

# Effect of Pig Iron Slag Particles on Soil Physico-Chemical, Biological and Enzyme Activities

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**Abstract:** The effect of pig iron slag particles on soil physico-chemical, biological and certain soil enzyme properties was studied. Contamination of iron slag particles altered physico-chemical, biological and enzyme properties of soil. While soil pH increased slightly, electrical conductivity, carbon, potassium and phosphorus contents increased significantly in polluted soil. Contamination of soil with pig iron slag caused drastic reduction in microbial population. Similarly, enzyme activities such as dehydrogenase and protease were ceased three fold in polluted soil when compared to the control.

Key words: Pig iron slag % Physico-chemical % Biological properties % Soil enzymes

# INTRODUCTION

Soil is a dynamic system in which continuous interaction takes place between soil minerals, organic matter and organisms. Each of these three major soil components influences the physico-chemical and biological properties of terrestrial system [1]. Metals are natural constituents of the earth's crust but anthropogenic activities have resulted in drastic alterations in their geochemical cycles causing pollution and biochemical imbalance in the biosphere. In recent decades the development of industry and agriculture and activities such as mining, smelting of metal ions, industrial emissions and application of agrochemicals and fertilizers contributed to elevated levels of metals including iron in soils of Rayalasema region of Andhra Pradesh, India. Although iron is required in trace amounts for living organisms, discharge and deposition of iron ore based industrial contaminants have become an environmental issue of great public concern owing to their adverse effects on soil flora and fauna and ultimately to mankind.

Since soil enzyme activities are very sensitive to pollution, enzymes have been suggested by some researchers as potential indicators or monitoring tools to assess soil quality [2-3] and bioremediation activities [4]. Numerous changes with respect to physico-chemical, biological and enzymatic parameters were observed in soils such as soils contaminated with effluents discharged from cotton ginning mill [5], alcohol and chemical industries [6] and pulp and paper industry [7]. Enzyme activities that influence functional processes occurring in a given soil play an important role in determination of soil profile and biological activity [8]. Pollution of the soil environment with heavy metals, their effect on the soil microorganisms and on their enzymatic activity depend, apart from other things, on the soil pH, the content of organic and mineral colloids and on the metal type and its chemical properties [9, 10].

In view of the increasing environmental pollution, decreasing soil fertility and adverse effects on soil flora and fauna, it is of utmost concern to study the effects of metal pollutants on soil characters and soil microorganisms which pave the way for greater understanding the direction of improving soil fertility or bioremediation. Since iron ore is one of the most extensively mined, smelted, processed and used metal for majority of purposes, iron ore based industrial discharge and contaminants have become a major issue of environmental concern. An attempt was made in the present study with an aim to determine the effect of iron ore slag of pig iron plant on soil physic-chemical, biological and enzymatic activities of dehydrogenase, protease, cellulase and amylase.

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### MATERIALS AND METHODS

**Soil Collection:** Soil samples were collected from dumpings of pig iron plant located at Srikalahasti, Chittoor District of Andhra Pradesh, India and this was treated as test soil and the soil sample collected away from the pig iron plant was taken as control. Prior to testing, the soils were air-dried, passed through a 2 mm sieve and stored at room temperature for further studies.

**Physico-Chemical Properties of Soil:** The percentage of coarse fragments was quantified from the weight of material retained after sieving through a 2 mm sieve [11]. Soil pH and electrical conductivity were determined by pH meter (Elico) and conductivity meter, respectively. Water holding capacity and organic carbon content were quantified by the method of Johnson and Ulrich [12], Walkely and Black [13]. Soil phosphorus and potassium contents were determined by the methods of Watanabe and Olsen [14] and Toth [15], respectively.

**Enumeration of Soil Microorganisms:** Soil microbial populations such as including bacterial and fungal populations were enumerated by serial dilution technique. One gram of each soil sample was serially diluted and 0.1 ml was spreaded with a sterile spreader on nutrient agar media (pH 7.2) and potato dextrose agar medium for the growth of bacteria and fungi, respectively. After incubation period, colonies formed on the surface of the medium were counted by colony counter.

**Assay of Dehydrogenase:** The dehydrogenase activity was determined by the modified procedure of Casida [16]. To five grams of soil in a test tube, 2.5 ml of sterile distilled water and 1ml of 3% aqueous solution of triphenyl tetrazolium chloride (TTC) was added. These tubes were mixed thoroughly and incubated at 30°C for 24 h. The liberated end product, triphenyl tetrazolium formazone was measured at 485 nm in spectrophotometer (Elico).

**Assay of Protease:** Five grams of soil sample was placed in 50 ml Erlenmeyer flasks and 6 ml of 0.2M acetate buffer (pH 5.5) containing 1% BSA was added and incubated for 6 h at 30°C. After incubation, 1ml of toluene was added and the suspension filtered through Whattman No.1 filter paper. The amount of protein content in the filtrate was determined by the method of Lowry *et al.* [17].

### RESULTS

**Physico-Chemical Properties of Soil:** The soil polluted with iron slag increased soil pH from 7.2 to 7.9, water holding capacity from 0.3 to 0.7 ml/g of soil and electrical conductivity from 0.6 to 1.13 Mmhos/cm, Similarly, higher organic carbon, phosphorus and potassium levels were observed in test soil (Table 1).

**Soil Microorganisms:** Polluted soil caused 2-3 fold decrease in bacterial and fungal population compared to control soil. For instance, the bacterial and fungal population in control and polluted soil were 142x10<sup>5</sup>, 86x10<sup>5</sup> cfu/g and 25x10<sup>5</sup> and 7x10<sup>5</sup> cfu/gram of soil, respectively.

**Soil Dehydrogenase Activity:** The dehydrogenase activity of polluted soil was drastically inhibited (8-9 fold) when compared to control soil. With increase in soil incubation period, dehydrogenase activity also increased up to 16<sup>th</sup> day and decreased beyond that (Fig. 1).

**Soil Protease Activity:** Like dehydrogenase, protease activity was also inhibited (3-4 fold) in polluted soil. With increase in soil incubation period, protease activity also increased up  $to16^{th}$  day incubation but decreased thereafter (Fig. 2).

Table 1: Physico-chemical properties of soil contaminated with/without slag particles discharged from pig iron plant

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Physico-chemical properties	Control soil	Test soil
Colour	Light Brown	Black
Odor	Normal	Unpleasant
Soil nature	High weight	Low Weight
pH	7.2	7.9
Electrical onductivity(Mmhos/cm)	0.60	1.13
Water holding capacity (ml/g of soil)	0.3	0.7
Organic carbon (%)	Low	Medium
Availability of phosphorus Kg/ha	21	30
Availability of potassium Kg/ha	73	130

Table 2: Microbial population (\*CFU) in soil contaminated with/without slag particles discharged from pig iron plant

Microorganisms	Control soil	Test soil
Bacteria	142x10 <sup>5</sup>	86x10 <sup>5</sup>
Fungi	25x10 <sup>5</sup>	7x10 <sup>5</sup>

\*Microbial populations measured in terms of colony forming units (CFU/g) of soil

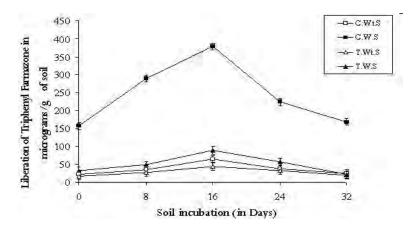


Fig. 1: Dehydrogenase activity of soil contaminated with/without slag particles discharged from pig iron plant, (Activity in terms of release of µg of Triphenyl formazone/g of soil)

C.Wt.S: Control without substrate; C.W.S: Control with substrate;

T.Wt.S: Test without substrate; T.W.S: Test with substrate

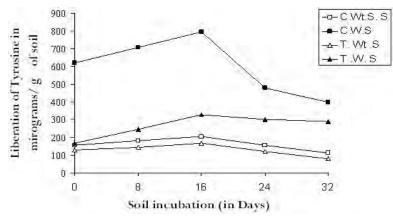


Fig. 2: Protease activity of soil contaminated with/without slag particles discharged from pig iron plant (Activity in terms of release of µg of tyrosine / g of soil)

C.Wt.S: Control without substrate; C.W.S: Control with substrate;

T.Wt.S: Test without substrate; T.W.S: Test with substrate

#### DISCUSSION

Our observations revealed that contamination with iron slag particles led to increase in pH of the test soil (Table 1). This indicates the alkalin nature of the disposed maste contaminants. Increase in soil pH was also reported by aluminium and heavy metal contaminated soils [18]. Increased water holding capacity and electrical conductivity observed in test soil reflects the altered chemical and biological characters of the polluted soil. Similar reports were observed in long term discharge of sewage effluents [19] and cotton ginning mill effluents [5]. In contrast, soils polluted with cement dust from cement industries had low water holding capacity and higher electrical conductivity [18-20]. The higher organic matter, phosphorus and potassium observed in polluted soil (Table 1) may be due to the effect of discharged effluents from pig iron plant. Similar results were noted in soils contaminated with effluents released from pesticide industries [21], with effluents from cotton ginning mills, with heavy metals content in vetiver grown on iron ore tailings and long term municipal waste disposed soils [5, 22, 23].

Reduced bacterial and fungal population in polluted soil may be due to the toxic effects of iron slag particles on microbial population. This may be because of oxidative stress caused by iron sledge particles or their interference in osmotic balance. The oligotrophic microbes' greater sensitivity to certain heavy metals and iron indicate that the growth of the microbial community is considerably limited in soils poor in organic matter and nutrient content [24]. Pollution of the soil environment with heavy metals also negatively influenced the soil microbial properties such as basal soil respiration rate and enzyme activities depending on the soil pH, organic matter content and other chemical properties [25].

Enzymes are strongly connected with important soil characteristics such as organic matter, physical properties, microbial activity or biomass. They are the sensitive indicators of soil quality.

In the present study, very significant inhibition of dehydrogenase and protease activities in polluted soil indicates the alterations in oxidation-reduction activities of enzymes released from microorganisms (Fig. 1 and 2). It is considered that iron also like other heavy metals, mainly inhibit enzymatic reactions through either their complexing with substrate or blocking the functional groups of enzymes or reacting with complex enzymesubstrate. Similar results were observed in dehydrogenases activity in flooded soils [10] and in herbicides contaminated soils [26]. Reduced protease activity was observed in soils contaminated with insecticides [27] and chlorothionil [28]. In contrast, the soil treated with tomato processing waste [29], cotton ginning mills [30] and dairy shed effluents [31] showed an increase in soil protease activity in comparison to the control soil.

#### CONCLUSION

The present study clearly indicates that dumping of pig iron slag on soil, altered its physico-chemical and biological properties and inhibited enzymatic activities such as dehydrogenase and protease.

## REFERENCES

- Bollag, J.M., J. Berthelin, D. Adriano and P.M. Huang, 2002. Impact of soil minerals-organic component-microorganisms interactions on restoration of terrestrial ecosystems. Poster presentation, paper no.861, Symposium no.47. 17<sup>th</sup> WCSS, Thailand.
- Dick, R.P., 1994. Soil enzyme activities as indicators of soil quality. (In J.W. Dpramm). Defining soil quality for a sustainable environment. SSSAS pec. Publ.35.SSSA, Madison, WI., pp: 107-124.
- Bandick, A.K. and R.P. Dick, 1999. Field management effects on soil enzyme activities. Soil Biol. Biochem., 31: 1471-1479.

- Margesin, R., A. Zimmer Bauer and F. Schinner, 2000. Monitoring of bioremediation by soil biological activities. Chemosphere, 40: 339-346.
- Narasimha, G., G.V.A.K. Babu and B. Rajasekhar Reddy, 1999. Physico-chemical and biological properties of soil samples collected from soil contaminated with effluents of cotton ginning industry. Asian J. Microbiol. Env. Biotechnol., 20: 235-239.
- 6. Monanmani, K., G. Chitraraju and K. Swaminathan, 1990. Effect of alcohol and chemical industrial effluents on physical and chemical biological properties of soil. Poll. Res., 9: 79-82.
- Kannan, K. and G. Oblisami, 1990. Influence of irrigation with pulp and papemill effluent on soil chemical and microbiological properties. Springer-Verlag GmbH., 10: 197-201.
- Amejkalova, M., O. Mikanova and L. Boruvka, 2003. Effects of heavy metal concentrations on biological activity of soil micro-organisms. Plant Soil and Environment, 49: 321-326.
- Kucharski, J. and J. Wyszkowska, 2004. Interrelationship between number of microorganisms or spring barley yield and degree of soil contamination with copper. Plant, Soil and Environment, 50: 243-249.
- Chandrayan, K., T.K. Adhya and N. Sethunathan, 1980. Dehydrogenase and invertase activity of flooded soils. Soil boil. Biochem., 2: 127-137.
- 11. Alexander M., 1980. Introduction to soil microbiology. New Delhi Wiley Estern Ltd.
- Johnson, C.M. and A. Ulrich, 1960. Determination of moisture in plant tissues. Calif. Agro. Bull., No. 766. (In Soil and Plant analysis for tree culture (Ed., S.A.Wilde 1960) Obortage publishing Co. Oxford, pp: 112-115.
- Walkley, A. and I.A. Black, 1934. An Examination of Degtjareff Method for Determining Soil Organic Matter and a Proposed Modification of the Chromic Acid Titration Method. Soil Sci., 37: 29.
- Watanabe, F.S. and S.R. Olsen, 1971. Test of an ascorbic acid method for determining phosphorus in water and NaHCO3 extracts from soil. Soil Sci. Soc. Am. Proc., 29: 677-678.
- Toth, S.J., A.L. Prince, A. Wallace and D.S. Mikklzson, 1948. Rapid quantitative determination of eight mineral elements in plant tissue by a systematic procedure involving use of a flame photometer. Soli Sci., 66: 459-466.
- Casida, L.E., D.A. Klein and T. Santoro, 1964. Soil dehydrogenase activity. Soil Sci., 98: 371-376.

- Lowry, O.H., N.J. Rosenbrough, A.L. Farr and R.J. Ramdall, 1951. Protein measurement with Folin Phenol Reagent, J. Bio. Chem., 193: 265-275.
- Shanthi, M., 1993. Soil biochemical processing industrially polluted areas of Cement industry. M. Phil dissertation submitted to Sri Krishnadevaraya University, Anantapur.
- Abdelnainm, E.M., M.S. Rao, T.M. Wally and E.M.B. Nashar, 1987. Effect of prolonged sewage irrigation on some physical properties of sandy soil. Bio.Wastes., 2: 269-274.
- Siva Kumar, S. and A. John De Brito, 1995. Effect of cement pollution on soil fertility. J. Ecotoxico Environ. Monit, 5: 47-149.
- Chuasavathi, T. and V. Trelo-ges, 2001. An important of yasothon soil fertility using municipal fermented organic compost and panicum maxium TD 58 grass. Pakistan Journal of Biological Sci., 4: 968-972.
- Anikew, M.A.N., 2002. Long term effect of municipal waste disposal on soil properties and productivity of sites used for urban agriculture in Abakaliki, Nigeria. Info Bioresource Technol., pp: 83.
- 23. Nualchavee, Roongtanakiat, Yongyuth Osotsapar and Charoen Yindiram, 2008. Effects of Soil Amendment on Growth and Heavy Metals Content in Vetiver Grown on Iron Ore Tailings. Kasetsart J. Nat. Sci., 42: 397-406.
- 24. Brookes, P.C., 1995. The use of microbial parameters in monitoring soil pollution by heavy metals. Biology and Fertility of Soils, 19: 269-279.
- 25. Szili-Kovacs, T., A. Anton and F. Gulya, 1999. Effect of Cd, Ni and Cu on some microbial properties of a calcareous chernozem soil. In J. Kubat, (Ed.), Procedures 2nd Symposium on the 'pathways and consequences of the dissemination of pollutants in the Biosphere, pp: 88-102.

- Pahwa, S.K. and K. Bajaj, 0000. Effect of Preemergence Herbicides on the Activity of "-Amylase and Protease Enzyme during Germination in Pigeonpea and Carpetweed, 3: 0253-8040.
- Omar, S.A. and M.A. Abd- Alla, 2000. Microbial populations and enzyme activities in soil treated with pesticides. Egypt. Water Air and Soil Pollute, 127: 49-63.
- Singh, B.K., W. Allan and J.W. Denis, 2002. Degradation of chlorpyrifos, fenamiphos and their effects on soil microbial activity. Environ. Toxicol. Chem., 21: 2600-2605.
- Sarada, R. and Joseph Richard, 1994. Characterization and enumeration of microorganisms associated with anaerobic digestion of tomato processing waste. Bioresource Techno., 49: 261-265.
- Narasimha, G., 1997. Effect of effluent of cotton ginning industry on soil microbial activities. M. Phil., Dissartation submitted to Srikrishnadevaraya University, Anantapur.
- 31. Zaman, M., H.J. Di and K.C. Cameron, 1999. A field study of gross rates of mineralization and Nitrification and their relationships of mineralization and nitrification and their relationships to microbial biomass and enzyme activities in soils treated with dairy effluent and ammonium fertilizer. Soil Use and Management, 15: 188-194.