

RESEARCH ARTICLE

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Screening of Bioactive Metabolites from the Starfish *Pentaceraster mammillatus* against Human Urinary Tract Infectious Pathogens

S. Mohamed Hussain*, M. Mohammed Muneesh, M. Sri Sathya, M. Durai

P.G. and Research Department of Zoology, Jamal Mohamed College (Autonomous), Affiliated to Bharathidasan University, Tiruchirappalli - 620020, Tamil Nadu, India

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ABSTRACT

Presence of different bacterial pathogens in the urinary tract results in the occurrence of infectious diseases among the human beings. The emerging multi drug resistance capacities of pathogens are the major constrain in the treatment of bacterial diseases. Researchers are trying to develop new antimicrobial agents from natural resources including marine organisms to combat the issue. This research work has carried out to investigate the anti-bacterial activity of sea star *Pentaceraster mammillatus* against ten common UTI pathogens. Four different polar solvents *viz;* methanol, acetonitrile, dichloromethane and ethanol were used for the experiment. At the lowest concentration $(250\mu g/ml)$, none of the above said extracts has showed any inhibitory effects in the pathogens; however the concentrations such as 500 and $1000\mu g/ml$ of extracts inhibited the bacterial growth. Acetonitrile and dichloromethane showed higher inhibitory effects on gram positive bacterial strains *Mycoplasma genetalium* (4.3 ± 0.23 mm) and *Staphylococcus aureus* (4.1 ± 0.21 mm) respectively. Ethanolic extract showed some good inhibitory effects on *M. genetalium* (3.5 ± 0.14 mm) and the methanolic extract demonstrated relatively lower inhibition.

Keywords: Starfish, Pentaceraster mammillatus, Antimicrobial activity, UTI.

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*Corresponding author: Dr. S. Mohamed Hussain

Address: P.G. and Research Department of Zoology, Jamal Mohamed College (Autonomous), Affiliated to Bharathidasan University, Tiruchirappalli - 620020, Tamil Nadu, India

Tel.: +91-9944283849

E-mail 🖂: mdhussainjmc@gmail.com

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INTRODUCTION

Sea stars or starfishes are diverse group of animals belonging to the class Asteroidea, comes under the phylum Echinodermata. They are prevalent among the coral reefs, sea grass meadows, deep-sea floor, intertidal zones, coastal areas and in rocky sea shores. ^[1-2] They feed on micro-organisms (algae) and certain small animals (bivalves, sponges, snails) and also absorb organic nutrients and fecal matters from the surrounding water. Starfishes are used for food preparation in China due to the abundance of nutritional metabolites. They are well known for their pharmacological and biological characteristics and they were employed in traditional medicines as a remedial measure for asthma, bronchitis, diabetes and heart problems. ^[3-4]. Presently they are also used as anticancer, anti-bacterial, anti- fungal and antifouling agents. ^[5] It was also reported that a number of novel marine natural products were elucidated from sea stars. ^[6] Studies revealed the presence of glycosylceramides ^[7], asterosaponins, polyhydroxysteroids ^[8] and polyhydroxysteroidal glycosides ^[9-10] in different sea stars across the world.

India is blessed with a rich coastal and marine biodiversity. ^[11] The Gulf of Mannar, which comprised of 140 km area from Rameshwaram island to Tuticorin is recognised as a marine national park and is noted for the rich diversity of various marine organisms having economic importance, that includes different starfishes, in which *Pentaceraster mammillatus* were considered as a relatively new one. ^[12-13] Only a few studies conducted earlier in the sea stars of this coastal area for the elucidation of potential compounds having desirable antimicrobial properties. ^[14-16] A previous study was conducted to purify and characterize a compound (lectins) from the digestive glands of *P. mammillatus* ^[17] and not any single work was performed on their antibacterial efficacy.

The higher incidence of Urinary tract infection (UTI) is a major threat for human beings and the consumption of antibiotics were found effective in controlling the pathogens. ^[18-20] Even though antibiotics at low-doses are effective in their treatment, increased antibiotic resistance in microorganisms is a major constrain for their control. ^[21] Hence the work was carried out to study the antibacterial efficacy of commonly available sea star *P. mammillatus* against UTI pathogens.

MATERIALS AND METHODS

Study area

The peninsular Tuticorin (Lat. 8° 81'N; Long. 78° 14' E) commonly known as 'Pearl City' is a major harbour in Tamil Nadu that lies in the South-east Indian coast. It is a part of Gulf of Mannar marine national park located in the Bay of Bengal. About 36,000 marine species including some rare floral and faunal groups exists in the study area.

Experimental animal and sample collection

Pentaceraster mammillatus is widely distributed in the East coast of Africa, including Madagascar and Mauritius, through Red Sea and Arabian Gulf to Sri Lanka and the Bay of Bengal into the tropical Indo-Pacific. They live in the subtidal on coarse sand or coral rubble. Live specimens of the starfish *P. mammillatus* were collected by scuba diving during January 2018. Using the sea water, the collected samples were cleaned and were immediately stored in ice box and transported to the Zoology laboratory of Jamal Mohamed College, Tiruchirappalli.

Extracts preparation

The standard methodology of Malla Reddy *et al.* ^[22] was followed for the extraction. Different polar solvents namely methanol, acetonitrile, dichloromethane and ethanol was used for extraction. Whole body of starfish samples were placed in the polar solvents and were kept at 22°C for a period of 72 hours. This was filtered through Whatman No. 1. filter paper and then the solvents were kept in rotary evaporator at 30°C. The aqueous suspension were further concentrated and the resultant residues were used as the crude extracts. The extracts were kept at 4°C until the commencement of the experiment.

Antibacterial susceptibility assay

The clinical isolates of selected human urinary tract infectious pathogens were obtained from the Govt. Medical College Hospital, Tiruchirappalli. Antibacterial efficacy of the starfish extracts were assessed by well diffusion technique. ^[23] The bacterial strains were enriched in nutrient broth overnight at 37°C. Then they were streaked over Mueller Hinton agar surface using sterile cotton swabs. Then wells were loaded with 50µl of different extracts at various concentrations (250µg, 500µg and 1000µg/ml). The positive control for the experiment was Streptomycin with the concentration of 400µl and distilled water was used as the negative control. The plates were then incubated for a period of 24 h at 37°C. Antimicrobial activities were determined after 72 h and results were expressed in millimeters.

RESULTS

The antibacterial efficacy of *Pentaceraster mammillatus* was examined against the selected human UTI pathogens on the basis of zone of inhibition. All the four extracts does not exhibited any kind of inhibitory effects at the lowest ($250\mu g/ml$) concentration, whereas an inhibition was recorded in the range of 0.8 to 4.3 mm for different extracts at the median and higher concentrations (500 and $1000\mu g/ml$), and the results were only below the level of the standard drug.

The acetonitrile and dichloromethane extracts showed relatively more inhibition against the gram positive bacterial strains like *Staphylococcus saprophyticus* (3.8 \pm 0.11 and 3.5 \pm 0.41 respectively), *Entercoccus faecalis* (3.1 \pm 0.18 and 3.0 \pm 0.10 respectively), *S. aureus* (3.5 \pm 0.41 and 4.1 \pm 0.21 respectively) and *Mycoplasma genitalium* (4.3 \pm 0.23 and 3.7 \pm 0.21 respectively) (Table 2 and 3).

Ethanolic extracts of the starfish exhibited good inhibitory activity against the gram positive bacterial strains like *S. saprophyticus* (3.0 ± 0.18) , *E. faecalis* (2.8 ± 0.30) , *M. genitalium* (3.5 ± 0.14) and gram negative bacterial strain *Escherichia coli* (2.5 ± 0.34) (Table 4).

Methanolic extracts showed minimal zones of inhibition than the other extracts against the gram positive bacterial strain *S. aureus* (2.5 ± 0.56) and gram negative bacterial strain *Enterobacter cloacae* (2.5 ± 0.40) (Table 1).

DISCUSSION

The available literatures revealed the pharmacological properties of marine derived compounds. ^[24] A number of divers novel bioactive secondary metabolites with potential pharmaceutical and therapeutic properties were obtained from marine organisms and are now being utilized as a source for new drug discovery. ^[25-27] Recent studies demonstrated that the Echinoderms have unique biological properties. ^[28]

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S. Mohamed Hussain et al. / Screening of Bioactive Metabolites from the Starfish Pentaceraster mammillatus.....

				Zone of inhibition	(mm)		
Type of Pathogens	Name of Pathogens	Concentrations of the extract µg/ml					
		250	500	1000	Standard	Control	
Gram positive	Staphylococcus Saprophyticus		1.8 ± 0.25	2.2 ± 0.26	20.0 ± 0.12		
	Enterococcus faecalis		1.8 ± 0.41	2.0 ± 0.18	5.30 ± 0.23		
	Staphylcoccus aureus		2.3 ± 0.51	2.5 ± 0.56	6.40 ± 0.35		
	Mycoplasma genitalium		2.1 ± 0.19	1.9 ± 0.31	10.1 ± 0.18		
Gram negative	Escherichia coli		1.5 ± 0.14	1.3 ± 0.16	18.2 ± 0.31		
Ū.	Klebsiella pneumonia		1.6 ± 0.34	2.4 ± 0.19	6.80 ± 0.36		
	Pseudomonas auroginosa		1.9 ± 0.24	2.3 ± 0.38	18.5 ± 0.16		
	Enterobacter cloacae		2.3 ± 0.37	2.5 ± 0.40	5.10 ± 0.19		
	Klebsiella oxytoca		2.0 ± 0.25	2.2 ± 0.19	6.70 ± 0.18		
	Proteus mirabilis		0.9 ± 0.29	1.3 ± 0.11	6.50 ± 0.35		

Table 1: Antibacterial efficacy of Pentaceraster mammillatus (Methanolic extract)

Table 2: Antibacterial efficacy of Pentaceraster mammillatus (Acetonitrile extract)

Type of Pathogens	Name of Pathogens	Zone of inhibition (mm) Concentrations of the extract µg/ml					
		Gram positive	Staphylococcus Saprophyticus		3.1 ± 0.18	3.8 ± 0.11	20.0 ± 0.12
Enterococcus faecalis			2.7 ± 0.21	3.1 ± 0.18	5.30 ± 0.23		
Staphylcoccus aureus			2.9 ± 0.28	3.5 ± 0.41	6.40 ± 0.35		
Mycoplasma genitalium			3.4 ± 0.12	4.3 ± 0.23	10.1 ± 0.18		
Gram negative	Escherichia coli			1.5 ± 0.51	18.2 ± 0.31		
	Klebsiella pneumonia			1.1 ± 0.42	6.80 ± 0.36		
	Pseudomonas auroginosa			1.3 ± 0.34	18.5 ± 0.16		
	Enterobacter cloacae			1.2 ± 0.14	5.10 ± 0.19		
	Klebsiella oxytoca			1.5 ± 0.32	6.70 ± 0.18		
	Proteus mirabilis			1.8 ± 0.14	6.50 ± 0.35		

Table 3: Antibacterial efficacy of Pentaceraster mammillatus (Dichloromethane extract)

Type of Pathogens	Name of Pathogens	Zone of inhibition (mm) Concentrations of the extract µg/ml					
		Gram positive	Staphylococcus Saprophyticus		2.3 ± 0.32	3.5 ± 0.41	20.0 ± 0.12
Enterococcus faecalis			2.1 ± 0.18	3.0 ± 0.10	5.30 ± 0.23		
Staphylcoccus aureus			2.8 ± 0.26	4.1 ± 0.21	6.40 ± 0.35		
Mycoplasma genitalium			2.7 ± 0.18	3.7 ± 0.21	10.1 ± 0.18		
Gram negative	Escherichia coli			1.5 ± 0.12	18.2 ± 0.31		
	Klebsiella pneumonia			0.8 ± 0.11	6.80 ± 0.36		
	Pseudomonas auroginosa			1.0 ± 0.12	18.5 ± 0.16		
	Enterobacter cloacae			0.9 ± 011	5.10 ± 0.19		
	Klebsiella oxytoca			0.9 ± 0.19	6.70 ± 0.18		
	Proteus mirabilis			0.9 ± 0.19	6.50 ± 0.35		

Table 4: Antibacterial efficacy of Pentaceraster mammillatus (Ethanolic extract)

Type of Pathogens	Name of Pathogens	Zone of inhibition (mm) Concentrations of the extract µg/ml					
		Gram positive	Staphylococcus Saprophyticus		2.5 ± 0.18	3.0 ± 0.18	20.0 ± 0.12
Enterococcus faecalis			2.1 ± 0.34	2.8 ± 0.30	5.30 ± 0.23		
Staphylcoccus aureus				1.1 ± 0.18	6.40 ± 0.35		
Mycoplasma genitalium			2.3 ± 0.41	3.5 ± 0.14	10.1 ± 0.18		
Gram negative	Escherichia coli			2.5 ± 0.34	18.2 ± 0.31		
	Klebsiella pneumonia			1.5 ± 0.25	6.80 ± 0.36		
	Pseudomonas auroginosa			1.3 ± 0.14	18.5 ± 0.16		
	Enterobacter cloacae			1.0 ± 0.28	5.10 ± 0.19		
	Klebsiella oxytoca			1.4 ± 0.61	6.70 ± 0.18		
	Proteus mirabilis				6.50 ± 0.35		

The work was carried out to understand antibacterial efficacy of *Pentaceraster mammillatus* extracts against ten human urinary tract pathogens. Previous reports were also revealed that the common bacterial pathogens of urinary tract can be controlled using different extracts of various sea stars such as *Astropecten indicus* ^[16], *Protoreaster linckii* ^[29], *Ophiocnemis marmorata* and *Stellaster equestris*. ^[30] Marine organisms including the Ascidians such as *Phallusia arabica* ^[31] and *Microcosmus*

exasperatus ^[32] extracts also exhibited antibacterial activity against UTI pathogens. In addition to this, various other marine species were also proven to be effective against the UTI pathogens. They include sea urchin such as *Echinometra mathaei* ^[33], *Tripneustes gratilla* ^[34] and sea sponge *Clathria indica*. ^[35] Previous studies have also described the antibacterial activity of marine algae *Sargassum whitti* ^[36], *Ulva lactuca, Laurencia optusa* and *Turbinaria triquatra*. ^[37]

The starfish Pentaceraster mammillatus were evaluated for their antibacterial potential against human Urinary tract infectious pathogens using the methanol, acetonitrile, dichloromethane and ethanol extracts. Results revealed that the acetonitrile and dichloromethane extracts were having relatively higher inhibitory effects against the pathogens. Hence further studies are required using the purified fractions of acetonitrile and dichloromethane extracts. Isolation, purification and characterization of the compound may recommended for greater effects on UTI pathogens with the minimal dose and this marine natural product will receive overwhelming response from the pharmaceutical industries in the coming future as an alternative method of novel drug development.

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REFERENCES

- Mah C, Nizinski M, Lundsten L. Phylogenetic revision of the Hippasterinae (Goniasteridae; Asteroidea): systematics of deep sea corallivores, including one new genus and three new species. Zoological Journal of the Linnean Society. 2010; 160(2): 266-301.
- Grzimek B, Neil S, Donna O. Grzimek's Animal Life Encyclopedia. Detroit: Gale, 2003.
- 3. Alves RR, Alves HN. The faunal drugstore: Animal-based remedies used in traditional medicines in Latin America. Journal of ethnobiology and ethnomedicine. 2011; 7(1): 9.
- Alves RR, Rosa IL. Zootherapy goes to town: The use of animal-based remedies in urban areas of NE and N Brazil. Journal of ethnopharmacology. 2007; 113 (3): 541-555.
- Sumithaa R, Banu N, Parvathi VD. Novel natural products from marine sea stars. Curr Trends Miomedical Eng Biosci. 2017; 2(4):1-5.
- 6. Walag AMP. Bioactivities of extracts from different marine organisms around the world (2000 to present). Clin Oncol. 2017; 2(11):355-61.
- Chludil HD, Seldes AM, Maier MS. Antifungal Steroidal Glycosides from the Patagonian Starfish Anasterias minuta: Structure– Activity Correlations. Journal of natural products. 2002; 65(2):153-7.
- Popov RS, Ivanchina NV, Kicha AA, Malyarenko TV, Dmitrenok PS, Stonik VA. LC-ESI MS/MS profiling of polar steroid metabolites of the Far Eastern starfish Patiria (= Asterina) pectinifera. Metabolomics. 2016; 12(2):21.
- Lu Y, Li H, Wang M, Liu Y, Feng Y, Liu K, Tang H. Cytotoxic polyhydroxysteroidal glycosides from starfish Culcita novaeguineae. Marine drugs. 2018; 16(3):92.
- Kang JX, Kang YF, Han H. Three new cytotoxic polyhydroxysteroidal glycosides from starfish Craspidaster hesperus. Marine drugs. 2016; 14(10):189.

- 11. Venkataraman K, Wafar M. Coastal and marine biodiversity of India. Indian Journal of Marine Sciences. 2005; 34 (1): 57-75.
- Murugan M, Rajendran N, Kasirajan S, Moorthy P, Balakrishnan G. Diversity assessment of echinoderms from Mudasalodai and Pazhayar in the southeast coast of India. Journal of Coastal Life Medicine. 2016; 4(2):108-13.
- Venkataraman K, Sivaperuman C, Raghunathan C, editors. Ecology and Conservation of Tropical Marine Faunal Communities. Springer; 2013.
- Ely R, Supriya T, Naik CG. Antimicrobial activity of marine organisms collected off the coast of South East India. Journal of experimental marine biology and ecology. 2004; 309(1):121-7
- Devi P, Wahidulla S, Kamat T, D'Souza L. Screening marine organisms for antimicrobial activity against clinical pathogens. Ind. J. Geo-Mar. Sci. 2011; 338-346.
- Chamundeeswari K, Saranya S, Rajagopal S. Exploration of potential antimicrobial activity of sea star astropecten indicus. Journal of Applied Pharmaceutical Science. 2012; 2(7):125.
- 17. Arokya Jothi Blessy A, Basil-Rose MR. Partial purification and characterization of a lectin from the digestive gland of the starfish, Pentaceraster mammillatus. International Journal of Pharmacy and Biological Sciences. 2018; 8(3): 438-444.
- Salvatore S, Salvatore S, Cattoni E, Siesto G, Serati M, Sorice P, Torella M. Urinary tract infections in women. European journal of obstetrics & gynecology and reproductive biology. 2011; 156 (2):131-6.
- Al-Achi A. An Introduction to Botanical Medicines: History, Science, Uses, and Dangers: History, Science, Uses, and Dangers. ABC-CLIO; 2008.
- Wan J, Kaplinsky R, Greenfield S. Toilet habits of children evaluated for urinary tract infection. The Journal of urology. 1995; 154(2):797-9. DOI: 10.1016/S0022-5347(01)67167-2.
- 21. Raz R. Urinary tract infection in postmenopausal women. Korean journal of urology. 2011; 52(12):801-8.
- Malla Reddy S, Srinivasulu M, Satyanarayana N, Kondapi AK, Venkateswarlu Y. New potent cytotoxic lamellarins alkaloids from Indian ascidian Didemnum obscurum. Tetrahedran. 2005; 61: 9242- 9247.
- 23. Reinheimer JA, Demkow MR, Condioti MC. Inhibition of coliform bacteria by lactic acid bacteria. Australian J. Dairy Technol. 1990; 45: 5-9.
- 24. Mayer AM, Rodríguez AD, Berlinck RG, Fusetani N. Marine pharmacology in 2007–8: Marine compounds with antibacterial, anticoagulant, antifungal, anti-inflammatory, antimalarial, antiprotozoal, antituberculosis, and antiviral activities; affecting the immune and nervous system, and other miscellaneous mechanisms of action. Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology. 2011; 153 (2):191-222.
- 25. Levina EV, Kalinovskii AI, Andriyashchenko PV, Dmitrenok PS. Evasteriosides A and B and other sulfated steroids from the Pacific starfish Evasterias retifera. Russian Chemical Bulletin. 2008; 57 (11):2431.
- 26. Kicha AA, Ivanchina NV, Huong TT, Kalinovsky AI, Dmitrenok PS, Fedorov SN, Dyshlovoy SA, Long PQ, Stonik VA. Two new asterosaponins, archasterosides A and B, from the Vietnamese starfish Archaster typicus and their anticancer properties. Bioorganic & medicinal chemistry letters. 2010; 20(12): 3826-30.
- 27. Kumaran RS, Jung H, Kim HJ. In vitro screening of taxol, an anticancer drug produced by the fungus, Colletotrichum capsici. Engineering in Life Sciences. 2011; 11(3):264-71.
- Marmouzi I, Ali K, Harhar H, Gharby S, Sayah K, El Madani N, Cherrah Y, Faouzi ME. Functional composition, antibacterial and antioxidative properties of oil and phenolics from Moroccan Pennisetum glaucum seeds. Journal of the Saudi Society of Agricultural Sciences. 2018; 17(3):229-34.
- Paul JV, Vinoliya J, Mary Mettilda Bai S. Antimicrobial Activity of various Tissue Extracts and Coelomic Fluid of Starfish, Protoreaster linckii (Blainville, 1830). Journal of

Global Trends in Pharmaceutical Sciences. 2018; 9(1): 4905-4916.

- Prabhu K, Bragadeeswaran S. Biological properties of brittle star Ophiocnemis marmorata collected from Parangipettai, Southeast coast of India. J Microbiol Antimicrob. 2013; 5(10):110-8. DOI: 10.5897/JMA2013.0270
- Ananthan G, Sivaperumal P, Hussain SM. Antibacterial potential of marine ascidian phallusia arabica against isolated urinary tract infections bacterial pathogens. Asian Journal of Animal Sciences. 2011; 5(3):208-12.
- Ananthan G, Iyappan K. Investigation of antibacterial potential of Ascidian, Microcosmus exasperatus (Heller, 1878) against human urinary tract pathogens. World Journal of Pharmacy and Pharmaceutical Sciences. 2013; 3(1): 396-403.
- 33. Kazemi S, Heidari B, Rassa M. Antibacterial and hemolytic effects of aqueous and organic extracts from different tissues of sea urchin Echinometra mathaei on pathogenic

streptococci. International Aquatic Research. 2016; 8(4):299-308.

- Abubakar LU, Mwangi CM, Uku JU, Ndirangu SN. Antimicrobial activity of various extracts of the sea urchin Tripneustes gratilla (Echinoidea). African Journal of Pharmacology and Therapeutics. 2012; 1(1):19-23.
- Ravichandran S, Solimabi W, Lisette D, Anbuchezhian R M. Antimicrobial Activity of Marine Sponge Clathria indica (Dendy, 1889). Bioorganicheskaia khimiia. 2011; 37: 483-9.
- 36. Sangeetha J, Gayathri S, Rajeshkumar S. Antimicrobial assessment of marine brown algae Sargassum whitti against UTI pathogens and its phytochemical analysis. Research Journal of Pharmacy and Technology. 2017; 10(6):1905.
- Deyab MA, Abou-Dobara MI. Antibacterial activity of some marine algal extracts against most nosocomial bacterial infections. Egypt. J. Exp. Biol. (Bot.). 2013; 9:281-6.

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