



Research Article

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Synthesis, Spectral and Molecular Characterization of Some Novel 2, 5-Disubstituted-1, 3, 4-Oxadiazole Derivatives and Evaluation of *in vivo* Antitumour Activity against HT 29 Cell Line

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ABSTRACT

Neoplasia is a type of abnormal and excessive growth of tissue. The growth of a neoplasia is uncoordinated with that of the normal surrounding tissue, and it persists growing abnormally, even if the original trigger is removed. This abnormal growth usually forms a mass. The main objective of the present research work was the synthesis, characterization and evaluation of *in vivo* antitumour activity of some novel 2, 5-disubstituted 1, 3, 4-oxadiazole derivatives. The *in vivo* antitumour activity of synthesized compounds was evaluated by HT 29 cell line induced malignant ascites on mouse model. The apoptosis of HT 29 cells was evaluated by using Gimsa and H33342 stain and the apoptosis ratios were analysed by FCM using AnnexinV-FITC/PI staining. The present experimental data displayed that the mortality was less in all groups except in tumour control group and all the synthesized compounds AB1-AB8 (100 mg/kg) significantly increased the PILS. While 5-FU increased the life span of 97.72%, and the PILS of synthesized compounds were found to be 45.45%, 59.09%, 68.18%, 56.81%, 38.63%, 84.09%, 77.27% and 90.90%. So the Synthesized compounds AB1-AB8 at the dose of 100 mg/kg significantly improved the overall survival of all treated animals and 5-FU was not significantly differed from each other in improving the overall survival of HT-29 cells. The apoptosis ratios of synthesized compounds were found as followed: AB1=26%; AB2=37.6%; AB3=43%; AB4=29%; AB5=24.1%; AB6=59.2%; AB7=48.2%; and AB8=63% respectively, while that of the Group-II (T. control) was 6.1%. When compared with standard drug 5-FU: 66.2%, it was indicated that compound AB8>AB6>AB7>AB3 were able to significantly induce HT-29 cells apoptosis.

Keywords: Neoplasia, antitumour, malignant ascites, HT 29, apoptosis, PILS.

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INTRODUCTION

Oxadiazoles are a class of heterocyclic aromatic chemical compound of the azole family; with the molecular formula $C_2H_2N_2O$. There are four isomers of oxadiazole depending on the position of nitrogen atom in the ring. [1] 1, 3, 4-oxadiazole is a five membered heterocyclic aromatic compound containing two nitrogen atom at position three and four and one oxygen atom present at position one. 1, 3, 4 oxadiazole is thermally stable than other oxadiazoles, these oxadiazole are very important compound in medicinal chemistry due to their biological activities, during last few years. [2] 1, 3, 4-oxadiazole is a liquid having boiling point 150°C . 2, 5-disubstituted-1, 3, 4-oxadiazole derivatives are colorless substances. The lower alkyl derivatives are liquids which distil without decomposition. Replacement of an alkyl residue by an aryl radical considerably raises the melting and boiling points. Usually the asymmetrical 1, 3, 4-oxadiazole derivatives melt and boil at lower temperature than the symmetrical compounds. The solubility of oxadiazoles in water varies with the substituents present: 2, 5-dimethyl-1, 3, 4-oxadiazole is miscible with water in all proportions whereas the solubility of 2, 5-diphenyl-1, 3, 4-oxadiazole in water is less. Electrophilic introduction of functional groups (for example nitro or sulphuric acid groups) into the nucleus is unusual. Electrophilic substitution occurs in aryl substituent. Halogenations are also difficult, but 2, 5-diaryl-1, 3, 4-oxadiazoles, afford complexes with halogens. A range of acylation and alkylation reactions of hydroxyl, thio and amino-1, 3, 4-oxadiazoles occur at the ring nitrogen. [3] Biologically active molecules containing oxadiazole moiety possessed a wide range of pharmacological activities such as antimicrobial, anticancer, anticonvulsant, anti-inflammatory and antiviral agents [4], antifungal [5], antimycobacterial [6] etc.

MATERIALS AND METHODS

Chemicals

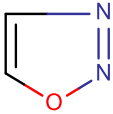
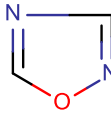
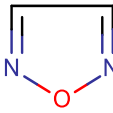
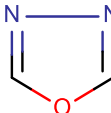
The solvents and other chemicals which were used for the synthesis and purification of target compounds provided by institutional store and were of LR and AR grade.

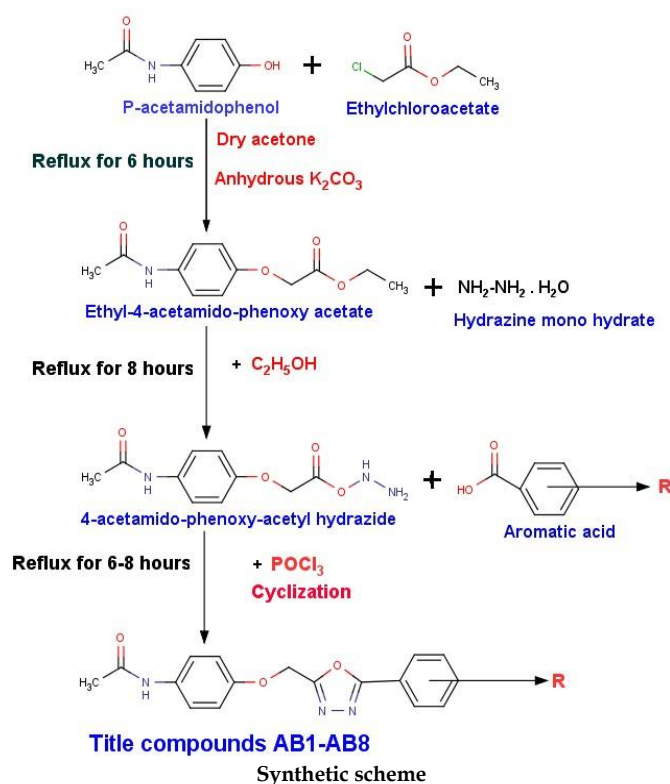
Instrumentation

The melting points of the synthesized compounds were determined by open capillary tube method. The IR spectra of the synthesized compounds were recorded on ABB Bomen FT-IR spectrometer MB 104 IR spectra recorded with potassium bromide pellets. The ^1H -NMR spectra of synthesized compounds were recorded on instrument BRUKER NMR spectrometer in DMSO. The Mass spectra of synthesized compounds were recorded JEOL GCmate. TLC method was used to determine the progress of the reaction. TLC plates are Pre-coated Silica gel (HF254-200 mesh) aluminium plates using ethyl acetate: n-hexane are used as solvent and visualized under UV- chamber. The IR, ^1H -NMR and

MASS spectra were used to assign the structure of synthesized compounds.

Table 1: Oxadiazole

	
1,2,3-oxadiazole	1,2,4-oxadiazole
	
1,2,5-oxadiazole	1,3,4-oxadiazole



Steps involved in the synthesis of target compounds [7-9]

Step 1: Ethyl-4-acetamido phenoxy acetate

A mixture of p-acetamido phenol (0.01 mol) and ethyl chloroacetate (0.01 mol) was refluxed by using dry acetone in presence of anhydrous potassium carbonate (K_2CO_3) for 6 hours. The reaction mixture was cooled and then poured in to crushed ice. The solid product obtained, these product was filtered, dried and recrystallized using ethanol.

Step 2: 4-Acetamido phenoxy acetyl hydrazide

A mixture of ethyl-4-acetamido phenoxy acetate (0.01 mol), hydrazine hydrate (0.01 mol) in ethanol (15 ml) was refluxed for 5-8 hours. The reaction mixture was cooled and then poured in to crushed ice. The solid product was obtained; this product was filtered, dried and recrystallized from ethanol.

Step 3: 2-(4-Acetamidophenoxy methyl) -5-aryl substituted - 1, 3, 4-oxadiazole

A mixture of 4-Acetamido phenoxy acetyl hydrazide (0.01 mol) and various aromatic acids (0.01 mol) in phosphorus oxychloride (10 ml) was refluxed for 6-8 hours. The completion of the reaction process was monitored by TLC plates. The contents were cooled and poured into the crushed ice and then neutralized the reaction mixture with sodium bicarbonate solution and the solid product was obtained; the product was filtered, dried and recrystallized from ethanol. [10]

Table 2: Physicochemical properties of synthesized compounds

S. No.	Compounds code	M. F	M. Wt	R _f value	m. p (°C)	Yield (%)
1.	AB1	C ₁₇ H ₁₆ N ₄ O ₃	324.33	0.77	116	74.5
2.	AB2	C ₁₇ H ₁₃ Cl ₂ N ₃ O ₃	378.20	0.74	180	69.9
3.	AB3	C ₁₇ H ₁₄ FN ₃ O ₃	327.30	0.75	189	74
4.	AB4	C ₁₇ H ₁₄ BrN ₃ O ₃	388.21	0.65	183	69
5.	AB5	C ₁₇ H ₁₃ BrN ₄ O ₅	433.21	0.64	166	60
6.	AB6	C ₁₇ H ₁₄ N ₄ O ₅	354.31	0.72	171	64
7.	AB7	C ₁₇ H ₁₃ N ₅ O ₇	399.31	0.68	204	78
8.	AB8	C ₁₇ H ₁₃ N ₅ O ₈	415.31	0.72	215	68

Spectral data of synthesized compounds**Compound AB1**

N-(4-[[5-(4-aminophenyl)-1,3,4-oxadiazol-2-yl]methoxy]phenyl) acetamide. IR (KBr) ν (cm⁻¹): 3393.16 cm⁻¹ (Ar-NH), 1633.67 cm⁻¹ (C=N), 1575.88 cm⁻¹ (C=C), 1069.05 cm⁻¹ (-C-O-C-), 3132.54 cm⁻¹ (Ar-CH), 1249.43 cm⁻¹ (Ar-NH₂), ¹H-NMR δ (ppm): 6.45-7.4 (s, 8H, Ar-H), 5.17 (s, 2H, -CH₂), 4.1 (s, 2H, -NH₂), 2.05 (s, 1H, -CH₃), 8.05 (s, 1H, -NH), Mass (m/e value) % relative abundance: 324.12 (M⁺) (5.1), 310.87 (4), 296.22 (8.25), 282.76 (2.2), 272.38 (2.32), 262.64 (7.3), 248.34 (11), 217.12 (15), 207.14 (7), 116.67 (18), 58.33 (B).

Compound AB2

N-(4-[[5-(2,4-dichlorophenyl)-1,3,4-oxadiazol-2-yl]methoxy]phenyl) acetamide. IR (KBr) ν (cm⁻¹): 3381.92 cm⁻¹ (Ar-NH), 1673.42 cm⁻¹ (C=N), 1545.03 cm⁻¹ (C=C), 1085.04 cm⁻¹ (-C-O-C-), 687.47 cm⁻¹ (C-Cl), 3115.62 cm⁻¹ (Ar-CH), ¹H-NMR δ (ppm): 6.6-7.82 (s, 8H, Ar-CH), 2.5 (s, 3H, -CH₃), 8.03 (s, 1H, -NH), 5.22 (s, 2H, -CH₂), Mass (m/e value) % relative abundance: 377.03 (M⁺) (2.8), 333.16 (1.5), 325.42 (2.7), 286.43 (2.6), 183.26 (6), 160.62 (7), 140.65 (16), 115.64 (33), 95.53 (B).

Compound AB3

N-(4-[[5-(4-fluorophenyl)-1,3,4-oxadiazol-2-yl]methoxy]phenyl) acetamide. IR (KBr) ν (cm⁻¹): 3392.09 cm⁻¹ (Ar-NH), 1617.53 cm⁻¹ (C=N), 1528.16 cm⁻¹ (C=C), 1093.52 cm⁻¹ (-C-O-C-), 1371.78 cm⁻¹ (C-F), 3114.61 cm⁻¹ (Ar-CH), ¹H-NMR δ (ppm): 2.21 (s, 1H, -CH₃), 8.09 (s, 1H, -NH), 5.21 (s, 1H, -CH₂), 6.7-8.01 (m, 8H, Ar-CH), Mass (m/e value) % relative abundance: 327.10 (M⁺) (6.3), 310.37 (2.3), 299.57 (3), 282.87 (3.9), 266.22 (5), 249.61 (1.2), 232.72 (4), 104.86 (8.1), 75.50 (B).

Compound AB4

N-(4-[[5-(2-bromophenyl)-1,3,4-oxadiazol-2-yl]methoxy]phenyl) acetamide. IR (KBr) ν (cm⁻¹): 3286.82 cm⁻¹ (Ar-NH), 1617.53 cm⁻¹ (C=N), 1528.16 cm⁻¹ (C=C), 1093.52 cm⁻¹ (-C-O-C-), 687.47 cm⁻¹ (C-Br),

3114.61 cm⁻¹ (Ar-CH), ¹H-NMR δ (ppm): 2.21 (s, 1H, -CH₃), 8.09 (s, 1H, -NH), 5.21 (s, 1H, -CH₂), 6.7-8.01 (m, 8H, Ar-CH), Mass (m/e value) % relative abundance: 387.02 (M⁺) (6.3), 310.37 (2.3), 299.57 (3), 282.87 (3.9), 266.22 (5), 249.61 (1.2), 232.72 (4), 104.86 (8.1), 75.60 (B).

Compound AB5

N-(4-[[5-(2-bromo,4-nitrophenyl)-1,3,4-oxadiazol-2-yl]methoxy]phenyl) acetamide. IR (KBr) ν (cm⁻¹): 3381.95 cm⁻¹ (Ar-NH), 1684.44 cm⁻¹ (C=N), 1586.2 cm⁻¹ (C=C), 1064.25 cm⁻¹ (-C-O-C-), 1365.57 cm⁻¹ (N=O), 619.89 cm⁻¹ (C-Br), 3130.43 cm⁻¹ (Ar-CH), ¹H-NMR δ (ppm): 6.74-8.36 (m, 7H, Ar-CH), 5.31 (s, 2H, -CH₂), 2.31 (s, 1H, -CH₃), 8.16 (s, 1H, -NH), Mass (m/e value) % relative abundance: 432.00 (M⁺) (4), 388.71 (8.1), 362.27 (4.2), 233.28 (5), 217.31 (8.9), 182.52 (5), 96.79 (7), 78.82 (B).

Compound AB6

N-(4-[[5-(4-nitrophenyl)-1,3,4-oxadiazol-2-yl]methoxy]phenyl) acetamide. IR (KBr) ν (cm⁻¹): 3382.43 cm⁻¹ (Ar-NH), 1703.01 cm⁻¹ (C=N), 1592.32 cm⁻¹ (C=C), 1088.54 cm⁻¹ (-C-O-C-), 1378.11 cm⁻¹ (N=O), 3112.69 cm⁻¹ (Ar-CH), ¹H-NMR δ (ppm): 6.41-7.8 (m, 8H, Ar-CH), 2.42 (s, 3H, -CH₃), 8.13 (s, 1H, -NH), 5.21 (s, 2H, -CH₂), Mass (m/e value) % relative abundance: 354.09 (M⁺) (3.8), 335.16 (4.8), 302.39 (3.1), 287.43 (3.7), 249.58 (7.1), 226.00 (5.8), 204.96 (6.7), 127.56 (13.1), 103.69 (9), 89.93 (B).

Compound AB7

N-(4-[[5-(3,5-dinitrophenyl)-1,3,4-oxadiazol-2-yl]methoxy]phenyl) acetamide. IR (KBr) ν (cm⁻¹): 3382.02 cm⁻¹ (Ar-NH), 1677.79 cm⁻¹ (C=N), 1530.6 cm⁻¹ (C=C), 1089.68 cm⁻¹ (-C-O-C-), 1372.45 cm⁻¹ (N=O), 1523.12 asym cm⁻¹ (N=O), 3117.5 cm⁻¹ (Ar-CH), ¹H-NMR δ (ppm): 6.83-8.42 (m, 8H, Ar-CH), 5.35 (s, 2H, -CH₂), 2.07 (s, 1H, -CH₃), 8.24 (s, 1H, -NH), Mass (m/e value) % relative abundance: 399.08 (M⁺) (5), 388.76 (13), 380.25 (8), 261.63 (8), 182.52 (5), 167.62 (17), 156.56 (19), 81.97 (B).

Compound AB8

N-(4-[[5-(2-hydroxy-3,5-dinitrophenyl)-1,3,4-oxadiazol-2-yl]methoxy]phenyl) acetamide. IR (KBr) ν (cm⁻¹): 3118.84 cm⁻¹ (Ar-NH), 1654.42 cm⁻¹ (C=N), 1541.89 cm⁻¹ (C=C), 1368.45 cm⁻¹ (N=O), 1528.45 asym. cm⁻¹ (N=O), 1090.01 cm⁻¹ (-C-O-C-), 3118.84 cm⁻¹ (Ar-CH), 3382.83 cm⁻¹ (Ar-OH), ¹H-NMR δ (ppm): 6.7-7.6 (s, 6H, Ar-CH), 2.11 (s, 1H, -CH₃), 8.00 (s, 1H, -NH), 5.12 (s, 1H, -CH₂), Mass (m/e value) % relative abundance: 415.07 (M) (11.1), 318.68 (16), 292.76 (7), 276.89 (20), 249.99 (8.2), 236.02 (28.1), 203.22 (76), 182.25 (8), 134.49 (32), 116.55 (B).

Experimental Pharmacology**Requirements**

Carboxymethyl cellulose sodium (CMC) was provided by institutional store. The standard drug 5-FU was purchased from local retail shop Apollo pharmacy, Jyothinagar, OMR road, Chennai. Female Swiss albino mice (20-30 g) were obtained from the central animal house of C. L. Baid Metha College of pharmacy, Jyothinagar, OMR, Chennai and they were maintained

under standard laboratory conditions throughout the study. The study protocol was approved by the

Institutional Animal Ethics Committee (IAEC) reference number: IAEC/XXIX/10/2017.

Table 3A: Molecular properties of synthesized compounds

Compound s codes	Polarisability				Conformation (K.cal/mol)			
	Molecular	a (XX)	a (YY)	a (ZZ)	Conf.1	Conf.2	Conf.3	Conf.4
AB1	35.00	19.00	30.00	55.2	55.94	56.59	58.16	58.35
AB2	37.89	21.96	33.05	58.66	59.01	69.11	60.66	60.72
AB3	33.54	18.46	29.65	52.52	55.29	55.42	55.85	38.13
AB4	36.32	51.74	32.75	24.47	57.31	57.62	58.25	59.42
AB5	38.96	56.70	35.70	24.49	64.29	64.51	65.36	66.64
AB6	35.90	32.78	20.88	84.03	62.01	62.63	63.65	63.79
AB7	38.32	35.03	23.00	56.14	69.64	69.92	70.56	71.87
AB8	38.71	36.06	25.11	54.95	73.62	73.94	75.98	76.27

Conf: Conformer

Table 3B: Molecular geometry of synthesized compounds

Compounds codes	Geometry								
	DE	MMFF94	Min.pa	Max. pa	Min.pr	Max.pr	Vdw.vol	Lp.max.a	Lp.min. a
AB1	57.23	99.59	40.44	103.65	4.17	10.08	280.38	5.55	19.92
AB2	59.01	98.71	35.35	109.22	4.46	10.74	297.21	6.67	21.4
AB3	55.42	105.21	34.19	102.79	4.03	10.48	274.33	5.69	20.77
AB4	57.31	110.43	34.34	105.77	4.17	10.00	287.82	5.70	19.91
AB5	64.51	148.45	34.38	112.98	4.18	10.79	310.7	5.75	21.57
AB6	62.53	156.94	37.72	107.67	4.62	10.66	292.24	7.19	21.16
AB7	69.64	172.16	41.05	115.79	4.94	10.61	315.35	6.14	21.12
AB8	73.62	186.39	42.39	118.30	4.97	10.58	324.00	6.18	21.10

DE: Dreiding energy (kcal/mol); MMFF94 energy (kcal/mol); Min. Pa: Minimal projection area; Max.pa: Maximal projection area; Min.pr: Minimal projection radius; Max.pr: Maximal projection radius; LP.max.a: Length perpendicular to the max area; Lp. mina: Length perpendicular to the min; Van der Waals volume

Table 3C: Molecular properties of synthesized compounds

Compounds codes	PSA(2D)	Vdw.SA(3D)	H-bonding				Refractivity
			Ds	Dc	As	Ac	
AB1	103.27	445.73	03	02	06	05	102.05
AB2	77.25	461.95	01	01	05	04	106.96
AB3	77.25	437.77	01	01	05	04	97.57
AB4	77.25	449.66	01	01	05	04	104.97
AB5	123.07	489.50	01	01	09	06	112.30
AB6	123.07	470.58	01	01	09	06	104.68
AB7	168.89	510.43	01	01	13	08	112.00
AB8	189.12	519.03	02	02	14	09	113.98

PSA(2D): Polar surface area; Vdw.SA(3D): Van der Waals surface area; Ds: Donor sites; Dc: Donor count; As: Acceptor sites; Ac: Acceptor count.

Determination of median lethal doses (LD₅₀)

In the present study acute oral toxicity of the synthesized compounds were performed by acute toxic class method according to OECD guideline-423. [9]

Cell culturing

HT 29 human colorectal cancer cells were purchased from Amala Cancer Research Centre, Thrissur, and Kerala.

Study design [10]

An investigational study was designed to evaluate the *in vivo* antitumor activity of synthesized compounds (AB1-AB8) on mouse tumour models. Study was carried out with HT 29 cell line induced malignant ascites on mouse models. The dose of synthesized compounds 100 mg/kg were chosen based on the results of a toxicity study done previously. The animals were divided into eleven groups (each group contain 6 mice) as follows:

A. Group I: Normal Control Group [only the vehicle (1 ml/kg/day of 1% CMC orally)]

B. Group II: T. Control (1% CMC orally + HT 29 = 2×10^6 *i.p.*)

C. Group III: Standard (HT 29 = 2×10^6 *i.p.* + 5-FU 25 mg/ml inj.)

D. Group IV-XI: AB1-AB8 (HT 29 = 2×10^6 *i.p.* + 100 mg/kg orally)

(Group IV: AB1, Group V: AB2, Group VI: AB3, Group VII: AB4, Group VIII: AB5, Group IX: AB6, Group X: AB7, Group XI: AB8)

Table 4: Designing of experiment [11]

Days	Activity was carried out	No. of mice / group
Day 1	Collection of 0.3 ml of blood sample	Group-I-XI
Day 2	Tumour cell injection, HT 29 = 2×10^6 <i>i.p.</i>	Group-II-XI
	Treatment of CMC	Group-II
Day 3-12	Treatment of std. drug 5-FU	Group-III
	Treatment of Synthesized compounds (AB1-AB8)	Group-IV -XI
Day 15	Collection of 0.3 ml of blood sample	Group-II-XI
Day 16-35 follow up	Observed till death/35 th day	Group-II-XI

Table 5A: The assessment of haematological parameters

Group	Treatment	Hb (g/dl) (MEAN \pm SEM)	RBCs ($1 \times 10^6/\text{mm}^3$) (MEAN \pm SEM)	WBCs ($1 \times 10^3/\text{mm}^3$) (MEAN \pm SEM)
I	N. Control	13.62 \pm 0.213	9.405 \pm 0.666	6.032 \pm 0.007
II	T. Control	7.275 \pm 0.259	6.19 \pm 0.096	8.455 \pm 0.348
III	5-FU***	12.7 \pm 0.204	9.353 \pm 0.183	6.17 \pm 0.001
IV	AB1	7.825 \pm 0.301	8.1 \pm 0.105	6.44 \pm 0.024
V	AB2**	10.35 \pm 0.246	8.55 \pm 0.0108	6.416 \pm 0.029
VI	AB3**	10.65 \pm 0.184	8.683 \pm 0.027	6.294 \pm 0.024
VII	AB4	8.05 \pm 0.225	8.365 \pm 0.022	6.725 \pm 0.195
VIII	AB5	7.625 \pm 0.239	8.45 \pm 0.167	6.957 \pm 0.295
IX	AB6***	11.88 \pm 0.946	8.843 \pm 0.028	6.173 \pm 0.019
X	AB7***	11.53 \pm 0.149	8.755 \pm 0.0202	6.284 \pm 0.032
XI	AB8***	12.43 \pm 0.179	8.955 \pm 0.055	6.168 \pm 0.007

Table 5B: The assessment of haematological parameters

Group	Treatment	Neutrophils (%) (MEAN \pm SEM)	Lymphocytes (%) (MEAN \pm SEM)	Platelets ($1 \times 10^5/\text{mm}^3$) (MEAN \pm SEM)
I	N. Control	14.1 \pm 0.294	13.33 \pm 0.125	4.499 \pm 0.0009
II	T. Control	86.24 \pm 0.745	7.856 \pm 0.021	12.85 \pm 0.248
III	5-FU***	14.47 \pm 0.211	12.14 \pm 0.016	4.574 \pm 0.006
IV	AB1	15.83 \pm 0.058	8.325 \pm 0.047	5.431 \pm 0.0326
V	AB2*	15.13 \pm 0.243	10.1 \pm 0.238	5.177 \pm 0.004
VI	AB3*	14.72 \pm 0.072	10.32 \pm 0.235	5.017 \pm 0.028
VII	AB4	15.07 \pm 0.125	8.52 \pm 0.033	5.253 \pm 0.003
VIII	AB5	15.24 \pm 0.099	8.538 \pm 0.209	5.367 \pm 0.005
IX	AB6**	14.78 \pm 0.346	11.25 \pm 0.159	4.943 \pm 0.004
X	AB7**	14.82 \pm 0.32	10.87 \pm 0.141	4.973 \pm 0.007
XI	AB8**	14.56 \pm 0.164	11.74 \pm 0.436	4.704 \pm 0.002

Table 6: Comparison PILS (%) in different treatment groups

Group	Treatment	MST(days) (MEAN \pm SEM)	PILS (%)
II	T. Control	4.4 \pm 0.04	----- \leq -----
III	5-FU***	8.725 \pm 0.025	97.72
IV	AB1	6.4 \pm 0.04	45.45
V	AB2*	7.05 \pm 0.064	59.09
VI	AB3*	7.45 \pm 0.028	68.18
VII	AB4	6.8 \pm 0.04	56.81
VIII	AB5	6.125 \pm 0.025	38.63
IX	AB6**	8.25 \pm 0.064	84.09
X	AB7**	7.75 \pm 0.028	77.27
XI	AB8***	8.475 \pm 0.047	90.90

$P < 0.001 = \text{***}$, highly significant. $P < 0.01 = \text{**}$, moderate significant $P < 0.05 = *$, significant. $P > 0.05 = \text{ns}$. Values are expressed as MEAN \pm SEM of 6 animals. The data were statistically analysed by ONE WAY ANOVA followed by Tukey Kramer multiple comparison test.

Determination of the percentage increase in life span (PILS): It is calculated from the mean survival time (MST) values. [12] The MST for each group was calculated as: MST (days) = Total number of days survived by all animals in the group/Number of animals in the group. For each group, Percent increase of lifespan (% ILS) was determined by the following formula: PILS (%) = [(MST of treated group/MST of control group) - 1] \times 100. The haematological parameters of all surviving animals such as haemoglobin, RBC, WBC, neutrophils, lymphocytes and platelets were assessed for all.

Apoptosis analysis

Cardinal morphological features of apoptotic cells are determined by Gimsa staining technique [13], H33342 staining technique, annexin V staining [14-16] by fluorescence microscopy.

RESULTS AND DISCUSSION

Assessment of haematological parameters

Effect on the haematological parameters

The present experimental data displayed that the mortality was less in all groups except in tumour control group. The haemoglobin and RBCs count were significantly lower in tumour control group compared to normal control group and significantly raise nearly to normal in all treatment groups when compared with control group. The WBC counts were significantly increased in tumour control and it came down to nearly normal range in all treatment groups. The neutrophils were increased and lymphocytes were decreased significantly in tumour control groups and significantly decreased neutrophils and increased lymphocytes in all treatment groups. The platelet count was significantly increased in tumour control (except Group-III to XI) group compared to normal group.

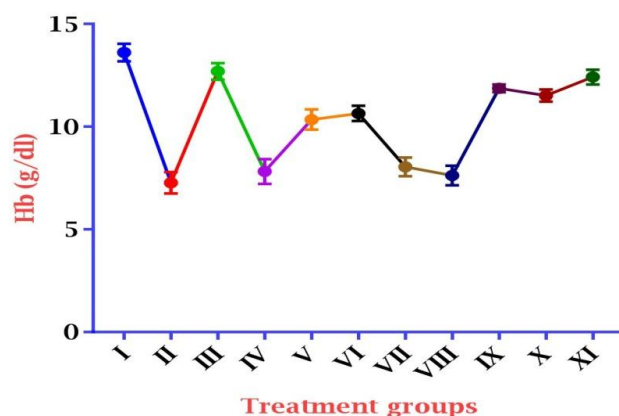


Fig. 1: Comparison of Hb level between different groups received HT29 tumour cells.

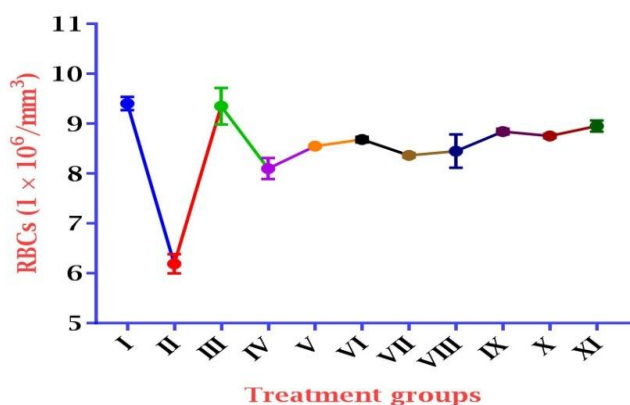


Fig. 2: Comparison of RBCs level between different groups received HT29 tumour cells.

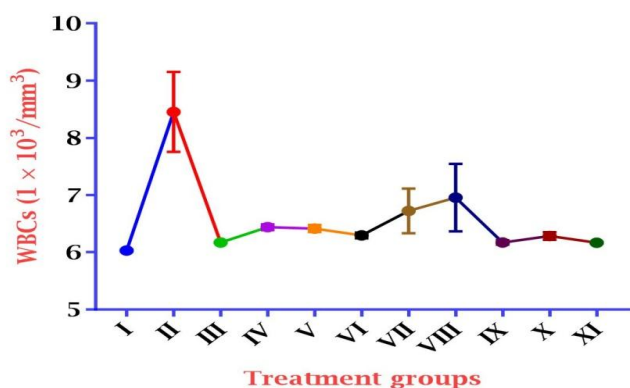


Fig. 3: Comparison of WBCs level between different groups received HT29 tumour cells.

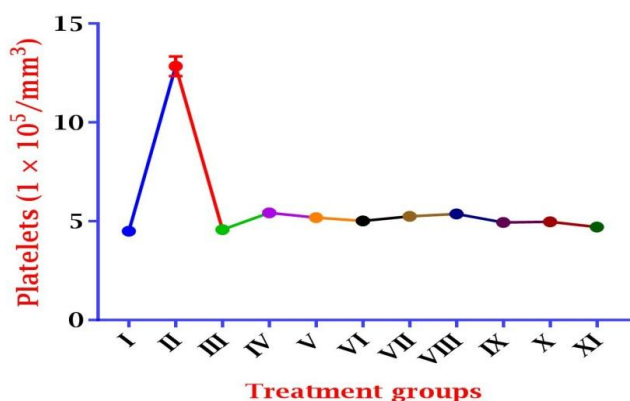


Fig. 4: Comparison of platelets count between different groups received HT29 tumour cells.

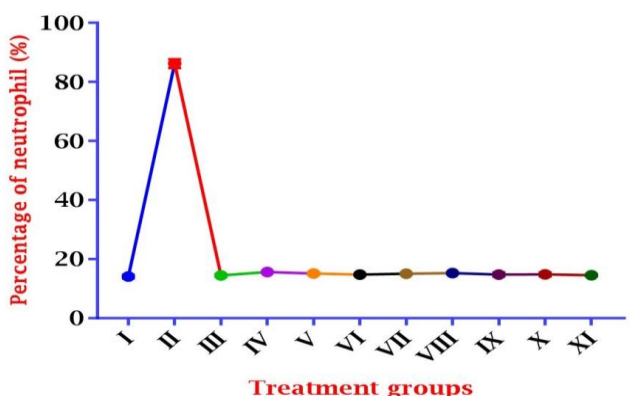


Fig. 5: Comparison of % of neutrophil between different groups received HT29 tumour cells.

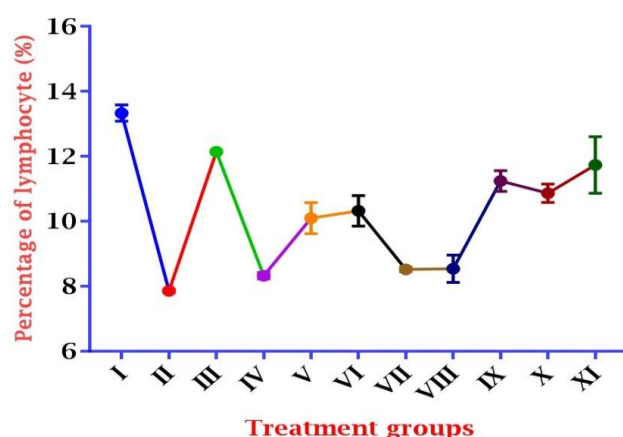


Fig. 6: Comparison of % of lymphocytes between different groups received HT29 tumour cells.

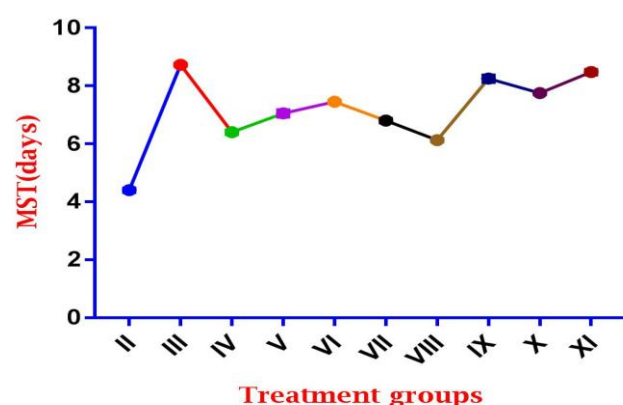


Fig. 7: Comparison of MST between different groups received HT29 tumour cells.

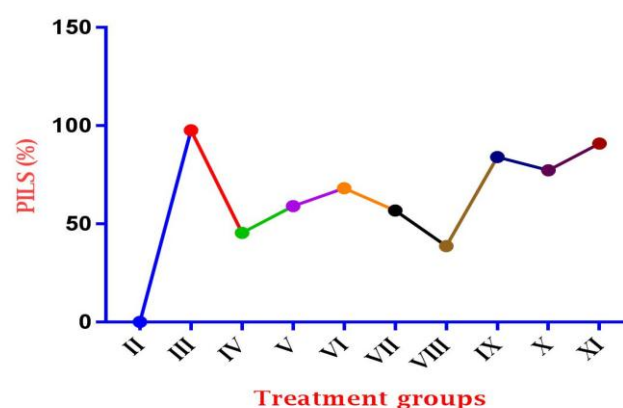


Fig. 8: Comparison of PILS between different groups received HT29 tumour cells.

Effect on the survival

All the synthesized compounds AB1-AB8 (100 mg/kg) significantly increased the PILS. While 5-FU increased the life span of 97.72%, and the PILS of synthesized compounds were found to be 45.45%, 59.09%, 68.18%, 56.81%, 38.63%, 84.09%, 77.27% and 90.90%. So the Synthesized compounds AB1-AB8 at the dose of 100 mg/kg significantly improved the overall survival of all treated animals and 5-FU was not significantly differed from each other in improving the overall survival of HT-29 cells.

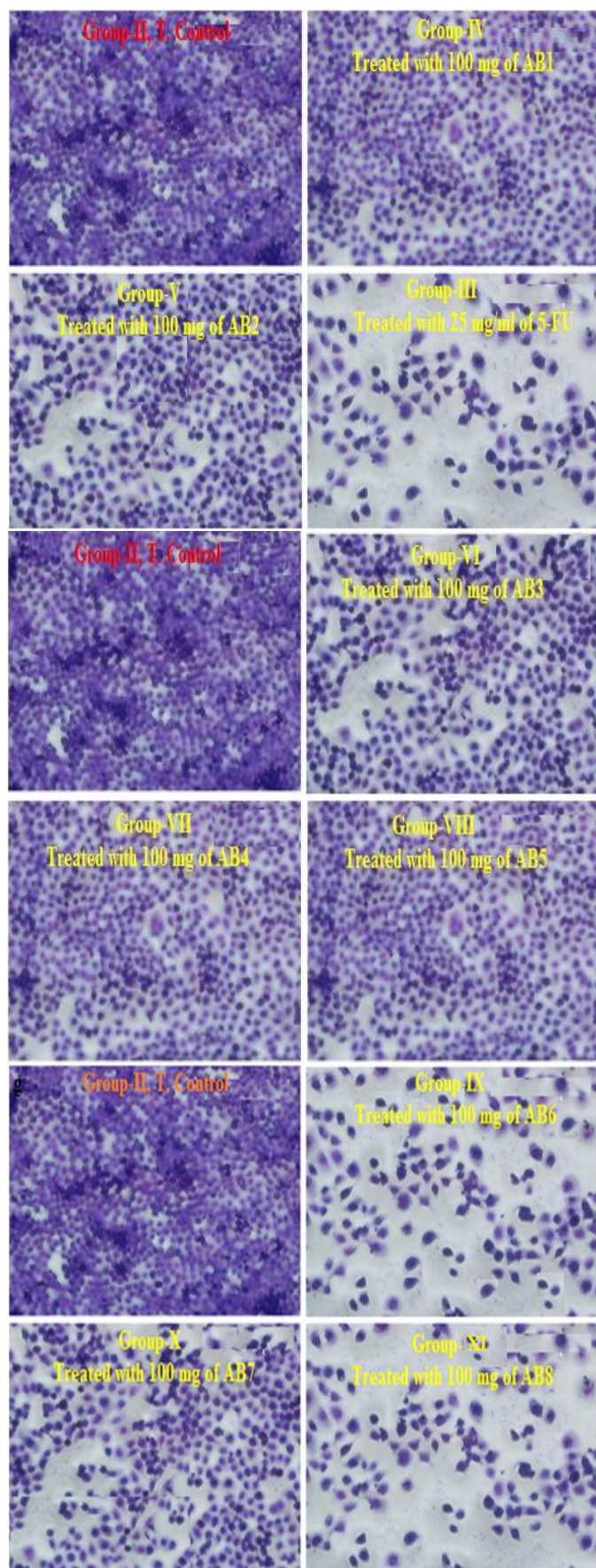


Fig. 9: Morphological changes observation of HT-29 cells by Giemsa staining (X200): Group-II: Tumour control; Group-III: Treated with standard drug and Group-IV-XI: Treated with 100 mg of synthesized compounds AB1-AB8. The cell morphology was observed and photographed under inverted phase-contrast microscope after Giemsa staining.

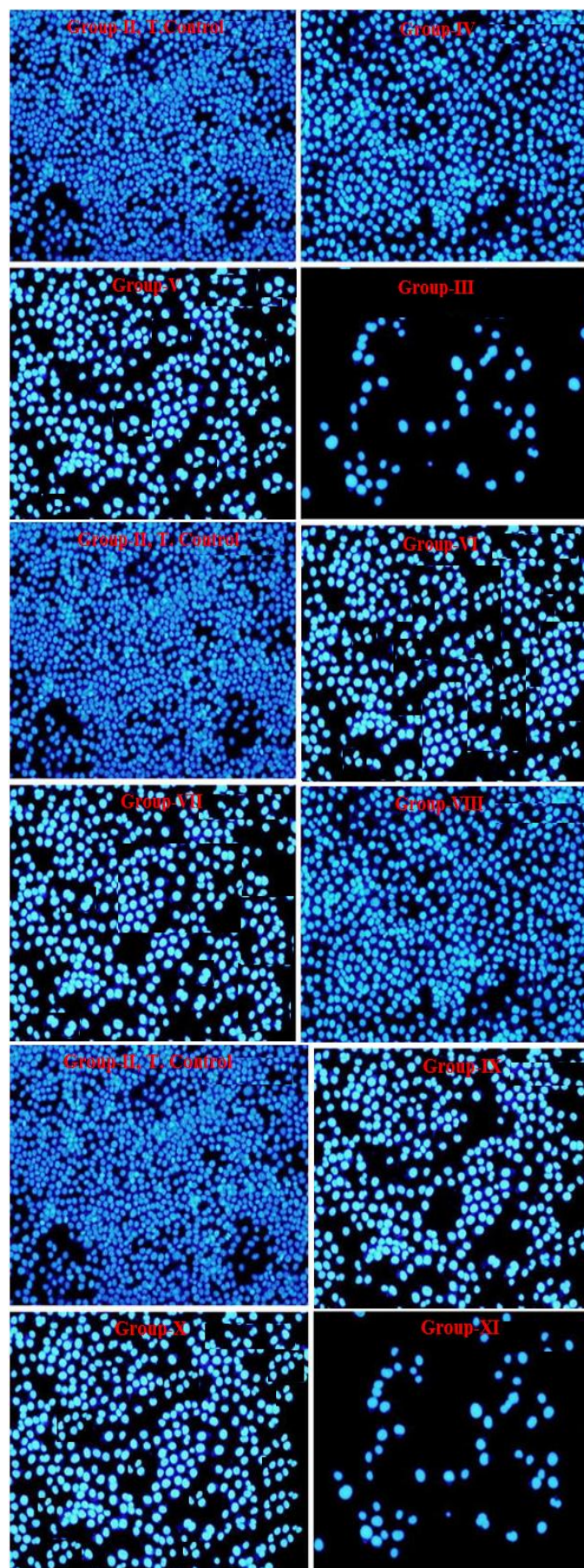


Fig. 10: Morphological changes observation of HT-29 cells by H33342 staining (X200): Group-II: T. Control; Group-III: Treated with standard drug 5-FU and Group-IV to Group-XI: Animals are treated with synthesized compounds (AB1-AB8).

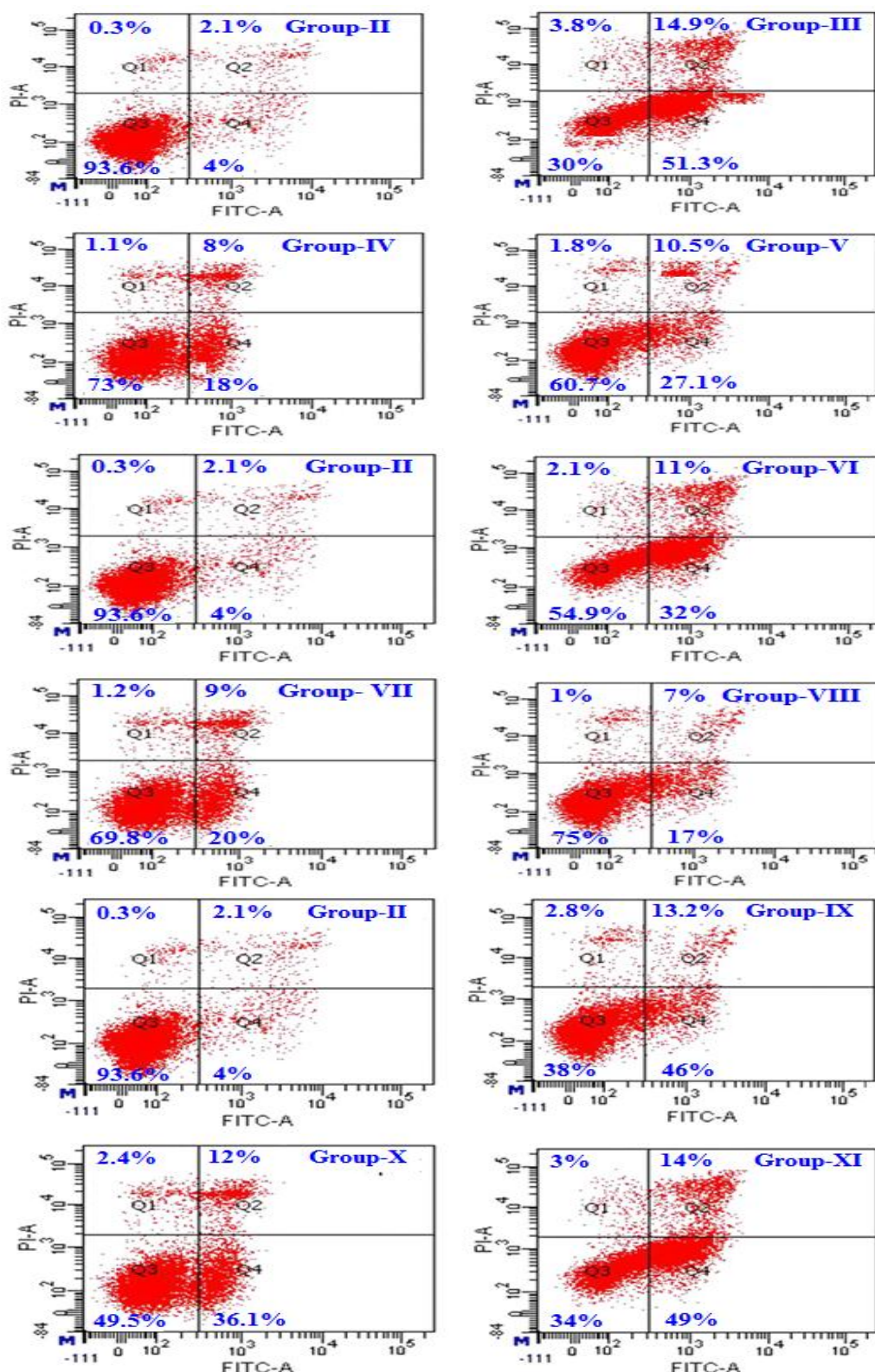


Fig. 11: Apoptosis analysis by FCM using AnnexinV-FITC/PI staining on the HT-29 cells treated by synthesized compounds; Group-II: Tumour control without compound and Group-III: Treated with standard drug 5-FU and Group-IV-XI: Treated with 100 mg of synthesized compounds: AB1-AB8.

Apoptosis analysis

Giemsa staining

To confirm whether the apoptotic morphological changes could be associated with the synthesized compounds, HT-29 cells were stained with Giemsa. The morphology changes were observed and photographed under inverted phase-contrast microscope at a

magnification of 200X. With the treatment of tests compounds, the process of cell loss, nuclei lysis, chromatin condensation and cytoplasmic shrinkage were aggravated.

H33342 staining on HT-29 cells

In the group-II i, e tumour control group, HT 29 cell nuclei displayed a normal and complete blue

appearance. By contrast, in HT 29 cells received groups i, e group-IV-XI which were treated with standard drug 5-FU (group-III), synthesized compounds (AB1-AB8), the cells displayed enhanced fragmentation or pyknosis of the nuclei, which were typical changes associated with cellular apoptosis. When compared with standard drug 5-FU, nuclear pyknosis and fragmentation in HT 29 cells were significantly increased by treatment with compounds AB8 < AB6 < AB7 < AB3 < AB2 and AB4 among the eight synthesized compounds.

Annexin V-FITC/propidium iodide (PI) assay

The effects of synthesized compounds on apoptosis in HT-29 cells were further determined by flow cytometric analysis. Cells were stained with both annexin V-FITC and PI. The flow cytometry observed four quadrant images: the Q1 area represented necrotic cells, the Q2 area represented late apoptotic cells, the Q3 area represented intact cells and the Q4 area represented the early apoptotic cells. The apoptosis ratios of different treatment groups were found as followed: Group-IV=26%; Group-V=37.6%; Group-VI=43%; Group-VII=29%; Group-VIII=24.1%; Group-IX=59.2%; Group-X=48.2%; and XI=63% respectively, while that of the Group-II (T. control) was 6.1%. When compared with Group-III (standard drug 5-FU: 66.2%), it was indicated that compound AB8 > AB6 > AB7 > AB3 were able to significantly induce HT-29 cells apoptosis.

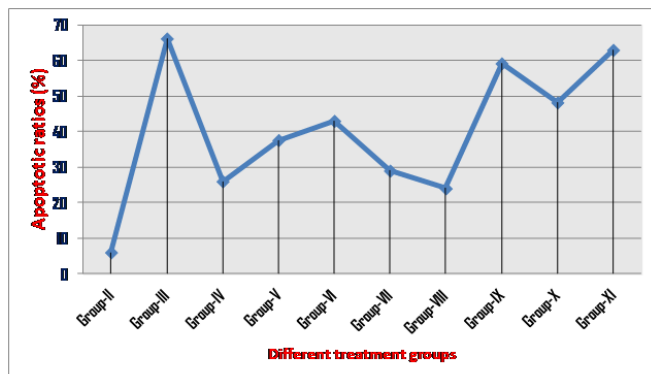


Fig. 12: Percentage of HT 29 cells death in different treatment groups.

In vivo experimental data showed that all the synthesized compounds possessed a mild to good antitumor activity against human colon cancer cell line HT29. Here it was found that among the all eight synthesized compounds the following compounds executed excellent antitumor activity against HT 29 cell line: AB2, AB3, AB6, AB7 and AB8 and apoptosis of HT 29 cell lines caused by synthesized compounds was evaluated by Giemsa staining and H33342 staining and

cells death of HT 29 was further confirmed by Annexin V-FITC/propidium iodide (PI) assay.

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