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Hepatoprotective activity of *Mentha arvensis* Linn. leaves against CCL₄ induced liver damage in rats

Kalpana Patil*, Alka Mall

Department of Pharmacognosy and Phytochemistry, K.L.E University's College of Pharmacy, Belgaum –590 010, Karnataka, India

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ABSTRACT

Objective: To study the Hepatoprotective activity of ethanol, chloroform and aqueous extracts of *Mentha arvensis* leaves against CCL₄ induced liver damage in rats. **Methods:** Hepatotoxicity was induced by CCL₄ and the biochemical parameters such as serum glutamate pyruvate transaminase (sGPT), serum glutamate oxaloacetate transaminase (sGOT), alkaline phosphatase (sALP), serum bilirubin (sB) and histopathological changes in liver were studied along with silymarin as standard Hepatoprotective agents. **Results:** The Phytochemical investigation of the extracts showed presence of flavonoids, steroids, triterpenoids, alkaloids, glycosides, carbohydrates, tannins, phenolic compounds. Treatment of the rats with chloroform, ethanol and aqueous extract with CCL₄ administration caused a significant reduction in the values of sGOT, sGPT, sALP and sB ($P < 0.01$) almost comparable to the silymarin. The Hepatoprotective was confirmed by histopathological examination of the liver tissue of control and treated animals. **Conclusions:** From the results it can be concluded that *Mentha arvensis* possesses Hepatoprotective effect against CCL₄ induced liver damage in rats.

1. Introduction

Mentha arvensis Linn (Lamiaceae) is distributed throughout the Western Himalayas and is cultivated throughout world for use as vegetable[1]. It is used as a carminative, anti-spasmodic, anti-peptic ulcer agent[2]. It contains 70–90% menthol[3]. It is used in oral products such as toothpaste and mouth freshners due to its physiological cooling effects, as fragrance component in soaps, detergents, cosmetic and perfumes, toothpastes[4]. It contains monoterpenes such as menthone, menthofuran, sesquiterpenes, flavonoids, triterpenes, carotenoids, tannins and minerals[5].

Liver is the key organ of metabolism and excretion is continuously and variedly exposed to xenobiotics because of its strategic placement in the body. The toxins absorbed from the intestinal tract gain access first to the liver resulting in a variety of liver ailments. Thus liver diseases remain one of the serious health problems[6]. Conventional or synthetic drugs used in the treatment of liver diseases are sometimes inadequate and can have serious adverse effects[7].

Carbon tetrachloride

(CCL₄) is a widely used hepatotoxic agent in rodents and its trichloromethyl radical (CCL₃) induced toxicity in rat's liver which closely resembles human cirrhosis. Hence, it is an acceptable animal model for analyzing hepatoprotective activity [8]. The survey of literature reveals that the *Mentha arvensis* are found to be used in the traditional system of medicine as a liver tonic. However Hepatoprotective activity of *Mentha arvensis* has not been scientifically investigated. Therefore, in the present study hepatoprotective effect of extracts of *Mentha arvensis* leaves have been evaluated against CCL₄ induced liver damage in the Wister albino rats.

2. Materials and Methods

2.1. Drugs and chemicals

Silymarin was obtained as a gift sample from Cadila Pharma Ltd, India. CCL₄ was obtained as gift sample from Thermo Fisher Scientific India Pvt. Ltd. Standard kit of serum glutamate pyruvate transaminase (sGPT), serum glutamate oxaloacetate trasaminase (sGOT) and alkaline phosphatase (sALP) were obtained from ERBA diagnostic Mannheim GmbH, Mallaustr, Mannheim/Germany. All other reagents used for the experiments were of high analytical grade.

*Corresponding author: Dr.Kalpana Patil, Professor Department of Pharmacognosy, KLEU's College of Pharmacy, Belgaum–590010, Karnataka, India.

Tel.: +91–9449024899

E-mail: kalpatil@yahoo.com

2.2. Plant material

The *Mentha arvensis* leaves were collected from the Miltops Exports, Jamnagar, Gujarat. The herb was authenticated by Dr. Harsha Hegde, Scientist B, Regional Medical Research Centre, ICMR, Belgaum. (Voucher No.RMRC–548).

2.3. Preparation of extracts

The fresh leaves were cleaned, shade dried and then coarse powdered. The dried coarse powder of the leaves was subjected to hot continuous extraction with chloroform, ethanol respectively and cold maceration with distilled water. After complete extraction, the extract was collected and the solvent was distilled off. It was concentrated to dry residue on water bath. The extracts were subjected to preliminary phytochemical investigation. Suspensions of each extract were prepared using 1% Tween 80. CCL₄ was mixed with olive oil in the ratio of 1:1 and subjected for Hepatoprotective activity against CCL₄ induced liver damage.

2.4. Experimental animals

Albino –Wistar rats weighing 150 to 180 g were housed in standard laboratory conditions of temperature [25±2 °C], 12 h light and dark places and with food and water ad libitum.

2.5. Acute oral Toxicity

The acute oral toxicity study was carried out as per the guidelines set by OECD, revised draft guidelines 423, received from CPCSEA, Ministry of Social Justice and empowerment, Govt. of India^[9].

2.6. Evaluation of Hepatoprotective activity

The method used for evaluation of hepatoprotectivity was according to Ranawat^[10] with some minor modifications. In the CCL₄ induced liver toxicity model, CCL₄ (0.5 ml/kg i.p.) was administered daily for 7 days to all animals except group 1. Silymarin (100 mg/kg, p.o) was used as standard. The animals were segregated in to six groups of six each. Group 1, which served as normal control and received 1% Tween 80. Group 2 received 1% Tween 80 p.o. for seven days and served as negative control. Group 3 received silymarin (100 mg/kg, p.o) for seven days. Group 4 received ethanolic extract (375 mg/kg, p.o.) for seven days. Group 5 received chloroform extract (375 mg/kg, p.o.) for seven days. Group 6 received aqueous extract (375 mg/kg, p.o.) for seven days.

The animals were sacrificed 24 h after last treatment under light anesthetic ether. Blood from each rat was withdrawn by retro orbital plexus under ether anesthesia for biochemical investigation i. e. sGOT, sGPT, sALT and bilirubin estimation. Blood was allowed to coagulate at 37°C for 30 min and the serum was separated by centrifugation at 2500 rpm for 15 min. The liver of one animal from each group was

removed and processed immediately for histopathological investigation.

2.7. Histopathological studies

One animal from the each group was utilized for this purpose. The liver specimens obtained from the control and treated groups of animals were fixed in 10% buffered formalin for 24 h. The formalin fixed liver samples were stained with haematoxylin–eosin for photomicroscopic observations of the liver histopathological architecture.

2.8. Statistical analysis

The data are presented as mean±SEM and analyzed by one way ANOVA, followed by Dunnett's 't' test. The results of all the extracts including the standard drug are compared with the result produced by control. And it is considered as significant as $P<0.05$.

3. Results

Preliminary Phytochemical investigation revealed the presence of flavonoids, steroids, triterpenoids in chloroform extract, alkaloids, flavonoids, glycosides, carbohydrates, triterpenoids, tannins, phenolic compounds in ethanolic extract, alkaloids, flavonoids, glycosides, carbohydrates, triterpenoids in aqueous extract.

3.1. Acute toxicity

The extracts were found to be safe in the dose used and there was no mortality up to a dose of 3000 mg/kg b.w. for all extracts. Hence 375 mg/kg b. w. p. o. were selected for the activity.

3.2. Hepatoprotective activity

Administration of CCL₄ induced a marked increase in the serum hepatic levels, sGOT, sGPT, sALP and SB as compared to normal controls indicating liver damage (centrilobular necrosis). Pretreatment of the rats with chloroform, ethanol and aqueous extract prior to CCL₄ administration caused a significant reduction in the values of sGOT, sGPT, sALP and sB ($P<0.01$) almost comparable to the silymarin (Table 1).

3.3. Histopathological results

The Hepatoprotective effect of *Mentha arvensis* leaves was confirmed by histopathological examination of the liver tissue of control and treated animals. The CCL₄ treated liver sections showed severe central vein congestion, sinus congestion, entrilobular degeneration and centrilobular necrosis along with moderate portal triaditis and inflammation. Fatty changes and ballooning of hepatocytes were also showed (Figure 1). The aqueous and chloroform extract treated group showed moderate

Table 1Biochemical assessment of CCL₄ induced liver injury.

Groups	SGPT (IU/L)	SGOT (IU/L)	ALP (IU/L)	Serum bilirubin (mg/dl)	Total bilirubin(mg/dl)
Normal	49.90±0.82	77.85±0.88	93.23±1.24	0.47±0.0045	1.46±0.0099
CCL ₄ induced	225.00±1.34	268.80±0.92	309.4±2.74	1.06±0.0055	4.30±0.012
Standard drug	66.04±0.50***	92.96±0.82***	107.00±0.60***	0.52±0.0062***	1.61±0.0084***
EE treated	74.62±0.73***	103.00±0.62***	121.60±0.96**	0.59±0.0086***	1.74±0.0065***
AQE treated	99.52±0.90**	124.8±0.59**	138.70±0.84**	0.69±0.0048*	2.21±0.0087*
CHE treated	135.90±0.63**	161.4±0.63**	190.70±0.97*	0.79±0.0054	2.62±0.0089

Data are presented as means±SEM (n=6), one way Anova followed by Dunnett's t test.

P<0.05, *P<0.01 vs CCL₄ treated group

central vein congestion, sinus congestion, centrilobular degeneration, centrilobular necrosis and mild inflammation, portal triaditis. The ethanolic extract treated group showed mild central vein congestion, sinus congestion, centrilobular degeneration, centrilobular necrosis but moderate inflammation and portal triaditis. Silymarin treated group had maintained the normal histology with minimal damage.

4. Discussion

CCL₄ metabolism begins with the trichloromethyl free radical (CCL₃) by the action of the mixed function of the cytochrome P450 oxygenase system. This free radical, which is initially formed as relatively unreactive, reacts very rapidly with oxygen to yield a highly reactive trichloromethyl peroxy radical (CCL₃O₂). Both radicals are capable of binding to proteins or lipids, or abstracting a hydrogen atom from an unsaturated lipid, thus, initiating lipid peroxidation^[11–14]. Lipid peroxidation may cause peroxidative tissue damage in inflammation. Therefore, inhibition of the cytochrome P450 dependent oxygenase activity could cause a reduction in the level of toxic reactive metabolites and a decrease in tissue injury. On the other hand, an elevation of plasma sGOT, sGPT, sGALP and bilirubin activities could be regarded as a sign of damage to the liver cell membrane.

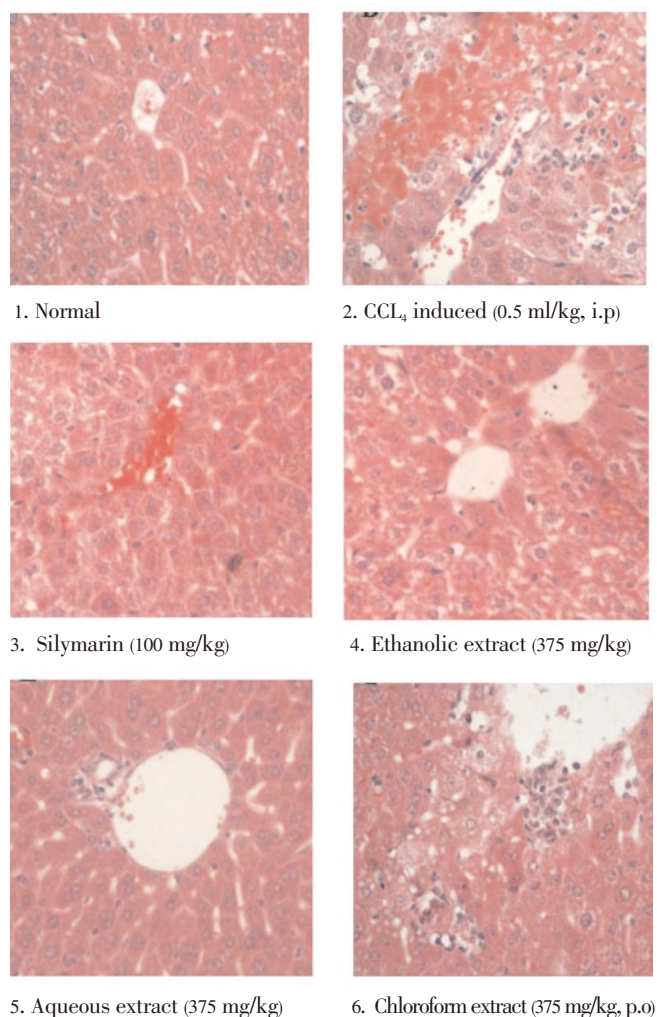
Many compounds known to be beneficial against carbon tetrachloride-mediated liver injury exert their protective action by toxin mediated lipid peroxidation either via a decreased production of CCL₄ derived free radicals or through antioxidant activity of the protective agent themselves^[15].

So in the present study, CCL₄ was employed as toxic agent and the protective effect of *Mentha arvensis* leaves against the CCL₄ induced hepatotoxicity was studied. The extent of toxicity was estimated by histopathological studies and biochemical enzyme markers like sGOT, sGPT, sALP and SB levels. The ethanol extract showed more significant result as compared to other extracts of *Mentha arvensis*. Report shows flavonoids and steroids are may be responsible for Hepatoprotective effect ^[16–19]. Perhaps flavonoids present in the *Mentha arvensis* leaves may be responsible for the marked Hepatoprotective effects, observed in the present study.

It can be concluded that *Mentha arvensis* leaves possess a protective effect against CCL₄ induced hepatotoxicity in rats as evidenced by the biochemical, histological parameters.

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**Figure 1.** Histopathological monograph of extract and standard.1:Normal; 2: CCL₄ (0.5ml/kg i.p) alone; 3:CCL₄+Silymarin(0.5ml/kg+100mg/kg)4: CCL₄+ethanolic extract (0.5ml/kg+375mg/kg); 5: CCL₄+Aqueous extract (0.5ml/kg+375mg/kg); 6: CCL₄+Chloroform extract (0.5ml/kg+375mg/kg)

Conflict of interest statement

We declare that we have no conflict of interest.

References

- [1] Akram M, Uzair M, Sarwer Naila, Asif HM. *Mentha arvensis* Linn.: A review article. *J Med Plants Res*. 2011; **5**(18):4499–503.
- [2] Kowati R, Satish BP, Harsha R, Dinesha R, Hareesh AR. In vitro antioxidant activity of leaves of *Mentha arvensis* Linn. *J Pharm Res*. 2010; **3**(8):1951–4.
- [3] Rastogi RP, Mehrotra BN. Compendium of Indian Medicinal Plants. Lucknow: Central Drug Research Institute; 1999. p.51.
- [4] Alvi MN, Ahmad S, Rehman K, Preparation of menthol Crystals from Mint (*Mentha arvensis*) *Int J Agri Biol*. 2001; **3**(4):527–8.
- [5] Liest, Hrhammer. Phytochemical Studies of Medicinal Plants. *Int J Plant Sci*. 1998; **68**:130–42.
- [6] Karan M, Vasisht K, Handa SS. Antihepatotoxic activity of *Swertia chirata* on carbon tetrachloride induced hepatotoxicity in rats. *Phytother Res* 1999; **13**:24–30.
- [7] Takate S, Pokharkar R, Chopad V. Hepatoprotective activity of the aqueous extract of *Launaea intybacea* Beauv against carbon tetrachloride induced hepatic injury in Albino rats. *J. Pharm Sci Tech* 2010; **2**(7):247–51.
- [8] Al-Shabanah OA, Alam K, Nagi M, Al-Rikabi A, Al-Bekairi A. Protective effect of aminoguanidine a nitric oxide synthetase inhibitor against CCL₄ induced hepatotoxicity in mice. *Life Sci* 2000; **66**:265–70.
- [9] Organization for Economic Co-operation and Development. Guidelines on acute oral Toxicity. Revised Document. October 2005.
- [10] Ranawat L S, Bhatt J, Patel J. Hepatoprotective Activity of Ethanolic Extracts of Bark of *Zanthoxylum aromatum* DC in CCL₄ induced Hepatic Damage in Rats. *J Ethnopharmacol*. 2010; **127**(3): 177–80.
- [11] Brattin WJ, Glende Jr. EA, Recknagel RO. Pathological mechanisms in carbon tetrachloride hepatotoxicity. *J Free Rad Biol Med*. 1985; **1**: 27–8.
- [12] Gosselin RE, Smith RP, Hodge HC. Carbon tetrachloride. Clinical Toxicology of Commercial Products. Williams and Wilkins, Baltimore 1984; 101–7.
- [13] Recknagel RO, Glende Jr. EA, Dolak JA, Waller RL. Mechanisms of carbon tetrachloride toxicity. *Pharmacol Ther*. 1989; **43**:139–54.
- [14] Lee KJ, Jeong HG, Protective effect of *Platycodi radix* on carbon tetrachloride induced hepatotoxicity. *Food chem Toxicol*. 2002; **40**:517–25.
- [15] Hewawasam RP, Jayatilaka KAPW, Pathirana C, Mudduwa LKB. Hepatoprotective effect of *Epaltes divaricata* extract on carbon tetrachloride induced hepatotoxicity in mice. *Indian J Med Res*. 2004; **120**: 30–4.
- [16] Panigrahi S, Panda PK, Patro VJ, Comparative Hepatoprotective activity of different extracts of spirulina against CCL₄ induced liver damage in rats *Inter J Pharma Sci Rev Res* 2010; **4**(2):200–2.
- [17] Pattanayak S, Nayak SS, Panda DP, dinda SC, Shende V, Jadav A. Hepatoprotective activity of crude flavonoids extract of *Cajanus scarabaeoides* (L) in paracetamol intoxicated albino rats. *Asian J Pharm Biol Res* 2011; **1**(1): 22–7.
- [18] Khatri A, Garg A, Agrawal SS. Evaluation of Hepatoprotective activity of aerial part Of *Tephrosia purpuria* L. and stem bark of *Tecomella undulate*. *J Ethnopharmacol* 2009; **122**:1–5.
- [19] Singh D, Gupta RS, Hepatoprotective activity of methanol extract of *Tecomella undulate* against alcohol and paracetamol induced hepatotoxicity in rats. *Life Sci Med Res* 2011.