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Histone Deacetylase Inhibitors: Current Status in Treatment of

Leukemia

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ABSTRACT

Leukemia, one of leading cancer occurs by the acquisition of gene mutations that confer deregulated proliferation and impaired differentiation. Several histone decaetylase inhibitors (HDACi) have exhibited significant protection against the growth of tumor cells in vitro as well as in vivo. Thus it is anticipated that some HDACi could be efficient anti-leukemic drugs. Here, we summarize the current status of different types of HDACi and their clinical role in therapy of different types of leukemia.

Keywords: Leukemia, HDACi, Hydroxamates, Benzamides, Cyclic peptides, Short chain fatty acids.

1. INTRODUCTION

The term leukemia represents a heterogeneous group of diseases described by the malignant clonal proliferation of blood progenitor cells. These cells primarily grow and expand in the bone marrow, and from there distribute to the entire body via the blood circulation. Based on the kinetics of disease onset and course, as well as the differentiation of the malignant clone, leukemia is classified into acute and chronic, and myeloid and lymphocytic, respectively.¹

Histones are small basic proteins that form nucleosomes leading to the compact structure of chromatin. These histones have long N-terminal extensions that have been known for decades to undergo epigenetic modifications such as acetylation, methylation, phosphorylation as well as ubiquitylation, sumoylation, and ADP-ribosylation. It is suggested that the balance between the actions of histone acetyltransferase (HAT) and histone deacetylases (HDAC) activity leads to a decreased level of acetylation and decreased expression of genes that control growth and development of various cancerous tissues. Discovery of chromatin-modifying enzymes, HDAC made a revolution in the development of a novel class of pharmacologic agents, HDAC inhibitors (HDACi), which are cytostatic agents that inhibit the proliferation of tumor cells in culture and in vivo by inducing cell cycle arrest, differentiation and/or apoptosis. Thus HDACi have emerged as a new class of chemotherapeutic drugs that regulate gene expression.^{2,3}

2. CLINICALLY USEFUL HISTONE DEACETYLASE INHIBITORS FOR LEUKEMIA

2.1 Short Chain Fatty Acid Derivatives

These drugs inhibit both class I and IIa HDAC, but usually show low potency due to the inability to make significant contact with the catalytic pocket of HDAC.

2.1.1 Sodium Phenylbutyrate

Sodium phenylbutyrate (PB) is an aromatic fatty acid initially developed for treatment of urea cycle disorders and thalassemia. It is able to induce hyperacetylation of histone proteins in leukemic cells at millimolar concentrations *in vitro*, but its clinical development has been impeded by its short half-life and adversity in achieving millimolar levels *in vivo*.^{4,5} In the phase I trial, continuous 7 day infusion of PB was employed (7/28 schedule: 7 days on, 21 days off schedule) in 11patients with MDS (myelodysplastic syndrome) and 16 patients with AML (acute myeloid leukemia) and a dose of 375 mg/kg/day was recognized as the maximally tolerated dose (MTD). Dose limiting toxicities (DLT) were neurological impediments and were reversible within 24–48 h of stopping the infusion.^{6,7}

2.1.2 Valproic Acid

Valproic acid (VPA, di-n-propylacetic acid) is a short chain fatty acid used as an antiepileptic and mood stabilizer. VPA has been shown to affect the growth of malignant cells *in vitro*, to prolong the G1 phase of the cell cycle.^{8,9} Generally, valproic acid is well tolerated. The reported leading toxicities are neurologic side effects such as dizziness, tremor, sedation, mild gastrointestinal side effects and hematologic toxicity, including pancytopenia and severe bone marrow hypoplasia. Liver failure and teratogenicity with neural-tube defects have been described.¹⁰ VPA is ineffective for the treatment of leukemia when used alone so it is further evaluated for its antileukemia activity in varied combination with cytarabine (Ara-C), etoposide, 5-azacitidine (5-AZA), decitabin, retinoic acid (ATRA).¹¹⁻¹⁶

Sodium N-butyrate and phenylacetate are also reported as fatty acid derivatives with HDAC inhibitory activity but having inadequate clinical efficacy and CNS toxicity during clinical trials.¹⁷

2.2 Cyclic Peptides

2.2.1 Depsipeptide or Romidepsin (FK228)

Despipepdide is a non-epoxyketone-containing bicyclic tetrapeptide isolated from *Chromobacterium violaceum* with antitumor activity in a broad variety of murine and human tumor cell lines both *in vitro* and *in vivo*. It is a pro-drug and the active moiety is a sulfhydryl group acting as the Zn^{2+} -chelator. It is a more selective inhibitor of the class I HDAC, preferentially blocking HDAC 1 and 2 versus HDAC 4 and 6.¹⁸

Phase I and II studies have established that depsipeptide has a favorable anticancer activity, particularly in patients with CTCL (cutaneous T-Cell Lymphoma) and peripheral T-cell lymphoma. It was given as an IV infusion at a dose of 13 mg/m² on days 1, 8, and 15 of a 28 day cycle. Generally, it is well tolerated with favorable toxicity profile. Common side effects include fatigue, nausea, vomiting, transient thrombocytopenia and neutropenia. Several ECG findings have been expressed during the treatment with depsipeptide, including ST and T wave abnormalities, QTc interval prolongation and cardiac arrhythmias. The Maximum Tolerated Dose (MTD) was found to be 17 mg/^{m2} on days 1 and 5 every 21 days.¹⁹⁻²¹

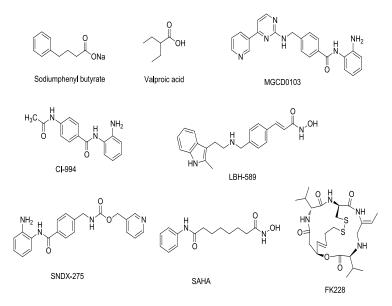


Fig 1: Clinically effective HDACi in leukemia

2.3 Benzamide Derivatives

2.3.1 MS-275 (Entinostat/SNDX-275)

MS-275 has demonstrated time and dose dependent growth inhibition of leukemia cell lines as well as primary leukemia blasts. It selectively inhibits HDAC 1, 2, and 3^{22}

Phase I clinical trial based on the study of MS-275 in solid tumors, a starting dose of 4 mg/m² weekly for 2 or 4 sequential weeks followed by 2-week washout was confirmed.²³ The maximum-tolerated dose was $8mg/m^2$ weekly for 4 weeks every 6 weeks and DLT included infections and neurologic toxicity indicating unsteady gait and somnolence. Other frequent non-DLTs were fatigue, anorexia, nausea, vomiting, hypoalbuminemia and hypocalcemia.²⁴

2.3.2 MGCD-0103 (Mocetinostat)

MGCD0103 is an isotype specific aminophenylbenzamide and has been shown to inhibit HDAC isotypes 1, 2, 3, and 11.²⁵ A dose escalation phase 1 study of oral MGCD0103 given three times a week in patients with AML and MDS has been performed. Doses of MGCD0103 ranged from 20–80mg/m² orally. Twenty-nine patients were treated (22 AML, 5 MDS, 1 ALL, 1 CML). Median age was 65 years and 83% of patients had taken previous chemotherapy. Fatigue, nausea, diarrhea, vomiting were the most often reported adverse events. MTD was established at 60 mg/m² orally three times a week.²⁶

A phase II clinical trial was carried out, starting at a dose of 85 mg/d, three times per week. MGCD0103 exhibited preclinical activity against CLL (Chronic lymphocytic leukemia) cells with a LC_{50} (concentration lethal to 50%) of 0.23 micromol/l and increased acetylation of the HDAC class I specific target histone H3. Twenty-one patients received a median of two cycles of MGCD0103 (range, 0-12). Grade 3-4 toxicity consisted of infections, thrombocytopenia, anaemia, diarrhoea and fatigue. HDAC inhibition was observed in six out of nine patients on day 8. Limited activity was observed with single agent MGCD0103 in high risk patients with CLL.²⁷

2.3.3 CI-994

N-acetyl-dinaline (CI-994) is an investigational anti-cancer drug which inhibits histone deacetylases. *In vitro*, CI-994 in combination with cytarabine (ara-C), daunorubicin and mitoxantrone, resulted in moderate synergism. *In vivo*, higher dosages of CI-994 induced complete remissions. The combinations of CI-994/daunorubicin and CI-994/mitoxantrone were also active.²⁸ CI-994 administration, inhibition of both histone deacetylation and cellular proliferation at the G1 to S transition phase of the cell cycle were noticed.²⁹ Its clinical studies for anti-leukemia activity have not yet been reported so far.

2.4 Hydroxamic Acid Derivatives

2.4.1 LBH589 (Panobinostat)

LBH 589, a potent HDACi is a cinnamic hydroxamic acid analogue which has been shown to induce apoptosis and histone acetylation in acute leukemia cells. LBH 589 effectively induce apoptosis in ATLL-related cell lines and primary ATLL cells.³⁰ Low nanomolar concentrations (IC₅₀: 5-20 nM) of LBH589 induces cell-cycle arrest, apoptosis and histone (H3K9 and H4K8) hyperacetylation.³¹

Phase I clinical trials shows that 15 patients (13 AML, 1 MDS, 1 ALL) treated with LBH589 at doses ranging from 4.8 to 14.0 mg/m² IV daily for 7 days. Reversible QTc prolongation was the dose limiting toxicity. The MTD (maximum tolerated dose) was not established, as 14 mg/m² exceeded it and the lower dose cohort of 11.5 mg/m² could not be expanded given the concern for QTc prolongation. 27% of patients were also noted to have grade 3–4 hypokalemia, however no relation was noted between QTc

prolongation and hypokalemia. Other toxicities included nausea, diarrhea, vomiting, loss of appetite and thrombocytopenia.³²

2.4.2 Vorinostat (Suberoylanilide hydroxamic acid, SAHA)

It is a hydroxamic acid multi-HDACi that blocks the enzymatic activity of both Class I (HDAC1, -2, and -3) and Class II (HDAC6) HDACs at low nanomolar concentrations (IC₅₀ <86 nM) by directly binding to the active site of these enzymes.³³ Vorinostat-induced DNA damage is accompanies by a G2-M arrest and ultimately apoptosis.³⁴

A phase I study of single agent vorinostat in patients with advanced leukemia and MDS was conducted for total of 41 patients, were treated (31 AML, 4 CLL, 3 MDS, 2 ALL, 1 CML) with a classical 3 + 3 dose escalation design. The starting dose was 100 mg orally three times daily for 2 weeks with 1 week washout. Twice daily and three times daily regimens were tested. DLTs included fatigue, nausea, vomiting, and diarrhea. The MTD was established as 200mg BID or 250mg TID daily for 14 days every 21 days. Of the 41 patients, 2 patients achieved CR and 2 CRi. Additionally, 7 patients had HI (>50% decrease in blast count). Median number of cycles to response/improvement was 2 (range, 1-8) and median response duration was 6 weeks. Transient acetylation of histone H3 was observed in all patients, regardless of the dose level or response.³⁵ The two stage phase 2 clinical trial examined in 37 patients for the toxicity and response rate concomitant with two treatment schedules of the HDACi, vorinostat in patients with relapsed acute myeloid leukemia. In both stages a total dose of 8400 mg of vorinostat was delivered in each 21-day cycle of treatment: in arm A the dose regimen was 400 mg daily whereas in arm B the dose regimen was 200 mg three times daily for 14 days followed by 1 week rest. In arm A (n=15), the confirmed full remission rate was 0% (95% CI, 0% to 23%); this arm was closed at the planned interim analysis. In arm B (n=22), the confirmed full remission rate was 4.5% (1 response; 95% CI, 0.4% to 24%), with a duration of response exceeding 398 days. The median time to treatment failure in arm A was 42 days (95% CI, 26 to 57); although a minimum of four cycles of treatment were planned, 11 patients (79%) received no more than two cycles. The median time to treatment failure in arm B was 46 days (95% CI, 20 to 71); 13 patients (59%) received no more than two cycles of treatment.³⁶

3. SUMMARY & CONCLUSION

Leukemia is a type of cancer caused by the acquisition of gene mutations that bestow deregulated proliferation, impaired differentiation. The HDAC has been found to interact with many partners through complex molecular mechanism leading to the regulation of gene expression; they have obtained the interest of a huge scientific community. Several HDACi have been confirmed to competently protect against the growth of tumor cells in vitro as well as in vivo. Now more effective and well tolerated HDACi, including depsipeptide, SAHA, LBH589, PDX101, MS-275, CI-994 and MGCD0103 are in clinical trials alone or in combination with other anticancer agents. Thus it is anticipated that some of the HDACi could be potential effective anti-leukemic drugs.

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REFERENCES

- Kohlschutter J, Michelfelder S, Trepel M. Drug delivery in acute myeloid leukemia. Expert Opin Drug Deliv. 2008; 5: 653-663.
- Bolden JE, Peart MJ, Johnstone RW. Anticancer activities of histone deacetylase inhibitors. Nature Rev Drug Discov. 2006; 5: 769-784.
- Ropero S, Esteller M. The role of histone deacetylases (HDACs) in human cancer. Mol. Oncol. 2007; 1: 19-25.
- Dover GJ, Brusilow S, Samid D. Increased fetal hemoglobin in patients receiving sodium 4-phenylbutyrate. N Engl J Med. 1992; 327: 569-570.
- Miller AA, Kurschel E, Osieka R, Schmidt CG. Clinical pharmacology of sodium butyrate in patients with acute leukemia. Eur J Med Chem. 1987; 23: 1283-1287.
- Di Giuseppe JA, Weng LJ, Yu KH, Fu S, Kastan MB, Samid D, Gore SD.
 Phenylbutyrate-induced G1 arrest and apoptosis in myeloid leukemia cells: structure-function analysis. Leukemia. 1999; 13: 1243-1253.
- Gore SD, Weng LJ, Zhai S, Figg WD, Donehower RC, Dover GJ, Grever M, Griffin CA, Grochow LB, Rowinsky EK, Zabalena Y, Hawkins AL, Burks K, Miller CB. Impact of the putative differentiating agent sodium phenylbutyrate on myelodysplastic syndromes and acutemyeloid leukemia. Clin Cancer Res. 2001; 7: 2330-2339.
- Gurvich N, Tsygankova OM, Meinkoth JL, Klein PS. Histone deacetylase is a target of valproic acid-mediated cellular differentiation. Cancer Res. 2004; 64: 1079-1086.
- Bacon CL, Gallagher HC, Haughey JC, Regan CM. Antiproliferative action of valproate is associated with aberrant expression and nuclear translocation of cyclin D3 during the C6 glioma G1 phase. J Neurochem. 2002; 83: 12-19.

- Kuendgen A, Gattermann N. Valproic acid for the treatment of myeloid malignancies. Cancer. 2007; 110: 943-954.
- Sanchez-Gonzalez B, Yang H, Bueso-Ramos C, Hoshino K, Quintas-Cardama A, Richon VM, Garcia-Manero G. Antileukemia activity of the combination of an anthracycline with a histone deacetylase inhibitor. Blood. 2006; 108: 1174-1182.
- Lane S, Gill D, McMillan NA, Saunders N, Murphy R, Spurr T, Keane C, Fan HM, Mollee P. Valproic acid combined with cytosine arabinoside in elderly patients with acute myeloid leukemiahas in vitro but limited clinical activity. Leuk Lymphoma. 2012; 53(6): 1077-1083.
- Corsetti MT, Salvi F, Perticone S, Baraldi A, De Paoli L, Gattoa S, Pietrasanta D, Pini M, Primon V, Zallio F, Tonso A, Alvaro MG, Ciravegna G, Levis A. Hematologic improvement and response in elderly AML/RAEB patients treated with valproic acidand low-dose Ara-C. Leukemia Res. 2011; 35: 991-997.
- 14. Raffoux E, Cras A, Recher C, Boelle PY, de Labarthe A, Turlure P, Marolleau JP, Reman O, Gardin C, Victor M, Maury S, Rousselot P, Malfuson JV, Maarek O, Daniel MT, Fenaux P, Degos L, Chomienne C, Chevret S, Dombret H. Phase 2 clinical trial of 5-azacitidine, valproic acid, and all-trans retinoic acid in patients with high-risk acute myeloid leukemia or myelodysplastic syndrome. Oncotarget. 2010; 1: 34-42.
- Hrebackova J, Hrabeta J, Eckschlager T. Valproic acid in the complex therapy of malignant tumors. Curr Drug Targets. 2010; 11: 361-379.
- 16. Yang H, Fang Z, Wei Y, Hu Y, Calin GA, Kantarjian HM, Garcia-Manero G. Levels of miR-29b do not predict for response in patients with acute myelogenous leukemiatreated with the combination of 5azacytidine, valproic acid, and ATRA. Am J Hematol. 2011; 86: 237-238.
- Monneret C. Histone deacetylase inhibitors for epigenetic therapy of cancer. Anticancer Drugs. 2007; 18: 363-370.
- Furumai R, Matsuyama A, Kobashi N, Lee KH, Nishiyama M, Nakajima H, Tanaka A, Komatsu Y, Nishino N, Yoshida M, Horinouchi S. FK228 (Depsipeptide) as a natural prodrug that inhibits class I histone deacetylases. Cancer Res. 2002; 62: 4916-4921.
- Byrd JC, Marcucci G, Parthun MR, Xiao JJ, Klisovic RB, Moran M, Lin TS, Liu S, Sklenar AR, Davis ME, Lucas DM, Fischer B, Shank R,

Tejaswi SL, Binkley P, Wright J, Chan KK, Grever MR. A phase 1 and pharmacodynamic study of depsipeptide (FK228) in chronic lymphocytic leukemia and acute myeloid leukemia. Blood. 2005; 105: 959-967.

- 20. Piekarz RL, Frye R, Turner M, Wright JJ, Allen SL, Kirschbaum MH, Zain J, Prince HM, Leonard JP, Geskin LJ, Reeder C, Joske D, Figg WD, Gardner ER, Steinberg SM, Jaffe ES, Stetler-Stevenson M, Lade S, Fojo AT, Bates SE. Phase II multi-institutional trial of the histone deacetylase inhibitor romidepsin as monotherapy for patients with cutaneous T-cell lymphoma. J Clin Oncol. 2009; 27: 5410-5417.
- Otterson GA, Hodgson L, Pang H, Vokes EE. Phase II study of the histone deacetylase inhibitor Romidepsin in relapsed small cell lung cancer (Cancer and Leukemia Group B 30304). J Thorac Oncol. 2010; 5: 1644-1648.
- Lucas DM, Davis ME, Parthun MR, Mone AP, Kitada S, Cunningham KD, Flax EL, Wickham J, Reed JC, Byrd JC, Grever MR. The histone deacetylase inhibitor MS-275 induces caspase-dependent apoptosis in Bcell chronic lymphocytic leukemia cells. Leukemia. 2004; 18: 1207-1214.
- 23. Ryan QC, Headlee D, Acharya M, Sparreboom A, Trepel JB, Ye J, Figg WD, Hwang K, Chung EJ, Murgo A, Melillo G, Elsayed Y, Monga M, Kalnitskiy M, Zwiebel J, Sausville EA. Phase I and pharmacokinetic study of MS-275, a histone deacetylase inhibitor, in patients with advanced and refractory solid tumors or lymphoma. J Clin Oncol. 2005; 23: 3912-3922.
- Gojo I, Jiemjit A, Trepel JB, Sparreboom A, Figg WD, Rollins S, Tidwell ML, Greer J, Chung EJ, Lee MJ, Gore SD, Sausville EA, Zwiebel J, Karp JE. Phase1andpharmacologic study of MS 275, a histone deacetylase inhibitor, in adults with refractory and relapsed acute leukemias. Blood. 2007; 109: 2781-2790.
- 25. Fournel M, Bonfils C, Hou Y, Yan PT, Trachy-Bourget MC, Kalita A, Liu J, Lu AH, Zhou NJ, Robert MF, Gillespie J, Wang JJ, Ste-Croix H, Rahil J, Lefebvre S, Moradei O, Delorme D, MacLeod AR, Besterman JM, Li Z. MGCD0103, a novel isotype-selective histone deacetylase inhibitor, has broad spectrum antitumor activity *in vitro* and *in vivo*. Mol Cancer Ther. 2008; 7: 759-768.
- Garcia-Manero G, Assouline S, Cortes J, Estrov Z, Kantarjian H, Yang H, Newsome WM, Miller Jr WH, Rousseau C, Kalita A, Bonfils C, Dubay M, Patterson TA, Li J, Besterman JM, Reid G, Laille E, Martell RE,

Minden M. Phase 1 study of the oral isotype specific histone deacetylase inhibitor MGCD0103 in leukemia. Blood. 2008; 112: 981-989.

- 27. Blum KA, Advani A, Fernandez L, Van Der Jagt R, Brandwein J, Kambhampati S, Kassis J, Davis M, Bonfils C, Dubay M, Dumouchel J, Drouin M, Lucas DM, Martell RE, Byrd JC. Phase II study of the histone deacetylase inhibitor MGCD0103 in patients with previously treated chronic lymphocytic leukaemia. Br J Haematol. 2009; 147: 507-514.
- Hubeek I, Comijn EM, Van der Wilt CL, Merriman RL,Padron JM, Kaspers GJL, Peters GJ. CI-994 (N-acetyl-dinaline) in combination with conventional anti-cancer agents is effective against acute myeloid leukemia in vitro and in vivo. Oncol Rep. 2008; 19: 1517-1523.
- Foster BJ, Jones L, Wiegand R, LoRusso PM, Corbett TH. Preclinical pharmacokinetic, antitumor and toxicity studies with CI-994 (correction of CL-994) (N-acetyldinaline). Invest New Drug. 1997; 15: 187-194.
- 30. Hasegawa H, Yamada Y, Tsukasaki K, Mori N, Tsuruda K, Sasaki D, Usui T, Osaka A, Atogami S, Ishikawa C, Machijima Y, Sawada S, Hayashi T, Miyazaki Y, Kamihira S. LBH589, a deacetylase inhibitor, induces apoptosis in adult T-cell leukemia/lymphoma cells via activation of a novel RAIDD-caspase-2 pathway. Leukemia. 2011; 25: 575-587.
- 31. Scuto A, Kirschbaum M, Kowolik C, Kretzner L, Juhasz A, Atadja P, Pullarkat V, Bhatia R, Forman S, Yen Y, Jove R. The novel histone deacetylase inhibitor, LBH589, induces expression of DNA damage response genes and apoptosis in Ph-acute lymphoblastic leukemia cells. Blood. 2008; 111: 5093-5100.
- 32. Giles F, Fischer T, Cortes J, Garcia-Manero G, Beck J, Ravandi F, Masson E, Rae P, Laird G, Sharma S, Kantarjian H, Dugan M, Albitar M, Bhalla K. A phase I study of intravenous LBH589, a novel cinnamic hydroxamic acid analogue histone deacetylase inhibitor, in patients with refractory hematologic malignancies. Clin Cancer Res. 2006; 12: 4628-4635.
- Richon VM, Garcia-Vargas J, Hardwick JS. Development of vorinostat: current applications and future perspectives for cancer therapy. Cancer Lett. 2009; 280: 201-210.
- Petruccelli LA, Dupere-Richer D, Pettersson F, Retrouvey H, Skoulikas S, Miller Jr. WH. Vorinostat induces reactive oxygen species and DNA damage in acute myeloid leukemia cells. PLoS One. 2011; 6: 20987.

- 35. Garcia-Manero G, Yang H, Bueso-Ramos C, Ferrajoli A, Cortes J, Wierda WG, Faderl S, Koller C, Morris G, Rosner G, Loboda A, Fantin VR, Randolph SS, Hardwick JS, Reilly JF, Chen C, Ricker JL, Secrist JP, Richon VM, Frankel SR, Kantarjian HM. Phase 1 study of the histone deacetylase inhibitor vorinostat (suberoylanilide hydroxamic acid [SAHA]) in patients with advanced leukemias and myelodysplastic syndromes. Blood. 2008; 111: 1060-1066.
- Schaefer EW, Loaiza-Bonilla A, Juckett M, DiPersio JF, Roy V, Slack J, Wu W, Laumann K, Espinoza-Delgado I, Gore SD. A phase 2 study of vorinostat in acute myeloid leukemia. Haematol.2009; 94: 1375-1382.