

Research Article

Feeding Efficiency of Green Lacewing, *Chrysoperla carnea* (Stephens) against Different Species of Aphid in Laboratory Conditions

Lok Bahadur Rana^{1*}, Ram Prasad Mainali², Homan Regmi² and Binayak Prasad RajBhandari¹

¹Himalayan College of Agricultural Sciences and Technology, Kalanki, Kathmandu, Nepal ²Nepal Agricultural Research Council, Lalitpur, Nepal

*Corresponding author email: lokrana222@gmail.com

Abstract

Green lacewing, *Chrysoperla carnea* Stephens (Neuroptera: Chrysopidae) is the most effective polyphagous predator of different species of aphids and is commonly known as "aphid lion". The experiment on feeding efficiency of green lacewing was studied in the laboratory of Entomology Division, NARC, Khumaltar, Lalitpur, Nepal from 21st, December 2015 to 26th, March 2016. The known number of predatory larva of green lacewings were fed with known number of seven different species of live aphid and frozen *Corcyra* eggs representing each treatment. The treatments were replicated four times. The predatory efficiency was calculated by counting the number of consumed host per day. The result revealed that the predatory efficiency of *C. carnea* larvae were increased from first to third instar and third instar were more voracious as compare to first two instars. It consumed significantly the highest rice moth, *Corcyra cephalonica* followed by *Aphis craccivora* and others aphid species, respectively. From this experiment, it is evident that the green lacewing is potent bio-agent against different aphid species and hence further research is required simultaneously in the farmer's field conditions.

Key words: Chrysoperla carnea; predatory efficiency; aphids; Corcyra cephalonica.

Introduction

Insects, diseases, weeds and nutritional factors are major constraints acting against the quality and quantity of crops yield. Out of many insect pests, aphids and mites are the most important and serious insect pests of crops (E.D., 2013). The aphids are one that damages the various crops in which they habitat. They damages crops by sucking sap from plant and transferring viral diseases to healthy plants. Aphids infest wide range of several agricultural crops in horticulture, cereal crops, oilseed crops etc.

Farmers are using more than one pesticide in alternating manner to suppress insect pest in their field (E.D., 2013). The average use of pesticides is 396 g a.i/ha in Nepal (PPD, 2015) and 500g a.i./ha in the world (Kodandaram *et al.*, 2013). Although, use of pesticides rate in Nepal is lower, but indiscriminate use of chemical pesticides in the agricultural crops have created many problems. Resulting that development of resistance to insecticides, pesticides residue on food, air, water and soil, pest resurgence, killing of natural enemies, harmful effect on non-target organisms including pollinators and disruption of ecosystem, hereby increasing the cost of production and hazard on human

beings and animals (Palikhe, 2002; Atreya, 2007; Neupane, 2010; Sharma *et al.*, 2012).

These negative impacts of chemical pesticides on human health and environment, have led to realize the need for alternative method, which is environmentally friendly, economically viable and sustainable method of insect pest management. It can be reduced or minimized through the development, dissemination and promotion of alternative method such as botanical pesticides (Akter, 2015; Kafle, 2015), biological pest control (Pinstrup-Andersen and Hazell, 1985) and IPM approach (Neupane, 2010). It is important to reduce the pesticides application on crops by using or conserving the biologically derived predator in the field such as Green lacewing, Chrysoperla carnea (Stephens) (Sarwar, 2014). The common green lacewing is an important generalist predator (Cheng et al., 2010; Jokar and Zarabi, 2012 and Sarwar, 2014) is best known as biocontrol agent (Menon et al., 2015). The larval stage are more voracious feeder of soft bodied insect such as aphid, whitefly, mealy bugs, thrips, mites, leaf hoppers, jassids, caterpillar and insect eggs (Ulhaq et al., 2006; Aryal and Giri, 2015; Sarwar and Salman, 2016) however, aphids are more preferred host (Solangi et al., 2013). The adults are free living and they only feed honey, pollen and water (Borah *et al.*, 2012; E.D., 2013; Nadeem *et al.*, 2014; E.D., 2015). The ability of *C. carnea* can be exploited as a biocontrol agent in IPM program (Bozsik *et al.*, 2009; Memon *et al.*, 2015). The application of the predator reduces the use of insecticides and save money spent on importing pesticides (Zia *et al.*, 2008).

After knowing the importance of *C. carnea* in agricultural systems, it is important to develop efficient pest management strategies that are simple, economical, sustainable and bio-friendly based on biological control. The objective of this study was to determine feeding efficiency of *C. carnea* on different species of aphid for effective management of aphids on agricultural crops.

Materials and Methods

The experiment was conducted on the feeding efficiency of C. carnea under controlled conditions at bio-agent insect rearing room, Entomology Division, NARC, Khumaltar, Lalitpur, Nepal during winter season from 21st December, 2015 to 26th March, 2016. The experiment was performed in a completely randomized design consisting of eight treatments, and each treatment was comprised of four replicates. The natural hosts were Aphis craccivora (Koch), Brevicorynae brassicae (Linnaeus), Myzus persicae (Sulzer), Eriosoma lanigerum (Hausman), Lipaphis erysimi (Kaltenbach), Aphis fabae (Scopoli), lanigera (Zehntner) Ceratovacuna and Corcyra cephalonica (Stainton) (Frozen eggs). The first seven hosts

were collected from field on daily basis. Eggs of *C*. *cephalonica* were taken from laboratory culture maintained for experimental purpose.

The five freshly hatched *C. carnea* larvae were kept in plastic bottles sized 8cm×6cm for each treatment and provided with 25 number of hosts per day. After providing hosts, bottles were covered at the top by piece of black muslin cloth and were fastened by rubber string. The number of each prey consumed by the predatory larvae was recorded by counting the live preys after every 24 hrs. Then, fresh aphids and frozen eggs were provided in each treatment. Similar, counting method was adopted by Chakraborty and Korat (2010); Gupta and Mohan (2012); Memon *et al.* (2015). The larvae were fed with aphids in bottles until pupation. To avoid cannibalism between newly hatched larvae, use of small piece of photocopy paper (10cm×8cm) folding 3 to 4 times in plastic bottles (Fig. 1).

All the recorded data were subjected to statistical analysis (one-way analysis of variance, ANOVA), all treatments means were compared by using Duncan's Multiple Range Test (DMRT) with the help of GenStat statistical package (VSN international Ltd.) as analyzing tool. Five percent significance level was considered for ANOVA. The result reported as Mean \pm S.D.



Fig. 1: Some photographs taken during experiment

			Mean n	umber of prey	Mean number of prey consumed by C. carnea±S.D.	carnea±S.D.					
Larval stage	A. craccivora	B. brassicae	M. persicae	M. persicae E. lanigerum L. erysimi	L. ervsimi	A. fabae	C. cephalonica C. lanigera	C. lanigera	L.S.D. 5%	C.V. (%)	F-value
181	7.7±2.51a	15.55±3.15d	11.9±0.33b	11.75±0.49b	16.37±2.20d	12.8±2.11bc	15.55±3.15d 11.9±0.33b 11.75±0.49b 16.37±2.20d 12.8±2.11bc 14.25±0.45bc 14.4±1.89bc	14.4±1.89bc	3.25	17	<0.001
2¤d	14.08±2.31a	16.73±1.69a	13.15±3.39a	12.99±1.62a	13.15±3.39a 12.99±1.62a 16.75±2.56a	13.87±3.77a	54±17.45b	20.42±5.36a	11.56	39.1	<0.001
3rd	110.43±35.69e	90.91±11.05cd	54.1±5.17ab	46.87±12.15a	89.66±8.59cd	56.33±13.26ab	110.43±35.69e 90.91±11.05cd 54.1±5.17ab 46.87±12.15a 89.66±8.59cd 56.33±13.26ab 166.87±36.09f 71.9±14.07ab	71.9±14.07ab	34.34	27.4	<0.001
Total	132.22±36.52e	123.20±9.78cd	79.15±5.22a	71.62±10.78a	122.78±9.78cd	83.01±14.68ab	235.12±43.32f	123.20±9.78cd 79.15±5.22a 71.62±10.78a 122.78±9.78cd 83.01±14.68ab 235.12±43.32f 106.72±18.08abd	38.1	21.9	<0.001

Table 1: Prey consumption of different larval instar of C. carned when reared on different preys species under no choice of feeding

Result and Discussion

Table 1 illustrates the prey consumption of different larval instar of C. carnea. The first instar larva of C. carnea was consumed significantly maximum number (16.37±2.20) of L. erysimi followed by B. brassicae, eggs of C. cephalonica, C. lanigera, A. fabae, M. persicae, E. lanigerum, A. craccivora, respectively. The second instar larva of C. carnea was consumed significantly maximum number (54±17.45) eggs of C. cephalonica followed by C. lanigera, B. brassicae, L. ervsimi, A. craccivora, M. persicae, A. fabae, E. lanigerum, respectively. Similarly, third instar larva was consumed significantly more number eggs of C. cephalonica (166.87±36.09) followed by A. craccivora, B. brassicae, L. erysimi, C. lanigera, A. fabae, M. persicae, E. lanigerum, respectively. The result was supported by the authors Aravind et al. (2012) who reported that the third instar larvae of C. carnea was consumed more number eggs of C. cephalonica.

The host consumption data revealed that the consumption rate of predator, *C. carnea* was increased with increased predatory stages: first, second and third larval instar in all the prey species. From the study, it was evident that the third instar larvae of *C. carnea* consumed maximum number of aphids and eggs of *Corcyra*. This is in conformity with the study of various authors Chakraborty and Korat (2010); Gupta and Mohan (2012); Solangi *et al.* (2013). It was observed that maximum food consumption (60-67%) by third instar larvae of *C. carnea* followed by second (20-24%) and first instar (10-17%) respectively (Yadav and Pathak, 2010).

Khan *et al.* (2013) reported that the predatory efficiency of first, second and third instar larva of *C. carnea* was 61 ± 1.97 , 113.6 ± 2.42 and 239.2 ± 6.87 number of aphids respectively. This result also supported our present finding as they concluded that the third instar larva of *C. carnea* was consumed maximum number of aphids than first and second respectively.

The total number food consumed by the predatory larva of *C. carnea* on the different hosts in the sequence of feeding potential in decreasing order eggs of *C. cephalonica*> *A. craccivora*> *B. brassicae*> *L. erysimi*>*C. lanigera*> *A. fabae*> *M. persicae*> *E. lanigerum* respectively. This decreasing order shows that the predatory larva of *C. carnea* consumed maximum on eggs of *C. cephalonica* and minimum fed on apple wooly aphid (*E. lanigerum*). According to Nandan *et al.* (2014) larvae of *Chrysoperla* was more preferred to feed eggs of *C. cephalonica* followed by different host *A. craccivora*, *B. brassicae*, *A. gossypii*, *M. persicae* and *L. erysimi* respectively.

Hassan (2014) reported that the *C. carnea* larva was fed on the different eggs masses of *C. cephalonica, Pectinophora gossypiella* and *Sitotroga cerealella* with an average 493.6 ± 50.32 , 654.3 ± 32.54 and 673.9 ± 31.52 numbers

respectively under no choice feeding conditions. However, the host preference (free Choice) data revealed that the predatory larva consumed 264.1 ± 68.8 , 111.2 ± 56 and 63.3 ± 47 numbers eggs of *C. cephalonica, P. gossypiella* and *S. cerealella,* respectively. This result shows that the most preferred host of *C. carnea* larvae was eggs of *C. cephalonica.*

Yadav and Pathak (2010) observed that the maximum predation rate of C. carnea larva on A. craccivora followed by A. gossypii, M. persicae and L. erysimi. However, in an another study, the total consumption of aphids by the predatory larva of C. carnea was 174.63±6.11, 143.40±8.70 and 131.80±6.62 numbers on L. ervsimi, A. craccivora, and B. brassicae, respectively, during whole larval period (Chakraborty and Korat, 2010). The results of present study and previous researcher showed different in the feeding efficiency of predatory larva of C. carnea in the different hosts. Previous authors (Chakraborty and Korat, 2010; Yadav and Pathak, 2010; Khan et al., 2013) reported that there are different factors that play important role in obtaining different results such as different stages (instars) of prey offered for feeding, size of prey species, different hosts species, preys populations and environmental conditions prevailing during the study period.

Conclusion

The present research finding demonstrates that the third instar larvae of *C. carnea* are more voracious as compare to 1^{st} and 2^{nd} instars. It is evident that the eggs of rice meal moth, *C. cephalonica* were more preferred host of *C. carnea* hence; it can be utilized as mass rearing diet of this predator. The predatory larvae fed on different aphid species and hence, these potential to utilize for biological control agent for management of the aphids. This result guides the entomologist to consider the *C. carnea* as efficient bio-control agent in eco-friendly management of aphids on agricultural crops and so, enhancing the potential of predators.

Acknowledgement

We would like to thank Entomology Division, NARC, Khumaltar, Lalitpur, Nepal for providing facilities to undergo research study. The co-operation and assistant made by HICAST, Purbanchal University team are also fully acknowledged.

References

- Akter A, Kabir MR, Roni MZK and Uddin AJ (2015) Control of mustard aphid (*Lipaphis erysimi*) using different botanical insecticides. *Bangladesh Res Pub J* 10(4): 298-303.
- Aravind J, Karuppuchamy P, Kalyanasundaran M and Boopath T (2012) Predatory potential of green lacewing, *Chrysoperla zastrowisillemi* (Esben-Petersen) (Neuroptera: Chrysopidae) on major sucking pests of

okra. Pest Management in Horticultural Ecosystems **18**(2): 231-232.

- Atreya K (2007) Pesticide Use in Nepal: Understanding Health Costs from Short-term Exposure. South Asian Network for Development and Environmental Economics (SANDEE), Kathmandu, Nepal.
- Borah T, Rahman A, Borthakur M and Barthakur BK (2012) Biology and predatory potential of a newly recorded Green lacewing, *Mallada* sp. (Neuroptera: Chrysopidae) on tea mosquito bug, *Helopeltis theivora* (water house). *Two and bud* **59**:60-62.
- Bozsik A, Ramon GR, and Baltasar HL (2009) Distribution of the *Chysoperla carnea* Complex in Southern Spain (Neuroptera: Chrysopidae). *In:* International Symposia Risk Factors for Environment and Food Safety & Natural Resources and Sustainable Development, Faculty of Environmental Protection ,November 6-7, Spain, pp: 71-78.
- Chakraborty D and Korat DM (2010) Feeding efficiency of green lacewing, *Chrysoperla carnea* (Stephens) on different species of aphids. *Karnataka J Agric Sci* **23**(5):793-794.
- Cheng LL, Nechols JR, Margolies DC, Campbell JF and Ping SY (2010) Assessment of prey preference by the massproduced generalist predator, *Mallada basalis* (Walker) (Neuroptera: Chrysopidae), when offered two species of spider mites, *Tetranychus kanzawai* Kishida and *Panonychus citri* (McGregor) (Acari: Tetranychidae), on papaya. *Biological Control* **53**: 267–272. DOI: <u>10.1016/j.biocontrol.2010.02.006</u>
- ED (2013) Annual report 2069/2070 (2012/13) Entomology Division, NARC, Khumaltar, Lalitpur, Nepal. 90p.
- ED (2015) Annual report 2071/72 (2014/15) Entomology Division, NARC, Khumaltar, Lalitpur Nepal. 100p.
- Gupta R and Mohan M (2012) Feeding efficiency of Chrysoperla carnea against aphids (L. erysimi and B. brassicae). An International Quarterly Journal of Life Sciences 7(3): 455-456.
- Hassan KA (2014) Feeding capacity and host preference of *Chrysoperla carnea* (Stephens) (Neuroptera: Chrysopidae) on three different insect prey under laboratory conditions. *J Plant prot path Mansoura Univ* 5(12): 1045-1051.
- Jokar M and Zarabi M (2012) Prominence of three diets on life table parameters for *Chrysoperla carnea* (Neuroptera: Chrysopidae) to mass rearing under laboratory conditions. *Archives of Phytopathology and Plant Protection* **45**(18): 2213-222. DOI: 10.1080/03235408.2012.724971
- Kafle K (2015) Management of mustard aphid, *Lipaphis erysimi* (Kalt.) (Homoptera: Aphididae). *International Journal Applied Sciences and Biotechnology* 3(3): 537-540.
 DOI: <u>10.3126/ijasbt.v3i3.13334</u>
- Khan MH, Ahmad N, Rashdi SMM, Ismail M, Rauf I and Tofique M (2013) Studies on the compatibility of neem

oil with predator, *C. carnea* for the management of aphids (Homoptera: Aphididae) in canola (*Brassica napus* L.). Journal of cereals and oilseeds 4(6): 85-88.

- Kodandaram MH, Saha S, Rai AB and Prakash S, Naik PS (2013) Compendium on pesticide use in vegetables. IIVR Extension Bulletin No. 50, Varanasi.
- Memon AS, Omar D, Muhamad R, Sajap AS, Asib N and Gilal AA (2015) Functional responses of green lacewing, *Chrysoperla nipponensis* (Neuroptera: Chrysopidae) reared on natural herb based artificial diet. *Journal of Entomology and Zoology studies* 3(6): 80-83.
- Nadeem S, Hamed M, Ishfaq M, Nadeem MK, Hasnain M, and Saeed NA (2014) Effect of Storage Duration And Low Temperatures On The Developmental Stages Of *Chrysoperla carnea* (Stephens) (Neuroptera: Chrysopidae). *The Journal of Animal & Plant Sciences* 24(5): 1569-1572.
- Nandan N, Korat DM and Dabhi MR (2014) Influence of different host insect (prey) on biological parameter of *Chrysoperla zastrowisillemi* (Esben-Peterson). *Insect environment* 20(2): 40-44.
- Neupane FP (2010) Crop pests and their management. 5th Eds., Sajha Prakashan, Lalitpur, Nepal. 584pp.
- Palikhe BR (2002) Challenges and options of pesticide use: In the context of Nepal. *Landschaftsökologie and Umweltforschung* **38**: 130-141.
- Paneru RB and Giri YP (2011) Management of Economically Important Agricultural and Household Pest of Nepal. Entomology Division, Khumaltar, Lalitpur, Nepal. 136pp.
- Pinstrup-Andersen P and Hazell PBR (1985) The Impact of the Green Revolution and Prospects for the Future. *Food Reviews International* 1(1): 1-25. DOI: 10.1080/87559128509540765

- PPD (2015) Annual report 2071/2072. Plant Protection Directorate, Hariharbhawan, Lalitpur, Nepal.
- Sarwar M (2014) The propensity of different larval stages of lacewing, Chrysoperla carnea (Stephens) (Neuroptera: Chrysopidae) to control aphid, Myzus persicae (Sulzer) (Homoptera: Aphididae) evaluated on Canola, Brassica napus L. Songklanakarin J Sci Technol 36(2): 143-148.
- Sarwar M and Salman M (2016) From Production to Field Application Methodology of Generalist Predator Green Lacewing, *Chrysoperla carnea* [Stephens] (Neuroptera: Chrysopidae). *International Journal of Zoology Research* 1(1):35-40.
- Sharma DR, Thapa RB, Manandhar HK, Shrestha SM, Pradhan SB (2012) Use of pesticides in Nepal and impacts on human health and environment. *The Journal of Agriculture and Environment* 13:67-74.
- Solangi AW, Lanjar AG, Baloch N, Rais MUN and Khuhro SA (2013) Population, host preference and feeding potential of *Chrysoperla carnea* (Stephens) on different insect host in cotton and mustard crops. *Sindh Univ Res Jour (Sci sre)* **45**(2): 213-218.
- Ulhaq MM, Sattar A, Salihah Z, Farid A, Usman A and Khattak SUK (2006) Effect of different artificial diet on the biology of adult green lacewing (*Chrysoperla carnea*: Stephens.). Songklanakarin J Sci Technol 28(1): 1-8.
- Yadav R and Pathak PH (2010) Effect of temperature on the consumption capacity of *Chrysoperla carnea* (Stephens) (Neuroptera: Chrysopidae) reared on four aphid Species. *Int Quat J Life Sci* **5**(2): 271 -274.
- Zia K, Hafeez F, Khan RR, Arshad M and Ullah UN (2008) Effectiveness of *Chrysoperla carnea* (Stephens) (Neuroptera: Chrysopidae) on the population of *Bemisia tabaci* (genn.) (Homoptera: Aleyrodidae) in different cotton genotypes. J Agri Soc Sci 4: 112–116.