



## Detection of Entamoeba in river water using multiplex-PCR

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### ABSTRACT

In the last 20 years, molecular technique based on 16S rRNA gene and other genetic markers have been developed for bacteriological and analysis of lakes, soil. PCR based pathogen detection resolved some of the problems encountered using conventional methods. PCR has a detection limit of <10 copies of specific gene present in mixed sample. PCR reaction are designed to either amplify a single product or to use several primer pairs as part of multiplex PCR.

**Keywords:** multiplex PCR, multiplex PCR, Entamoeba

### INTRODUCTION

Entamoeba is a genus of phylum Amoebozoa. It contains many species, six of which – *Entamoeba histolytica*, *Entamoeba dispar*, *Entamoeba moshkovskii*, *Entamoeba polecki*, *Entamoeba coli* and *Entamoeba hartmanni* resides in the human intestinal lumen. Among these the only species responsible for the pathological sequelae in humans is *Entamoeba histolytica*, others as non-pathogenic parasites. These parasites have a global distribution and the disease it causes is usually characterized by diarrhea. (Fotedar *et al.*, 2007).

Though *E. histolytica*, *E. dispar* and *E. moshkovskii* are morphology identical, they can be differentiated at molecular level. (Tanyuksel and Petri 2003; Fotedar *et al.*, 2007).

The morphological similarities among the pathogenic *E. histolytica*, non-pathogenic *E. dispar* and free-living *E. moshkovskii* has made the differential diagnosis of the three species by PCR necessary.

PCR – based approaches are the best method to achieve desired result for clinical and epidemiological studies in the developed countries. (Calderaro, *et al.*, 2006; Hamzah, *et al.*, 2006, Haque, *et al.*, 2006 and Zaki, *et al.*, 2002). PCR assay has been utilized to increase sensitivity and specificity of Entamoeba diagnosis in a variety of clinical specimens including fecal and liver abscess pus samples. (Anane and Khalid, 2005; Paul, *et al.*, 2007; Kurt, *et al.*, 2008).

PCR by amplifications of small subunit r RNA gene (SSU – rDNA) was reported to be 100 times more sensitive than ELISA for *E. histolytica* detection. (Mirelaman, *et al.*, 1997; Fotedar, *et al.*, 2007).

Small subunit r RNA gene (SSU – rDNA) is widely used as a target for the differential detection of Entamoeba species, as this target is present in multi copies when compared to DNA target present in single - copy gene, present on extra – chromosomal plasmids. (Bhattacharya, *et al.*, 1989).

Several Molecular Diagnostic tests including Conventional nested and real – time PCR has been developed for the detection and differentiation of *Entamoeba histolytica*, *Entamoeba dispar*, *Entamoeba moshkovskii* in clinical samples. (Fotedar, *et al.*, 2007). Fode – Vaughan, *et al.*, (2003) applied a simple and rapid Direct PCR (DPCR) approach for the detection of *E. coli* 0157: H7 in environmental water samples and milk using the untreated samples directly as a template in PCR and eliminating the steps of cell recovery or DNA extraction prior to PCR.

Santos, *et al.*, (2010) developed a PCR procedure followed by the DNA sequencing analysis of specific regions of 18S r RNA gene to differentiate Entamoeba species commonly found in human’s stools.

#### PCR BASED DETECTION OF *E. COLI*:

Escherichia coli are gram-negative coliform bacteria which grow abundantly in lower intestine of warm-blooded animals, expelled through feces and are easily culture-able (Winfield and Groisman, 2003).

*E. coli* is a clonal species including both commensal and pathogenic strains. As a pathogen *E. coli* is best known for its ability to cause intestinal diseases, Urinary Tract Infection in the brain stem.

Six classes or virotypes of *E. coli* that caused diarrheal diseases and now recognized and classified by the method of pathogenesis as:

1. Enteropathogenic *E. coli* (EPEC)
2. Enterotoxigenic *E. coli* (ETEC)
3. Enteroinvasive *E. coli* (EIEC)
4. Enterohaemorrhagic *E. coli* (EHEC)
5. Enteroaggregative *E. coli* (EAaggEC)
6. Diffusely Adherent *E. coli* (DAEC)

Each class falls within a serological subgroup and manifests distinct features in pathogenesis. (Nataro and Kaper, 1998).

*E. coli* is a bacterium commonly found in the intestine of human and other mammals. Though most of the *E. coli* strains are harmless commensal, some strains can cause severe diseases. These pathogens are transmitted through consumption contaminated drinking water and foods such as raw or undercooked ground meat products, raw milk and even vegetables. (Kaper, *et al.*, 2004).

Among the indicators of fecal contamination *E. coli* is considered the most reliable one, as its presence directly relates fecal contaminations with its implied threat of the occurrence of enteric disease. (Rice, *et al.*, 1991).

Molecular methods such as “Polymerase Chain Reaction” (PCR) and molecular probes have been extensively used to detect *E. coli* involved in gastrointestinal infections. (Natro and Kaper, 1998).

Sandra (2004) used repetitive element anchored PCR to evaluate the genetic profile of *E. coli* isolated from surface water contaminated with urban storm water, sanitary sewage and gull feces to determine if strains found in environment samples reflect the strain composition of *E. coli* obtained from host sources.

Md. Shahedur Rahman, *et al.*, (2011) explored the antimicrobial activity of extracts of natural spices on multiple drug resistant enteric pathogen like Escherichia coli isolated from drinking water and suggested the possibility of their use as new and effective herbal medicines to treat and prevent diarrheal diseases.

Diarrheagenic *E. coli* strains were among the first pathogens for which molecular diagnostic approaches were developed.

Molecular Approach like PCR showed ability to amplify and detect diarrheagenic *E. coli* and can be implemented in the diagnosis water, food-borne outbreaks related to *E. coli*. (Ahmed, *et al.*, 2008).

Multiplex PCR (mPCR) has been developed for the detection of multiple bacterial species in a single test by amplification of multiple gene targets. A number of studies have reported on m PCR detection of multiple

waterborne pathogens in marine water samples. (Kong, *et al.*, 1999; Kong, *et al.*, 1995 and Kong, *et al.*, 2002).

**SEROTYPING OF *E. COLI*:**

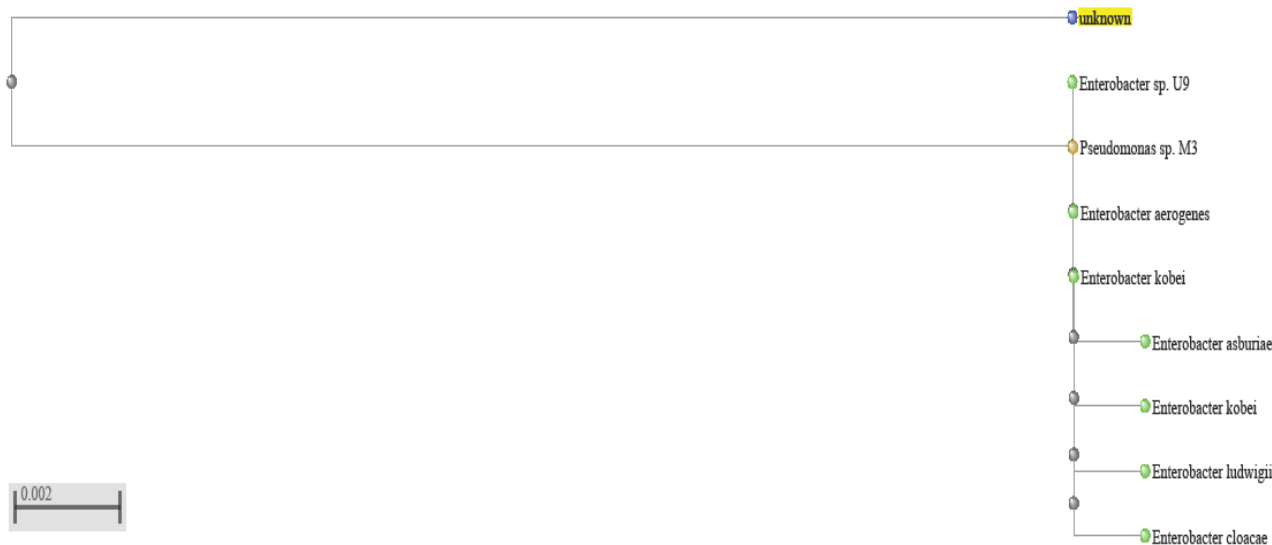
Serotyping with the 'O' antigen and the 'H' yields important epidemiological information.

Serotyping occupies a central place in the differentiation of pathogenic and non-pathogenic *E. coli* strains as specific Serogroups are associated with certain clinical syndromes. (Kaper, *et al.*, 2004).

There are over 700 serotypes of *E. coli* found in humans, of which a small number of strains cause disease and are distinguished only by their 'O', 'H' and 'K' antigens. (Todar, 2002).



**S3 Phylogenetic Tree**



**S4 Phylogenetic Tree**

**>S3 Sequences**

CACCGTGGCATTCTGATCCACGATTACTAGCGATTCCGACTTCATGGAGTCGAGTTGCAGACTCCAATCCGGACTACGACGCAC  
 TTTATGAGGTCCGCTTGCTCTCGCGAGGTCGCTTCTCTTTGTATGCGCCATTGTAGCACGTGTGTAGCCCTGGTCGTAAGGGCC  
 ATGATGACTTGACGTCATCCCCACCTTCCCTCCAGTTTATCACTGGCAGTCTCCTTTGAGTTCCCGGCCGGACCGCTGGCAACAA  
 AGGATAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATTTTCAACAACACGAGCTGACGACAGCCATGCAGCACCTGTCTCACG  
 GTTCCCGAAGGCACATTCTCATCTCTGAAAACCTCCCGTGGATGTCAAGACCAGGTAAGGTTCTTCGCGTTGCATCGAATTA AAC  
 CACATGCTCCACCGCTTGTGCGGGCCCCCGTCAATTCATTTGAGTTTTAACCTTGC GGCCGTA CTCCCAGGCGGTGCGACTTAA  
 CGCGTTAGCTCCGGAAGCCACGCCTCAAGGGCACAACCTCCAAGTCGACATCGTTTACGGCGTGGACTACCAGGGTATCTAATC  
 CTGTTTGCTCCCCACGCTTTCGCACCTGAGCGTCAGTCTTCGTCCAGGGGGCCGCTTCGCCACCGGTATTCTCCAGATCTCT  
 ACGCATTTACCGCTACAC

**>S4 Sequences**

GCCCTCCCGAAGGTTAAGCTACCTACTTCTTTTGCAACCCACTCCCATGGTGTGACGGGCGGTGTGTACAAGGCCGGGAACGT  
 ATTCACCGTAGCATTCTGATCTACGATTACTAGCGATTCCGACTTCATGGAGTCGAGTTGCAGACTCCAATCCGGACTACGACG  
 CACTTTATGAGGTCCGCTTGCTCTCGCGAGGTCGCTTCTCTTTGTATGCGCCATTGTAGCACGTGTGTAGCCCTACTCGTAAGG  
 GCCATGATGACTTGACGTCATCCCCACCTTCCCTCCAGTTTATCACTGGCAGTCTCCTTTGAGTTCCCGGCCAACCCTGGCAA  
 CAAAGGATAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATTTTCAACAACACGAGCTGACGACAGCCATGCAGCACCTGTCTC  
 AGAGTTCCCGAAGGCACCACCGCATCTCTGATAAGTTCCGTGGATGTCAAGAATAGGTAAGGTTCTTCCCGTTGCATCCAATTA  
 AACCACATGGTCCACCGCTTGTGCGGGCCCCCGTCAATTCATTTGAGTTTTAACCTTGC GGCCGTA CTCCCAGGCGGTGCGACT  
 TAACGCGTTTGCTTCCGAAGCCACGGCTTCAGGGCACAACCTTCTAAGTCCACCTCGTTTACGGGGTGGACTAACAAGGTA ACT  
 AATCCTGGTTGCTCCCCACGCTTTCACCTGAACGTTAATCTTTGTCCAGGGGGCCGCTTC

When analyzed by serotyping, significant differences have been observed between fecal *E. coli* isolates from human and wildlife sources. (Parveen, *et al.*, 2001).

As coliforms *E. coli* and Entamoeba histolytica were found in the water samples, hence precautions should be taken in usage of this water.

Mohamed Hatha, *et al.*, (2004), analyzed the prevalence of diarrheagenic serotypes of *E. coli* in the Cochin estuary, by serotyping the strains and worked out the seasonal variations in the prevalence levels of these organisms.

The results add to the increasing evidence of the occurrence of diarrheal pathogens in surface water bodies that may represents a threat to human beings exposed to water-borne pathogens and related consequences.

The following are the results of characterization of Pathogens from the positive samples (S3 & S4) and gene sequences.

The present investigation may be worthwhile for the clinical laboratories of the surrounding locality to characterize *E. coli* serotypes isolated from the diarrheal patients, so as to get a clear picture of the emergence of pathogenic strains of *E. coli* in the study area.

**CONCLUSION:**

The deterioration of the quality of surface water is alarming and regular monitoring and preventive measures are required.

Enumeration of culture able bacteria using count plates, isolated colonies with distinct morphologies from each plate were characterized by 16S r RNA gene sequencing using extracted genomic DNA from water samples.

The present study warranted to safeguard ourselves against emerging water-borne pathogens.

PCR has been suggested as a sensitive and rapid alternative to traditional growth based methods for detection of bacteria in water.

During the study it was found that maximum number of physico-chemical parameters within the desirable limits (excepts alkalinity, total hardness and calcium), as suggested by WHO (1971) and BIS (1991).

PCR provides a useful molecular approach for the detection of diarrheal pathogens in environmental

samples. This approach is an alternative to the existing technique or may be used in conjugation with these methods. This can also be useful for the detection of other pathogens.

Serotyping is a common method used for the characterization of *E. coli* isolates and has a broad use in epidemiological and medical diagnosis. The existence association between serotype and pathotype makes this method a valuable tool for typing *E. coli* and other bacterial species.

**Conflicts of interest:** The authors stated that no conflicts of interest.

## REFERENCES

- Absalon D and Matysik M (2007) Changes in water quality and runoff in the upper Oder River Basin. *Geomorphology*, 92 (3-4), 106 – 118.
- Adefemi SO and Awokunmi EE (2010) Determination of Diarrheal – Chemical Parameters and heavy metals and water samples from Itaogbolu area of Ondo-State, Nigeria. *African Journal of Environmental Science and Technology*, 4(3), PP 145 – 148.
- Agarwal Animesh and Manish Saxena (2011) Assessment of Pollution by Physico-chemical Water parameters using Regression Analysis: A case study of Ganga River at Moradabad – India. *Advances in Applied Science Research*, 2(2), PP 185 – 189.
- APHA (1991, 1998) Standard Methods for Examination of Water and Wastewater, American Public Health Association, Washington D.C.
- Archana T (2012) Water quality and quantity analysis in Sikkim, North Eastern Himalaya. *Current Science*, Vol. 103, No. 1.
- Bej AK, Dicesare JL, Haff L and Atlas RM (1991) Detection of *Escherichia coli* and *Shigella* spp. in water by using the Polymerase Chain Reaction and gene probes for uid, *Applied and Environmental Microbiology*, Vol. 57, No. 4, PP. 1013 – 1017.
- Gatica EA, Almeida CA, Mallea MA, Gonzalez P and del Corigliano MC (2012) Water quality assessment by statistical analysis, on rural and urban areas of Choeancharava River (Rio Cuarto), Cordoba, Argentina. *Environ Monit Assess*, 184: 7257 – 7274.
- Gupta DS, Sunita and Sahara JP (2009) Physico Chemical Analysis of Ground water of selected Area of Kaithal city (Haryana) India. *Researcher*, 1(2), PP 1-5.
- Milacron Marketing Co., The Effects of Water Impurities on water – based metal working fluids. Technical Report No. J/N 96/47.
- Nataro JP and Kaper, JB (1998) Diarrheogenic *Escherichia coli*. *Clinical Microbiology Reviews*, Vol. 11, No. 1, PP. 140 – 201.
- Navneet K, Sinha DK (2010) Drinking Water Quality Management through correlation studies among various diarrheal - chemical parameters: A case study. *International Journal of Environmental Sciences*, 1(2), PP 253 – 259.
- Patil PN, Sawant DV and Deshmukh RN (2012) Diarrheal, Chemical Parameters for tests of water-A-review, *International Journal of Environmental Sciences*, Vol. B, No 3.
- Premlata Vikal (2009) Multivariate analysis of drinking water quality parameters of lake Picchola in Udaipur, India. *Biological Forum, Biological Forum – An International Journal*, 1 (2), PP 97 – 102.
- Pruss Ustun A, Kay D, Fewtell E and Bartram J (2004) Unsafe water, Sanitation and Hygiene in Comparative Quantification of Health Risk: Global and Regional Burden of Disease Attributable to selected Major Risk Factor, M. Ezzati, *et al.*, WHO, Geneva, 2004, Vol. 2. , PP. 1321 – 1352.
- Rokade PB, Ganeshwade RM, (2005) Impact of pollution on Water Quality of Salim Ali Lake at Aurangabad Uttar Pradesh. *Journal of Zoology*, 25 (2), PP 219 -220.
- Sukumaran DP, Srinivasan D and Abdulla MH (2012) Antibiotic Resistance of *Escherichia coli* Serotypes from Cochin Estuary. *Interdisciplinary prospective on Infectious Diseases*, Vol. 2012, Article ID 124879, 7 pages.
- Trivedi RC (2010) Water quality of the Ganga River – An overview. *Aquatic Eco system Health and Management*, 13 : 4, 347 – 351.
- Veg M, Pardo R, Barrado E and Peban L (1998) Assessment of seasonal and polluting effects on the quality of river water by exploratory data analysis. *Water Research*, 32 (12), 3581 – 3592.
- Wang Q, Ruan X, Wei D (2010) Development of a Serogroup Specific Multiplex PCR assay to detect a set of *Escherichia coli* Serogroups based on the identification of their O-antigen gene clusters, *Molecular and Cellular probes*, Vol. 24, No. 5, PP. 286 – 290.
- Water Quality Control (2007) Parameters of Natural Water.
- Zhang GZ, Liu H and Jia DW (2010) River basin management based on the mechanisms of water rights trading. *Procedia Environmental Sciences*, 2, 665 – 673