

Cadmium contamination of drinking water and its treatment using biological chelators

Amiri A, PhD^{1*}, Mirhoseiny Z, Msc²

1- Assistant Prof., Dept. of Chemistry, Payame Noor University, P.O. Box, 19395-3697, Tehran, Iran. 2- MSc in Chemistry, Department of Chemistry, Payame Noor University, P.O. Box, 19395-3697, Tehran, Iran

Abstract

Received: July 2016, Accepted: October 2016

Background: Cadmium (Cd) is an extremely toxic metal and environmental exposure to Cd has been particularly problematic in the world. Oral exposure to Cd may result in adverse effects on a number of tissues, the immune system, and the cardiovascular system. Compounds containing Cd are also carcinogenic. The presence of Cd in drinking water resources in the southeastern region of Rafsanjan plain (Iran) at concentrations greater than acceptable limits may result in various adverse health effects. The aim of this research was to test the chelation potency of deferasirox (DFS or ICL670), a tridentate metal chelator, and deferiprone (L₁), a bidentate metal chelator, in the mobilization of Cd in Cd-exposed rats as a biological model.

Materials and Methods: Male Wistar rats were exposed to 40 mg/kg body weight of cadmium chloride in drinking water for 6 weeks, followed by treatment with DFS (100 mg/kg body weight, oral, once daily) and L₁ (100 mg/kg body weight, oral, once daily), alone or in combination, for 7 consecutive days. After chelation therapy, the rats were anesthetized by ether vapor and immobilized by cervical dislocation. Then, their heart, liver, kidneys, intestine, and blood were sampled for clinical hematological variables and determination of Cd and Fe concentration by inductively coupled plasma optical emission spectrometry (ICP-OES). The data were subjected to statistical analysis using Student's t-test. All P values of less than 0.05 were considered significant.

Results: The results show that both chelators (DFS and L₁) increase the removal of Cd from the tissues. No effects of Cd or any of the two treatments (L₁ or DFS) were observed on white blood cell (WBC) count, red blood cell (RBC) count, and hemoglobin (Hb) level.

Conclusions: The comparison of single and combined therapy showed that the combined chelation therapy (DFS + L₁) was more effective in depleting Cd concentration in soft tissues.

Keywords: Chelation Therapy, Cadmium, Drinking Water, Rats

Introduction

Cadmium is a toxic metalloid widely present around the world particularly in soil, water, or contaminated food. Cd is primarily used in metal plating and coating operations, including transportation equipment, machinery and baking enamels, photography, and television phosphors. It is also used in nickel-cadmium and solar batteries, and in pigments (1). Cd is regularly found in ores together with zinc, copper, and lead. Therefore, volcanic activity is one natural reason for a temporary increase in environmental Cd concentrations. Phosphate fertilizers also show a high Cd load. Acute exposure to Cd fumes may cause flu-

like symptoms including chills, fever, and muscle ache sometimes referred to as "the cadmium blues". Symptoms may resolve after a week if there is no respiratory damage. More severe exposures can cause tracheobronchitis, pneumonitis, and pulmonary edema. Symptoms of inflammation may start hours after the exposure and include coughs, dryness and irritation of the nose and throat, headache, dizziness, weakness, fever, chills, and chest pain. Ingestion of any significant amount of Cd causes immediate poisoning and

* **Corresponding author:** Asghar Amiri, Dept. of Chemistry, Payame Noor University, P.O. Box, 19395-3697, Tehran, Iran.
Email: a.amiri@pnu.ac.ir

damage to the liver and the kidneys. The bones become soft (osteomalacia), lose bone mineral density (osteoporosis), and become weaker. This causes pain in the joints and back and also increases the risk of fractures (2, 3).

The concentrations of Cd in drinking water supplies of villages located in the southeastern region of Rafsanjan plain (Iran) exceed the standard limit permitted by the World Health Organization (WHO) guidelines (0.010 mg/l). About 10.4% of residents of this area were exposed to arsenic. Furthermore, the results revealed that 66.6% and 46.7% of the residents of the study area had, respectively, been exposed to high levels of lead and Cd. The heavy metals, such as Cd, contamination of drinking water resources in Rafsanjan plain is linked to both natural presence of sulfide veins in this area and manmade pollution due to the presence of the main road and agricultural use of pesticides (4, 5).

Detoxification of Cd is possible with ethylenediaminetetraacetic acid (EDTA) and other chelators and has been shown to be therapeutically beneficial in humans and animals when done using the established protocols. It is clear that EDTA and meso 2, 3-dimercaptosuccinic acid (DMSA) increase urinary excretion of Cd. In clinical use, EDTA is credited with an anecdotal report of rheumatoid arthritis relief, reduction of oxidative stress, and reduction of general metal toxicity. Subsequent human trials in West Bengal (India) with DMSA failed to provide clinical recoveries in patients chronically exposed to arsenic and some heavy metals (6-10). Clinical investigations of the use of some chelators for the removal of toxic metals in rats have been previously published (11, 12).

Deferasirox {4-[3,5-bis(2-hydroxyphenyl)-1,2,4-triazol-1-yl]-benzoic acid (ICL670 or DFS)} (Figure 1) is a tridentate chelator with high selectivity for Fe^{3+} . In 2005, DFS became the first Food and Drug Administration (FDA) approved oral alternative for treatment of Fe overload, and subsequently, was approved in

the EU in 2006 (8). Its comparatively long half-life before excretion allows a once-daily dosage and good overall patient compliance, as well as cost-effectiveness. DFS possesses a pFe^{3+} value of 22.5, can penetrate membranes easily, and possesses good oral availability. Indeed, when orally administered to hypertransfused rats, DFS promotes the excretion of chelatable iron from hepatocellular iron stores four to five times more effectively than desferrioxamine (13). Another developed orally active chelating agent is deferiprone (1,2-dimethyl-3-hydroxypyrid-4-one or L_1), which is rapidly absorbed in the gastrointestinal tract and normally appears in serum a few minutes after oral administration. The main excretion route is the kidneys. L_1 is a bidentate iron chelator forming a 3:1 complex with Fe and it is likely to act intracellular (14, 15). The presence of Cd in drinking water resources of Rafsanjan plain at concentrations greater than acceptable limits may result in various adverse health effects. The therapeutic efficacy of DFS and L_1 in reducing Cd concentration in Cd-exposed rats as a biological model was investigated in the present study.

Material and Methods

All the chemicals used in this work were of either analytical grade or of extra pure grade of the highest purity available locally. Cadmium chloride, deferiprone, and other materials were purchased from Merck Chemicals Co. (Germany) and deferasirox was purchased from Novartis Co. (Basel, Switzerland). Male Wistar rats were obtained from the animal house facility of Kerman Neuroscience Research Center (Iran). The animals were kept under a controlled light/dark (12/12 hours) schedule. The rats were divided randomly into control and experimental groups. They were treated in groups and were housed in well-cleaned sterilized cages in an air-conditioned room with the temperature maintained at 22 ± 2 °C and 50% humidity. The Animal Ethical Committee of Payame Noor University and

Kerman Neuroscience Research Center approved the protocols for the experiments.

Experimental Groups: In order to evaluate the efficacy of chelators (DFS and L₁) in removing Cd in Cd-exposed rats, experiments were performed on Wistar male rats (220 ± 12 g). The animals were classified into 2 groups. The control group (n = 5) was given normal food and distilled water to drink. The concentrations of Cd and Fe in the control group rats were compared with the groups that received Cd and chelators. The toxic groups (n = 25) were given water containing 40 mg/kg body weight Cd²⁺ as cadmium chloride for 42 days. In order to compare the Cd and Fe concentrations in tissues, before and after chelation therapy, one group was selected (Vehicle Cd and Vehicle Fe) and sacrificed before chelation therapy. Other Cd-exposed animals were divided into 4 sub-groups of 5 rats each and given the following treatment for 7 consecutive days:

- Group control chelator (No treatment)
- Group L₁ (100 mg/kg body weight, oral, once daily)
- Group DFS (100 mg/kg body weight, oral, once daily)
- Group DFS+L₁ (50 and 50 mg/kg body weight, respectively, oral, once daily)

All animals of each group were sacrificed under light ether anesthesia, 48 hours after the last dosing. Kidneys, heart, liver, and intestine samples were weighed, dried, and collected for determination of Cd and Fe concentration. The samples were placed in an oven at 60 °C for 3 days. Then, 1 g of each sample was digested

by 1 ml of HNO₃ (10 M). After digestion, the solutions were vaporized with the addition of 0.5 ml of H₂O₂ (30%) under a hood. Subsequently, the fragments were diluted with distilled water to 10 ml volume. Determination of Cd and Fe in samples was performed using inductively coupled plasma optical emission spectrometry (ICP-OES) (Vista-MPX, Varian Inc., CA, USA). The values are expressed as mean values (at least three separate determinations) ± standard error of the mean (SEM). The data were subjected to statistical analysis using Student's t-test. All P values of less than 0.05 were considered significant.

Clinical hematological variables: Blood was collected through cardiac puncture in heparinized tubes and the hemoglobin (Hb) level, platelet (PLT) count, red blood cell (RBC) count, and white blood cell (WBC) count were measured using a hematology analyzer (model K4500, Sysmex Corp., Kobe, Japan).

Results

Oral exposure to Cd may result in adverse effects on a number of tissues, including kidney, liver, bone, testes, the immune system, and the cardiovascular system. In humans, death is usually due to excessive fluid loss as a result of vomiting and diarrhea. Lethal doses in humans have been reported to range from 1,500 to 8,900 mg, corresponding to doses of about 20 to 130 mg/kg in a 70 kg adult (16). The effects of DFS and L₁ chelators on Cd concentration in various tissues are presented in table 1.

Table 1: Concentration of cadmium (mg/kg) in cadmium-intoxicated rats before and after chelation therapy

Group	Control	Vehicle Cd	DFS	L ₁	DFS + L ₁
Heart	0.12 ± 0.01	6.31 ± 0.32*	4.52 ± 0.37 [†]	5.04 ± 0.12*	4.02 ± 0.19 [†]
Kidney	0.72 ± 0.05	42.31 ± 0.21*	9.06 ± 0.24*	8.01 ± 0.32 [†]	7.07 ± 0.24 [†]
Liver	0.55 ± 0.02	15.11 ± 0.32*	6.62 ± 0.34*	7.02 ± 0.29 [†]	4.12 ± 0.25 [†]
Intestine	0.32 ± 0.01	6.96 ± 0.03*	4.21 ± 0.32 [†]	5.97 ± 0.23	3.01 ± 0.19 [†]

Values are presented as mean ± SEM (n = 5); *Significant at P < 0.05 when compared with control; [†]Significant at P < 0.05 when compared with vehicle Cd

Table 2: Concentration of iron (mg/kg) in cadmium-intoxicated rats before and after chelation therapy

Group	Control	Vehicle Fe	DFS	L ₁	DFS + L ₁
Heart	5.02 ± 0.42	4.76 ± 0.21	4.13 ± 0.29	3.06 ± 0.27*	3.01 ± 0.13*
Kidney	7.51 ± 0.23	5.85 ± 0.34*	6.01 ± 0.39	6.12 ± 0.29	5.12 ± 0.22*
Liver	9.03 ± 0.31	6.98 ± 0.27*	5.21 ± 0.31*	4.12 ± 0.21*	3.01 ± 0.21*
Intestine	5.02 ± 0.24	4.16 ± 0.33	4.03 ± 0.22	4.01 ± 0.20	4.01 ± 0.23

Values are presented as mean ± SEM (n = 5); *Significant at P < 0.05 when compared with control

The maximum amount of Cd accumulation was observed in the kidneys and liver, respectively. In order to investigate the spontaneous elimination of Cd from the body by the biological system, the control chelator group was treated without chelation therapy and the removal of Cd by the biological system in this group was not noticeable. After the chelation therapy, the obtained results indicated that Cd concentration had significantly reduced in all tissues. There was a statistical difference between DFS and L₁ in reducing the amount of Cd in various tissues. As single therapy efficiencies of chelators were compared in this study, it was found that

DFS was more effective in decreasing Cd level in all tissues, whereas L₁ was more effective in reducing Cd level in the kidneys. The results of Fe concentrations before and after chelation therapy are provided in table 2.

Fe concentration had significantly decreased after the chelation therapy. Thus, consumption of Fe tablets is recommended for returning the Fe level to its normal state. Combination of DFS + L₁ shows more efficiency in decreasing Fe level. The effects of exposure to Cd and treatment with chelators either individually or in combination on some hematological variables are shown in table 3.

Table 3: Hematological variables in the blood of cadmium-intoxicated rats before and after chelation therapy

Group	Control	Vehicle Cd	DFS	L ₁	DFS + L ₁
WBC	12.01 ± 2.11	12.01 ± 3.12	15.44 ± 2.21	10.11 ± 1.10	11.61 ± 1.29
RBC	8.22 ± 0.54	9.08 ± 0.11	8.55 ± 0.21	8.51 ± 0.12	8.65 ± 0.34
Hb	133.7 ± 11.4	120.7 ± 3	126.7 ± 1.3	121.5 ± 6	122.2 ± 4.5
PLT	679.6 ± 55.5	260.7 ± 88.7	553.6 ± 101.1*	221.7 ± 98.1	701.7 ± 56.4*

WBC: White blood cell count as $\times 10^3/\mu\text{l}$; RBC: Red blood cell count as $\times 10^6/\mu\text{l}$; Hb: Hemoglobin as g/l; PLT: Platelet as $\times 10^3/\mu\text{l}$; Values are presented as mean ± SEM (n = 5); *Significant at P < 0.05 when compared with vehicle Cd

Discussion

Through the survey of corrosion indexes, it was recognized that drinking water from wells and aqueducts in rural areas adjacent to Rafsanjan plain has scaling tendency. Thus, it is necessary to stop economical loss and hygienic harms in order to maintain water quality stabilization. Water corrosion leads to increase in toxic metals concentration, such as arsenic, copper, lead, Cd, zinc, nickel, iron, and manganese in water. Toxic metals have acute health risks for water consumers. Scaling water leads to economic and technical problems (16). After absorption, Cd is

transported throughout the body, usually bound to a sulfhydryl group-containing protein like metallothionein. About 30% deposits in the liver and 30% in the kidneys, with the rest distributed throughout the body. The half-life of Cd in the blood has been estimated at 75 to 128 days (16). It was found that as Cd concentration increased in tissues, Fe concentration decreased in the kidneys and liver, which is probably due to interference by Cd in the Fe uptake mechanism.

The combined therapy procedure can likely increase metal excretion, target specific metal tissues, minimize side effects, and improve compliance. Many studies have now reported

the high absorption/distribution, long-term efficacy, and safety of DFS and L₁ in removing some toxic metal ions and treating Fe overload in patients with β -thalassemia major (8-10). After administration of chelators, the Cd content was reduced. The results show that both chelators (DFS and L₁) increase the removal of Cd from the tissues. L₁ is able to redistribute Fe in mammals (17). DFS, by virtue of its small size and the ability to penetrate cells, is efficient in scavenging excess toxic Cd (16, 18). The comparison of single and combined therapy showed that the combined chelation therapy (DFS + L₁) is more effective in reducing Cd concentration in all tissues. No effects of Cd or any of the two treatments (L₁ or DFS) were observed on WBC, RBC, and Hb and PLT count showed a decrease in the Cd-exposed rats. Cd had no effect on these variables in the present study. This can be attributed to the short duration of Cd exposure. Treatment with DFS and DFS + L₁ provided significant recovery in terms of PLT counts.

Conclusion

Chelation therapy is one of the most effective ways to remove toxic metals from the biological system.

The comparison of the results indicates that the combined therapy (DFS + L₁) enhanced the removal of Cd from rat organs considerably. Each of the chelators (DFS and L₁) has a different target tissue. Therefore, their combination can effectively help the removal Cd from various tissues. This study might be effective for preliminary testing of the efficacy of chelating agents in the removal of Cd. Therefore, after essential preclinical experiments, the same study can be suggested for human administration.

Acknowledgments

The authors would like to thank Professor V. Sheibani from Kerman Neuroscience Research Center for his assistance.

Conflict of Interest: None declared.

References

1. Nordberg GF. Biomarkers of exposure, effects and susceptibility in humans and their application in studies of interactions among metals in China. *Toxicol Lett* 2010; 192(1):45-9.
2. Zalups RK, Ahmad S. Molecular handling of cadmium in transporting epithelia. *Toxicol Appl Pharmacol* 2003; 186(3):163-88.
3. Matovic V, Buha A, Bulat Z, Dukic-Cosic D. Cadmium toxicity revisited: focus on oxidative stress induction and interactions with zinc and magnesium. *Arh Hig Rada Toksikol* 2011; 62(1):65-76.
4. Malakootian M, Darabi-Fard Z, Amirmahani N, Nasiri A. Evaluation of arsenic, copper, lead, cadmium, and iron concentration in drinking water resources of central and southern Bardsir plain, Iran, in 2014. *Journal of Kerman University of Medical Sciences* 2015; 22(5): 542-54.
5. Malakootian M, Mobini M, Sharifi I, Haghghi Fard A. Evaluation of corrosion and scaling potential of wells drinking water and aqueducts in rural areas adjacent to Rafsanjan fault in during october to december 2013. *Journal of Rafsanjan University of Medical Sciences* 2014; 13(3):293-304.
6. Waters RS, Bryden NA, Patterson KY, Veillon C, Anderson RA. EDTA chelation effects on urinary losses of cadmium, calcium, chromium, cobalt, copper, lead, magnesium, and zinc. *Biol Trace Elem Res* 2001; 83(3):207-21.
7. Kelley C. Cadmium therapeutic agents. *Curr Pharm Des* 1999; 5(4):229-40.
8. Tandon SK, Prasad S, Singh S. Chelation in metal intoxication: influence of cysteine or N-acetyl cysteine on the efficacy of 2, 3-dimercaptopropane-1-sulphonate in the treatment of cadmium toxicity. *J Appl Toxicol* 2002; 22(1):67-71.
9. Bamonti F, Fulgenzi A, Novembrino C, Ferrero ME. Metal chelation therapy in rheumatoid arthritis: a case report. Successful management of rheumatoid arthritis by metal chelation therapy. *Biometals* 2011; 24(6):1093-8.
10. Piga A, Galanello R, Forni GL, Cappellini MD, Origa R, Zappu A, et al. Randomized phase II trial of deferasirox (Exjade, ICL 670), a once-daily, orally administered iron chelator, in comparison to deferoxamine in thalassemia patients with transfusional iron overload. *Haematologica* 2006; 91(7):873-80.

11. Amiri A, Fatemi SJ, Fatemi SN. Removal of thallium by combining desferrioxamine and deferiprone chelators in rats. *Biometals* 2007; 20(2):159-63.
12. Fatemi SJ, Tubafard S, Nadi B. Evaluation of the effect of cadmium on rat organs and investigation of diethyl carbamate as an oral drug in treatment of cadmium toxicity. *Med Chem Res* 2009; 18(3):179-86.
13. Voskaridou E, Christoulas D, Terpos E. Successful chelation therapy with the combination of deferasirox and deferiprone in a patient with thalassaemia major and persisting severe iron overload after single-agent chelation therapies. *Br J Haematol* 2011; 154(5):654-6.
14. Hershko C, Konijn AM, Nick HP, Breuer W, Cabatchik ZI, Link G. ICL670A: a new synthetic oral chelator: evaluation in hypertransfused rats with selective radio iron probes of hepatocellular and reticuloendothelial iron stores and in iron-loaded rat heart cell in culture. *Blood* 2001; 97(4):1115-22.
15. Steinhauser S, Heinz U, Bartholoma M, Weyhermuller T, Nick H, Hegetschweiler K. Complex formation of ICL670 and related ligands with Fe-III and Fe-II. *Berichte der deutschen chemischen Gesellschaft* 2004; 21:4177-92.
16. Luparello C, Sirchia R, Longo A. Cadmium as a transcriptional modulator in human cells. *Crit Rev Toxicol* 2011; 41(1):75-82.
17. Neufeld EJ. Oral chelators deferasirox and deferiprone for transfusional iron overload in thalassaemia major: new data, new questions. *Blood* 2006; 107(9):3436-41.
18. Wan L, Zhang H. Cadmium toxicity: effects on cytoskeleton, vesicular trafficking and cell wall reconstruction. *Plant Signal Behav* 2012; 7(3):345-8.