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# Phytochemical evaluation and quantification of beta-sitosterol in geographical variation of *Withania coagulans* Dunal by HPTLC analysis

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#### Abstract

The recent global resurgence of interest in herbal medicines, has led to an increase in demand for herbal drugs and consequently a decline in their quality, particularly due to a lack of adequate evidence proof data for assessing the quality of drug. The dried seeds of Withania coagulans Dunal of Solanaceae family, play a major role in indigenous system of medicine for the treatment of ulcers, dyspepsia, rheumatism, dropsy, etc. Organoleptic parameters are not much reliable in establishing the standards of herbal drugs for which an attempt was made through analytical analysis, providing a more concrete picture regarding the qualitative and quantitative aspects which were widely accepted in the quality assessment of herbal drugs such as TLC and HPTLC studies. In the present study,  $\beta$ -sitosterol has been quantified in methanol and ethyl acetate extracts of Withania coagulans Dunal from different states or regions, showing the variation of  $\beta$ -sitosterol content in the drug due to geographical variation. TLC carried with mobile phase Toluene: Ethyl acetate: Glacial Acetic acid (6:1.5:0.5 (v/v)) on Precoated aluminium silica gel plates (Merck) and densitometric determinations was done at 254 nm. Calibration curve was prepared and the amount of  $\beta$ -sitosterol estimated in the extracts by comparing the respective peak areas with that of the standard. A faster, reliable and sensitive HPTLC method has been developed and validated for the analysis of  $\beta$ -sitosterol in seeds of Withania coagulans. Other parameters studied such as phytochemical screening, morphology, heavy metals, aflatoxin contamination and fluorescence behaviour to lay down the standard for the genuine drug.

Key words: Phytochemical screening, Safety evaluation, HPTLC studies, β-sitosterol, Quantification

# Introduction

The traditional medicines are increasingly solicited through the traditional practitioners and herbalists in the treatment of infectious diseases. Medicinal plants play a vital role for the development of new drugs. The bioactive extract should be standardized on the basis of active compounds (Mathur and

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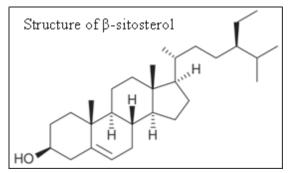
Agrawal, 2011). Herbal drug technology is used for converting botanical materials into medicines, where standardization and quality control with proper integration of modern scientific techniques and traditional knowledge is important. Seeds of *Withania coagulans* Dunal used in Unani System, known as Tukm-e-Hayath which is a small genus of shrubs, popularly known as Indian cheese maker. It is common in Iran, Afghanistan and East India, and also used in folk medicine. It is known differently in various languages such as English - Vegetable rennet; Persian - Arusaka, Paneer-bad; Urdu -Paneerband, Tukme-Hayath; Telugu - Panneru-gadda. It is fairly common in dry hot and stony places up to 1700 m, found in North-west India, Shimla, Punjab, Gujarat, Garhwal

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and Kumaon. Withania coagulans is commercially important for its ability to coagulate milk, in the treatment of ulcers, rheumatism, dropsy, consumption and sensile debility. The seeds are reported to be sedative, emetic and stomachic, a blood purifier and febrifuge, an alternative, diuretic and bitter tonic in dyspepsia as well as a growth promoter in infants (Watt, 1972; Kirtikar and Basu, 1996; Nadkarni, 1976 and Hemlatha et al., 2004). It is well known in the indigenous system of medicine for the treatment of ulcers, dyspepsia, rheumatism, dropsy, consumption and sensile debility (Hemalatha et al., 2008). In indian sub-continent, the berries are used as a blood purifier. The twigs are chewed for cleaning of teeth and the smoke of the plant is inhaled for relief in toothache (Dymock et al., 1972 and Anonymous, 1969). High percentage of  $\beta$ -sitosterol and Linoleic acid are the factors reported to be responsible for the hypocholesterolemic effect (Anonymous, 1982).

 $\beta$ -sitosterol is a phytosterols which is ubiquitous throughout the plant kingdom and reported to possess broad range of biological activities.  $\beta$ -sitosterol is a main phytosterol, found in numerous plants including rice, wheat, corn, nut, peanut etc.  $\beta$ -sitosterol has recorded an amazing health benefits as an hepatoprotective (Shailajan et al., 2005), antioxidant and antipyretic (Ahmed et al., 2001 and Ali, 1967), inflammatory disorders and immunomodulatory (Karl, 1997), antiinflammatory (Gupta et al., 1980), rheumatoid arthritis (Bouic et al., 1996), colon cancer (Awad et al., 2000a), benign prostatic hypertrophy (Awad et al., 2001 and 2000b), and breast cancer (Awad et al., 2000c). Preliminary phytochemical screening, TLC fingerprinting and co-TLC studies (with â-sitosterol) of seeds of Withania coagulans revealed the presence of  $\beta$ -sitosterol and an identical spot as that of standard  $\beta$ -sitosterol was observed. Further, it was confirmed by Rf values comparison and multiwavelength scanning.



Previously,  $\beta$ -sitosterol has been quantified by liquid chromatography and tandem mass spectrometry, using atmospheric pressure photoionization (APPI– LC–MS–MS) (Lembcke, 2005) while there are other reports, using liquid chromatography (LC) with evaporative light scattering detection (ELSD) (Nair and Hoogmartens , 2006), online liquid chromatography- gas chromatography (LC–GC) (Kamm *et al.*, 2002), and gas chromatography (Sorenson *et al.*, 2006).

Considering the wide therapeutic applications of  $\beta$ -sitosterol, as well as an alternative quantification technique of marker constituent was generated to ensure identity and quality of the selected plants. This is a sensitive, specific and reproducible HPTLC method for the quantification of  $\beta$ -sitosterol from the seeds of *Withania coagulans* (Misar *et al.*, 2010).

*Chemical constituents:* Dihydrostigmasterol and  $\beta$ -sitosterol, Withanolides, Esterases, Fatty oil, Essential oil, Triacontane, and Amino acids.

# **Materials and Methods**

#### Collection of material

*Withania coagulans* Dunal seeds were procured from the local market of different states or regions *i.e.*, Andhra Pradesh, Bhopal, Delhi, Aligarh and Mumbai and are authenticated with the help of a Botanist, Dr. V.C. Gupta at Central Research Institute of Unani Medicine before carrying out the study.

All solvents were of HPLC grade.  $\beta$ -sitosterol (98% purity) was procured from Sigma-Aldrich, Bangalore for reference standard.

The present investigation includes parameters such as morphology, physicochemical analysis, TLC and HPTLC fingerprint, phytochemical screening, fluorescence study, safety evaluation, quantification of active principle etc. Physicochemical parameters were determined according to the methods described in 'The Unani Pharmacopoeia of India' (Anonymous, 2009). Fluorescence analysis was carried out as per the method described by Trease and Evans (1972) and GBC-908 AA model Atomic Absorption Spectrophotometer (AAS) was used to determine the concentration of heavy metals. Microbial load and aflatoxins contamination were analyzed as per the methods described in WHO guidelines (Anonymous, 1998). Phytochemical screening was carried out in methanol and ethyl acetate extracts of drug as per the methods described by Trease and Evans (1972) to know the nature of phytoconstituents present in the drug.

# Preparation of the sample drug extract

Five grams of powdered drug each of *Withania coagulans* from different states or regions were macerated in 100 ml of methanol and ethyl acetate separately, in a stoppered 250ml conical flask and was kept for 2 hours while shaking at regular intervals. Later the contents were filtered through Whattmann No. 41 paper and evaporate the solution to 20 ml. The solution, thus, obtained was used as sample for the determination of components.

#### Preparation of the standard solution of $\beta$ -sitosterol

Stock solution (100 $\mu$ g/ml) of  $\beta$ -sitosterol was prepared by dissolving with methanol in 10ml standard volumetric flask

and different aliquots were prepared. For standard drug, methanol is used and for sample drug, methanole and ethyl acetate is used to check in which solvent standard concentration is more.

### Chromatography

HPTLC was performed on 20 cm × 10 cm Precoated Aluminium Sheets of Silica Gel 60  $F_{254}$  (Merck). Samples solution of about 10µl were applied as 6 mm width bands using automatic TLC applicator system of the DESAGA Sarstedt Gruppe (Germany). A linear ascending development with Toluene: Ethyl acetate: Glacial Acetic acid (6:1.5: 0.5) (v/v) as mobile phase was carried out in a twin trough glass chamber previously saturated with mobile phase vapour for 20 min. at room temperature ( $25 \pm 2^{\circ}$ C). The development of solvent distance was 85 mm. After development, plates were airdried. Scanning was performed using densitometer of DESAGA Sarstedt Gruppe (Germany) at 254nm and 366nm wavelength and operated by ProQuant 1.06 version software. The source of radiation was a deuterium lamp emitting a continuous UV spectrum in the range 190-400 nm. The slit dimensions were 4 mm × 6 mm.

## **Development of HPTLC technique**

After the development, TLC plate was then removed dried completely and detected with under UV Cabinet system for detection of spots. Further, it is scanned with the Densitometer CD60 of DESAGA Sarstedt Gruppe system under the UV range of 254nm, and 366nm as shown in the figures. A corresponding densitogram was then obtained in which peaks are appeared for the corresponding spots being detected in the densitometer while scanning and the peaks area under the curve corresponds to the concentration of the component in the sample for the concentration that applied on the TLC plate.

## **Results and Discussion**

## Organoleptic characters

The crude drug consists of the seeds of *Withania coagulans* Dunal. of Solanaceae family, having yellow colour.

#### Morphology of Withania coagulans Dunal

*Macroscopic*: Seeds: Dark brown, ear shaped, glabrous, pulp brown, having yellow, berry, globose, 1.5-1 cm long, 0.7-1 cm width, seeds oval to rounded, yellowish brown, 41-59 in number, 0.1-0.3cm long, 0.2-0.3cm wide, dotted (Figure 1).

*Microscopic*: The T. S. section of seeds shows a single layer of epidermis, followed by a layer of highly flattened thin walled sub epidermal cells. Under the sub-epidermis, there is a layer of highly lignified palisade like cells having narrow lumen. The epidermis of the seed coat inner comprise of 1-2 layer of thin-walled parenchymatous cells which are at places are collapsed showing hyaline-like structure. The endosperm is represented by cells showing strong cellulosic thickening filled with aluerone grains without any globoide. The cotyledon shows thin walled radially elongated cells enclosing a wide zone of round to oval to polyhedral parenchymatous cells.

# Quantitative estimation of $\beta$ -sitosterol by HPTLC analysis

TLC of the  $\beta$ -sitosterol along with methanol and ethyl acetate extracts of *Withania coagulans* developed with the mobile phase solvent system and Rf values for the  $\beta$ -sitosterol is 0.45±0.02 and the corresponding spot at the same Rf value was obtained in the other drug extracts at 254nm and under UV 366nm of chromatogram is shown in the Figure 4 and an overlay of densitometric scan of *Withania coagulans* Dunal, collected from different regions and  $\beta$ -sitosterol in fluorescence 254nm was shown in Figure 2.

The Rf value  $0.45\pm0.02$  of  $\beta$ -sitosterol is in correspndance with all the spots at the same Rf value in other drug extracts and were tabulated with respect to position, area percentage and the  $\beta$ -sitosterol concentration corresponds to each peak in the solution applied on the TLC plate are shown in the Table 7 in drugs extracts of different states or region. Further, in Table 8, amount of  $\beta$ -sitosterol in g% with respect to drug is mentioned. A graph illustrated for the amount of  $\beta$ -sitosterol in g% with respect to *Withania coagulans* Dunal, collected from different states or regions has been depicted in the Figure 3.



Figure 1: Macroscopic feature of seeds of Withania coagulans Dunal

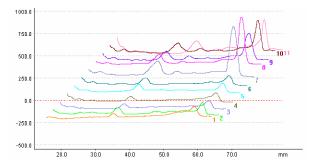


Figure 2: An overlay of densitometric scan of seeds of Withania coagulans Dunal collected from different regions and β-sitosterol in fluorescence at 254nm

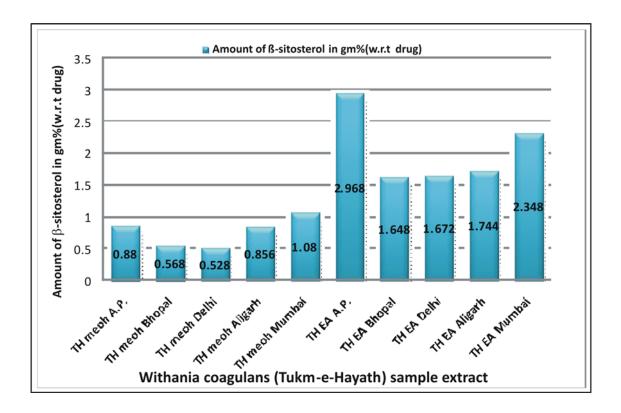
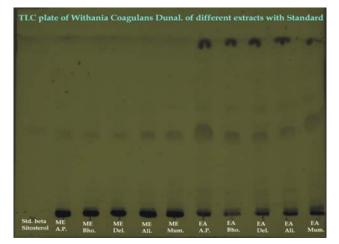


Figure 3: A graph showing the amount of  $\beta$ -sitosterol in g% with respect to Withania coagulans Dunal collected from different states or regions

TLC plate of Tukhme-Hayath of different extracts at U.V. 254nm



TLC plate of Tukhme-Hayath of different extracts at U.V. 366nm

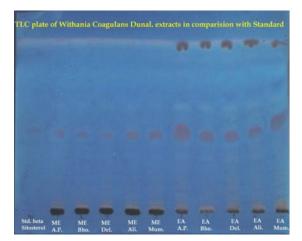


Figure 4: TLC chromatogram of methanol and ethyl acetate seed extracts of *Withania coagulans* Dunal collected from different regions and β-sitosterol at UV 254nm and 366nm

#### **Physicochemical studies**

Physicochemical data of *Withania coagulans* such as total ash, acid insoluble ash, water soluble ash, alcohol soluble

matter, water soluble matter, pH of 1% and 10% aqueous solution, loss of weight on drying at 105°C and volatile oil are summarized in Table 1.

| S.No. | Parameter                         | Results found    |
|-------|-----------------------------------|------------------|
| 1.    | Total ash                         | 3.95 - 4.803g%   |
|       | Acid insoluble ash                | 0.04 - 0.06 g%   |
|       | Water soluble ash                 | 1.58 - 1.93 g%   |
| 2.    | Alc. sol .matter                  | 10.6 - 11.12 g%  |
|       | water sol. matter                 | 25.52 - 26.58 g% |
| 3.    | pH values                         |                  |
|       | a. 1% Aqueous solution            | 4.36-4.43        |
|       | b. 10% Aqueous solution           | 4.39 - 4.41      |
| 4.    | Loss of weight on drying at 105°C | 9.17 g%          |
| 6.    | Moisture content                  |                  |
|       | (by Karlfischer Titrator Method)  | 0.3301%          |
| 7.    | Volatile oil                      | Nil              |

Table 1: The physicochemical parameters data expressed here as mean values of the three readings calculated

Phytochemical screening of *Withania coagulans* Dunal was carried out in different solvent such as ethanol, methanol, chloroform, aqueous, ethyl acetate, petroleum ether extracts to know the nature of compounds present. Qualitative tests for phytoconstituents were carried out for alkaloids, carbohydrates, resin, glycosides, phenols, saponins, proteins, starch, steroids, tannins and flavonoids which are tabulated in Table 2.

| S.No. | Phytoconstituents                  | Ethanol | Methanol | Chloroform | Aqueous | Ethyl acetate | Pet ether |
|-------|------------------------------------|---------|----------|------------|---------|---------------|-----------|
|       | Alkaloids                          | +       | +        | +          | +       | +             | +         |
|       | 1. Dragendroff's reagent           | +       | +        | +          | +       | +             | +         |
| 1.    | 2. Mayer's test                    | +       | +        | +          | +       | +             | +         |
|       | Carbohydrates                      | +       | +        | -          | +       | +             | -         |
|       | 1.Benedict's test                  |         |          |            |         |               |           |
|       | 2. Molisch's test                  | +       | +        | -          | +       | +             | -         |
| 2.    | Resinifiedvolatile oils            | -       | -        | -          | -       | -             | -         |
| 3.    | Glycosides                         | +       | +        | +          | +       | +             | +         |
| 4.    | Phenols: 1. FeCl <sub>3</sub> test | +       | +        | +          | +       | +             | +         |
| 5.    | Saponins                           | +       | +        | -          | +       | +             | +         |
| 6.    | Proteins: 1. Millon's test         | +       | +        | -          | +       | +             | +         |
| 7.    | Starch                             | +       | +        | -          | +       | +             | +         |
| 8.    | Phytosterols(Steroids)             |         |          |            |         |               |           |
|       | 1. Salkowski reactiontest          | +       | +        | +          | +       | +             | +         |
| 9.    | Tannins: 1. Ferric chloride test:  | +       | +        | +          | +       | -             | +         |
| 10.   | Flavanoids: 1. Shinoda test        | +       | +        | +          | +       | +             | +         |

Table 2: Phytochemical screening of the nature of compounds present in different solvent extracts of Withania coagulans Dunal

Fluorescence analysis of powdered drug reaction with chemicals ordinary light and UV light were observed and reported in Table 3. Fluorescence analysis of powdered drug extracts in different solvents were observed and reported in Table 4. Heavy metals and aflatoxins contamination along with their permissible limits were given in Tables 5 and Table 6, respectively.  $\beta$ -sitosterol detection in correspondence with

all the spots at the same Rf value in other drug extracts were tabulated with respect to position, area percentage and the  $\beta$ -sitosterol concentration corresponds to each peak in the solution applied on the TLC plate are shown in the Table 7 in drugs extracts of different states or region. Further, in Table 8 amount of  $\beta$ -sitosterol in g% with respect to drug is mentioned.

Table 3: Fluorescence analysis of powdered drug

|      |   | UV light    |                |               |
|------|---|-------------|----------------|---------------|
| S.No | Reagents                                  | Short 254nm | Long 366nm     | Visible light |
| 1.   | Powder as such                            | Brown       | Black          | Dark brown    |
| 2.   | Powder treated with 1N NaOH in methanol   | Black       | Dark brown     | Dark brown    |
| 3.   | Powder treated with 1N NaOH in water      | Black       | Dark brown     | Dark brown    |
| 4.   | Powder treated with 1N HCl                | Black       | Light brown    | Black         |
| 5.   | Powder treated with 50% $HNO_3$ aqueous   | Black       | Blackish green | Dark brown    |
| 6.   | Powder treated with 50% $H_2SO_4$ aqueous | Black       | Black          | Black         |
| 7.   | Powder treated with glacial acetic acid   | Black       | Pale yellow    | Dark brown    |

Table 4: Fluorescence analysis of powdered drug extracts in different solvents

|       |                         | UV light       |             |               |
|-------|-------------------------|----------------|-------------|---------------|
| S.No. | Extraction solvent      | Short 254nm    | Long 366nm  | Visible light |
| 1.    | Acetone extract         | Black          | Pale yellow | Yellow        |
| 2.    | Alcoholic extract       | Greenish black | Light green | Light brown   |
| 3.    | Chloroform extract      | Black          | Pale yellow | Brown         |
| 4.    | Petroleum ether extract | Black          | Pale yellow | Yellow        |
| 5.    | Methanol extract        | Greenish black | Pale yellow | Dark brown    |
| 6.    | Ethyl acetate extract   | Greenish black | Light blue  | Yellow        |
| 7.    | Aqueous extract         | Black          | Pale yellow | Dark brown    |

Table 5: Heavy metal analysis

| S.No. | Parameter analyzed | Results | Permissible limits as per WHO |
|-------|--------------------|---------|-------------------------------|
| 1     | Arsenic            | Nil     | Not more than 3.0 ppm         |
| 2     | Cadmium            | Nil     | Not more than 0.3 ppm         |
| 3     | Lead               | Nil     | Not more than 10.0 ppm        |
| 4     | Mercury            | Nil     | Not more than 1.0 ppm         |

Table 6: Aflatoxin contamination

| S.No | Parameter analyzed | Results | Permissible limits as per WHO |
|------|--------------------|---------|-------------------------------|
| 1    | B1                 | Nil     | Not more than 0.50 ppm        |
| 2    | B2                 | Nil     | Not more than 0.10 ppm        |
| 3    | Gl                 | Nil     | Not more than 0.50 ppm        |
| 4    | G2                 | Nil     | Not more than 0.10 ppm        |

| Sample Name     | Y-Pos   | Area (%) | Concentration |
|-----------------|---------|----------|---------------|
| TH meoh A.P.    | 40.8 mm | 156.608  | 22.0 µg       |
| TH meoh Bhopal  | 40.3 mm | 105.676  | 14.2 µg       |
| TH meoh Delhi   | 40.0 mm | 99.625   | 13.2 µg       |
| TH meoh Aligarh | 40.4 mm | 155.310  | 21.4 µg       |
| TH meoh Mumbai  | 40.3 mm | 169.451  | 27.0 µg       |
| TH EA A.P.      | 42.5 mm | 346.539  | 74.2 µg       |
| TH EA Bhopal    | 40.1 mm | 236.844  | 42.1 µg       |
| TH EA Delhi     | 40.1 mm | 230.391  | 41.8 µg       |
| TH EA Aligarh   | 42.1 mm | 228.992  | 43.6 µg       |
| TH EA Mumbai    | 42.9 mm | 291.142  | 58.7 μg       |

**Table 7:** Concentration of  $\beta$ -sitosterol in  $\mu$ g present in different solvent extracts of *Withania coagulans* Dunal collected from different states or regions

**Table 8:** Amount of  $\beta$ -sitosterol in g%(w.r.t drug)present in different solvent extracts of *Withania coagulans* Dunal collected from different states or regions

| Sample name     | Concentration | Amount of β-sitosterol<br>in gm%(w.r.t drug) |
|-----------------|---------------|--|
| TH meoh A.P.    | 22.0 µg       | 0.880 %                                      |
| TH meoh Bhopal  | 14.2 µg       | 0.568 %                                      |
| TH meoh Delhi   | 13.2 µg       | 0.528 %                                      |
| TH meoh Aligarh | 21.4 µg       | 0.856 %                                      |
| TH meoh Mumbai  | 27.0 µg       | 1.080 %                                      |
| TH EA A.P.      | 74.2 µg       | 2.968 %                                      |
| TH EA Bhopal    | 42.1 µg       | 1.648 %                                      |
| TH EA Delhi     | 41.8 µg       | 1.672 %                                      |
| TH EA Aligarh   | 43.6µg        | 1.744 %                                      |
| TH EA Mumbai    | 58.7 µg       | 2.348 %                                      |

# Conclusion

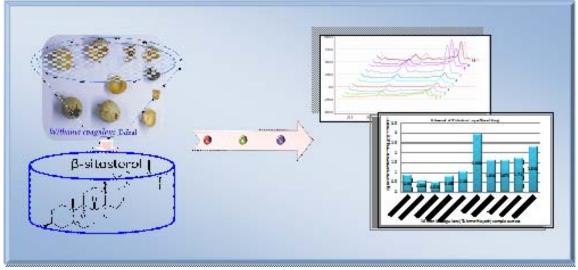
The drug under study was subjected for physicochemical analysis, which is very much supportive in establishing the standard along with the other parameters such as macroscopic, microscopic, fluorescence behavior as reported in the present investigation including heavy metals, aflatoxins contamination found nil, allowing in the permissible limits of WHO. HPTLC studies were thoroughly studied in methanol and ethyl acetate extract and  $\beta$ -sitosterol successfully quantified and observed the variation of content with respect to the drug. Amount of  $\beta$ -sitosterol content found maximum in ethylacetate extract of A.P. with 2.96 g%. On the basis of these data, the drug was broughtup in determining and

ascertaining its quality and standardization of drug. Thus, the study is likely to help in the quality assurance of drug with HPTLC technique which is cost effective, less time consuming and fast analysis to lead formulation with the more therapeutic efficacy due to chemical constituents present in it.

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## **Graphical abstract**



- A faster, reliable and sensitive HPTLC method has been developed and validated for the analysis of β-sitosterol in seeds of Withania coagulans Dunal.
- Standardization and quantification of  $\beta$ -sitosterol.
- Used as a reference standard in quality control of *Withania coagulans* in marketed samples. Active marker species content can be compared within the market products.

## References

Mathur, D. and Agrawal, R.C. (2011). *Withania coagulans*: a review on the morphological and pharmacological properties of the shrub. World Journal of Science and Technology, 1(10): 30-37.

Watt, G.A. (1972). "Dictionary of the economic products of India"; Cosmo Publication, Delhi, India; Vol.6, pp: 309.

Kirtikar, K.R. and Basu, B.D. (1996). Indian Medicinal Plants, III, International Book Distributors, Allahabad, pp: 22-47.

Nadkarni, K.M. (1976). Indian Materia Medica, Vol 1, III ed., pp: 1241-1291

Hemlatha, S.; Wahi, A.K.; Singh, P.N. and Chaurasia, J.P.N. (2004). Hypoglycemic activity of *Withania coagulans* Dunal in Streptozotocin induced diabetic rats. J. Ethnopharmacol., **93**:261.

Hemalatha, S.; Kumar, R. and Kumar, M. (2008). *Withania coagulans* Dunal: A Review. Phcog. Rev., **2**: 351-358.

Dymock, W.; Waden C.J.H. and Hopper, D. (1972). Pharmacographia Indica, reprinted by Institute of Health and TB Research, Karachi. pp:306.

Anonymous (1969). *The Wealth of India*. Publication Information Directorate, CSIR, New Delhi. pp:582.

Anonymous (1982). "The Wealth of India", PID, CSIR, New Delhi, Volume X, pp:581.

Shailajan, S.; Naresh, C.; Sane R.T. and Sasikumar, M. (2005). Effect of *Asteracantha longifolia* Nees. against  $CCl_4$  induced liver dysfunction in rat. Indian Journal of Experimental Biology, **43**:68-75.

Ahmed, S.; Rahman, A.; Mathur, M.; Athar, M. and Sultana, S. (2001). Antitumor promoting activity of *Asteracantha longifolia* against experimental hepatocarcinogenesis in rats. Food and Chemical Toxicology, **39**:19-28. Ali, M.A. (1967). Chemical investigation on the seeds of *Hygrophila spinosa*. Pakistan Journal of Industrial Research, **10**:82-83.

Karl, H.P. (1997). The importance of sitosterol and sitosterolin in human and animal nutrition. South African Journal of Science, **93**:263-268.

Gupta, M.B.; Nath, R.; Srivastava, N.; Kishor, K. and Bhargava, K.P. (1980). Anti-inflammatory and antipyretic activities of β-sitosterol. Planta Medica, **39**:157-163.

Bouic, P.J.D.; Etsebeth, S.; Liebenberg, R.W.; Albrecht, C.F.; Pegel, K. and Van Jaarsveld P.P. (1996). The effect of Beta-sitosterol and beta-sitosterol glucoside stimulate human peripheral blood lymphocyte proliferation: Implications for their use as an immunomodulatory vitamin combination. International Journal of Immunopharmacology, **1**:693-700.

Awad, A.B.; Chan, K.C.; Downie, A.C. and Fink, C.S. (2000a). Phytosterols as Anticancer Dietary Components: Evidence and Mechanism of Action, Nutrition and Cancer, **36**:74-78.

Awad, A.B.; Williams, H. and Fink, C.S. (2001). Phytosterols reduce *in vitro* metastatic ability of MDA-MB-231 human breast cancer cells. Nutrition and Cancer, **40**(2):157-64.

Awad, A.B.; Downie, A.; Fink, C.S. and Kim, U. (2000b). Dietary phytosterol inhibits the growth and metastasis of MDA-MB-231 human breast cancer cells grown in SCID mice. Anticancer Research, **20**:821-824.

Awad, A.B. and Fink, C.S. (2000c). Phytosterols as anticancer dietary components: Evidence and mechanism of action. Journal of Nutrition, **130**:21-27.

Lembcke, J.; Ceglarek, U.; Fiedler, G.M.; Baumann, S.; Leichtle, A. and Thiery, J. (2005). Accumulation of Dietary Cholesterol in Sitosterolemia Caused by Mutations in Adjacent ABC Transporters. Journal of Lipid Research, **46**:21-26.

Nair, V.D.P.; Kanfer, I. and Hoogmartens, J. (2006). "Determination of stigmasterol, beta-sitosterol and stigmastanol in oral dosage forms using HPLC with evaporative light scattering detection". Journal of Pharmaceutical and Biomedical Analysis, **41**(3):731-737.

Kamm, W.; Dionisi, F.; Hischenhuber, C.; Schmarr, H.G. and Engel, K.H. (2002). Rapid detection of vegetable oils in milk fat by on-line LC-GC analysis of â-sitosterol as marker. European Journal of Lipid Science and Technology, **104**(11):756-761.

Sorenson, W.R. and Sullivan, D. (2006). Determination of Campesterol, Stigmasterol, and beta-Sitosterol in Saw Palmetto Raw Materials and Dietary Supplements by Gas Chromatography: Single-Laboratory Validation. Journal of AOAC International, **89**(1):22-34.

Misar, A.; Mujumdar, A.M.; Ruikar, A. and Deshpande, N.R. (2010). Quantification of  $\beta$ - sitosterol from barks of three *Acacia* species by HPTLC. Journal of Pharmacy Research, **3**(11):2595-2596.

Anonymous (2009). The Unani Pharmacopoeia of India, Part-I, Vol.VI, Ministry of Health and Family Welfare, Govt. of India, New Delhi, pp:119-135.

Trease, G.E. and Evans, W.C. (1972). Pharmacognosy  $10^{\rm th}$  edn. Edn. Bailliere Tindel, London.

Anonymous (1998). Quality Control Methods for Medicinal Plant Materials. World Health Organization, Geneva, pp: 25-28.