RESEARCH AND REVIEWS: JOURNAL OF MICROBIOLOGY AND BIOTECHNOLOGY

Evaluation of Antimicrobial Effects of Three Medicinal Plants in South of Iran against the Staphylococcus Aureus, Pseudomonas Aeruginosa and Escherichia Coli.

Mustafa Baloch¹*, and Reza Ranjbar².

¹Doctor of Veterinary Medicine, Medical Baghiyatallah University - Molecular Biology Research Center. ²Assistant Professor, Medical Baghiyatallah University - Molecular Biology Research Center.

Research Article

Received: 13/07/2014 Revised: 26/07/2014 Accepted: 29/07/2014

*For Correspondence

Doctor of Veterinary Medicine, Medical Baghiyatallah University -Molecular Biology Research Center.

Keywords: Antibacterial activity, Origanum Vulgare, Mentha longifolia, Ziziphora tenuior, and South of Iran

ABSTRACT

Nowadays, natural treatment of bacterial diseases using substances which are herbal is of utmost importance. In the present study effects of three antibacterial medicinal plants in south of Iran against the separated Staphylococcus aureus, Pseudomonas aeruginosa and Escherichia coli from Clinical samples and standard strains was investigated. Hydro-alcoholic extracts of Ziziphora tenuior, Origanum Vulgare and Mentha longifolia were prepared after being dried at shade. In the following the antibacterial activity of the extracts against separated Staphylococcus aureus, Pseudomonas aeruginosa and Escherichia coli from clinical samples of the Shiraz hospitals and standard samples using disk agar diffusion method was investigated and the drug resistance in the studied samples was conducted using the standard Kirby-Bauer method. The growth of the all studied Staphylococcus aureus and Escherichia coli was stopped by hydro-alcoholic extracts of Mentha longifolia. Ziziphora tenuior and Origanum Vulgare but Pseudomonas aeruginosa showed growth zone of inhibition only against Origanum Vulgare. From statistical viewpoint Origanum Vulgare had also the most antimicrobial effect against Staphylococcus aureus, Pseudomonas aeruginosa and Escherichia coli in the all studied samples (P<0.05). Extracts of Mentha longifolia, Origanum Vulgare and Ziziphora tenuior have a potent antibacterial effect against the gram-positive and gram-negative. But on the nosocomial pathogen. Pseudomonas aeruginosa, only Origanum Vulgare has growth inhibitory effect.

INTRODUCTION

The belief that some plants have the treatment capability goes back many years. Some of these natural products contain Substances that we commonly know them as antimicrobial agents and use them ^[1, 2].On the other hand, strength of bacteria to antibiotics is increasing every day which it causes to think more about replacing effective antimicrobial agents with fewer side effects instead of antimicrobial substance with less and unexpected side effects.

Ziziphora tenuior plants belong to the Lamiaceae family. This plant is a bush with height of 20 to 50 cm and has small leaves, cross, more or less lanceolate form and is sessile. It also has small flowers and perfect which are white, pink and purple. Treatment of digestive disorders such as diarrhea and nausea can be named as its medicinal properties ^[5]. Additionally Ziziphora tenuior has antibacterial effects ^[6, 7], antioxidant ^[8, 9], disinfection of intestine ^[5, 6, and 10], expectorant and anti-cold ^[8].

Origanum Vulgare is a plant of the Labiatae family and series of Plantae ^[11]. This plant is seen in large parts of Europe, especially in South of this continent, North of Africa and large parts of Asia can be seen as well. Also in Iran has been scattered in the most North and North West parts and in the southern warm regions cannot be found. Origanum Vulgare is used in the traditional medicine as an antiseptic, antispasmodic, carminative, anti-worms and is used to eliminate liver and gall bladder pain ^[13, 14, 15].

Marjoram containing one percent essence, major part of it constitutes of phenols, monoterpene hydrocarbons and alcohols. Marjoram essence generally contains 25 compounds such as 26.9% thymol, 40.7% carvacrol, 7.3 percent gamma terpinene ^[13, 14, 16].

Mentha longifolia is one of Labiatae family species, which includes 20 species which are distributed around the world. This medicinal plant is one of the Mint species that typically is known as Pennyroyal. The natural habitat of this plant is in Europe, North of Africa and Asia Minor and the Middle East ^[17]. Aerial parts including Mentha longifolia flowers normally are used as effective substance in treating colds, sinusitis, cholera, food poisoning, inflammation of the bronchi, and tuberculosis as well [18]. This medicinal plant also contains properties such as carminative, expectorant and diuretic, antitussive and anti-menstruation ^[19]. Antimicrobial medicinal plant powder and oil of Mentha longifolia has been determined ^[20].

Methicillin-resistant Staphylococcus aureus (MRSA) is the most common bacteria that spread via infection. This bacterium is found in the skin and other body parts, like nose of people. MRSA bacterium enters the body through a wound or cut. MRSA bacterium resists many antibiotics therefore it is very difficult to treat infections caused by the bacteria. MRSA is one of the most common infections that a person will suffer from during hospitalization ^[21,22]. Pseudomonas aeruginosa is hospital pathogens. This organism is entered to hospital via fruits, vegetables, visitors and patients who are transferred from other wards. Development and dissemination of the disease to other patients via hospital staff hands and direct contact of the patient with infected sources such as contaminated drinking water and food occurs as well ^[23, 24].

Increasing expansion of antibiotic-resistant to Staphylococcus aureus, Pseudomonas aeruginosa and Escherichia species is one of the problems that medicals and veterinaries are dealing with todays. And due to the rise of antibiotic-resistant strains of the bacteria, the number of antibiotics available for treatment of these infections is reduced daily. Pseudomonas aeruginosa and Escherichia cause Sepsis, Wound infections, Gastroenteritis and Neonatal Meningitis in hospitals as well, respectively. Staphylococcus aureus, Pseudomonas aeruginosa and Escherichia are opportunistic nosocomial pathogens ^[25, 26, 27].

Due to the irregular use of antibiotics in the region, the study was performed in order to investigate Antimicrobial effects of three medical plants in south of Iran against the Staphylococcus aureus, Pseudomonas aeruginosa and Escherichia.

METHOD AND MATERIALS

Origanum Vulgare, Mentha longifolia and Ziziphora extraction:

Origanum Vulgare, Mentha longifolia and Ziziphora tenuior plants has been gathered nearby mountains of Shiraz and Kerman in the spring and dried shade outdoors. After drying and cleaning the plant, it has been powdered by electric mill, then per each 100 grams of powdered Origanum Vulgare 1000 CC hydro-alcoholic 50% was added and about 72 hours has been kept in percolator in the lab temperature. After 72 hours the faucet of percolator has been opened and droplets of extract has been gathered and by separator funnel droplets of hydro-alcoholic was added simultaneously until the obtained extract does not have the same color of plant. The obtained extract has been condensed and the dense extract for drying has been kept in Desiccator for 24 hours which per each 100 grams plants, 21 grams dried crystal extract obtained ^[24].

Method for determining the antimicrobial effect

In this study, 60 microbial diseases samples were taken from at Shiraz's hospitals. After diagnosis and separation Staphylococcus aureus, Pseudomonas aeruginosa and Escherichia, determination of their drug resistance was performed by standard Kirby – Bauer method. Simultaneously standard strains

Pseudomonas aeroginosa (PTCC 1430), Staphylococcus aureus (PTCC 1298) and E.coli (ATCC25922) were used.

In order to determine the antimicrobial effects of Origanum Vulgare, Mentha longifolia and Ziziphora tenuior plants, Staphylococcus aureus, Pseudomonas aeruginosa and Escherichia have been grew within 10-15 ml of medium Mueller Hinton Broth (MHB) in sterile condition and have been kept in Incubator at 35-37 centigrade degree for 24 hours till turbidity proportion to 0.5 Mcfarland pipe were obtained.

In the next step the obtained extracts have been injected into 6 mm sterile paper discs (Schleicher No 2668Germany) using 20 micrometer volume sampler under sterile condition ^[30]. By microbial suspension linear growth on the Mueller Hinton agar medium was provided and discs including medical plant extracts have been placed on the medium by sterilized pence and then prepared plates have been kept under 35-37 centigrade degree for 24 hours. After finishing the Incubation time, plates of plant extract discs and also antibiotic discs were brought out from incubator and the diameter of growth inhibition zone around the discs using a ruler were written down.

Statistical analysis of data

The obtained data using SPSS software and variation analysis were analyzed and to investigate the difference among groups, Tukey test was used with signification level p<0.05. The all results were presented by standard deviation.

RESULTS

The mean of the obtained results of Antibacterial effects investigation of Origanum Vulgare, Mentha longifolia and Ziziphora tenuior against separated bacterium from Clinical samples using Agar diffusion method by disc are in tables 1, 2 and 3.

Antibiotics and studied plants	growth inhibition zone diameter	growth inhibition zone diameter (standard)
Penicillin	15	29
Ampicillin	20	29
Gentamicin	14	15
Erythromycin	18	23
Tetracycline	17	19
Chloramphenicol	16	19
Origanum Vulgare (10 micro l/disk)	21	-
Ziziphora tenuior (10 micro l/disk)	16	-
Mentha longifolia (10 micro l/disk)	14	-

Table 2: mean of growth inhibition zone diameter of separated Pseudomonas aeruginosa samples from patients

Antibiotics and studied plants	growth inhibition zone diameter	growth inhibition zone diameter (standard)
Penicillin	0	0
Ampicillin	0	0
Gentamicin	10	15
Erythromycin	0	0
Tetracycline	14	19
Chloramphenicol	15	18
Origanum Vulgare (10 micro l/disk)	10	-
Ziziphora tenuior (10 micro l/disk)	0	-
Mentha longifolia (10 micro l/disk)	0	-

Separated Staphylococcus aureus from clinical samples had antibiotic resistance to penicillin and ampicillin and Origanum Vulgare extract had the most effect (p<0.05). But separated Escherichia coli in addition to penicillin, ampicillin and Erythromycin, was fairly resistant to Ziziphora tenuior. For

Pseudomonas aeruginosa also clinical samples had antibiotic resistance to penicillin, ampicillin, Erythromycin, Mentha longifolia and Ziziphora tenuior extracts and Origanum Vulgare extract had the most growth inhibition effect (p<0.05). The analyzed results of antimicrobial effects investigation of Origanum Vulgare, Mentha longifolia and Ziziphora tenuior plants against standard strains using Agar diffusion method by disc are in tables 4, 5 and 6.

Antibiotics and studied plants	growth inhibition zone diameter	growth inhibition zone diameter (standard)
Penicillin	5	15
Ampicillin	3	17
Gentamicin	14	15
Erythromycin	9	23
Tetracycline	20	18
Chloramphenicol	21	19
Origanum Vulgare (10 micro I/disk)	20	-
Ziziphora tenuior (10 micro I/disk)	14	-
Mentha longifolia (10 micro l/disk)	3	-

Table 3: mean of growth inhibition zone diameter of separated Escherichia coli samples from patients

Table 4: mean of growth inhibition zone diameter of Staphylococcus aureus samples standard strain

Antibiotics and studied plants	growth inhibition zone diameter	growth inhibition zone diameter (standard)
Penicillin	29	29
Ampicillin	29	29
Gentamicin	16	15
Erythromycin	21	23
Tetracycline	20	19
Chloramphenicol	20	19
Origanum Vulgare (10 micro l/disk)	32	-
Ziziphora tenuior (10 micro l/disk)	28	-
Mentha longifolia (10 micro l/disk)	27	-

Table 5: mean of growth inhibition zone diameter of Pseudomonas aeruginosa samples standard strain

Antibiotics and studied plants	growth inhibition zone diameter	growth inhibition zone diameter (standard)
Penicillin	0	0
Ampicillin	0	0
Gentamicin	15	15
Erythromycin	0	0
Tetracycline	20	19
Chloramphenicol	19	18
Origanum Vulgare (10 micro I/disk)	16	-
Ziziphora tenuior (10 micro I/disk)	0	-
Mentha longifolia (10 micro l/disk)	0	-

Table 6: mean of growth inhibition zone diameter of Escherichia coli samples standard strain

Antibiotics and studied plants	growth inhibition zone diameter	growth inhibition zone diameter (standard)
Penicillin	14	15
Ampicillin	17	17
Gentamicin	16	15
Erythromycin	22	23
Tetracycline	20	19
Chloramphenicol	18	18
Origanum Vulgare (10 micro l/disk)	25	-
Ziziphora tenuior (10 micro l/disk)	18	-
Mentha longifolia (10 micro l/disk)	15	-

DISCUSSION

Variety of geographic and climatic conditions has resulted in existence of a diverse and powerful source of plant species in our country. Some of these plants have medical properties such as antibacterial activity ^[2]. In this study antimicrobial effect of three plants (Origanum Vulgare, Mentha longifolia, Ziziphora tenuior) was investigated in south of Iran. The results indicated that the investigated plants had the most growth inhibition effect on studied separated Staphylococcus aureus and Escherichia coli from clinical samples and standard strains respectively. But just Origanum Vulgare had appropriate growth inhibition properties on Pseudomonas aeruginosa. Quance's study in turkey showed the native Ziziphora tenuior of that region has effect on Gram-negative and Gram-positive bacteria but did not has effect on Pseudomonas aeruginosa ^[33]. The result of Salehi and et.al shows that Ziziphora tenuior is able to prevent from Gram-positive bacteria growth which is in agreement with our results. Mentha longifolia has a strong antibacterial effect [35 and 36]. The methanol extract of Mentha longifolia has shown antibacterial and Anti-Fungal effects on a wide range of Gram-negative and Gram-positive bacteria and fungus as well ^[37]. There are typically Peritoneum (60-80%), Beta caryophyllene (5-15%), 1 and 8 Cineol (2-7%) and Hesperidin and quercetin Flavonoids in hydro-alcoholic Mentha longifolia extract ^[38].

In our study Mentha longifolia extract had antibacterial activity on Staphylococcus aureus and Escherichia coli but it did not have effect on Pseudomonas aeruginosa. According to the study of Taba tabaii nejad and et.al in 2002 and the study of Farahani and et.al on the extract and essence of Origanum Vulgare, it did not have significant antibacterial effect on Pseudomonas aeruginosa, is different ^[39,40]. The obtained results indicates that Origanum Vulgare had 10 mm growth inhibition zone diameter in Pseudomonas aeruginosa clinical samples and 16 mm growth inhibition zone diameter in standard Pseudomonas aeruginosa which this difference could depend on the place and the time of gathering the plant, soil of the region and the site of taking bacterial sample. Even though our results show that Origanum Vulgare has the most antibacterial effect on three studied bacteria (Staphylococcus aureus, Pseudomonas aeruginosa, Escherichia coli). So it is proposed to be done more study on Origanum Vulgare, its antimicrobial effect combination, growing place and condition.

ACKNOWLEDGMENT

The authors would like to thank the Department of Microbiology, Faculty of Veterinary Kazeroon, medical Baghiyatallah University, Dr. Ali Ghorbani Ranjbari, Dr. Toba periov and research ward of shahid Beheshty hospital for their support of this study.

REFERENCES

- 1. Rios JL, Recio MC. Medicinal plants and antimicrobial activity. J Ethnopharmacol. 2005;100:80– 84.
- 2. Marjorie MC. Plant Products as Antimicrobial Agents Clin Microb Rev. 1999; 12: 564-582.
- 3. Laxminarayan R, Heymann DL. Challenges of drug resistance in the developing world. BMJ. 2012; 344: e1567.
- 4. Laxminarayan R, Klugman KP. Communicating trends in resistance using a drug resistance index. BMJ Open 2011; 1: e000135.
- 5. Naghibi F, Mosaddegh M, Mohammadi Motamed S, Ghorbani A. Labiatae family in folk medicine in Iran from ethnobotany to phamacology. Iranian J Pharm Res. 2005; 2; 63-79.
- Sonboli A, Mujalili MH, Hadian J, Nejad Ebrahimi S, Yousefaade M. Antibacterial activity and composition of the essential oil of Ziziphora clinopodioides subsp. Bungeana (Juz) Rech ffrom Iran. Z. Natureforsch. 2006; 61: 677-680.
- 7. Economou KD, Oreopoulou V, Thomopoulos CD. Antioxidant activity of some plant extracts of the family labiatae. J Am Chem Soc. 1991; 68: 109-113.
- 8. Salehi P, Sonboli A, Eftekhar F, Nejad Ebrahimi S, Yousefzadi M. Essential oil composition, antibacterial and antioxidant activity of the oil and various extracts of Ziziphora clinopodioides subsp. rigida (Boiss.) Reech ffrom Iran. Biol Pharm Bull. 2005; 28: 1892-1896.
- 9. Konyalioglu S, Qzturk B, Elgin MG. Comparison of chemical compositions and antioxidant activities of the essential oils of two Ziziphora taxa from Anatolia. Phann Biol. 2006; 44: 121-126.
- 10. Ozturk S and Ercisli S. Antibacterial activity and chemical constitutions of Ziziphora clinopodioides. Food Control. 2007; 18: 535-540.
- 11. El-Ashmawy IM, Amal S and Salama OM. Alex J Pharm. 2007; 21: 29-35.

- 12. Bremness L. The Complete Book of Herbs: A Practical Guide to Growing and Using Herbs 5th ed. Studio, Seattle Goodwill, Washington, USA 1994.
- 13. Novak J, Bitsch C, Langbehn J, Pank F, Skoula M, Gotsiou Y and Franz CM. Biochem System Ecol. 2000; 28(7): 697-704.
- 14. Fabio A, Corona A, Forte E and Quaglio P. New Microbiol. 2003; 26, 115-120.
- 15. Hazzit M, Baaliouamer A, Leonor-Faleiro M and Graca MM. J Agric Food Chem. 2006; 54(17); 6314-6321.
- 16. Ernst E and Pittler MH. Herbal Medicine. Med Clin North Am. 2002; 86: 149-61.
- 17. Chalchat, J. C., M. S. Gorunovlc, Z. A. Maksimovlc and S. D. Petrovlc. Essential oil of wild growing Mentha pulegium L from Yugoslavia. J. Essential Oil Res. 2000; 12: 598–600.
- 18. Cuppett, S. L and C. A. Hall. Antioxidant activity of Labiatae. Adv Food Nutr Res. 1998; 42: 245–271.
- 19. Mahboubi, M and G. Haghi. Antimicrobial activity and chemical composition of Mentha pulegium L. essential oil. J Ethnopharmacol. 2008; 119: 325-327.
- 20. Perić, L., D. Žikić and M. Lukić. Aplication of alternative growth promoters in broiler production. Biotechnology in Animal Husbandry, Belgrade-Zemun 2009; 25: 387-397.
- 21. Beam JW, Buckley B. Community-Acquired Methicillin-Resistant Staphylococcus aureus: prevalence and risk Factors. J Athletic Training. 2006; 41(3): 337-340.
- 22. Weichhart T, Horky M. Functional Selection of Vaccine Candidate Peptides from Staphylococcus aureus Whole-Genome Expression Libraries In Vitro. J Inf Immunity. 2003; 71(8): 4633-4641.
- 23. Arora D, Jindal N, Kumar R, Romit. Emerging Antibiotic Resistance In Pseudomonas-A Challenge. Int J Pharm Pharm Sci. 2011;3(2):8284.
- 24. [24] Viedma E, Juan C, Villa J, Barrado L, Orellana MA, Sanz F, et al. VIM-2 producing Multidrug-Resistant Pseudomonas aeruginosa ST175 Clone, Spain. Emerg Infect Dis. 2012; 18(8):1235-1241.
- 25. Hota S, Hirji Z, Stockton K, Lemieux C, Dedier H, Wolfaardt G, et al. Outbreak of multidrug-resistant Pseudomonas aeruginosa colonization and infection secondary to imperfect intensive care unit room design. Infect Control Hosp Epidemiol. 2009;30(1):25-33.
- Lindsay JA. Genomic Variation an Devolution of Staphylococcus aureus. Int J Med Microbiol. 2010; 300(2-3): 98-103.
- 27. Astal ZE. Increasing ciprofloxacin resistance among prevalent urinary tract bacterial isolates in the Gaza Strip. Singapor Med J. 2005;46(9):457-59.
- 28. Younes R.N. Varella A.D. Suffradini I.B. Extract of plants brazilizn, Acta On cologica Brazilian2000; Vol. 20pp: 15-90.
- 29. Vitkauskienė A, et al. Characteristics of carbapenem- resistant Pseudomonas aeruginosa strains in patients with ventilator-associated pneumonia in intensive care units. Medicina (Kaunas). 2011;47(12):652-656.
- 30. NCCLS. Performance Standards for Antimicrobial Susceptibility Testing; Sixteenth Informational Supplement seventh Edition NCCLS document M2-A7. Pennsylvania, Wayne 2006; 26.
- 31. Elgayyar, M. Draughon, F.A. Golden D. A. Antimicrobial activity of essential oils from plants against selected pathogenic and saprophytic microorganisms. J Food Protect. 2001;64:1019-1024.
- 32. Zargari A. Iranian medicinal plants. Tehran university press. Tehran. Iran 1995; (4): 103-104.
- 33. Baser K, Sezik E, Tumen G. Composition of the essential oil of Ziziphora clinopodioides Lam. J Essential Oil Res. 1991; 3(4): 237-239.
- 34. Salehi P, Sonboli A, Eftekhar F, Nejad- Ebrahimi S, Yousefzadi M. Essential oil composition, antibacterial and antioxidant activity of the oil and various extracts of Ziziphora clinopodioides subsp. rigida (BOISS.) RECH. F. from Iran. Biol. Pharm. Bull. 2005; 28: 1892-1896.
- 35. Najifi, P and M. Torki. Performance, blood metabolities and immunocompetance of broiler chicks fed diets included essential oils of medicinal herbs. J Anim Vet Adv. 2010; 9: 1164-1168.
- 36. Mimica-Dukic N, Bozin B, Sokovic M, Mihajlovic B, Matavul JM: Antimicrobial and antioxidant of three Mentha species Essential oils. J Agric Food Chem. 2007;19;55:7879-7885.
- 37. Gulluce M, Sahin F, Sokmen M, Ozaer H, Daferera D, Sokmen A, et al: Antimicrobial and antioxidant properties of the essential oil and methanol extract from Mentha longifolia. ssp. Longifolia. Food Chem. 2007;103:1449-1456.
- 38. La Gow B: PDR for herbal medicine. 3rd Ed. USA: Thamson 2005; 698–779,899–901.
- 39. Tabatabai Nejad, S., Antimicrobial Effect of Zataria Multiflora on Psuedomonas Auruginoza. (2003). The 3rd Congress of Medical Plant, Shahed University, Tehran 2007.
- 40. Farahanikia, B., Khanavi, M., Samadi, N., Janbakhsh, S., and Hadijiakhoondi, A, Antibacterial Activity of Five Thymus Species Essential Oils. The 3rd Congress of Medical Plant, Shahid University, Tehran 2007.