Synchronous luminescence spectroscopy for determination of possibility of medicinal and food values of plants

Suryawanshi AD¹, Padampalle AS², Sanap VB³, Suryawanshi DD⁴, Pawar BH⁵

¹Dept of Physics, B.J. College, Ale (Pune),²Dept. of Physics, S.C.S. College, Omerga (Osmanabad),³Dept. of Physics, Y. C. College, Sillod (Aurangabad),⁴Dept of Chemistry, B.A.M. University, Aurangabad'⁵ Ex. Head, Dept. of Physics, S.G.B. Amravati University, Amravati. Email: ajitdsuryawanshi@rediffmail.com

Manuscript Details

Available online on http://www.irjse.in ISSN: 2322-0015

Editor: Dr. Arvind Chavhan

Cite this article as:

Suryawanshi AD, Padampalle AS, Sanap VB, Suryawanshi DD, Pawar BH. Synchronous luminescence spectroscopy for determination of possibility of medicinal and food values of plants, Int. Res. Journal of Science & Engineering, January 2018; Special Issue A2: 172-175.

© The Author(s). 2018 Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

ABSTRACT

The study of the effect of various stresses like light stress, temperature stress, pollutant stress etc. may be performed using various spectroscopic techniques like absorption spectroscopy, fluorescence spectroscopy, fluorescence kinetics, Fourier Transform Infrared spectroscopy etc. In addition to these techniques the synchronous luminescence technique may be successfully employed to study the effect of stresses on the plant health. In this technique the fluorescence signal is recorded by simultaneously scanning both the excitation and emission wavelengths at same speed with a fixed wavelength interval between the excitation and emission wavelengths.

Key words: Photosynthesis, Deuterium/Halogen source, SL spectra.

INTRODUCTION

In this technique the fluorescence signal is recorded by simultaneously scanning both the excitation and emission wavelengths at same speed with a fixed wavelength interval between the excitation and emission wavelengths. Since it takes the advantage of the absorption as well as emission properties of the molecules, it leads to considerable amount of simplification in the measured fluorescence spectral profile. In the present attempt we are going to use the

synchronous luminescence spectroscopy for the study of plant health and classification. As per our information the type of measurements made by us is the first report of this kind. It is seen that more information can be obtained from the analysis of synchronous luminescence spectra of the plant leaves.

METHODOLOGY

Sample preparation:

The plants that are selected for the experiment have been grown in the campus of the nursery at Amravati. The plants are developed in the natural condition of light and temperature. Plants are well watered and regularly nourished and developed in the good environmental conditions. Typical and healthy leaf from a plant has been chosen for the study of the synchronous luminescence spectra.

Experimental Method:

A block diagram for recording synchronous luminescence spectrum is shown in fig 1.

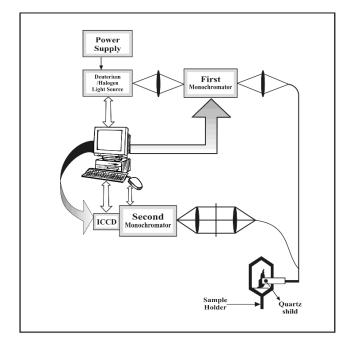


Figure 1 Experimental arrangement for recording synchronous luminescence spectra of plants leaves. Commercial spectra physics fluorometer (SPEX, USA, Fluorolog –II) was used to record the synchronous luminescence spectra.

The spectra were recorded by scanning both the excitation and emission monochromator simultaneously at the same speed of 5 nm/s with a fixed wavelength separation between them. For these studies wavelength difference between excitation and fluorescence emission was chosen to be 20 nm since it lead to the most resolved spectra. The band pass of both the excitation and emission monochromator was kept 2 nm wide. A xenon lamp of 45 W is used as the excitation source. The light from xenon lamp was incident perpendicular to the sample surface to a spot of size approximately 2mm X 4 mm and the emitted light was collected at approximately 20⁰ angle with respect to the direction of the excitation light. Excitation intensity varied with wavelength but was always less than 40 W/mm².

A bundle of seven optical fibers is used as light transporting system. A single central fiber carries light from the source to the sample and six fibers surrounding the centro fiber collect the fluorescence signal given by sample.

RESULTS AND DISCUSSION

The synchronous luminescence spectra of many plants belonging to different family have been recorded using SPEX USA fluorolong–II spectrophotometer and results are displayed in figure 2 (a-c).

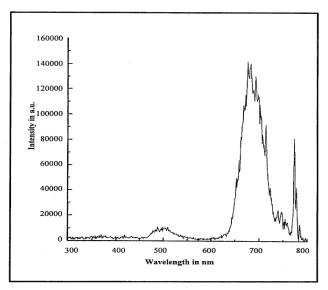


Figure 2 (a) Synchronous luminescence spectra of plants of Fiscus benghalensis [Family-Moraceae].

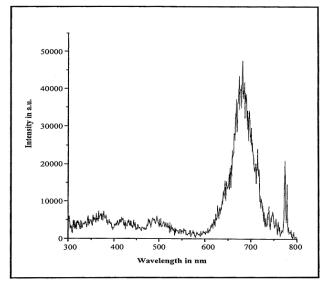


Figure 2 (b) Synchronous luminescence spectrum of leaf of Fiscus religiosa [family-Moraceae].

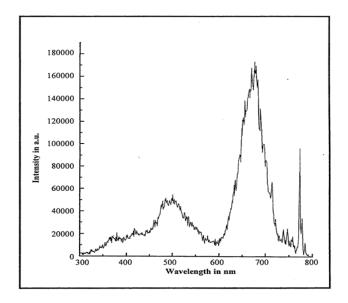


Figure 2 (c) Synchronous luminescence spectrum of leaf of Hibiscus rosasinensis [Family-Malvaceae].

All the spectra recorded by us exhibit novel structure difference. The number of peaks observed in the spectra and the intensity of peaks differ from plant to plant.

All the synchronous luminescence spectra of plant leaves were characterized by 2 to 5 narrow spectral bands with peak around 366, 416, 500, 677 and 772 nm. A prominent peak around 677 nm exhibits some extra structure of few plants. Significant differences can be seen in the intensities of different spectral band of different plants. The peak position of the different spectral bands characteristics of the different intrinsic fluorophores provide a rapid qualitative estimate of the relevant contains of the fluorophores present in the leaf cells. For example the peak at 500 nm spectral band is similar in case of all the plants. From the analysis of the peak structure one would expect no significant difference in the concentration of pigments in the plant cells. However, significant difference in the peak positions in the spectra would suggest that there is significant difference in the quantity of the different pigments present in the cells.

The synchronous luminescence spectra contains main peak and few side peaks. The main peak in the emission spectrum is wide and appears around 680 nm. However, there is a slight difference in the position of the other peaks depending upon the species selected for the studies. For getting better understanding about the structure and density of the pigments in the leaf heights of the peaks in the spectra are normalized to the height of the prominent peak near 680 nm.

It should be noticed that position of the main peak lies between 650 to 683 nm and the width of the main peak is between 40 to 70 nm FWHM (full width at half maximum) for all samples under investigation.

It should be noticed from the analysis of the spectra that all plants on the longer wavelength slope of the main peak exhibit peaklets. It is also clear that lot of information about the plant health may be extracted from the position and properties of peaklets with respect to main peak.

The synchronous luminescence spectra of all the plants exhibit a narrow peak near 680 nm. The relative height of this peak shows the considerable difference depending upon the families of the plant. The structure of peak carry a lot of physiological information about the plants as the related height of the peak changes from 0.2 to 0.3 with respect to the height of main peak.

Depending upon the plant type we get the different structure such as prominent peak of maximum height, double peak structure near the main peak or extra structure at main peak, widths of the different peaks, exact position of the peak, number of peaks in the spectrum, position of peaklets near the main peak.

Thus several excitation and emission spectra are required to obtain the information on fluorophores concentration, the same information can be obtained more conveniently in the single step by recording synchronous luminescence spectra. Since synchronous luminescence spectrum provides more information compared to emission spectrum at single excitation wavelength. The study of synchronous luminescence spectra may lead to better discrimination in the plant synchronous physiology. The study of the luminescence spectra is helpful in the investigation of band structure of the pigments present in the plant leaves.

CONCLUSION

It is cleared that each sample of plant leaves from various families and plant types gives the different feature. An attempt can be made to record the synchronous luminescence spectra of all possible samples having some medicinal value or special characteristics such as oil producing plant, poisoning plant etc. and then by differentiating and classifying according to various parameters such as peaklets position, total number of peaks, position and intensity of various peaks in the spectra, full width at half maximum of each peak, the ratio of different peak heights. Some conclusion may be drawn about the position of the plant.

An unknown sample may be identified as a medicinal and nonmedicinal plant. The synchronous luminescence spectroscopy is very much useful in the identification of various plants, which belong to different families, species etc.

It is obvious that from the study of synchronous luminescence spectra large amount of information may be provided to the branch of bio-informatics.

Synchronous luminescence spectral study also gives good information about leaf structure, degree of stresses and leaf contents. Thus the synchronous luminescence spectroscopy is very powerful tool for investigation of plant physiology.

Conflicts of interest: The authors stated that no conflicts of interest.

REFERENCES

- 1. Gupta P.K. and Bhawalkar D. D. Current Sci., vol.,77, No 7, 1999.
- Wolbarsht M. L., "Laser applicatipon in medicine and biology." Pelenum press NewYork, Vol.1-5, 1991.
- 3. Gupta P. K., Pradhan A. S., Bhatnagar P. K. Reddy A. R. 'Medical Physics for Human Health Care'.Scientific Publisher, Jodhapur.
- 4. Servik-Muraca E, and Benaron D. " Trends in optics and photonics on Biomedical optical spectroscopy and Diagnostics," Optical society of America, Washington DC, Vol. 3, 1996.
- Majumdar S. K., Mohanty S. K., Ghosh N., Gupta P. K. and et al "A Pilot Study on the use of Autofluorescence Spectroscopy for Diagnosis of the cancer Of Human Oral Cavity," Current Science, Vol. 79, No. 8, pp 1089-1094, 2000.
- Lang Michael, Lichtenthaler H. K., "Changes in the Blue –Green and Red Fluorescence Emission Spectra of Beech Leaves During the Autumnal Chlorophyll Breakdown," J. Plant Physiol, Vol. 138, pp. 550-553, 1991.
- Krause G. H. and Wies E. 'Chlorophyll Fluorescence and Photosynthesis: The Basics', Annual Review Plant Physiology, Vol. 42, pp. 313-349, 1991.
- Gitelson A. A. and Marzlyak M. N., "Signature Analysis of Leaf Reflectance Spectra: Algorithm Development for remote sensing of chlorophyll", J. Plant Physiol, 148, 494-500, 1996.
- Gitelson A. A., Buschmann C., Lichtenthaler H. K., "Leaf chlorophyll Fluorescence Corrected for Reabsorption by Means of Absorption and reflectance Measurements", J. Plant physiology, Vol, 152, pp. 283-296,1998.

© 2018 | Published by IRJSE