

Role of microRNA in the regulation of mitochondrial functions

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Abstract

Mitochondria supply eukaryotic cells with energy and perform other essential cellular functions. Mitochondrial functions are regulated by the products of both nuclear and mitochondrial genomes, which include micro (mi)RNAs, the 18- to 24-nucleotide non-coding RNAs that provide post-transcriptional inhibition of target gene expression. Recent studies have shown that miRNAs are expressed in mitochondria and regulate mitochondrial energy metabolism, apoptosis, and biogenesis. This review discusses the role of miRNAs in the regulation of mitochondrial functions. We firstly provide an overview of miRNA expression in mitochondria and regulation of mitochondrial metabolism. This review also addresses the role of miRNAs in the mitochondrial apoptosis pathway and mitochondrial dynamic equilibrium. The overview of the current state of knowledge regarding the role of miRNAs in mitochondrial autophagy and cancer will provide basis for the future research.

Keywords: Mitochondria, miRNAs, mitochondrial equilibrium, metabolism.

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Introduction

Mitochondria provide the cell with energy, and also participate in oxidative phosphorylation reactions, regulate intracellular calcium balance, act as a trigger for apoptosis, and modulate other basic cellular processes [1, 2]. Mitochondrial function is regulated by the products of both nuclear and mitochondrial genomes. The latter encodes 37 genes, two ribosomal (r)RNAs, 22 transfer (t)RNAs, and 13 mitochondrial oxidative phosphorylation complex subunits. Most mitochondrial proteins (~98%) including those involved in mitochondrial DNA replication, transcription, translation, are encoded by nuclear genes and transported to mitochondria.

Micro (mi)RNAs are conserved, 18- to 24-nucleotide (nt) non-coding RNAs with temporally and spatially restricted expression patterns that bind to specific sites in the 3' untranslated region (UTR) of mRNAs, thereby targeting them for degradation or inhibiting their translation and thus providing negative regulation of target gene expression at the post-transcriptional level [3, 4]. MiRNAs regulate cell proliferation, differentiation, apoptosis, fat metabolism, and oxidative stress, and are implicated in normal physiological processes as well as human diseases. Recent studies have shown that miRNAs are expressed in mitochondria and regulate mitochondrial energy metabolism, apoptosis, and biogenesis [5-7].

miRNA expression in mitochondria

The miRNAs are encoded by nuclear DNA and are cut into small fragments in the cytoplasm before being transported to mitochondria by an unknown

mechanism. A microarray analysis of highly purified mitochondria showed enrichment of specific miRNAs [5]. The number of miRNA-specific mitochondria varies by tissue and cell type; for instance, the adult mouse liver expresses 15 mitochondrial miRNAs, including miR-122, -805, and -609 [6], whereas mitochondria in human muscle cells contain more than 20 miRNAs [8]. HeLa cells express 13 miRNAs, of which three have miR-1947, -177, and -1978, target the mitochondrial tRNA genes *TRNE* and *TRNN* and the rRNA gene *RNR1* [9]. We have previously shown that the renal cortex of a normal mouse expresses several mitochondrial miRNAs that were present in the lumen at the concentrations > 50-fold higher or, in other cases, 30-fold lower than in the cytoplasm. *In situ* hybridization experiments have shown that pre-mir-302a and pre-let-7b expressed in mitochondria are encoded by the mitochondrial genome [10]. A group of miRNAs known as myo-miRNAs, miR-181a, -16-1, -133a, -181b, and -206 were found to promote muscle cell proliferation [11, 12], although those are encoded by the nuclear rather than the mitochondrial genome [9].

In addition to miRNAs, key components of miRNA regulation are present in mitochondria. MiRNA regulation of target gene expression is achieved by complementary binding of target mRNAs with the RNA-induced silencing complex, of which the key component Argonaute (Ago)2 [13] is present in mitochondria [6, 9]. MiR-181c, which is encoded by the nuclear genome, regulates the expression of the mitochondrial gene *mt-cytochrome*

C oxidase (COX)I and thereby modulates energy metabolism. Berray et al. identified potential target loci for 169 miRNAs in the mitochondrial genome; for instance, NADH dehydrogenase (*ND1*), 4, 4L, and 6, *cytochrome B*, and *COX1* each have dozens of miRNA-binding sites, whereas *let-7b* targets ATP synthase F0 subunits 6 and 8, COX2, and ND5 [14], substantiating the “many-to-one and one-to-many” rule of miRNA-mRNA interaction [8]

A link between miRNAs and mitochondrial dysfunction has been established in animal models. For example, in a mouse model of streptozotocin-induced type I diabetes mellitus, which is associated with mitochondrial dysfunction, the expression of the miRNAs miR-494, -202-5p, -134, and -155 in hepatic mitochondria was significantly upregulated while that of miR-705 and -122 was reduced with respect to healthy control mice. Yuan et al. in a study found that aldosterone-induced kidney damage in mice occurs simultaneously with mitochondrial dysfunction and is accompanied by a significant change (up- or down regulation) of renal cortex mitochondrial miRNA [15]. These lines of evidence implicate miRNAs in mitochondrial dysfunction, although the detailed mechanisms remain to be elucidated.

miRNA regulation of mitochondrial metabolism

Mitochondria are the main site of oxidative metabolism of sugars, fats, and amino acids in eukaryotic cells, and provide energy for the cell by generating ATP through oxidative phosphorylation reactions. The majority of mitochondrial proteins is encoded by the nuclear genome [16]. COXIV is a component of mitochondrial respiratory chain that is encoded by nuclear DNA and is involved in the mitochondrial synthesis of ATP; changes in COXIV expression level can therefore affect mitochondrial function. It has been reported that miR-338, a neuron-specific miRNA, can bind the 3' UTR of the *COXIV* gene and inhibit expression of the mRNA and protein; therefore, suppressing the endogenous expression of miRNA-338 improves mitochondrial oxygen consumption and activity and ATP production [17].

In mitochondria, glutamine is converted to glutamic acid, which then enters the Krebs's cycle. MiR-23a and -23b inhibit mitochondrial function by suppressing glutaminase expression [18]. Other studies have shown that MDA-MB231, MCF7, HT29, and HCT116 human tumor cells express high levels of miR-210 under hypoxic conditions [19], and that miR-210 inhibition of iron-sulfur enzyme cluster

expression blocks the mitochondrial respiratory chain [20].

Some miRNAs regulate the expression, biosynthesis, and secretion of insulin [21]. In mouse pancreatic β -cells, miR-15a inhibits the expression of endogenous uncoupling protein 2, an inner mitochondrial membrane transport protein that promotes the synthesis of insulin and can uncouple electron transport and phosphorylation, thereby impeding ATP production [22]. Insulin resistance can lead to mitochondrial dysfunction, although the underlying mechanisms are poorly understood [23]. It was recently reported that miR-126 targets insulin receptor substrate-1 to alter insulin resistance. These findings indicate that miRNAs should be explored as potential therapeutic agents in the treatment of diabetes mellitus.

MiR-696 regulates fatty acids metabolism and mitochondrial biogenesis by targeting peroxisome proliferator-activated receptor-gamma co-activator 1- α [10], which promotes aerobic metabolism and mitochondrial function in skeletal muscle. When overexpressed, miR-696 inhibits fatty acid oxidation and reduces mitochondrial DNA copy number, whereas inhibiting miR-696 has the opposite effect [10, 24].

Role of miRNAs in the mitochondrial apoptosis pathway

Apoptosis is a tightly regulated process of cell death that plays an important role in cell growth, development, and differentiation as well as in pathological states. Apoptosis is triggered by extrinsic and intrinsic factors; the former involves tumor necrosis factor α and the binding of Fas ligand to its receptor, which is mediated by Fas-associated death domain proteins. This leads to the accumulation of procaspase-8 in the cytoplasm and the formation of the death-inducing signaling complex [25]. The intrinsic or mitochondrial apoptotic pathway involves DNA damage-induced activation of tumor suppressor genes such as *p53*, which in turn stimulates the expression of pro-apoptotic molecules of the B cell lymphoma (*Bcl*-2 family, such as *Bcl*-2-associated X protein and *Bcl*-2-associated death promoter, thereby inducing the release of pro-apoptotic molecules [26] such as cytochrome C [27] and second mitochondria-derived activator of caspase and direct IAP-binding protein with low PI, also known as Smac and DIABLO, respectively [28]. Cytochrome c, cytoplasmic apoptotic protease-activating factor 1, and procaspase-9, mediate apoptosis by activating

caspase-9 which, along with caspase-8 induces apoptosis.

MiR-15a and -16-1 regulate the mitochondrial apoptotic pathway by activating oncogenes such as Bcl-2 and myeloid cell leukemia 1 and inducing cytochrome c release from mitochondria, thereby disrupting mitochondrial membrane stability and function [29]. Under normal circumstances, miR-143 is specifically expressed by colon cells, but the level is reduced in colon cancer patients. MiR-143 targets extracellular signal-regulate kinase 5 [30] to modulate mitochondrial function and trigger apoptosis [31]. The miR-1 is expressed in skeletal muscle cells; when overexpressed, it triggers the release of cytochrome c and undermines mitochondrial membrane stability, leading to apoptosis [32].

miRNA regulation of mitochondrial dynamic equilibrium

Mitochondrial biogenesis comprising fusion and fission is a dynamic process that occurs throughout the life cycle of a cell, ensuring the stability of mitochondrial structure and function [33]. Biogenesis involves mitochondrial DNA replication and an increase in mitochondrial matrix volume; however, mitochondrial DNA is not replicated during the fission process, which ultimately generates small mitochondria that later mature [33, 34]. Mitochondrial fusion and fission inhibit and trigger apoptosis, respectively [35], while excessive fission is implicated in diseases such as diabetic nephropathy and brain and skeletal muscle disorders [36–38].

The miRNAs also regulate mitochondrial structure. For instance, stimulating myocardial cells with hydrogen peroxide reduces the expression of three members of the miR-30 family, miR-30a, -30b, and -30d that are highly expressed in the heart. Dynamin-related protein (DRP)1 plays a key role in mitochondrial fission, specifically in the fragmentation of the mitochondrial outer membrane; *p53*, a target gene of the miR-30 family, promotes *Drp1* transcription while triggering apoptosis. Mitochondrial fission and apoptosis are thus regulated via targeting of P53 and DRP1 by miR-30 members [39]. The miR-499, which is encoded in an intron of the myosin gene, is enriched in myocardial cells and inhibits apoptosis via the calcineurin/Drp1 signaling pathway [40, 41]. Some studies have reported the regulation of miR-499 transcription by P53 [42], while *Drp1* regulation by miRNAs affects mitochondrial dynamics and apoptosis. The

underlying mechanisms are complex, but may involve the fission 1 protein [43]. Additional studies are required to clarify the regulation of mitochondrial dynamics by miRNAs.

MiRNAs in mitochondrial autophagy

Autophagy is the lysosome-mediated degradation of intracellular proteins and organelles involving changes to cell membrane structure [44, 45]. This dynamic process balances cellular anabolism and catabolism, thereby stabilizing the intracellular environment and promoting cell survival. Autophagy can be triggered by cell starvation, absence of growth factors, hypoxia, and a variety of pathological conditions. During autophagy, cellular organelles and other components are packed within a phagosome that is transported via microtubules to fuse with the lysosome, leading to substrate degradation. Mitochondrial autophagy or mitophagy is a process by which the cell removes damaged mitochondria and maintains homeostasis [46]; this regulates mitochondrial number so that the biological and metabolic demands of the cell can be balanced [47]. During mitochondrial membrane depolarization in mammalian cells, phosphatase and tensin homolog (PTEN)-induced putative kinase (PINK)1 triggers the transport of Parkin proteins from the cytoplasm to mitochondria and the subsequent ubiquitination of mitochondrial proteins, leading to the formation of autophagosomes containing damaged mitochondria [48].

MiR-101, -204, and -30a regulate autophagy by targeting autophagy-related proteins [49–51]. Regulation by miR-34b/c is thought to be an early factor in Parkinson's disease, although whether it involves mitochondrial dysfunction has yet to be established [52]. The up-regulation of miR-21 expression in many types of human tumors and the regulation of PTEN expression by miR-21 has been demonstrated [53, 54]. PTEN in turn regulates PINK-1, although further study is required to determine how this controls the mitophagy pathway.

Role of miRNAs in cancer

The Warburg effect postulates that cancer cells use glycolysis instead of oxidative phosphorylation to generate ATP, even in the presence of oxygen [55, 56]; this enables tumor cells to sustain their rapid growth, which involves many factors including miRNAs [57]. It has been proposed that miRNAs can serve as a tool in tumor detection and treatment [58]. For example, miR-200a was found to reduce the growth of tumors in liver [59] and breast [60]

cancers. Many miR-200a target genes have been identified [61, 62], including mitochondrial transcription factor A (TFAM), a major transcription factor in mitochondria [63]. Changes in TFAM expression have been linked to tumor progression and the development of chemoresistance [64]; this may be due to miRNA regulation, since overexpressing miR-200a inhibits TFAM level [60].

The failure of apoptosis is considered as a major cause of therapeutic resistance in several cancers, including non-small cell lung cancer [65]. This program is controlled by many factors, including the activation of P53 as part of the DNA damage response. P53 stimulates the transcription of genes containing P53 binding sites. The miRNAs maintain cellular functions that are dysregulated in cancer cells, such as proliferation, differentiation, and apoptosis [66-68]. Given that they are directly activated by P53 [69], miR-34 family members play a crucial role in tumor suppression, possibly through modulation of mitophagy [69].

The miR-126 acts as a tumor suppressor in many types of human cancer, including oral squamous cell cancer (OSCC), bladder, lung and colorectal cancers [70-73]. In OSCC cells, the miR-126 expression is significantly lower than in adjacent normal tissues [70]. The miR-126 and acts as a tumor suppressor in malignant mesothelioma (MM), and also regulates mitochondrial function by modulating energy production and respiration and increasing glycolysis in MM cells. These findings provide evidence of the key role of miRNAs in cancer, and suggest that miRNAs can serve as tumor biomarkers and as agents or targets for cancer treatment.

Summary

Mitochondria are essential organelles in eukaryotic cells that provide energy to the cell and also play an important role in apoptosis and other cellular processes. The mitochondrial dysfunction is the basis of many diseases. miRNAs mediate post-transcriptional gene regulation and thereby regulate mitochondrial metabolism, the mitochondrial apoptotic pathway, and mitophagy. The discovery of mitochondria-specific miRNAs underscores the interaction between the nuclear and mitochondrial genomes in the regulation of miRNA expression and presents new avenues for studying the regulation of mitochondrial function. Moreover, research on mitochondrial dysfunction and the role of miRNAs can provide new insight into the

pathogenic mechanism of diseases such as cancer as well as potential treatment strategies.

References

- [1] Suen DF, Norris KL, Youle RJ. Mitochondrial dynamics and apoptosis. *Genes Dev* 2008; 22(12):1577-1590.
- [2] Trenker M, Malli R, Fertschai I, Levak-Frank S, Graier WF. Uncoupling proteins 2 and 3 are fundamental for mitochondrial Ca²⁺ uniport. *Nat Cell Biol* 2007; 9(4):445-452.
- [3] Lee Y, Ahn C, Han J, Choi H, Kim J, Yim J, Lee J, Provost P, Rådmark O, Kim S, Kim VN. The nuclear RNase III Drosha initiates microRNA processing. *Nature* 2003; 425:415-419.
- [4] He L, Hannon GJ. MicroRNAs: small RNAs with a big role in gene regulation. *Nat Rev Genet* 2004; 5(7):522-531.
- [5] Kren BT, Wong PY, Sarver A, Zhang X, Zeng Y, Steer CJ. MicroRNAs identified in highly purified liver-derived mitochondria may play a role in apoptosis. *RNA Biol* 2009; 6(1):65-72.
- [6] Bian Z, Li LM, Tang R, Hou DX, Chen X, Zhang CY, Zen K. Identification of mouse liver mitochondria-associated miRNAs and their potential biological functions. *Cell Res* 2010; 20(9):1076-1078.
- [7] Li N, Bates DJ, An J, Terry DA, Wang E. Up-regulation of key microRNAs, and inverse down-regulation of their predicted oxidative phosphorylation target genes, during aging in mouse brain. *Neurobiol Aging* 2011; 32(5):944-955.
- [8] Barrey E, Saint-Auret G, Bonnamy B, Damas D, Boyer O, Gidrol X. Pre-microRNA and mature microRNA in human mitochondria. *PLoS One* 2011; 6(5):e20220.
- [9] Bandiera S, Rübberg S, Girard M, Cagnard N, Hanein S, Chrétiën D, Munnich A, Lyonnet S, Henrion-Caude A. Nuclear outsourcing of RNA interference components to human mitochondria. *PLoS One* 2011; 6(6):e20746.
- [10] Aoi W, Naito Y, Mizushima K, Takanami Y, Kawai Y, Ichikawa H, Yoshikawa T. The microRNA miR-696 regulates PGC-1(alpha) in mouse skeletal muscle in response to physical activity. *Am J Physiol Endocrinol Metab* 2010; 298(4):E799-E806.
- [11] Naguibneva I, Ameyar-Zazoua M, Polesskaya A, Ait-Si-Ali S, Groisman R, Souidi M, Cuvellier S, Harel-Bellan A. The microRNA miR-181 targets the homeobox protein Hox-A11 during mammalian myoblast differentiation. *Nat Cell Biol* 2006; 8(3):278-284.
- [12] Mccarthy JJ, Esser KA. MicroRNA-1 and microRNA-133a expression are decreased during skeletal muscle hypertrophy. *J Appl Physiol* 2007; 102(1):306-313.
- [13] Parker R, Sheth U. P bodies and the control of mRNA translation and degradation. *Mol Cell* 2007; 25(5):635-646.
- [14] Gidrol X, Barrey E, Damas D, Saint-Auret G, Boyer O, Bonnamy B. Pre-microRNA and mature microRNA in human mitochondria. *PLoS One* 2011; 6(5):e20220.
- [15] Yuan Y, Huang S, Wang W, Wang Y, Zhang P, Zhu C, Ding G, Liu B, Yang T, Zhang A. Activation of peroxisome proliferator-activated receptor-gamma coactivator 1alpha ameliorates mitochondrial dysfunction and protects podocytes from aldosterone-induced injury. *Kidney Int* 2012; 82(7):771-789.
- [16] Cannino G, Di Liegro CM, Rinaldi AM. Nuclear-mitochondrial interaction. *Mitochondrion* 2007; 7(6):359-366.
- [17] Aschrafi A, Schwechter AD, Mameza MG, Natera-Naranjo O, Gioio AE, Kaplan BB. MicroRNA-338 regulates local cytochrome c oxidase IV mRNA levels and oxidative phosphorylation in the axons of sympathetic neurons. *J Neurosci* 2008; 28(47):12581-12590.
- [18] Gao P, Tchernyshyov I, Chang TC, Lee YS, Kita K, Ochi T, Zeller KI, De Marzo AM, Van Eyk JE, Mendell JT, Dang CV. c-Myc suppression of miR-23a/b enhances mitochondrial

- glutaminase expression and glutamine metabolism. *Nature* 2009; 458(7239):762-765.
- [19] Kulshreshtha R, Ferracin M, Wojcik SE, Garzon R, Alder H, Agosto-Perez FJ, Davuluri R, Liu CG, Croce CM, Negrini M, Calin GA, Ivan M. A microRNA signature of hypoxia. *Mol Cell Biol* 2007; 27(5):1859-1867.
- [20] Rouault TA, Tong WH. Iron-sulfur cluster biogenesis and human disease. *Trends Genet* 2008; 24(8):398-407.
- [21] Poy MN, Eliasson L, Krutzfeldt J, Kuwajima S, Ma X, Macdonald PE, Pfeffer S, Tuschl T, Rajewsky N, Rorsman P, Stoffel M. A pancreatic islet-specific microRNA regulates insulin secretion. *Nature* 2004; 432(7014):226-230.
- [22] Bordone L, Motta MC, Picard F, Robinson A, Jhala US, Apfeld J, McDonagh T, Lemieux M, McBurney M, Szilvasi A, Easlon EJ, Lin SJ, Guarente L. Sirt1 regulates insulin secretion by repressing UCP2 in pancreatic beta cells. *PLoS Biol* 2006; 4(2):e31.
- [23] Lowell BB, Shulman GI. Mitochondrial dysfunction and type 2 diabetes. *Science* 2005; 307(5708):384-387.
- [24] Wu H, Kanatous SB, Thurmond FA, Gallardo T, Isotani E, Bassel-Duby R, Williams RS. Regulation of mitochondrial biogenesis in skeletal muscle by CaMK. *Sci* 2002; 296(5566):349-352.
- [25] Li P, Jayarama S, Ganesh L, Mordi D, Carr R, Kanteti P, Hay N, Prabhakar BS. Akt-phosphorylated mitogen-activated kinase-activating death domain protein (MADD) inhibits TRAIL-induced apoptosis by blocking Fas-associated death domain (FADD) association with death receptor 4. *J Biol Chem* 2010; 285(29):22713-22722.
- [26] Cleland MM, Norris KL, Karbowski M, Wang C, Suen DF, Jiao S, George NM, Luo X, Li Z, Youle RJ. Bcl-2 family interaction with the mitochondrial morphogenesis machinery. *Cell Death Differ* 2011; 18(2):235-247.
- [27] Susin SA, Lorenzo HK, Zamzami N, Marzo I, Snow BE, Brothers GM, Mangion J, Jacotot E, Costantini P, Loeffler M, Larochette N, Goodlett DR, Abersold R, Siderovski DP, Penninger JM, Kroemer G. Molecular characterization of mitochondrial apoptosis-inducing factor. *Nature* 1999; 397(6718):441-446.
- [28] Verhagen AM, Ekert PG, Pakusch M, Silke J, Connolly LM, Reid GE, Moritz RL, Simpson RJ, Vaux DL. Identification of DIABLO, a mammalian protein that promotes apoptosis by binding to and antagonizing IAP proteins. *Cell* 2000; 102(1):43-53.
- [29] Gao SM, Chen C, Wu J, Tan Y, Yu K, Xing CY, Ye A, Yin L, Jiang L. Synergistic apoptosis induction in leukemic cells by miR-15a/16-1 and arsenic trioxide. *Biochem Biophys Res Commun* 2010; 403(2):203-208.
- [30] Akao Y, Nakagawa Y, Naoe T. MicroRNAs 143 and 145 are possible common onco-microRNAs in human cancers. *Oncol Rep* 2006; 16(4):845-850.
- [31] Nakagawa Y, Iinuma M, Naoe T, Nozawa Y, Akao Y. Characterized mechanism of alpha-mangostin-induced cell death: caspase-independent apoptosis with release of endonuclease-G from mitochondria and increased miR-143 expression in human colorectal cancer DLD-1 cells. *Bioorg Med Chem* 2007; 15(16):5620-5628.
- [32] Chen JF, Mandel EM, Thomson JM, Wu Q, Callis TE, Hammond SM, Conlon FL, Wang DZ. The role of microRNA-1 and microRNA-133 in skeletal muscle proliferation and differentiation. *Nat Genet* 2006; 38(2):228-233.
- [33] Berman SB, Pineda FJ, Hardwick JM. Mitochondrial fission and fusion dynamics: the long and short of it. *Cell Death Differ* 2008; 15(7):1147-1152.
- [34] Tatsuta T, Langer T. Quality control of mitochondria: protection against neurodegeneration and ageing. *EMBO J* 2008; 27(2):306-314.
- [35] Cassidy-Stone A, Chipuk JE, Ingerman E, Song C, Yoo C, Kuwana T, Kurth MJ, Shaw JT, Hinshaw JE, Green DR, Nunnari J. Chemical inhibition of the mitochondrial division dynamin reveals its role in Bax/Bak-dependent mitochondrial outer membrane permeabilization. *Dev Cell* 2008; 14(2):193-204.
- [36] Yang Y, Ouyang Y, Yang L, Beal MF, McQuibban A, Vogel H, Lu B. Pink1 regulates mitochondrial dynamics through interaction with the fission/fusion machinery. *Proc Natl Acad Sci USA* 2008; 105(19):7070-7075.
- [37] Yang Y, Gehrke S, Imai Y, Huang Z, Ouyang Y, Wang JW, Yang L, Beal MF, Vogel H, Lu B. Mitochondrial pathology and muscle and dopaminergic neuron degeneration caused by inactivation of *Drosophila* Pink1 is rescued by Parkin. *Proc Natl Acad Sci U S A* 2006; 103(28):10793-10798.
- [38] Edwards JL, Quattrini A, Lentz SI, Figueroa-Romero C, Cerri F, Backus C, Hong Y, Feldman EL. Diabetes regulates mitochondrial biogenesis and fission in mouse neurons. *Diabetologia* 2010; 53(1):160-169.
- [39] Li J, Donath S, Li Y, Qin D, Prabhakar BS, Li P. miR-30 regulates mitochondrial fission through targeting p53 and the dynamin-related protein-1 pathway. *PLoS Genet* 2010; 6(1):e1000795.
- [40] Kim HJ, Cui XS, Kim EJ, Kim WJ, Kim NH. New porcine microRNA genes found by homology search. *Genome* 2006; 49(10):1283-1286.
- [41] van Rooij E, Quiat D, Johnson BA, Sutherland LB, Qi X, Richardson JA, Kelm RJ Jr, Olson EN. A family of microRNAs encoded by myosin genes governs myosin expression and muscle performance. *Dev Cell* 2009; 17(5):662-673.
- [42] Wang JX, Jiao JQ, Li Q, Long B, Wang K, Liu JP, Li YR, Li PF. miR-499 regulates mitochondrial dynamics by targeting calcineurin and dynamin-related protein-1. *Nat Med* 2011; 17(1):71-78.
- [43] Bian Z, Li LM, Tang R, Hou DX, Chen X, Zhang CY, Zen K. Identification of mouse liver mitochondria-associated miRNAs and their potential biological functions. *Cell Res* 2010; 20(9):1076-1078.
- [44] Mortensen M, Ferguson DJ, Edelman M, Kessler B, Morten KJ, Komatsu M, Simon AK. Loss of autophagy in erythroid cells leads to defective removal of mitochondria and severe anemia in vivo. *Proc Natl Acad Sci U S A* 2010; 107(2):832-837.
- [45] Yang Z, Klionsky DJ. Eaten alive: a history of macroautophagy. *Nat Cell Biol* 2010; 12(9):814-822.
- [46] Kim I, Rodriguez-Enriquez S, Lemasters JJ. Selective degradation of mitochondria by mitophagy. *Arch Biochem Biophys* 2007; 462(2):245-253.
- [47] Kundu M, Lindsten T, Yang CY, Wu J, Zhao F, Zhang J, Selak MA, Ney PA, Thompson CB. Ulk1 plays a critical role in the autophagic clearance of mitochondria and ribosomes during reticulocyte maturation. *Blood* 2008; 112(4):1493-1502.
- [48] Geisler S, Holmström KM, Skujat D, Fiesel FC, Rothfuss OC, Kahle PJ, Springer W. PINK1/Parkin-mediated mitophagy is dependent on VDAC1 and p62/SQSTM1. *Nat Cell Biol* 2010; 12(2):119-131.
- [49] Frankel LB, Wen J, Lees M, Høyer-Hansen M, Farkas T, Krogh A, Jäättelä M, Lund AH. microRNA-101 is a potent inhibitor of autophagy. *EMBO J* 2011; 30(22):4628-4641.
- [50] Xiao J, Zhu X, He B, Zhang Y, Kang B, Wang Z, Ni X. MiR-204 regulates cardiomyocyte autophagy induced by ischemia-reperfusion through LC3-II. *J Biomed Sci* 2011; 18:35.
- [51] Zhu H, Wu H, Liu X, Li B, Chen Y, Ren X, Liu CG, Yang JM. Regulation of autophagy by a beclin 1-targeted microRNA, miR-30a, in cancer cells. *Autophagy* 2009; 5(6):816-823.
- [52] Miñones-Moyano E, Porta S, Escaramís G, Rabionet R, Iraola S, Kagerbauer B, Espinosa-Parrilla Y, Ferrer I, Estivill X, Martí E. MicroRNA profiling of Parkinson's disease brains identifies early downregulation of miR-34b/c which modulate

- mitochondrial function. *Hum Mol Genet* 2011; 20(15):3067-3078.
- [53] Meng F, Henson R, Wehbe-Janek H, Ghoshal K, Jacob ST, Patel T. MicroRNA-21 regulates expression of the PTEN tumor suppressor gene in human hepatocellular cancer. *Gastroenterol* 2007; 133(2):647-658.
- [54] Zhang JG, Wang JJ, Zhao F, Liu Q, Jiang K, Yang GH. MicroRNA-21 (miR-21) represses tumor suppressor PTEN and promotes growth and invasion in non-small cell lung cancer (NSCLC). *Clin Chim Acta* 2010; 411(11-12):846-852.
- [55] Warburg O. On respiratory impairment in cancer cells. *Sci* 1956; 124:269-270
- [56] Zheng J. Energy metabolism of cancer: Glycolysis versus oxidative phosphorylation. *Oncol Lett* 2012; 4:1151-1157.
- [57] Fogg VC, Lanning NJ, Mackeigan JP. Mitochondria in cancer: At the crossroads of life and death. *Chin. J. Cancer* 2011; 30:526-539.
- [58] Krell J, Frampton AE, Stebbing J. MicroRNAs in the cancer clinic. *Front Biosci* 2013; 5:204-213.
- [59] Radojicic J, Zaravinos A, Vrekoussis T, Kafousi M, Spandidos DA, Stathopoulos EN. MicroRNA expression analysis in triple-negative (ER, PR and Her2/neu) breast cancer. *Cell Cycle* 2011; 10:507-517.
- [60] Yao J, Zhou E, Wang Y, Xu F, Zhang D, Zhong D. microRNA-200a inhibits cell proliferation by targeting mitochondrial transcription factor A in breast cancer. *DNA Cell Biol* 2014; 33:291-300.
- [61] Roybal JD, Zang Y, Ahn Y-H, Yang Y, Gibbons DL, Baird BN, Alvarez C, Thilaganathan N, Liu DD, Saintigny P et al. miR-200 inhibits lung adenocarcinoma cell invasion and metastasis by targeting Flt1/VEGFR1. *Mol Cancer Res* 2011; 9:25-35.
- [62] Rasheed SAK, Teo CR, Beillard EJ, Voorhoeve PM, Casey PJ. MicroRNA-182 and microRNA-200a control G-protein subunit α -13 (GNA13) expression and cell invasion synergistically in prostate cancer cells. *J Biol Chem* 2013; 288:7986-7995.
- [63] Bonawitz ND, Clayton DA, Shadel GS. Initiation and beyond: Multiple functions of the human mitochondrial transcription machinery. *Mol Cell* 2006; 24:813-825.
- [64] Han B, Izumi H, Yasuniwa Y, Akiyama M, Yamaguchi T, Fujimoto N, Matsumoto T, Wu B, Tanimoto A, Sasaguri Y, et al. Human mitochondrial transcription factor A functions in both nuclei and mitochondria and regulates cancer cell growth. *Biochem Biophys Res Commun* 2011; 408:45-51.
- [65] Chakraborty S, Mazumdar M, Mukherjee S, Bhattacharjee P, Adhikary A, Manna A, Chakraborty S, Khan P, Sen A, Das T. Restoration of p53/miR-34a regulatory axis decreases survival advantage and ensures Bax-dependent apoptosis of non-small cell lung carcinoma cells. *FEBS Lett.* 2014; 588:549-559.
- [66] Wu N, Liu X, Xu X, Fan X, Liu M, Li X, Zhong Q, Tang H. MicroRNA-373, a new regulator of protein phosphatase 6, functions as an oncogene in hepatocellular carcinoma. *FEBS J* 2011; 278:2044-2054.
- [67] Annibali D, Gioia U, Savino M, Laneve P, Caffarelli E, Nasi S. A new module in neural differentiation control: Two microRNAs upregulated by retinoic acid, miR-9 and -103, target the differentiation inhibitor ID2. *PLoS One* 2012; 7:e40269.
- [68] Chan JA, Krichevsky AM, Kosik KS. MicroRNA-21 is an antiapoptotic factor in human glioblastoma cells. *Cancer Res* 2005; 65:6029-6033.
- [69] He L, He X, Lim LP, de Stanchina E, Xuan Z, Liang Y, Xue W, Zender L, Magnus J, Ridzon D et al. microRNA component of the p53 tumour suppressor network. *Nature* 2007; 447:1130-1134.
- [70] Yang X, Wu H, Ling T. Suppressive effect of microRNA-126 on oral squamous cell carcinoma *in vitro*. *Mol Med Rep* 2014; 10:125-130.
- [71] Jia AY, Castillo-Martin M, Bonal DM, Sánchez-Carbayo M, Silva JM, Cordon-Cardo C. MicroRNA-126 inhibits invasion in bladder cancer via regulation of ADAM9. *British J Cancer* 2014; 110:2945-2954.
- [72] Kim MK, Jung SB, Kim JS, Roh MS, Lee JH, Lee EH, Lee HW. Expression of microRNA miR-126 and miR-200c is associated with prognosis in patients with non-small cell lung cancer. *Virchows Arch* 2014; 465(4):463-471.
- [73] Liu Y, Zhou Y, Feng X, An P, Quan X, Wang H, Ye S, Yu C, He Y, Luo H. MicroRNA-126 functions as a tumor suppressor in colorectal cancer cells by targeting CXCR4 via the AKT and ERK1/2 signaling pathways. *Int. J. Oncol* 2014; 44:203-210.