

Pitfalls in Diagnostic Myopathology of Sporadic Inclusion Body Myositis (SIBM)

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ABSTRACT

Sporadic inclusion body myositis (SIBM) is the most common idiopathic inflammatory myopathy in Caucasians over the age of 50 years. The prevalence of SIBM in the Asian population was initially thought to be very low, although the recent study showed that the prevalence of SIBM in Japan is in fact similar to the prevalence in Australia and the USA. SIBM is a refractory myositis with associated myodegenerative features mimicking the neuropathology of Alzheimer's disease. The diagnosis of definite SIBM requires the typical clinical features and the presence of an autoaggressive inflammatory reaction, rimmed vacuoles, and congophilic deposits or tubulofilamentous inclusions. About one-fourth of patients with the typical clinical features of SIBM did not fulfill the pathological criteria of the definite SIBM. These canonical biopsy features may be absent in the early stage of the disease. Here we review the typical findings and the diagnostic pitfalls of the muscle biopsy in SIBM.

Keywords: Amyloid- β , PM/IBM, PM-Mito, SIBM, sporadic inclusion body myositis

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Immune-mediated myopathy refers to a clinically and pathologically heterogeneous group of myopathies due to the dysimmune process. Immune-mediated myopathies are divided into 2 groups: non-inflammatory (necrotizing) and inflammatory myopathies. Inflammatory myopathies are characterized by the biopsy findings of inflammatory changes and necrotic and regenerating fibers. Inclusion body myositis (IBM), polymyositis (PM) and dermatomyositis (DM) are 3 major categories of idiopathic inflammatory myopathy.¹ These inflammatory myopathies are clinically and pathologically distinct (Table 1). Idiopathic inflammatory myopathy sometimes occurs in patients with other well-characterized connective tissue disease forming overlap syndromes or in patients suffering from viral infections especially HIV. Despite the clinical differences among various types of inflammatory myopathies and the recent knowledge on myositis-related antibodies, muscle biopsy remains the essential diagnostic procedure. Some muscular dystrophies or myopathies may

have biopsy findings mimicking inflammatory myopathy. A history of affected family members and distribution of muscle weakness should guide the clinicians to reach a correct diagnosis (Table 2).²⁻¹¹

IBM is a unique inflammatory myopathy with myodegenerative features mimicking the neuropathology of Alzheimer's disease. It is the most common idiopathic inflammatory myopathy in Caucasians over the age of 50 years and accounts for nearly one-third of all inflammatory myopathies.¹² The prevalence of IBM in the Asian population including Thailand and other non-Caucasian ethnic groups has not been reported, but it is considered to be uncommon.¹³ These racial and ethnic variations were thought to be due to the low frequency of the IBM susceptible alleles, HLA-DRB1*0301 (DR3) in the non-Caucasian population.¹³ However, the recent preliminary study from Japan showed that the prevalence of IBM in Japan in 2005 was 11.8 per million which is similar to the prevalence of IBM in the USA and Australia.^{13,14} In Japan, there was a 4.7-times increase in the prevalence over a 10-year period (1995-2005).¹⁴

SIBM is a slowly progressive, refractory inflammatory myopathy. Early atrophy and weakness of quadriceps (Fig 1A) and finger flexors are the typical clinical features of IBM. The canonical biopsy findings include autoaggressive inflammatory reaction, rimmed vacuoles, and congophilic

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TABLE 1. Idiopathic inflammatory myopathies.

| Characteristics | Sporadic inclusion body myositis | Polymyositis | Dermatomyositis |
|---------------------------------------|--|--|--|
| Onset | Late adulthood (> 50 years) | Adulthood | Childhood, juvenile or adulthood |
| Clinical presentations | Symmetrical or asymmetrical proximal and distal weakness with predominant quadriceps and finger flexors involvement; dysphagia | Symmetrical limb-girdle weakness; dysphagia | Symmetrical limb-girdle weakness; dysphagia |
| Serum CK level | - | Myocarditis; interstitial lung disease | Rash; myocarditis; interstitial lung disease |
| Electrodiagnostic study | Normal to mildly elevated | Moderately to markedly elevated | Normal to markedly elevated |
| Response to immunosuppressive therapy | Variable but mixed myopathic and neurogenic MUPs with fibrillations are typical. | Myopathic MUPs with fibrillations | Myopathic MUPs with fibrillations |
| Associated conditions | None or minimal | Yes | Yes |
| | Malignancy | Yes | Yes (adult-onset) |
| | Others | Connective tissue diseases | Connective tissue diseases or vasculitides |
| Light microscopic pathology | Site of inflammation | Predominant endomyrial exudates | Predominant perivascular perimyrial exudates |
| | Autoaggressive exudates | Common | Uncommon |
| | Lymphocyte subtypes | CD8-positive T cells | CD4-positive and B cells |
| | Perifascicular atrophy and other changes | - | + |
| | COX-negative fibers | - | + |
| | Vacuolated fibers | - | + |
| | (scattered fibers) | - | + |
| | (rimmed vacuoles in scatted fibers) | - | + |
| | (non-rimmed vacuoles in rare perifascicular fibers) | - | + |
| Immunohistochemical analysis | Congophilic inclusions | - | - |
| | MHC class I | + | + |
| | QB-crystallin | + | + |
| | (scattered fibers) | + | + |
| | (scattered fibers) | + | + |
| | Normal | Normal | Capillary depletion and C5b-9 complex deposits in the remaining capillaries |
| Electron microscopic analysis | Tubulofilamentous inclusions (16-20 nm) in myonuclei and sarcoplasm | - | - |
| | Tubuloreticular inclusions in capillary endothelium | - | + |
| Myopathies with similar pathology | Hereditary inclusion body myopathy and myofibrillar myopathy | Anti-synthetase syndrome, polymyositis with mitochondrial pathology and PM/IBM | Anti-synthetase syndrome, anti-SRP myopathy and necrotizing myopathy with pipsesem capillaries |

CK, creatine kinase; COX, cytochrome C oxidase; IBM, inclusion body myositis; MUP, motor unit potential; PM, polymyositis; SRP, signal recognition particle

TABLE 2. Myopathy or muscular dystrophy with pathologic features resembling myositis.

| Diagnosis | Mode of inheritance | Gene | Onset | Phenotypes |
|---|---------------------|--|---|---|
| 1. Dystrophinopathies | XR | DMD | Childhood to adulthood onset | Duchenne or Becker muscular dystrophy, manifesting carrier, muscle cramps or X-linked dilated cardiomyopathy |
| 2. Merosinopathy | AR | LAMA2 | Neonatal or infantile onset | Congenital muscular dystrophy (MDC1A) |
| 3. Alpha-dystroglycanopathies | AR | FKTN, FKRP, POMT1, POMT2, POMGnT1, and LARGE | Neonatal to early adulthood onset | Congenital muscular dystrophy with neuronal migration defects or LGMD2 |
| 4. Calpainopathy* | AR | CAPN3 | Childhood or early adulthood onset | LGMD2A or presymptomatic hyperCKemia |
| 5. Dystferlinopathy | AR | DYSF | Early adulthood > infantile onset | Congenital muscular dystrophy, distal myopathy with anterior tibial onset, LGMD2B, Miyoshi myopathy, presymptomatic hyperCKemia or rigid spine syndrome |
| 6. Hereditary inclusion body myopathy | AR | MYH2 | Neonatal to early adulthood onset | Congenital transient joint contracture, childhood onset ophthalmoparesis and adult-onset limb-girdle or scapuloperoneal weakness |
| | AR | GENE** | Early adulthood onset | Distal myopathy with anterior tibial onset (distal myopathy with rimmed vacuoles or DMRY) |
| | AD | VCP | Early adulthood onset | Limb-girdle, scapuloperoneal or axial weakness associated with Paget disease of bone and frontotemporal dementia (inclusion body myopathy with Paget disease of bone and frontotemporal dementia or IBMPFD) |
| 7. Facioscapulohumeral muscular dystrophy | AD | D4Z4 contraction or hypomethylation | Early adulthood > childhood > infantile onset | Axial myopathy, camptocormia or scapuloperoneal myopathy with or without facial involvement. |
| 8. Myofibrillar myopathies | AD | BAG3 | Childhood onset | Congenital muscular dystrophy with cardiomyopathy, neuropathy and rigid spine syndrome |
| | AR | PLEC1 | Childhood onset | Epidermolysis bullosa simplex with congenital myasthenic syndrome, congenital muscular dystrophy, LGMD or pyloric atresia or isolated LGMD, |
| | X-linked | FHL1 | Childhood onset | EDMD, reducing body myopathy, rigid spine syndrome, and scapuloperoneal myopathy |
| | AD>AR | CRYAB, DES, FLNC, MYOT, and ZASP | Childhood to late adulthood onset | Distal myopathy or LGMD with associated cardiomyopathy, neuropathy or cataracts (CRYAB) |

AD, autosomal dominant; AR, autosomal recessive; EDMD, Emery-Dreifuss muscular dystrophy; LGMD, limb-girdle muscular dystrophy; MDC, congenital muscular dystrophy; *, biopsy resembling eosinophilic myositis; **, biopsy showing inflammatory changes in some patients

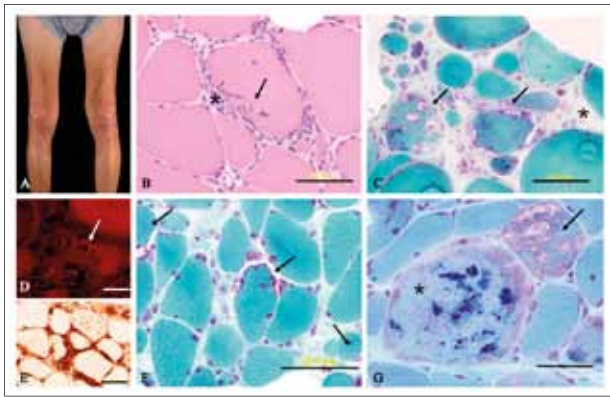


Fig 1. Clinical and pathologic features of sporadic inclusion body myositis and its mimickers.

(A-E) Sporadic inclusion body myositis (SIBM). (A) Photograph of definite SIBM patient showing atrophy of quadriceps. (B) H&E stained section revealed a marked variation in fiber size and an inflammatory exudate concentrated at the endomyosial site (asterisk), where it invaded non-necrotic muscle fibers (arrow). (C) Trichromatically stained section showed rimmed vacuoles (arrow) and endomyosial fibrosis (asterisk). (D) Congo-red stained section viewed under rhodamine optics displayed congophilic deposits in vacuolated fiber (arrow). (E) MHC-I immunoreactivity is present at the sarcolemma of non-necrotic muscle fibers. (F) Inclusion body myopathy with Paget disease of bone and frontotemporal dementia (IBMPFD). Trichromatically stained section revealed rimmed vacuoles (arrow) which were identical to those seen in SIBM. (G) Myofibrillar myopathy. Trichromatically stained section displayed a fiber containing rimmed vacuoles (arrow) and another fiber harboring amorphous hyaline material and multiple small cytoplasmic bodies (asterisk). Bar, 50 μ m

deposits or 16- to 20-nm tubulofilamentous inclusions.¹² Griggs and colleague defined definite IBM patients as having all canonical biopsy findings regardless of the clinical presentations (Table 3).¹⁵ These biopsy features were originally thought to be IBM-specific, but they are also present in other myopathies including hereditary inclusion body myopathy (HIBM) and myofibrillar myopathy (MFM). HIBM is a group of clinically and genetically heterogeneous, hereditary myopathies with the common pathological features which include rimmed vacuoles, congophilic deposits and tubulofilamentous inclusions. The onset of HIBM is typically earlier than IBM and the pattern of muscle weakness is also different between HIBM and IBM. An inflammatory exudate is normally absent in HIBM, but rare patients with the *GNE* variant of HIBM were reported with inflammatory changes mimicking IBM.^{16,17} To avoid the confusion, IBM is sometimes referred to as sporadic inclusion body myositis (SIBM), although in rare occasions SIBM occurs in the same family.^{13,18} For myofibrillar myopathy, the authors have described it in detail elsewhere.¹⁹

SIBM patients typically respond poorly to immunosuppressive therapy.¹² One of the possible explanations is that most SIBM patients were diagnosed late in their clinical course.^{12-14,20} In the Japanese SIBM cohort, only 38% of patients were initially diagnosed as SIBM. The mean time to diagnosis was 4.6 years after the onset.¹⁴ To achieve the correct diagnosis early, it is vital for clinicians and pathologists to be familiar with the typical clinicopathological features of SIBM. Diagnosis of SIBM can be

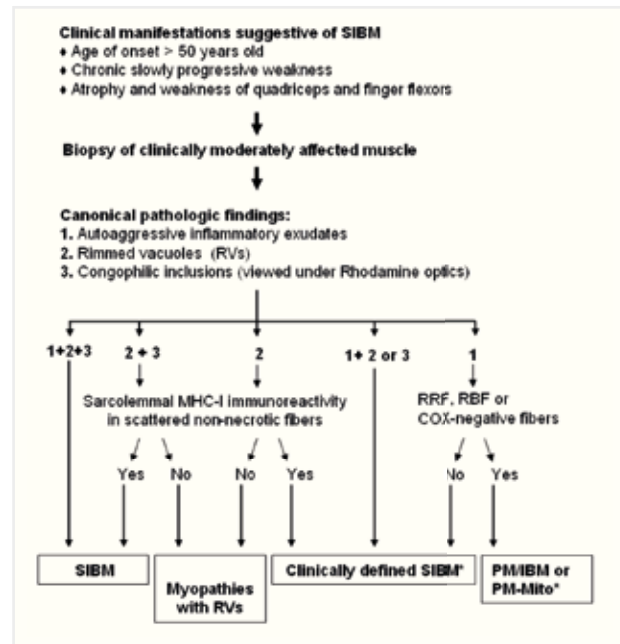


Fig 2. Diagnostic algorithm of the muscle biopsies in patients with clinical features of sporadic inclusion body myositis (SIBM). COX, cytochrome c oxidase; PM-Mito, polymyositis with mitochondrial pathology; RBF, ragged blue fibers; RRF, ragged red fibers; RV, rimmed vacuoles; SIBM, sporadic inclusion body myositis; * PM/IBM, PM-Mito and clinically defined SIBM may be the same entity and represent an early stage of SIBM (see text).

made easily when all characteristic pathological features are present, and difficulties arise when some of the typical changes are absent and it requires clinicopathological correlation, special immunohistochemical studies or even a repeat biopsy to reach the correct diagnosis. Herein, we review the typical findings and the diagnostic pitfalls of muscle biopsy in SIBM.

Pathology of Sporadic Inclusion Body Myositis (SIBM)

As mentioned earlier, SIBM is pathologically characterized by an autoaggressive inflammatory reaction, rimmed vacuoles, congophilic deposits and 16- to 20-nm tubulofilamentous, nuclear or sarcoplasmic inclusions.²¹ Each pathological feature individually is nonspecific, although a combination of all 4 features increases the specificity toward SIBM. It is important to keep in mind that the inflammatory and myopathic changes in inflammatory myopathy can be focal and often these may be absent at an early stage or in patients who receive immunosuppressive therapy.

I. Autoaggressive inflammatory reaction:

Invasion of non-necrotic muscle fibers (Fig 1B) by CD8+ T cells indicates autoaggressive inflammatory changes.²² The similar autoaggressive feature also presents in PM, but the number of invaded fibers per unit biopsy area was significantly lower than in SIBM.²¹ The distribution and degree of inflammatory changes in inflammatory myopathies vary from muscle to muscle, therefore careful selection of the biopsied muscle is the first step to avoid a false-negative biopsy. A clinically moderately affected muscle or a muscle showing moderate signal abnormality offers the proper choice for the biopsy. Open biopsy is

TABLE 3. Diagnostic criteria of sporadic inclusion body myositis.

| <i>Diagnostic criteria</i> | <i>Clinical and laboratory features</i> | <i>Pathological features</i> |
|--------------------------------------|---|---|
| Griggs et al, 1995 ¹⁵ | <p>Definite SIBM</p> <p>None of clinical and laboratory features are mandatory if biopsy features are diagnostic</p> <p>Possible SIBM</p> <p>Fulfill all clinical and laboratory characteristics of SIBM including</p> <p>(1) age of onset > 30 years, (2) duration of weakness > 6 months, (3) predominant wrist flexors, finger flexors or quadriceps weakness, (4) CK < 12 times normal, and (5) EMG findings consistent with inflammatory myopathy</p> | <p>Pathological features</p> <p>Fulfill all pathological characteristics of SIBM including (1) autoaggressive inflammatory exudates, (2) rimmed vacuoles and (3) either congophilic deposits or tubulofilamentous inclusions</p> <p>Only autoaggressive inflammatory exudate</p> |
| Needham et al, 2007 ²⁵ | <p>Definite SIBM</p> <p>Fulfill all clinical and laboratory characteristics of SIBM including</p> <p>(1) age of onset > 30 years, (2) duration of weakness > 6 months, (3) predominant finger flexors and quadriceps weakness, (4) normal or high CK, and (5) EMG showing myopathic or mixed MUPs but the distribution of weakness can be atypical if biopsy findings are diagnostic</p> | <p>Fulfill all pathological characteristics of SIBM including</p> <p>(1) autoaggressive inflammatory exudates, (2) rimmed vacuoles, (3) COX-negative fibers, (4) upregulation of MHC-I expression and (5) amyloid deposits or tubulofilamentous inclusions</p> |
| Benveniste et al, 2010 ²⁰ | <p>Probable SIBM</p> <p>Fulfill all clinical and laboratory characteristics of SIBM</p> | <p>Incomplete biopsy criteria</p> |
| | <p>Possible SIBM</p> <p>Pathologically defined SIBM</p> <p>Atypical pattern of weakness</p> <p>Consistent with SIBM</p> | <p>Incomplete biopsy criteria</p> <p>Fulfill all pathological characteristics of SIBM including (1) autoaggressive inflammatory exudate, (2) rimmed vacuoles and (3) amyloid deposits (Congo-red or crystal violet) or hyperphosphorylated tau (SMI-31 or p62) or TDP-43-positive aggregates or tubulofilamentous inclusions</p> <p>Autoaggressive inflammatory exudate or upregulation of MHC-I without congophilic deposits or SMI-31-positive aggregates or tubulofilamentous inclusions</p> |
| | <p>Clinically defined SIBM</p> <p>Fulfill all of clinical and laboratory characteristics of SIBM including</p> <p>(1) age of onset > 30 years, (2) duration of weakness > 6 months, (3) predominant finger flexors and quadriceps weakness, and (4) EMG findings consistent with inflammatory myopathy</p> <p>Possible SIBM</p> <p>Similar to clinically defined SIBM but weakness predominantly involves either finger flexors or quadriceps</p> | <p>Similar to Clinically defined SIBM</p> |

CK, creatine kinase; COX, cytochrome c oxidase; MUP, motor unit potential; SIBM, sporadic inclusion body myositis

preferable over needle biopsy because it provides a larger sample.

Major histocompatibility complex class-I (MHC-I) is normally expressed in intramuscular blood vessels, inflammatory cells and the sarcoplasm of necrotic or regenerating fibers.²³ Sarcolemmal immunoreactivity of otherwise morphologically healthy muscle fibers is a key feature of inflammatory myopathies (Fig 1E) (Table 2) and is present even when the inflammatory exudates are subtle or absent.^{24,25} However, this aberrant expression of MHC-I should be interpreted in the appropriate clinical context, because it also presents in other myopathies, muscular dystrophies and in some patients with statin-induced myopathy.²³⁻²⁶

II. Rimmed vacuoles and congophilic inclusions:

On H&E stained sections, rimmed vacuoles appear empty and mostly located at the subsarcolemmal areas. A vacuole is typically rimmed by hematoxyphilic material, hence the name rimmed vacuole. It is best seen on a trichromatically stained section (Fig 1C). Rimmed vacuoles were also reported in HIBM, MFM, oculopharyngeal muscular dystrophy (OPMD), reducing body myopathy, Marinesco-Sjogren syndrome, some variants of distal myopathies or limb-girdle muscular dystrophies and in rare patients with facioscapulohumeral muscular dystrophy (FSHD) or X-linked Emery-Dreifuss muscular dystrophy (EDMD).²⁷⁻³¹ In SIBM, only about 3% of muscle fibers harbor rimmed vacuoles.³²

SIBM differs from PM and DM because it features not only an inflammatory reaction, but also a myodegenerative process characterized by ectopic expression of several proteins in muscle fibers resembling the brain pathology of Alzheimer's diseases.³³ Abnormal protein aggregates are a composite of misfolded proteins (e.g. amyloid- and phosphorylated tau) and key molecules of unfolded protein response (UPR) and endoplasmic reticulum associated degradation (ERAD) (e.g. ubiquitin and proteasome).^{33,34} Amyloid aggregate is visualized by Congo-red (Fig 1D), thioflavin or crystal violet, but it is often that these special stains are not included in the routine set of biopsy analysis. Moreover, these amyloid deposits can easily be missed if examined only through polarizing filters. Amyloid deposits often are very small and typically present in only 60-80% of vacuolated fibers or about 0.75% of all myofibers.^{20,35} Viewing a Congo-red stained section under rhodamine optics greatly facilitates identification of amyloid deposits.³⁶ Amyloid aggregate itself is non-specific and has been reported in a variety of myopathies.^{11,28,36-41} The intramuscle fiber amyloid deposits observed in SIBM, HIBM, MFM, OPMD, and in rare patients with X-linked EDMD differ from the interstitial or vascular amyloid aggregates observed in dysferlinopathy and immune-mediated or hereditary systemic amyloidosis.^{11,28,36-41}

III. Nuclear or sarcoplasmic tubulofilamentous inclusions:

The characteristic ultrastructural feature of SIBM are 16- to 20-nm tubulofilamentous, nuclear or sarcoplasmic inclusions resembling paired helical filaments (PHFs) of neurofibrillary tangles seen in Alzheimer's disease brain.³³ Similar to the tangles, these inclusions contain phosphorylated tau. PHFs are present in 2-6% of myonuclei.³³ The similar tubulofilamentous inclusions were also reported in other myopathies, muscular dystrophies, neurogenic disorders and congenital myasthenic syndromes.^{23,42} PHFs

differ from amyloid- aggregates which are 6- to 10-nm filaments or flocculomembranous or amorphous material.³³

IV. Other pathological features of SIBM:

There are other pathological features of SIBM that are important, but not diagnostic. These encompass type 1 fiber preponderance (Liewluck T, unpublished data), with the pathological alterations suggestive of mitochondrial dysfunction (ragged-red, ragged-blue and cytochrome c oxidase-negative fibers), and small angular fibers over-reactive for nonspecific esterase indicative of recent denervation.²⁰ The myopathological finding of denervation atrophy is consistent with the electrodiagnostic feature of mixed neurogenic and myopathic motor unit potentials and with the axonal degeneration seen in the nerve biopsy of some SIBM patients.⁴³⁻⁴⁵

Recent developments in diagnostic myopathology of SIBM

SIBM was initially recognized as a specific subset of inflammatory myopathies based on the identification of rimmed vacuoles and tubulofilamentous inclusions in patients with steroid-unresponsive PM.⁴⁶ It was later realized that SIBM patients had a unique distribution of muscle weaknesses different from PM patients and the pathological changes were not as specific as initially thought.¹⁵ In clinical practice, about 1/4 of patients with characteristic clinical presentations of SIBM do not fulfill the pathological diagnostic criteria of definite SIBM based on Griggs criteria (Table 3).¹⁵ This may reflect sampling error or the currently recognized canonical pathologic alterations are late features of SIBM, and not the sensitive biomarkers of the disease. An attempt has been made to identify a new specific marker for SIBM that is more sensitive than rimmed vacuoles, amyloid deposits or tubulofilamentous inclusions in order to identify SIBM patients at an early stage of the disease.

Salajegheh et al., reported sarcoplasmic TDP-43 immunoreactivity as the most sensitive histologic marker of SIBM identified in nearly 25% of myofibers in definite SIBM cases.³² However, the amount of TDP-43 may vary from patients to patients as suggested by the lower sensitivity of TDP-43 immunoreactivity observed in other series of SIBM patients.^{20,47} SMI-31 and p62 antibodies detect phosphorylated tau and a shuttle protein in ERAD, respectively. Both SMI-31 and p62-positive inclusions occur in the similar proportion of fibers harboring rimmed vacuoles.^{20,35,47} Some experts consider electron microscopy as a nonessential study, because the presence of SMI-31 or p62-positive inclusions is equally sensitive and specific, and is more practical in routine laboratories.

Banwell et al., detected enhanced expression of α B-crystallin, a small heat shock protein, in approximately 10% of myofibers in definite SIBM patients.³⁵ Most of these α B-crystallin-positive fibers did not harbor other structural changes. The similar fibers were also present in PM, DM, MFMs and critical illness myopathy but less frequent. Although α B-crystallin is the most sensitive histologic marker for SIBM to date, its lack of specificity does not permit distinction between SIBM and PM. Currently, the list of aggregated proteins found in SIBM continues to expand and this may provide the future direction to identify the more sensitive and specific histologic marker for SIBM.³⁴

Clinically defined SIBM, PM/IBM and PM-Mito

As described earlier, about 25% of patients with typical clinical features of SIBM demonstrate only the

autoaggressive inflammatory changes without rimmed vacuoles or amyloid deposits.²¹ This group of patients was originally designated as “possible SIBM” based on Griggs criteria.¹⁵ Currently, it is known that some of possible SIBM patients behaved clinically like definite SIBM patients and a repeat biopsy a few years later revealed the canonical pathological findings of SIBM.²⁰ Therefore it is recommended to classify these possible SIBM patients as “clinically defined SIBM” or “PM/IBM”.^{20,21}

In 1997, Blume et al., reported a new entity of inflammatory myopathy, PM with mitochondrial pathology (PM-Mito).⁴⁸ Characteristic distribution of muscle weakness as in SIBM and PM/IBM were reported in 60% of PM-Mito patients.⁴⁹ All PM-Mito patients, either with or without predominant quadriceps or finger flexors weakness, shared the similar clinical course to SIBM and PM/IBM patients.⁴⁹ It is known that mitochondrial pathology is common in SIBM and PM/IBM,²¹ therefore PM-Mito may in fact be the same entity as PM/IBM and both reflect the early stage of definite SIBM. To date, all randomized clinical trials recruited only definite SIBM patients, hence patients with an advanced disease, and all drugs tested appear to be ineffective.²⁰ Several experts postulated that a clinical trial in early SIBM patients (e.g. PM/IBM or PM-Mito) may yield a better result.²⁰ Fig 2 showed the diagnostic algorithm of muscle biopsies in patients with clinical features of SIBM.

CONCLUSION

SIBM has unique clinical and pathological features. It is a refractory inflammatory myopathy with associated myodegenerative features. Some SIBM patients were not initially diagnosed as definite SIBM, because the biopsies lacked some of the canonical pathological findings. In order to diagnose SIBM at the early stage of the disease, it is important to find the specific histologic markers that have higher sensitivity than rimmed vacuoles, congophilic deposits and tubulofilamentous inclusions. Clinically defined SIBM, PM/IBM and PM-Mito may represent the early stage of definite SIBM and should be included in the study of any new histologic markers or novel therapeutic agents in the future.

REFERENCES

- Dalakas MC. Pathophysiology of inflammatory and autoimmune myopathies. *Presse Med.* 2011 Apr;40(4 Pt 2):e237-47.
- Spencer MJ, Tidball JG. Do immune cells promote the pathology of dystrophin-deficient myopathies? *Neuromuscul Disord.* 2001 Sep;11(6-7):556-64.
- Pegoraro E, Mancias P, Swerdlow SH, Raikow RB, Garcia C, Marks H, et al. Congenital muscular dystrophy with primary laminin alpha 2 (merosin) deficiency presenting as inflammatory myopathy. *Ann Neurol.* 1996 Nov;40(5):782-91.
- Godfrey C, Escolar D, Brockington M, Clement EM, Mein R, Jimenez-Mallebrera C, et al. Fukutin gene mutations in steroid-responsive limb girdle muscular dystrophy. *Ann Neurol.* 2006 Nov;60(5):603-10.
- Darin N, Kroksmark AK, Ahlander AC, Moslemi AR, Oldfors A, Tulinius M. Inflammation and response to steroid treatment in limb-girdle muscular dystrophy 2l. *Eur J Paediatr Neurol.* 2007 Nov;11(6):353-7.
- Biancheri R, Falace A, Tessa A, Pedemonte M, Scapolan S, Cassandrini D, et al. POMT2 gene mutation in limb-girdle muscular dystrophy with inflammatory changes. *Biochem Biophys Res Commun.* 2007 Nov 30;363(4):1033-7.

- Krahn M, Lopez de Munain A, Streichenberger N, Bernard R, Pecheux C, Testard H, et al. CAPN3 mutations in patients with idiopathic eosinophilic myositis. *Ann Neurol.* 2006 Jun;59(6):905-11.
- Amato AA. Adults with eosinophilic myositis and calpain-3 mutations. *Neurology.* 2008 Feb 26;70(9):730-1.
- Gallardo E, Rojas-Garcia R, de Luna N, Pou A, Brown RH, Jr, Illa I. Inflammation in dysferlin myopathy: immunohistochemical characterization of 13 patients. *Neurology.* 2001 Dec 11;57(11):2136-8.
- Arahata K, Ishihara T, Fukunaga H, Orimo S, Lee JH, Goto K, Nonaka I. Inflammatory response in facioscapulohumeral muscular dystrophy (FSHD): immunocytochemical and genetic analyses. *Muscle Nerve.* 1995;2:S56-66.
- Selcen D, Ohno K, Engel AG. Myofibrillar myopathy: clinical, morphological and genetic studies in 63 patients. *Brain.* 2004 Feb;127(Pt 2):439-51.
- Amato AA, Barohn RJ. Inclusion body myositis: old and new concepts. *J Neurol Neurosurg Psychiatry.* 2009 Nov;80(11):1186-93.
- Mastaglia FL. Sporadic inclusion body myositis: variability in prevalence and phenotype and influence of the MHC. *Acta Myol.* 2009 Oct;28(2):66-71.
- Suzuki N, Aoki M, Tateyama M, Izumi R, Warita H, Itoyama Y, et al. Prevalence of inclusion body myositis (IBM) in Japanese population. *Neuromuscul Disord.* 2010;20:631. Abstract.
- Griggs RC, Askanas V, DiMauro S, Engel A, Karpati G, Mendell JR, et al. Inclusion body myositis and myopathies. *Ann Neurol.* 1995 Nov;38(5):705-13.
- Yabe I, Higashi T, Kikuchi S, Sasaki H, Fukazawa T, Yoshida K, et al. GNE mutations causing distal myopathy with rimmed vacuoles with inflammation. *Neurology.* 2003 Aug 12;61(3):384-6.
- Krause S, Schlotter-Weigel B, Walter MC, Najmabadi H, Wiendl H, Muller-Hocker J, et al. A novel homozygous missense mutation in the GNE gene of a patient with quadriceps-sparing hereditary inclusion body myopathy associated with muscle inflammation. *Neuromuscul Disord.* 2003 Dec;13(10):830-4.
- Amato AA, Shebert RT. Inclusion body myositis in twins. *Neurology.* 1998 Aug;51(2):598-600.
- Liewluck T, Kintarak J, Sangruchi T, Selcen D, Kulankrakorn K. Myofibrillar myopathy with limb-girdle phenotype in a Thai patient. *J Med Assoc Thai.* 2009 Feb;92(2):290-5.
- Benveniste O, Hilton-Jones D. International Workshop on Inclusion Body Myositis held at the Institute of Myology, Paris, on 29 May 2009. *Neuromuscul Disord.* 2010 Jun;20(6):414-21.
- Chahin N, Engel AG. Correlation of muscle biopsy, clinical course, and outcome in PM and sporadic IBM. *Neurology.* 2008 Feb 5;70(6):418-24.
- Engel AG, Arahata K. Monoclonal antibody analysis of mononuclear cells in myopathies. II: Phenotypes of autoinvasive cells in polymyositis and inclusion body myositis. *Ann Neurol.* 1984 Aug;16(2):209-15.
- Hewer E, Goebel HH. Myopathology of non-infectious inflammatory myopathies - the current status. *Pathol Res Pract.* 2008;204(9):609-23.
- Emslie-Smith AM, Arahata K, Engel AG. Major histocompatibility complex class I antigen expression, immunolocalization of interferon subtypes, and T cell-mediated cytotoxicity in myopathies. *Hum Pathol.* 1989 Mar;20(3):224-31.
- Needham M, Mastaglia FL. Inclusion body myositis: current pathogenetic concepts and diagnostic and therapeutic approaches. *Lancet Neurol.* 2007 Jul;6(7):620-31.
- Needham M, Fabian V, Knezevic W, Panegyres P, Zilko P, Mastaglia FL. Progressive myopathy with up-regulation of MHC-I associated with statin therapy. *Neuromuscul Disord.* 2007 Feb;17(2):194-200.
- Goto Y, Komiyama A, Tanabe Y, Katafuchi Y, Ohtaki E, Nonaka I. Myopathy in Marinesco-Sjogren syndrome: an ultrastructural study. *Acta Neuropathol.* 1990;80(2):123-8.
- Fidzianska A, Rowinska-Marcinska K, Hausmanowa-Petrusewicz I. Coexistence of X-linked recessive Emery-Dreifuss muscular dystrophy with inclusion body myositis-like morphology. *Acta Neuropathol.* 2004 Mar;107(3):197-203.
- Neudecker S, Krasnianski M, Bahn E, Zierz S. Rimmed vacuoles in facioscapulohumeral muscular dystrophy: a unique ultrastructural feature. *Acta Neuropathol.* 2004 Sep;108(3):257-9.
- Liewluck T, Hayashi YK, Ohsawa M, Kurokawa R, Fujita M, Noguchi S, et al. Unfolded protein response and aggressive formation in hereditary reducing-body myopathy. *Muscle Nerve.* 2007 Mar;35(3):322-6.
- Schoser B. Physiology, pathophysiology and diagnostic significance of autophagic changes in skeletal muscle tissue—towards the enigma of rimmed and round vacuoles. *Clin Neuropathol.* 2009 Jan-Feb;28(1):59-70.
- Salajegheh M, Pinkus JL, Taylor JP, Amato AA, Nazareno R, Baloh RH, Greenberg SA. Sarcoplasmic redistribution of nuclear TDP-43 in inclusion body myositis. *Muscle Nerve.* 2009 Jul;40(1):19-31.
- Askanas V, Engel WK, Nogalska A. Inclusion body myositis: a degenerative muscle disease associated with intra-muscle fiber multi-protein aggregates, proteasome inhibition, endoplasmic reticulum stress and decreased lysosomal degradation. *Brain Pathol.* 2009 Jul;19(3):493-506.
- Askanas V, Engel WK. Sporadic inclusion-body myositis: Conformational multifactorial ageing-related degenerative muscle disease associated with proteasomal and lysosomal inhibition, endoplasmic reticulum stress, and accumulation of amyloid-42 oligomers and phosphorylated tau. *Presse Med.* 2011 Apr;40(4 Pt 2):e219-35.
- Banwell BL, Engel AG. AlphaB-crystallin immunolocalization yields new insights into inclusion body myositis. *Neurology.* 2000 Mar 14;54(5):1033-41.
- Askanas V, Engel WK, Alvarez RB. Enhanced detection of congo-red-positive amyloid deposits in muscle fibers of inclusion body myositis and brain of Alzheimer's disease using fluorescence technique. *Neurology.* 1993 Jun;43(6):1265-7.

37. Oldfors A. Hereditary myosin myopathies. *Neuromuscul Disord.* 2007 May; 17(5):355-67.
38. Leclerc A, Tome FM, Fardeau M. Ubiquitin and beta-amyloid-protein in inclusion body myositis (IBM), familial IBM-like disorder and oropharyngeal muscular dystrophy: an immunocytochemical study. *Neuromuscul Disord.* 1993 Jul;3(4):283-91.
39. Spuler S, Emslie-Smith A, Engel AG. Amyloid myopathy: an underdiagnosed entity. *Ann Neurol.* 1998 Jun;43(6):719-28.
40. Spuler S, Carl M, Zabojszcza J, Straub V, Bushby K, Moore SA, et al. Dysferlin-deficient muscular dystrophy features amyloidosis. *Ann Neurol.* 2008 Mar;63(3):323-8.
41. Kiuru-Enari S, Somer H, Seppalainen AM, Notkola IL, Haltia M. Neuromuscular pathology in hereditary gelsolin amyloidosis. *J Neuropathol Exp Neurol.* 2002 Jun;61(6):565-71.
42. Fidzińska A, Ryniewicz B, Shen XM, Engel AG. IBM-type inclusions in a patient with slow-channel syndrome caused by a mutation in the AChR epsilon subunit. *Neuromuscul Disord.* 2005 Nov;15(11):753-9.
43. Lotz BP, Engel AG, Nishino H, Stevens JC, Litchy WJ. Inclusion body myositis. Observations in 40 patients. *Brain.* 1989 Jun;112 (Pt 3):727-47.
44. Joy JL, Oh SJ, Baysal AI. Electrophysiological spectrum of inclusion body myositis. *Muscle Nerve.* 1990 Oct;13(10):949-51.
45. Lindberg C, Oldfors A, Hedstrom A. Inclusion body myositis: peripheral nerve involvement. Combined morphological and electrophysiological studies on peripheral nerves. *J Neurol Sci.* 1990 Nov;99(2-3):327-38.
46. Yunis EJ, Samaha FJ. Inclusion body myositis. *Lab Invest.* 1971 Sep;25(3):240-8.
47. D'Agostino C, Nogalska A, Engel WK, Askanas V. In sporadic inclusion body myositis muscle fibres TDP-43-positive inclusions are less frequent and robust than p62 inclusions, and are not associated with paired helical filaments. *Neuropathol Appl Neurobiol.* 2011 Apr;37(3):315-20.
48. Blume G, Pestronk A, Frank B, Johns DR. Polymyositis with cytochrome oxidase negative muscle fibres. Early quadriceps weakness and poor response to immunosuppressive therapy. *Brain.* 1997 Jan;120 (Pt 1):39-45.
49. Temiz P, Wehl CC, Pestronk A. Inflammatory myopathies with mitochondrial pathology and protein aggregates. *J Neurol Sci.* 2009 Mar 15; 278(1-2):25-9.