9): 182–192.
- [11]Jeeva S, Kingston C, Kiruba S, Kannan D. Sacred forests—treasure trove of medicinal plants: a case study from south Travancore. In: Trivedi PC. (ed.) *Indigenous medicinal plants*. Jaipur: Pointer Publishers; 2007, p. 262–274.
- [12]Ajila CM, Bhat SG, Prasada Rao UJS. Valuable components of raw and ripe peels from two Indian mango varieties. *Food Chem* 2007; **102**: 1006–1011.
- [13]Mohamed EM, Girgis AY. Utilization of mango seed kernels for improving stability of some oils and biscuit production. *J Agri Sci Mansoura University* 2005; **30**(8): 4625–4636.
- [14]Abdalla AEM, Darwish SM, Ayad EHE, El-Hamahmy RM. Egyptian mango by-product 1. Compositional quality of mango seed kernel. *Food Chem* 2007; **103**: 1134–1140.
- [15]Berardini N, Knodler M, Schieber A, Carle R. Utilization of mango peels as a source of pectin and polyphenolics. *Inno Food Sci Emerg Tech* 2005; **6**(4): 442–452.
- [16]Sahu S, Das BK, Pradhan J, Mohapatra BC, Mishra BK, Sarangi N. Effect of *Mangifera indica* kernel as a feed additive on immunity and resistance to *Aeromonas hydrophila* in Labeo rohita fingerlings. *Fish Shellfish Immun* 2007; **23**: 109–118.
- [17]Engels C, Knodler M, Zhao YY, Carle R, Ganzle MG, Schieber A. Antimicrobial activity of gallotannins isolated from mango (*Mangifera indica* L.) kernels. *J Agric Food Chem* 2009; **57**(17): 7712–7718.
- [18]Engeles C, Ganzle MG, Schieber A. Fractionation of gallotannins from mango (*Mangifera indica* L.) kernels by high-speed counter-current chromatography and determination of their antibacterial activity. *J Agric Food Chem* 2010; **58**(2): 775–780.
- [19]Engels C, Schieber A, Ganzle MG. Inhibitory spectra and modes of antimicrobial action of gallotannins from Mango kernels (*Mangifera indica* L.). *App Environ Microbio* 2011; **77**(7): 2215–2223.
- [20]Sowmiya S, Soundarapandian P, Rajan S. Bioactive studies of *Mangifera indica* against bacteria isolated from urine samples. *Curr Res J Biol Sci* 2009; **1**(3): 139–143.
- [21]Gupta C, Garg AP, Gupta S. Antimicrobial and phytochemical studies of fresh ripe pulp and dried unripe pulp of *Mangifera indica* (AMCHUR). *Middle-East J Sci Res* 2010; **5**(2): 75–80.
- [22]Ribeiro SMR, Barbosa LCA, Queiroz JH, Knodler M, Schieber A. Phenolic compounds and antioxidant capacity of Brazilian mango (*Mangifera indica* L.) varieties. *Food Chem* 2008; **110**(3): 620–626.
- [23]Soong YY, Barlow PJ. Quantification of gallic acid and ellagic acid from longan (*Dimocarpus longan* Lour.) seed and mango (*Mangifera indica* L.) kernel and their effects on antioxidant activity. *Food Chem* 2006; **97**(3): 524–530.
- [24]Nithitanakool S, Pithayanukul P, Bavovada R. Antioxidant and hepatoprotective activities of Thai mango seed kernel extract. *Planta Medica* 2009; **75**(10): 1118–1123.
- [25]Barreto JC, Trevisan MTS, Hull WE, Erben G, de Brito ES, Pfundstein B, et al. Characterization and quantitation of polyphenolic compounds in bark, kernel, leaves, and peel of mango (*Mangifera indica* L.). *J Agric Food Chem* 2008; **56**(14): 5599–5610.
- [26]Kim Y, Lounds-Singleton AJ, Talcott ST. Antioxidant phytochemical and quality changes associated with hot water immersion treatment of mangoes (*Mangifera indica* L.). *Food Chem* 2009; **115**(3): 989–993.
- [27]Rivera DG, Balmaseda IH, León AÁ, Hernández BC, Montiel LM, Garrido GG, et al. Anti-allergic properties of *Mangifera indica* L. extract (Vimang) and contribution of its glucosylxanthone mangiferin. *J Pharm Pharmacol* 2006; **58**(3): 385–392.
- [28]Nithitanakool S, Pithayanukul P, Bavovada R, Saparpakorn P. Molecular docking studies and anti-tyrosinase activity of Thai mango seed kernel extract. *Molecules* 2009; **14**: 257–265.
- [29]Maisuthisakul P, Gordon MH. Antioxidant and tyrosinase inhibitory activity of mango seed kernel by product. *Food Chem* 2009; **117**(2): 332–341.
- [30]Noratto GD, Bertoldi MC, Krenek K, Talcott ST, Stringheta PC, Mertens-Talcott SU. Anticarcinogenic effects of polyphenolics from mango (*Mangifera indica*) varieties. *J Agric Food Chem* 2010; **58**(7): 4104–4112.
- [31]Daud NH, Aung CS, Hewavitharana AK, Wilinon AS, Pierson JT, Roberts-Thomson SJ, et al. Mango extracts and the mango component mangiferin promote endothelial cell migration. *J Agric Food Chem* 2010; **58**(8): 5181–5186.
- [32]Masibo M, He Q. Major mango polyphenols and their potential significance to human health. *Comprehen Rev Food Sci Food Saf* 2008; **7**: 309–319.
- [33]Ezeigbo II, Ezeja MI, Madubuike KG, Ifenkwe DC, Ukwani IA, Udeh NE, et al. Antidiarrhoeal activity of leaf methanolic extract of *Rauwolfia serpentina*. *Asian Pac J Trop Biomed* 2012; **2**(6): 430–432.
- [34]Sharma P, Vidyasagar G, Bhandari A, Singh S, Bhadoriya U, Ghule S, et al. A pharmacological evaluation of antidiarrhoeal activity of leaves extract of *Murraya koenigii* in experimentally induced diarrhoea in rats. *Asian Pac J Trop Dis* 2012; **2**(3): 230–233.