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## RESEARCH ARTICLE



# HEPATOPROTECTIVE EFFECT OF SAGE (SALVIA OFFICINALIS L.) LEAVES HYDRO-METHANOLIC EXTRACT AGAINST ASPERGILLUS PARASITICUS AFLATOXIN-INDUCED LIVER DAMAGE IN MALE RATS

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Different parts of the Salvia officinalis L. are used to treat liver disorders, traditionally. The aim of the study is to evaluate the protective effect of Salvia officinalis against Aspergillus parasiticus aflatoxin induced hepatotoxicity in rats. Various biochemical parameters like serum alanine amino transferase (ALT), aspartate amino transferase (AST), alkaline phosphatase (AP) and total protein (TP) levels were determined. The treatment of aflatoxin at dose 450 µg/kg increased serum ALT, AST and AP levels, while decreased total protein levels in contaminated rats in comparison to control normal rats. Treatment of sage extract at doses 25, 50, 100 and 150 mg/kg body weight decreased the raised serum AST, ALT and AP levels and increased serum total protein level in treated rats in comparison to control rats. This study demonstrated the hepatoprotective activity of Salvia officinalis and thus scientifically supports the usage of this plant for treatment of liver disorders.

Key words: Aflatoxin, Aspergillus parasiticus, Sage, Salvia officinalis, Hepatotoxicity, Hepatic enzymes.

# INTRODUCTION

Liver is the organ for metabolism and detoxification of various components entering into the body. It is involved in wide range of functions and hence it is exposed to toxic substances and drugs absorbed from the intestine.

Aflatoxins are a group of closely related compounds with small differences in chemical composition (Cullen and Newberne, 1993). Aflatoxins were first isolated about 40 years ago after outbreaks of disease and death in turkeys (Blount, 1961) and of cancer in rainbow trout (Rucker *et al* 2002) fed on rations formulated from peanut and cottonseed meals. The toxins are produced as secondary metabolites by *Aspergillus flavus* and *Aspergillus parasiticus* fungi. The fungi responsible are ubiquitous and can affect many of the developing-country dietary staples of rice, corn, cassava, nuts, peanuts, chillies, and spices (Rucker *et al* 2002). Aflatoxicosis is the poisoning that results from ingesting aflatoxins. Two forms of aflatoxicosis have been identified: the first is acute severe intoxication, which results in direct liver damage and subsequent illness or death, and the second is chronic subsymptomatic exposure. The symptoms of severe aflatoxicosis include



hemorrhagic necrosis of the liver, bile duct proliferation, edema, and lethargy. Aflatoxin  $B_1$  is the most prevalent form and also the most potent of these toxins (Cullen and Newberne, 1993).

Globally, plant based drugs like *Silybum marianum* (Luper, 1998), *Picrorhiza kurroa* (Chander *et al* 1992), *Phyllanthus emblica* (Gulati *et al* 1995) etc. are widely and successfully used in the treatment of liver disorders.

The genus *Salvia* L. (Lamiaceae) comprises about 900 species, spread throughout the world, some of which with great economic value since they are used as spices and flavouring agents by perfumery and cosmetic industries (Longaray Delamare *et al* 2007). *Salvia officinalis* (sage, garden sage, or common sage) is a perennial, evergreen subshrub, with woody stems, greyish leaves, and blue to purplish flowers. It is native to the Mediterranean region, being currently cultivated in various countries (Raal *et al* 2007).

Since ancient times, plants are well known for their pharmacological potential (Jain *et al* 2011; Jenny *et al* 2012; Deb *et al* 2013). The predominant medicinally valuable metabolites of sage are monoterpenes (e.g.,  $\alpha$ - and  $\beta$ -thujone, 1, 8-cineole, camphor), diterpenes (e.g. carnosic acid) triterpenes (oleanoic and ursolic acids), and phenolic compounds like rosmarinic acid (Cuvelier *et al* 1994).

Salvia sp. has also been used for a long time in folk medicine as medication against fever, rheumatism, perspiration, sexual debility, and in the treatment of chronic bronchitis, as well as mental and nervous diseases (Raal *et al* 2007). Sage leaves and its essential oil possess carminative, antispasmodic, antiseptic, astringent, antioxidant and antihidrotic properties (Cuvelier *et al* 1994; Kamatou *et al* 2005).

There is no report about hepatoprotective effect of sage extract against *Aspergillus parasiticus* aflatoxin-induced liver damage in male rats. So, in the present study, we evaluated the protective effect of methanolic extract of *Salvia officinalis* L. against aflatoxin induced hepatotoxicity in rats.

# MATERIALS AND METHODS Plant material

Sage leaves (*Salvia officinalis* L.) were purchased from Karaj in June 2014, identified by Department of Botany, Science and Research Branch, Islamic Azad University (Voucher number: 037420, Director: Dr. Ali Mazooji). The plant was cleaned, shed dried at 25°C, and the dried leaves of the plant were ground with a blender, and the powder was kept in nylon bags in a deep freezer until the time of experiments. Dried and ground leaves (about 100 g) were submitted to extraction with 300 ml methanol (80%) in a soxhlet apparatus for 48 h. After extraction, the solvent was filtered and then evaporated by rotavapor. The obtained hydromethanolic extract was stored at -20°C until being used (**Figure 1**).



Fig. 1. Leaves of Salvia officinalis L. (Sage)

# Animal

In this study, male Wistar rats weighing 200-250 g were housed in clean cages with temperature (22–24°C), 12h light/12h dark cycle and relative air humidity 40–60%. Rats had continuous access to food and tap water. Permission for the study was obtained from the Pastour institute, Tehran, IRAN. In the present experiment, 48 rats (40 contaminated, 8 intact rats) divided into six groups were used. To group 1, 0.5 ml of saline as aflatoxin vehicle was administered intraperitoneally to normal control rats (intact) every week. To group 2, 0.5 ml of aflatoxin at dose 450  $\mu$ g/kg was administered intraperitoneally to control rats every week. To groups 3-6, 0.5 ml of aflatoxin intraperitoneally and sage methanolic extract orally at doses 25, 50, 100 and 150 mg/kg body wt were co administered daily for 8 weeks.

# **Biochemical measurements**

After 8 weeks of treatment, weight of each rat was measured. Then, the animals were anesthetized by ether and blood samples were drawn from heart. Serum total protein, aspartate amino transferase (AST), alanine amino transferase (ALT) and alkaline phosphatase levels were determined by kit (Parsazmoon, Iran). Also, their livers removed and weighed. Then, liver coefficients were measured as liver weight divided to body weight for each animal.

## Statistical analysis

All the data were expressed as mean $\pm$ S.E.M. Statistical analysis was carried out using oneway ANOVA followed by Tukey post hoc test. The criterion for statistical significance was p<0.05.

## **RESULTS AND DISCUSSION**

There were significant elevations in serum ALT (p<0.001), AST (p<0.001) and AP (p<0.001) levels in contaminated rats, while significant

attenuation in serum total protein level in the contaminated control rats in comparison with control normal rats (p<0.001).

The present results showed that treatment of sage leaves extract decreased serum ALT (p<0.001), AST (p<0.001) and AP (p<0.001), while increased serum total protein level (p<0.001) in treated contaminated rats in comparison to control rats. Also, treatment of aflatoxin increased liver coefficient in contaminated control rats (p<0.001) and sage extract decreased liver coefficient in treated rats in comparison to contaminated control rats (p<0.001) and sage extract decreased liver coefficient in treated rats in comparison to contaminated control rats (p<0.001) (Table 1).

**Table 1.** Effect of sage extract on liver coefficient and serum parameters in Aspergillus parasiticusaflatoxin-induced liver damage in male rats.

Group parameter	Intact	Control	Extract (mg/kg)			
			25	50	100	150
AST (U/l)	161±3	196±1***	191±2***	185±1***	180±1***	172±1***
			+++	+++	+++	+++
ALT (U/l)	79±4	108±1 ***	101±1 ***	95±1***	93±1***	90±2***
			+++	+++	+++	+++
AP (U/l)	190±4	230±1***	221±1***	211±3***	201±1***	199±0.3***
			+++	+++	+++	+++
Total protein	7.8±0.056	6.5±0.045***	6.8±0.03***	7.2±0.07***	7.2±0.02***	7.4±0.03***
(g/dl)			+++	+++	+++	+++
Liver	0.031±0.001	0.054±0.0004***	0.041±0.001***	0.038±0.002***	0.037±0.0007***	0.035±0.0004***
coefficient			+++	+++	+++	+++

\*\*\*p<0.001 difference from intact group. +++p<0.001 difference from control group.

Fungal infections may discolour grains, change its chemical and nutritional characteristics, reduce germination and most importantly, contaminate it with mycotoxins, such as aflatoxins which are highly toxic to man and animals (Paster *et al* 1993). *Aspergillus parasiticus* is one of the major storage fungi found regularly in important cereals cultivated in the world, which produces aflatoxins such as aflatoxin B1, B2, G1, G2 (Paster, 1995).

Aflatoxin is predominantly perceived as an agent promoting liver cancers, although lung cancer is among workers also а risk handling contaminated grain (Kelly et al 1997). The risk of cancers due to exposure to the various forms of aflatoxin is well established (Gorelick et al 1993) and is based on the cumulative lifetime dose. The International Cancer Research Institute identifies aflatoxin as a Class 1 carcinogen, resulting in the regulation of this toxin to very low concentrations in traded commodities (Henry et al 1999). The leaves extract of sage (Salvia officinalis L.) have been documented to have wide range of biological effects (Cuvelier et al 1994). The present results showed that weight of liver in rats with liver damage by aflatoxin was more than normal rats. So, their liver coefficients were higher than control group. Treatment of sage extract decreased liver coefficient in treated animal and improved liver inflammation. The hepatic cells consist of higher concentrations of AST and ALT in cytoplasm and AST in particular exists in mitochondria (Wells, 1988). Due to the damage caused to hepatic cells, the leakage of plasma (Zimmerman and Seef, 1970) caused an increased levels of hepatospecific enzymes in serum. The elevated serum enzyme levels like AST and ALT are indicative of cellular leakage and functional integrity of cell membrane in liver (Drotman and Lawhorn, 1978). The hepatoprotective index of a drug can be evaluated by its capability to reduce the injurious effects or to preserve the normal hepatic physiological mechanisms, which have induced by a hepatotoxin. been The measurement of serum AST, ALT and ALP levels serves as a means for the indirect assessment of condition of liver.

Hepatic enzymes included ALT, AST and AP in serum increased in rats with liver injury which

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are markers of liver damage. Administration of *Salvia officinalis* extract attenuated serum ALT, AST and AP levels, significantly.

The total protein including albumin levels depressed in hepatotoxic conditions due to defective protein biosynthesis in liver (Clawson, 1989). The aflatoxin causes disruption and dissociation of polyribosomes on endoplasmic reticulum and thereby reducing the biosynthesis of protein.

The pre-treatment of sage extract well restored the proteins synthesis by protecting the polyribosomes. Serum total protein level decreased in rats with liver damage, because of liver dysfunction. Treatment of sage elevated total protein level in treated animal. The therapeutic effect of sage extract also showed the antioxidant activity and removed reactive

## REFERENCES

Blount WP. Turkey "X" disease. J. Br. Turk. Fed. 1961;9:52-4.

- Chander R, Kapoor NK, Dhawan BN. Effect of picroliv on glutathione metabolism in liver and brain of *Mastomys natalensis* infected with *Plasmodium berghei*. *Indian J. Exp. Biol*. 1992;30(8):711-4.
- Clawson GA. Mechanism of carbon tetrachloride hepatotoxicity. *Pathol. Immunopathol. Res.* 1989;8(2): 104-12.
- Cullen JM, Newberne PM. Acute Hepatotoxicity of Aflatoxins. In: Eaton DL, Groopman JD (eds). The Toxicology of Aflatoxins: Human Health, Veterinary, and Agricultural Significance. Academic Press: London, 1993; 1-26.
- Cuvelier ME, Berset C, Richard H. Antioxidant constituents in sage (*Salvia officinalis*). *J. Agric. Food Chem.* 1994;42(3):665-9. [DOI: 10.1021/jf00039a012]
- Deb L, Bhattacharjee C, Shetty SR, Dutta A. Evaluation of anti-diabetic potential of the *Syzygium cuminii* (linn) skeels by reverse pharmacological approaches. *Bull. Pharm. Res.* 2013;3(3):135-45.
- Drotman RB, Lawhorn GT. Serum enzymes as indicators of chemically induced liver damage. *Drug Chem. Toxicol.* 1978;1(2):163-71. [DOI: 10.3109/01480547809034433]
- Gorelick NJ, Bruce RD, Hoseyni MS. Human Risk Assessment Based on Animal Data: Inconsistencies and Alternatives. In: Eaton D, Groopman JD (eds.). The Toxicology of Aflatoxins: Human Health, Veterinary, and Agricultural Significance. Academic Press: London: 1993; 508-11.

Gulati RK, Agarwal S, Agrawal SS. Hepatoprotective studies on *Phyllanthus emblica* Linn. and quercetin. *Indian J. Exp. Biol.* 1995;33(4):261-8.

- Henry SH, Bosch FX, Troxell TC, Bolger PM. Reducing liver cancer-global control of aflatoxin. *Science* 1999;286 (5449):2453-4. [DOI: 10.1126/science.286.544 9.2453]
- Jain RA, Agarwal RC, Pandey A, Jain R. Evaluation of *Argemone mexicana* fruits extract using micronucleus assay in mouse bone marrow cells. *Bull. Pharm. Res.* 2011;1(2):22-4.
- Jenny A, Saha D, Paul S, Dutta M, Uddin MZ, Nath AK. Antibacterial activity of aerial part of extract of

oxygen species (Cuvelier et al 1994).

#### CONCLUSION

The hydro-methanolic extract of *Salvia officinalis* could effectively control serum AST, ALT, AP and total protein levels and increased the protein levels in the protective studies. The protective effect of sage extract may be attributed to the reduced lipid peroxidation and improved defence of the hepatocytes against the reactive oxygen species. Therefore, the study scientifically supports the usage of plant in traditional medicines for liver disorders.

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Elephantopus scaber Linn. Bull. Pharm. Res. 2012;2(1): 38-41.

- Kamatou GPP, Viljoen AM, Gono-Bwalya AB, van Zyl RL, van Vuuren SF, Lourens ACU, Başer KHC, Demirci B, Lindsey KL, van Staden J, Steenkamp P. The *in vitro* pharmacological activities and a chemical investigation of three South African *Salvia* species. *J. Ethnopharmacol.* 2005;102(3):382-90. [DOI: 10.1016/j.jep.2005.06.034]
- Kelly JD, Eaton DL, Guengerich FP, Coulombe Jr RA. Aflatoxin B1 activation in human lung. *Toxicol. Appl. Pharmacol.* 1997;144(1):88-95. [DOI: 10.1006/taap.1997. 8117]
- Longaray Delamare AP, Moschen-Pistorello IT, Artico L, Atti-Serafini L, Echeverrigaray S. Antibacterial activity of the essential oils of *Salvia officinalis* L. and *Salvia triloba* L. cultivated in South Brazil. *Food Chem.* 2007;100(2):603-8. [DOI: 10.1016/j.foodchem.2005.09.078]
- Paster N. Fungi in stored grain and animal feeds: their occurrence and harm caused to animals. *Isr. J. Vet. Med.* 1995;50(2):49-53.
- Paster N, Droby S, Chalutz E, Menasherov M, Nitzan R, Wilson CL. Evaluation of the potential of the yeast *Pichia guilliermondii* as a biocontrol agent against *Aspergillus flavus* of stored soya beans. *Mycol. Res.* 1993;97(10):1201-6.
- Raal A, Orav A, Arak E. Composition of the essential oil of *Salvia officinalis* L. from various European countries. *Nat. Prod. Res.* 2007;21(5):406-11.
- Rucker RR, Yasutake WT, Wolf H. Trout hepatoma A preliminary report. *Prog. Fish Cult.* 2002;23:3-7.
- Luper S. A review of plants used in the treatment of liver disease: Part 1. *Altern. Med. Rev.* 1998;3(6):410-21.
- Wells FE. Tests in Liver and Biliary Tract Disease. In: Gowenlock HA (ed.). Varley's Practical Clinical Biochemistry. CRC Press, Florida: 1988; 93-110.
- Zimmerman HJ, Seef L.B. Enzymes in Hepatic Disease. In: Goodly EI (ed.). Diagnostic Enzymology. Lea and Febiger, Philadelphia: 1970; 118-35.
- http://en.shram.kiev.ua/health/travnik/salvia-officinalisl.shtml