

# **Original Article**

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In vitro efficacy of new synthetic benzimidazole-related compounds against Schistosoma mansoni adult worms

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# ABSTRACT

**Objective:** To evaluate the *in vitro* antischistosomal activity of two new synthetic benzimidazole-related compounds: NBTP-OH and NBTP-F.

**Methods:** *Schistosoma* adult worms were recovered from mice infected with *Schistosoma mansoni* cercaria, washed and then incubated in the culture media with different concentrations of compounds NBTP-OH and NBTP-F up to 72 h. Scanning electron microscopy was conducted to report morphological changes.

**Results:** Incubation of adult *Schistosoma mansoni* with 10 µg/mL of NBTP-OH for 48 h killed 81.25% of worms. The calculated  $LC_{50}$  and  $LC_{90}$  72 h post-incubation were 6.8 µg/mL and 9.8 µg/mL, respectively. Exposure of worms to 10 µg/mL of NBTP-F killed 89.5% of worms after 48 h, mostly males (83.3%), the  $LC_{50}$  and  $LC_{90}$  after 72 h of incubation were 4.8 µg/mL and 6.9 µg/mL, respectively. Worms incubated for 72 h with these compounds revealed swelling and deformity of oral sucker, disorganization and erosion of the tegument when examined with scanning electron microscopy.

**Conclusions:** NBTP-OH and NBTP-F possess *in vitro* antischistosomal activities; however, *in vivo* studies should be conducted to examine their antischistosomal effects..

**KEYWORDS:** Schistosoma mansoni; Benzimidazole; Tubulin; Tegument; In vitro

## 1. Introduction

Schistosomiasis caused by helminthes of the genus Schistosoma is a

chronic debilitating disease afflicting more than 290 million people in 78 countries most of whom live in Africa. The disease annual loss is approximately 4.5 million disability adjusted life years (DALYs), which are expected to increase in the coming days[1]. *Schistosoma (S.) mansoni*, causing intestinal schistosomiasis, is associated with a wide range of chronic pathology including hepatosplenomegaly, portal hypertension and gastrointestinal varices[2].

To date, there is no vaccine available for schistosomiasis and controlling the disease depends solely on praziquantel (PZQ), which interferes with calcium haemostasis, causing spastic paralysis of worms; however, the appearance of PZQ-resistant strains in laboratory and field raised major concerns and called for development of novel antischistosomal drugs[3]. One important rationale to develop a new schistosomicidal agent is to target a protein that interferes with the worm vital function as feeding and locomotion.

Tubulin is a protein composed mainly of two homologous polypeptides  $\alpha$  and  $\beta$ , both assembled together to form heterodimer in microtubules, the major component of eukaryotic cytoskeleton. In addition, microtubules play a vital role in structural support, as well as cell division, motility, transport and signaling mechanisms<sup>[4]</sup>.

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In trematodes, the cytoskeleton has vital roles in maintaining the tegumental structure, transportation of tegumental granules from tegumental cells to the syncytium, thus it is directly involved in the synthesis and turnover of the surface membrane and its outer coat<sup>[5]</sup>. Previous research recorded the presence of tubulin in the tegument cytoskeleton of schistosomes. Within the syncytium, tubulin appeared as vertical lines passing throughout its thickness, in the cell bodies, their cytoplasmic processes, tubercles and parenchymal cells<sup>[6]</sup>. It is worthy of noting that tubulin is a target of several agents such as anticancers, antigouts, antifungal and antihelmintics<sup>[7,8]</sup>.

 $\beta$  tubulin is the target of benzimidazoles, upon binding, these drugs interfere with microtubules assembly leading to disruption of spindle formation during cell division and interruption of the movement of subcellular components and metabolites within the cytoplasm[7].

Benzimidazoles as albendazole, thiabendazole, and flubendazole, have wide-spectrum activity against cestodes and various nematodes affecting human. In addition to triclabendazole, the drug was used in the treatment of fascioliasis[9].

The antischistosomal effects of triclabendazole and flubendazole have been reported experimentally by several researchers[10–13]. More recently, a novel benzimidazole derivative named BTP-Iso showed promising *in vivo* and *in vitro* effects on adult *S. mansoni*[14,15] as significant reductions in immature ova, hepatic

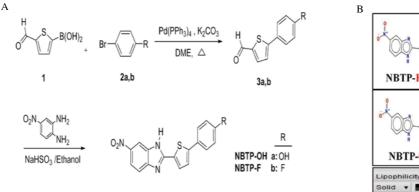
lesions, and recovered adult worms particularly females, as well as marked tegumental changes were seen by scanning electron microscopy. These data highlight the potential antischistosomal activity of benzimidazole-derived compounds, making them appropriate alternatives to PZQ.

In this *in vitro* study, we tested the effect of two benzimidazolederived compounds: NBTP-OH and NBTP-F on adult *S. mansoni*. The efficacy was assessed based on the percentage of dead worms after treatment and morphological changes recorded by scanning electron microscopy.

# 2. Materials and methods

#### 2.1. Compounds and synthetic procedure

Compounds NBTP-OH: 4-[5-(6-nitro-1H-benzimidazol-2-yl) thiophen-2-yl]phenol and NBTP-F: 2-[5-(4-Fluorophenyl)thiophen-2-yl]-6-nitro-1H-benzimidazole are novel benzimidazole derivatives synthesized as shown in (Figure 1A). Physical and chemical properties of the two compounds are shown in (Table 1). The calculated lipophilicity of NBTP-OH and NBTP-F are 4.9 and 5.7 respectively (Figure 1B).



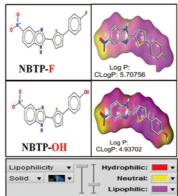


Figure 1. (A) Synthetic pathway of compounds NBTP-OH and NBTP-F; (B) Structure and calculated lipophilicity (clogP) of both compounds.

Table 1. Some physical and chemical properties of NBTP-OH and NBTP-F.

Items	NBTP-OH	NBTP-F
Structure	O <sub>2</sub> N N S OH	O <sub>2</sub> N N S
Chemical name	4-[5-(6-Nitro-1H-benzimidazol-2-yl)thiophen-2-yl]phenol	2-[5-(4-Fluorophenyl)thiophen-2-yl]-6-nitro-1H-benzimidazole
Physical characters	Ruddy crystals	Ruddy crystals
Melting point (Mp) °C	> 300	295-297
Yield (%)	73	71
Chemical formula	$C_{17}H_{11}N_3O_3S$	$C_{17}H_{10}FN_{3}O_{2}S$
Molecular weight (g)	337.35	339.34
Lipophilicity (ClogP)	4.937 02	5.707 56 (more lipophilic)
Molarity (mole/L)	337.35 g/1 000 mL	339.34 g / 1 000 mL

For synthesis of the two compounds, we adopted Suzuki-Miyaura reaction<sup>[16]</sup> and 5-formyl thiophen-2-ylboronic acid (1) was coupled with the 1-bromo-4-hydroxy benzene (2a) to provide 5-(4-hydroxyphenyl) thiophene-2-carbaldehyde (3a)<sup>[17]</sup>. Likewise, 5-(4-fluorophenyl) thiophene-2-carbaldehyde (3b) was synthesized by Bussolari and Rehborn<sup>[18]</sup>. In the present work, the appropriate aldehydes 3a and 3b were reacted with 4-nitrobenzene-1,2-diamine in ethanol in the presence of sodium bisulfite to afford 4-[5-(6-nitro-1*H*-benzimidazol-2-yl)thiophen-2-yl]phenol (NBTP-OH) and 2-[5-(4-Fluorophenyl)thiophen-2-yl]-6-nitro-1H-benzimidazole (NBTP-F) respectively. PZQ used in the experiments was purchased from Sigma-Aldrich.

### 2.2. Parasites and animals

Subcutaneous injection of freshly shed cercariae from infected *Biomphalaria glabrata* snails were used to infect CD1 mice using 400 cercariae/mouse. Adult *S. mansoni* worms (Egyptian strain) were obtained by portal perfusion 42 d post-infection. Recovered worms were washed in phosphate buffered saline (PBS) to remove host blood cells then incubated in the culture media.

Infected snails and mice used in the study were obtained from the Schistosome Biological Supply Centre (SBSC), Theodor Bilharz Research Institute (TBRI), Giza, Egypt. Animals were maintained and experiments were carried out at the SBSC/TBR and Mansoura Faculty of Medicine.

## 2.3. In vitro schistosomicidal activity

The *in vitro* schistosomicidal assay was performed on adult worms, according to the method of Yousif *et al.*[19] with a few modifications. Roswell Park Memorial Institute medium (RPMI-1640), supplemented with L-glutamine, 10 mM HEPES, 20% heat-inactivated fetal bovine serum (FBS), 300 U/mL penicillin, 300 µg/mL streptomycin, and 160 µg/mL gentamycin (all from Gibco) were used for adult worms incubation in 24-well culture plates (Costar), each well contained 5 pairs of worms. Adult *S. mansoni* were incubated for 2 h in the culture medium to be allowed for adaptation.

Compounds NBTP-OH and NBTP-F were dissolved in dimethylsulfoxide (DMSO) and added to wells to give concentrations from 2 to 40 µg/mL. Controls used include worms incubated with 10 µg/mL PZQ (reference drug) and those incubated with RPMI-1640 only or with 0.8% DMSO (highest concentration of vehicle used). Each assay was performed at least three times in duplicate. Plates were incubated in a humidified atmosphere of 5%  $CO_2$  at 37 °C for 3 d.

Before incubation with drug and immediately after incubation, all

worms were observed under dissecting microscope. Then, treated and control worms were examined every 24 h under a dissecting microscope to record motility, integument damage, pairing changes and dead worms. Treated worms that did not exhibit motility for two minutes were considered dead. Based on the observations, mortality rates, and the  $LC_{50}$  and  $LC_{90}$  of each compound against adult *S. mansoni* were calculated using SPSS version 16.

#### 2.4. Scanning electron microscopy

For ultrastructural analysis of the compounds activity on adult *S. mansoni*, scanning electron microscopy was used. Control and treated adult worms were washed with PBS, fixed in 2.5% glutaraldehyde-phosphate buffer for 24 h at 4 °C, then post fixed in osmium tetroxide buffer (1%) for 1 h. Dehydration of adult worms were done through a graded series of ethanol (from 25% to 100%, for 10 min in each ethanol grade), and critical-point dried through carbon dioxide then worms were embedded in epon resin, coated with gold particles before examination with electron microscopy (Inspect S, FEI Company, Holland).

## 2.5. Ethics statement

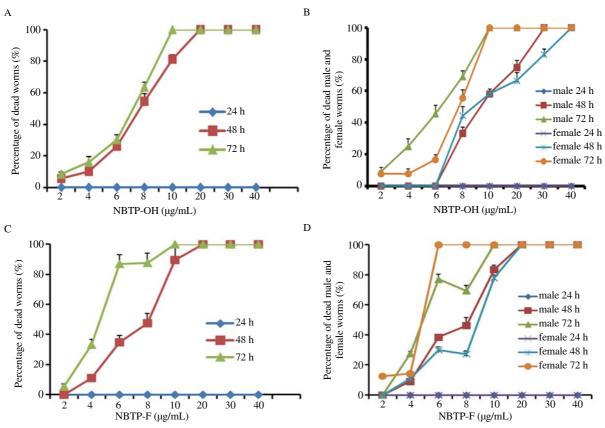
All the study procedures were approved by Mansoura Faculty of Medicine Institutional Research Board (MFM-IRB approval number: R.19.12.701), Mansoura University, Mansoura, Egypt and complied with the international valid animal ethics guidelines.

## **3. Results**

### 3.1. In vitro schistosomicidal activity

Worms incubated in media containing 0.8% of DMSO for 72 h survived; male and female schistosomes showed normal tegumental appearance and motor activity characterized by wavy movement along the body axis. While PZQ-treated worms presented with muscle contraction immediately and with incubation, all worms died in 24 h.

Treatment of adult schistosomes with different concentrations (2-40  $\mu$ g/mL) of compound NBTP-OH for 24 h resulted in changing the worm behavior in the form of coiling and slow locomotion. After 48 h exposure to 8  $\mu$ g/mL, 54.6% of the worms died (Figure 2A), female worms (44.4%) were more affected than males (Figure 2B). With increasing the incubation time for additional 24 h, 63.6% of worms died but mostly males (Figure 2B). At 10  $\mu$ g/mL, 81.25% of worms died after 48 h of incubation and the remaining worms died over the next 24 h (Figure 2B). A 20  $\mu$ g/mL of compound



**Figure 2.** Dose- and time- response curves of adult *Schistosoma mansoni* treated with either NBTP-OH or NBTP-F at concentrations of 2–40 µg/mL for 72 h. (A) The average percentage of total dead worms exposed to NBTP-OH; (B) The average percentage of dead male and female schistosomes exposed to NBTP-OH. (C) The average percentage of total dead worms exposed to NBTP-F; (D) The average percentage of dead male and female schistosomes exposed to NBTP-F. Death of worms was documented after 2 min of no movement under the dissecting microscope. Note that the intervals on the X-axis are not equal.

NBTP-OH killed all worms after 48 h. The calculated  $LC_{50}$  and  $LC_{90}$  of compound NBTP-OH after 72 h incubation were 6.8 µg/mL and 9.8 µg/mL, respectively.

Exposure of worms to 8 µg/mL of compound NBTP-F for 48 h resulted in death of 47.5% of worms and the death rate was higher in male than female worms (Figure 2C and 2D), while 10 µg/mL resulted in 89.5% death rate at the same time point (Figure 2C), the mortality rate in male worms was 83.3% (Figure 2D). All worms died after 48 h at 20 µg/mL, and at 10 µg/mL after 72 h incubation (Figure 2D). Moreover, NBTP-F compound  $LC_{50}$  and  $LC_{90}$  were 4.8 µg/mL and 6.9 µg/mL, respectively, while PZQ  $LC_{50}$  and  $LC_{90}$  were 0.2 µg/mL and 0.3 µg/mL, respectively. NBTP-F showed a more potent schistosomicidal activity than NBTP-OH against *S. mansoni* worms, although not as potent as PZQ as detected from  $LC_{50}$  and  $LC_{90}$ .

Separation of paired worms was detected after 24 h incubation with both compounds. The percentages of separated worms were 46.7% and 86.7% for NBTP-OH and NBTP-F, respectively.

## 3.2. Scanning electron microscopy results

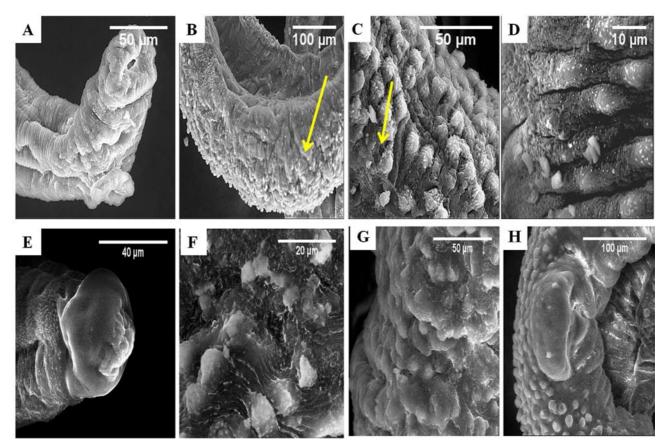
To report the possible morphological changes caused by compounds NBTP-OH and NBTP-F, adult *S. mansoni* worms treated with

DMSO or 8 µg/mL of each compound for 48 h were examined using SEM. Worms treated with compound NBTP-OH revealed deformity of the oral sucker and anterior part of male worms, shrunken body with swellings and furrows (Figure 3A). Deformity of the dorsal aspect of the worm and gynaecophoric canal with presence of blebs, erosions and vesicles, as well as swelling of tubercles and ridges with peeling of the tegument (arrow) and few or even absence of spines were also seen (Figure 3B, 3C and 3D).

Male worms exposed to compound NBTP-F showed swollen, shrunken and deformed oral sucker and anterior part of the body (Figure 3E). Moreover, erosion, disorganization, blebs and fissures along with swelling and fusion of tegumental folds, in addition to fusion and collapse of tubercles, and deformity of the gynaecophoric canal were recorded (Figure 3F, 3G and 3H).

Adult male exposed to culture media only has normal surface membrane with no apparent destruction and normal body structure (Figure 4A), while DMSO-treated worms, the tegument is intact showing many tubercles with spines (Figure 4B).

Unfortunately, we did not examine PZQ-treated worms as very few worms were recovered from PZQ-treated mice and nearly all of them were destructed and/or lost during preparation for scanning electron microscopy examination.



**Figure 3.** Worms exposed to NBTP-OH. (A) Deformity of oral sucker and the anterior part of male, shrunken body with swelling. (B) Deformity of the dorsal aspect and gynaecophoric canal with erosions. (C, D) Swelling of tubercles and tegument erosions (arrow) with few spines on the tubercles. (E) Worms exposed to NBTP-F showed deformed male oral sucker and anterior body part. (F) Erosion, disorganization, blebs and fissures of tegument. (G) Swelling of tegumental folds and collapse of tubercles. (H) Disorganization of male tegument and deformity of the gynaecophoric canal.

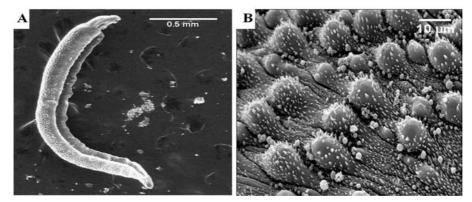


Figure 4. (A) Adult male exposed to media only has normal surface membrane with no apparent destruction and normal body structure. (B) DMSO-treated worms, the tegument is intact showing many tubercles with spines. NO PZQ-treated worms were examined as very few worms were recovered and nearly all lost and/or destructed during preparation for examination by scanning electron microscopy.

#### 4. Discussion

Schistosomiasis is a major neglected public health disease that comes second after malaria in many tropical and subtropical countries. To date, the cornerstone for control of human schistosomiasis depends on PZQ; therefore, it is urgent to identify new alternatives.

Globally, many studies have scientifically assessed numerous natural and synthetic compounds on *S. mansoni* worm[14,20–22], however, a lot of them still remain unused, pending their efficacy

on the parasite biology is confirmed. It is realized that the study of parasite biology is fundamental to potential drug discovery and development.

In this *in vitro* study, we assessed the effectiveness of 2-40 µg/mL of two structurally-related synthetic benzimidazoles compounds: NBTP-OH, and NBTP-F, on *S. mansoni* adult worms. The results clearly recorded the evidence for their promising schistosomicidal effects. NBTP-F showed a more potent schistosomicidal activity than NBTP-OH against *S. mansoni* worms, although not as potent as PZQ as detected from  $LC_{50}$  and  $LC_{90}$ .

In this work, we observed that male worms were more sensitive to the tested compounds especially 48 h post-treatment. Male schistosomes were more susceptible than females as described in some *in vitro* studies[21,23]. Others reported that females were more susceptible[20,24], whereas other researchers did not record any differences between male and female worms sensitivity to the tested drugs[25]. The greater sensitivity of male worms to both compounds in spite of their thick tegument in comparison to females might be due to a sex-specific interference of the drug with the target or presence of different targets in males compared to females[26].

The thick tegument is a major target for antischistosomal agents because of its essential roles for schistosomes. In order to test the effect of these new agents on the tegument and surface membrane of the worm, we used scanning electron microscopy. The main change induced by treatment with the two compounds was destruction of the tegument.

Upon disruption of the tegumental coat, the drug can deeply penetrate into the internal structures to bind another target as muscles of the parasite or causing disturbance in the normal physiological and biochemical process of the worm. Similar results have been previously reported by many researchers when testing the *in vitro* effect of several compounds on schistosomes such as PZQ, oxamniquine, artemether, mefloquine, miltefosine and artemisininbased combination therapies[14,22,27,28].

In a previous study, we tested the compound BTP-Iso, another new synthetic benzimidazole compound<sup>[15]</sup>, which showed a lethal effect nearly similar to compound NBTP-OH, although it is more lipophilic. The *in vitro* schistosomicidal activity of the three compounds against *S. mansoni* adult worms ranked as NBTP-F, followed by BTP-Iso, and then NBTP-OH.

Results obtained from the two compounds tested in this study go with the calculated lipophilicity as compound NBTP-F, which is more lipophilic, was found more effective. Lipophilic agents are more able to penetrate the membranes including tegument. The tegument of schistosomes is a normal plasma membrane overlain by a membrane-like secretion known as the membranocalyx<sup>[29]</sup>.

Herein, the precise mechanism of action of these compounds has not yet been clarified. But we can hypothesize that these benzimidazole compounds can interfere with tegument structure and function (based on scanning electron microscopy results) through penetration of membranocalyx to interact with tubulin in syncytium, cell bodies and the subtegumental structures including microtubulelined cytoplasmic channels, which are responsible of transporting secretory inclusions synthesized in the cell bodies to the tegument to form the secreted membranocalyx[29].

The tegument is a known essential structure for schistosomes subsistence, nutritional support, lipid metabolism, motility, proliferation, and host immune evasion[22,30]. Therefore, interference of one of these mechanisms could be lethal to the adult worm.

Also, deformity and damage in both oral and ventral suckers, particularly in male worms were observed with exposure to both compounds. If this effect is the same *in vivo*, both compounds will affect worm nutrition and subsequent interference of worm maturation and female oviposition besides elimination of the worm from the circulation<sup>[31]</sup>.

Another possible mechanism could be considered as the tested compounds are structurally-related to benzimidazoles, thus the mode of action could be alike. Benzimidazoles are well-known microtubule inhibitors; upon their binding to  $\beta$ -tubulin, they cause depolymerization of cytoplasmic microtubules, instability of the muscular skeleton and disturbance of microtubule-based process in helminthic parasites in the suppression of mitosis in vitelline and spermatogenic cells[7].

Separation of paired worms was detected after incubation with compound NBTP-OH, but was observed more with NBTP-F. This effect could be due to loss of some tubulin functions, as tubulin is one of the components of the worm musculature. Muscles are located in the gynaecophoric canal to retain mating between males and females worms. Separated female schistosomes are incapable of eggs production, the key factor in schistosomiasis sequelae.

Generally, *in vitro* studies do not cover anthelmintic activities of the tested compounds, particularly with respect to pharmacological and immunological interaction with the host; however, they provide the first evidence of anthelmintic activity and insight into the mode of action of these agents[32]. Docking studies and molecular dynamics simulation predicted NBTPs bind tubulin with high stability (unpublished data). Therefore, it would be worthwhile to test for this hypothesis; also it is interesting to investigate "based on modeling results" whether these compounds could exert such lethal effect *in vivo* on different *Schistosoma* species and on other trematodes and nematodes as well.

## **Conflict of interest statement**

The authors declare that there are no conflicts of interest.

## **Authors' contributions**

AT initiated the idea, participated in the experimental work and wrote the manuscript. SB, MT and MA conduced the experimental work, read and approved the final manuscript. BM prepared the two tested compounds, read and approved the final manuscript. FS and SNB participated in the experimental work, revised and edited the manuscript.

#### References

 WHO. Schistosomiasis fact sheet. 2020. [online]. Availavle from: http:// www.who.int/en/news-room/fact-sheets/detail/schistosomiasis [Accessed on 31 March 2020].

- [2] McManus DP, Dunne DW, Sacko M, Utzinger J, Vennervald BJ, Zhou XN. Schistosomiasis. *Nat Rev Dis Primers* 2018; 4(1): 13.
- [3] Doenhoff MJ, Hagan P, Cioli D, Southgate V, Pica-Mattoccia L, Botros S, et al. Praziquantel: Its use in control of schistosomiasis in sub-Saharan Africa and current research needs. *Parasitology* 2009; **136**(13): 1825-1835.
- [4] Singh P, Rathinasamy K, Mohan R, Panda D. Microtubule assembly dynamics: An attractive target for anticancer drugs. *IUBMB Life* 2008; 60(6): 368-375.
- [5] Van Hellemond JJ, Retra K, Brouwers JFHM, van Balkom BWM, Yazdanbakhsh M, Shoemaker CB, et al. Functions of the tegument of schistosomes: Clues from the proteome and lipidome. *Int J Parasitol* 2006; **36**(6): 691-699.
- [6] Tansatit T, Sahaphong S, Riengrojpitak S, Viyanant V, Sobhon P. Immunolocalization of cytoskeletal components in the tegument of the 3-week-old juvenile and adult *Fasciola gigantica*. *Vet Parasitol* 2006; 135(3-4): 269-278.
- [7] Lacey E. The role of the cytoskeletal protein, tubulin, in the mode of action and mechanism of drug resistance to benzimidazoles. *Int J Parasitol* 1988; **18**(7): 885-936.
- [8] Pullagura MKP, Avdhut Kanvinde S, Raja S. Potent biological agent benzimidazole-a review. Int J Pharm Pharm Sci 2016; 8(12): 22-33.
- [9] Taman A, Azab M. Present-day anthelmintics and perspectives on future new targets. *Parasitol Res* 2014; **113**: 2425-2433.
- [10]el Sayed MH, Allam AF. Effect of triclabendazole on the tegument of Schistosoma mansoni: A scanning electron microscopic study. J Egypt Soc Parasitol 1997; 27(1): 143-152.
- [11]Khalil SS. On the schistosomicidal effect of triclabendazole an experimental study. J Egypt Soc Parasitol 2000; 30(3): 799-808.
- [12]Nessim NG, Hassan SI, William S, el-Baz H. Effect of the broad spectrum anthelmintic drug flubendazole upon *Schistosoma mansoni* experimentally infected mice. *Arzneim Forsch* 2000; **50**(12): 1129-1133.
- [13]William S, Guirguis F, Nessim NG. Effect of simultaneous and/ or consecutive administration of the broad spectrum anthelmintic flubendazole together with praziquantel in experimental *Schistosoma mansoni* infection. *Arzneim Forsch* 2003; **53**(7): 532-537.
- [14]El Bialy SA, Taman A, El-Beshbishi SN, Mansour B, El-Malky M, Bayoumi WA, et al. Effect of a novel benzimidazole derivative in experimental *Schistosoma mansoni* infection. *Parasitol Res* 2013; **112**(12): 4221-4229.
- [15]Taman A, El-Beshbishi SN, Bardicy SE, Tadros M, Ayoub M, Mansour B, et al. *In vitro* screening of BTP-Iso on *Schistosoma mansoni* and its intermediate host *Biomphalaria alexandrina*. *Asian Pac J Trop Dis* 2016; 6(12): 946-951.
- [16]Miyaura N, Yamada K, Suzuki A. A new stereospecific cross-coupling by the palladium-catalyzed reaction of 1-alkenylboranes with 1-alkenyl or 1-alkynyl halides. *Tetrahedron Lett* 1979; **20**(36): 3437-3440.
- [17]Costa SPG, Batista RMF, Cardoso P, Belsley M, Raposo MMM.2-Arylthienyl-substituted 1,3-Benzothiazoles as new nonlinear optical

chromophores. Eur J Org Chem 2006. doi: org/10.1002/ejoc.200600059.

- [18]Bussolari JC, Rehborn DC. Preparation of 5-Aryl furfurals and Aryl thiophene-2-carboxaldehydes via palladium-catalyzed C-C bond formation in aqueous media. *Org Lett* 1999; 1(7): 965-967.
- [19]Yousif F, Hifnawy MS, Soliman G, Boulos L, Labib T, Mahmoud S, et al. Large-scale *in vitro* screening of Egyptian native and cultivated plants for schistosomicidal activity. *Pharm Biol* 2007; **45**(6): 501-510.
- [20]Manneck T, Haggenmuller Y, Keiser J. Morphological effects and tegumental alterations induced by mefloquine on schistosomula and adult flukes of *Schistosoma mansoni*. *Parasitology* 2010; **137**(1): 85-98.
- [21]de Melo NI, Magalhaes LG, de Carvalho CE, Wakabayashi KA, de PAG, Ramos RC, et al. Schistosomicidal activity of the essential oil of *Ageratum conyzoides* L. (Asteraceae) against adult *Schistosoma mansoni* worms. *Molecules* 2011; **16**(1): 762-773.
- [22]Bertão HG, da Silva RA, Padilha RJ, de Azevedo Albuquerque MC, Radis-Baptista G. Ultrastructural analysis of miltefosine-induced surface membrane damage in adult *Schistosoma mansoni* BH strain worms. *Parasitol Res* 2012; **110**(6): 2465-2473.
- [23]Sanderson L, Bartlett A, Whitfield PJ. In vitro and in vivo studies on the bioactivity of a ginger (Zingiber officinale) extract towards adult schistosomes and their egg production. J Helminthol 2002; 76(3): 241-247.
- [24]Barth LR, Fernandes APM, Ribeiro-Paes JT, Rodrigues V. Effects of Goyazensolide during *in vitro* cultivation of *Schistosoma mansoni*. *Mem Inst Oswaldo Cruz* 1997; 92(3): 427-429.
- [25]Moraes J, Nascimento C, Lopes PO, Nakano E, Yamaguchi LF, Kato MJ, et al. *Schistosoma mansoni: In vitro* schistosomicidal activity of piplartine. *Exp Parasitol* 2011; **127**(2): 357-364.
- [26]Keiser J, Chollet J, Xiao SH, Mei JY, Jiao PY, Utzinger J, et al. Mefloquine-an aminoalcohol with promising antischistosomal properties in mice. *PLoS Negl Trop Dis* 2009; **3**(1): e350.
- [27]Keiser J, Manneck T, Vargas M. Interactions of mefloquine with praziquantel in the Schistosoma mansoni mouse model and in vitro. J Antimicrob Chemother 2011; 66(8): 1791-1797.
- [28]Kato K, Miura M, Mitsui Y. In vitro effects of amodiaquine on paired Schistosoma mansoni adult worms at concentrations of less than 5 μg/mL. Mem Inst Oswaldo Cruz 2013; 108(2): 192-196.
- [29]Braschi S, Borges WC, Wilson RA. Proteomic analysis of the schistosome tegument and its surface membranes. *Mem Inst Oswaldo Cruz* 2006; 101(Suppl 1): 205-212.
- [30]Faghiri Z, Skelly PJ. The role of tegumental aquaporin from the human parasitic worm, *Schistosoma mansoni*, in osmoregulation and drug uptake. *FASEB J* 2009; 23(8): 2780-2789.
- [31]Halton DW, Maule AG. Flatworm nerve-muscle: Structural and functional analysis. *Can J Zool* 2004; 82(2): 316-333.
- [32]Halferty L, O'Neill JF, Brennan GP, Keiser J, Fairweather I. Electron microscopical study to assess the *in vitro* effects of the synthetic trioxolane OZ78 against the liver fluke, *Fasciola hepatica*. *Parasitology* 2009; **136**(11): 1325-1337.