

Diagnostic difficulties in c-kit negative gastrointestinal stromal tumors: report of four cases

Andreea C. Iorgescu¹, Vlad Herlea^{1,2}, Elena Stoica Mustafa¹, Catalin Pechianu^{1,2}, Alexandru Procop¹, Maria Sajin^{3,4}, Irinel Popescu^{1,2}

Abstract: *Introduction: The gastrointestinal stromal tumors (GIST) are dominated by KIT and PDGFRA mutation. The immunohistochemical detection of CD117, a protein expressed by KIT gene, is essential for the diagnosis and those that are negative always represented a diagnostic challenge*

Case reports: In this article we present a series of 4 cases of CD117 negative GIST tumors, diagnosed and surgically resected in Fundeni Clinical Institute and an overview of the histogenesis, diagnostic problems and management of c-kit negative GIST. All patients were males and the tumors were located in the stomach and small bowel.

Conclusion: It is important for the pathologists to beware of the fact that a CD117 negative in the context of a typical morphological appearance does not exclude a GIST tumor and also the oncologist must be aware not to exclude the therapy with imatinib based on the negativity of CD117.

Keywords: GIST, gastrointestinal stromal tumors, c-kit negative, CD117, DOG1, immune-histochemistry

INTRODUCTION

GISTs are rare tumors accounting for less than 1% of all gastrointestinal tumors but they are the most common mesenchymal tumors of the GI tract.[1] The incidence in Europe and USA is 7-10 cases/1,000,000 [2] and in Korea is 16-22/1,000,000 [3]. Large population based studies from Iceland, Netherlands and Sweden found the incidence to be 11, 12.7 and 14.5 cases/1,000,000 [4,5,6]. However there are studies that explain this increased incidence by a better diagnosis using modern diagnosis criteria rather than a real increase [7].

The most common age of the diagnosis is between 50 to 60 years old (a mean age of 65) and a peak incidence between 70 and 79, before 20 years old being quite

rare ≈ 1% [7,8]

GIST affects both sexes equally [6] though some authors found them to be more common in men [7,9]. They arise most often in the stomach (50-60%) and small intestine (30-35%) but it can occur in any segment of the gastrointestinal tract only with a lower frequency.[10]

Gain of function mutation in KIT or PDGFRA oncogenes lead to ligand-independent kinase activation [11] which gives rise to the majority of GIST. Immunohistochemistry is useful for diagnosis and

Corresponding author: Vlad Herlea MD, PhD
herlea2002@yahoo.com

¹ Fundeni Clinical Institute, Bucharest, Romania

² Titu Maiorescu University - Faculty of Medicine, Bucharest, Romania

³ University Emergency Hospital, Bucharest, Romania

⁴ Carol Davila University of Medicine and Pharmacy, Bucharest, Romania

typically detects c-kit protein expression, but if the tumor is negative for c-kit, a final diagnosis can be challenging.

In this article we present a series of 4 cases of c-kit negative GIST tumors, diagnosed and surgically resected in Fundeni Clinical Institute and an overview of the histogenesis, diagnostic problems and management of c-kit negative GIST.

CASE REPORTS

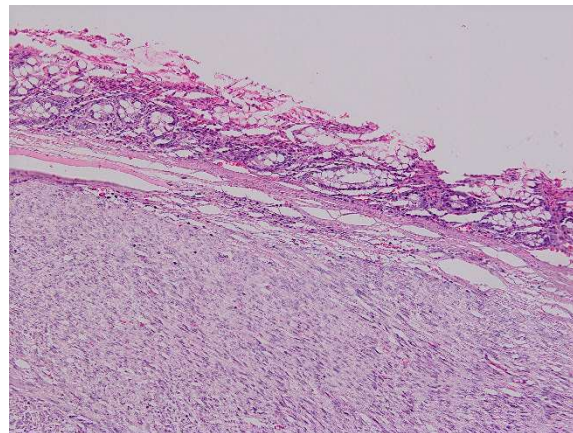
In the pathology archives of Fundeni Clinical Institute, between 2004 and 2017 we have traced four cases of c-kit negative GIST. We have examined available clinical data from the hospital database and also HE slides. Immunohistochemically investigations were performed with biotin-streptavidin method [12] in order to assess the phenotype and provide a definitive diagnosis.

Antibody suppliers were Novocastra, and Labvision/Thermo Fisher Scientific as previously reported [13] and the dilutions respected the manufacturer recommendation. The stratification risk according to Miettinen&Lasota (2006) was used [2]. For the mitotic rate we counted the number of mitosis on 50 high power fields (HPF). Tumor cellularity was divided in three categories: spindle, epithelioid and mixed.

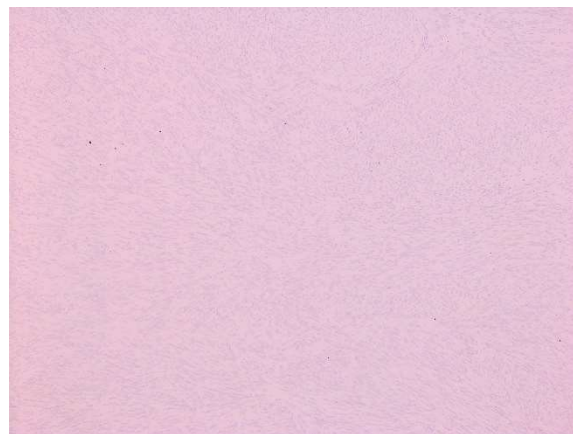
Case no. 1

A 47 years old male who presented with lower gastrointestinal bleeding, was investigated with capsule-endoscopy procedure and found with a tumor on the ileum, close to the ileocecal valve. A segmental enterectomy was performed. The surgical specimen was 12 cm long and presented centrally a well circumscribed white nodule of 4/3/4 cm, expanded in the submucosa and muscularis propria with mucosal ulceration. Histologic examination revealed a tumor consisting of spindle cells with a mitotic rate of 1-2/50 hpf. Immunohistochemical stains showed negativity for CD34, CD117, PDGFRA, Desmin, a weak positivity for SMA and a diffuse cytoplasmic positivity for DOG1. The prognostic group was 2, and 7 years later the patient is alive with no recurrent disease.

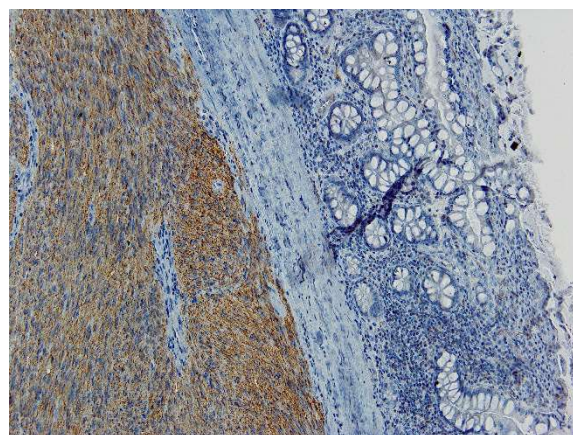
Figures case 1



HEx100



CD117x100



DOG1x100

Case no. 2

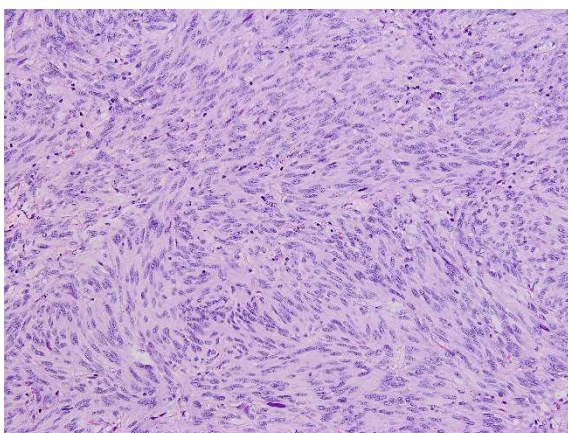
A 69 years old male presented with abdominal pain, nausea and vomiting for 12 hours before admission. The abdominal CT scan revealed a pelvic tumoral mass on the left paramedian side, developed behind the

abdominal wall that involved a small intestinal loop. A segmental enterectomy was performed. The surgical specimen was 30 cm long and presented a solid white tumor with irregular contour 8.5/3.5/2cm with predominantly intramural and subserosal development and focal mucosal ulceration.

Microscopic examination reveal a mixt morphology with spindle and epithelioid cells, with mild pleomorphism, intratumoral hemorrhagic foci and a mitotic rate of 2/50 HPF.

Immunohistochemical stains were negative for SMA, Desmin and CD117 and positive for DOG1. CD34 and PDGFRA were not done.

Figures case 2



HEX100



CD117x100



DOG1x100

On the bases of these results the tumor was diagnosed as a c-kit negative GIST with moderate risk (group 3a) and survival from the time of the diagnosis until death was 33 months.

Case no. 3

A 67 years old male was admitted to the hospital with melena, fatigue and weight loss. An esophagogastro-duodenoscopy was performed, revealing a 1.5 cm lesion in the gastric fornix.

The tumor biopsy showed a stromal gastric hemorrhagic proliferation associated with a chronic gastritis with Helicobacter Pylori infection.

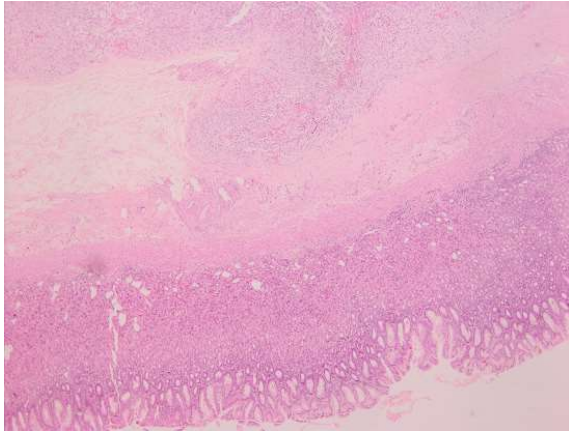
A local excision was carried out and on the surgical specimen was found a firm rubbery white intramural nodule with a diameter of 3/3/3 cm.

Microscopy revealed a tumor proliferation with spindle cell morphology, without mucosal involvement, with 1-2 mitotic figure/50HPF.

Immunohistochemistry showed negativity for CD117, Actin, Desmin and positivity for DOG1. CD34 and PDGFRA were not done.

The diagnosis of c-kit negative GIST with low risk (group 2) was established and the patient is alive with no recurrence.

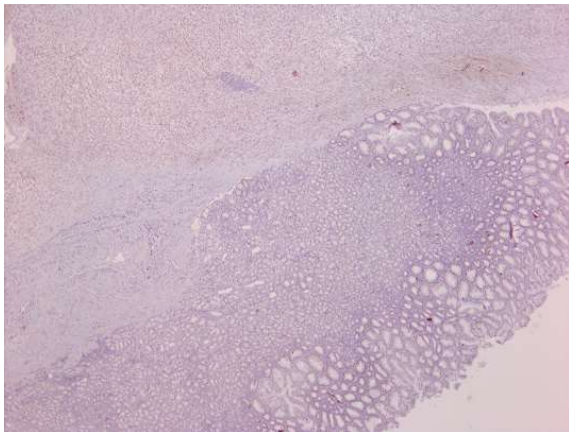
Figures case 3



HEx100



CD117x100



DOG1x100

Case no. 4

59 years old male with loss of appetite, abdominal pain, weight loss, is admitted to the hospital for investigations.

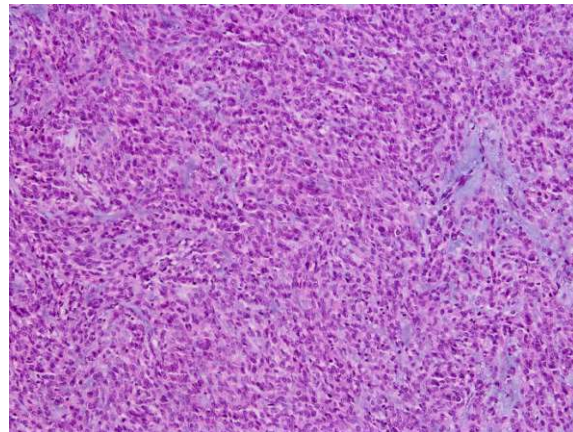
Esophagogastroduodenoscopy revealed an ulcerated

tumoral proliferation on the body of the stomach and an ultrasound discovered multiple localized proliferations in left and right hepatic lobes.

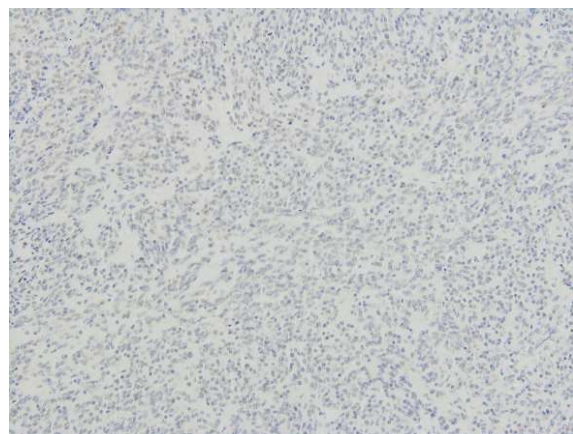
A total gastrectomy was performed and the surgical specimen showed a 12 cm white grey tumor with intramural development and cystic degeneration. On the microscopic examination it showed an epithelioid and spindle cell morphology with fascicles that intersect and intertwine at various angles, with more than 10 mitotic figures/50 hpf Immunohistochemistry showed negativity for CD117 and Desmin, and positivity for SMA and DOG1.

A diagnosis of GIST with high risk of recurrence (group 6b) was established and the patient survived time from the initial diagnosis was for 9 months.

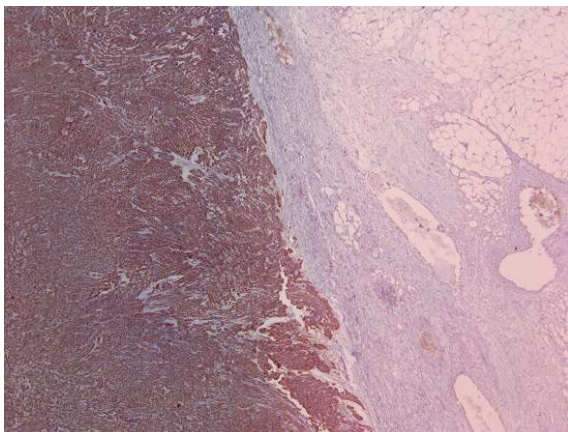
Figures case 4



HEx100



CD117x100



DOG1x100

DISCUSSION

Arising in the interstitial cell of Cajal, GIST tumors are an entity dominated by mutations in KIT receptor tyrosine kinase which accounts for almost 80% of all cases. The histopathological diagnosis of such tumors is built upon the microscopic morphology and immunohistochemically stains.

The detection of CD117, a protein expressed by KIT gene, is essential for the diagnosis and we can find it in approximately 95% of GIST [8, 14].

Table 1: Clinical-pathological characteristics of the four cases presented

| Case no. | Age | Location | Metastasis | Tumor diameter | Mitosis/HPF | Cell type | Group | Death | Survival months | Resection |
|----------|-----|----------|---------------------|----------------|-------------|-----------|-------|-------|-----------------|-----------|
| 1 | 47 | SI | no | 4.0 | <5 | fusiform | 2 | no | 84 | R0 |
| 2 | 69 | SI | peritoneum lung | 8.5 | <5 | mixt | 3a | yes | 33 | R1 |
| 3 | 67 | S | no | 3.0 | <5 | fusiform | 2 | no | 135 | R0 |
| 4 | 60 | S | liver peritoneum | 12.0 | >5 | mixt | 6b | yes | 9 | R1 |

Table 2

| | CD117 | DOG1 | Desmine | SMA |
|-----------|----------|----------|----------|----------|
| Clone | T595 | K9 | DE-R-11 | αsm-1 |
| Dilution | 1:40 | 1:100 | 1:100 | 1:50 |
| Case no.1 | negative | positive | negative | positive |
| Case no.2 | Negative | positive | negative | negative |
| Case no.3 | Negative | positive | negative | positive |
| Case no.4 | negative | positive | negative | positive |

However, there are 4-5% of GISTs that are negative for CD117 and in these cases immunomarkers like DOG1 (ANO1) are very useful for the confirmation of the diagnosis [9,15,16].

DOG1 shows higher prevalence of positivity in gastric epithelioid GISTs, which are often KIT negative.[17]

Depending of the antibody used, the positivity of DOG1 in KIT-negative GISTs varies: with clone DOG1.1, the positivity is 36% [18] and with clone K9, the positivity is 50–76%. [17,19]. We also used K9 clone in our cases, which we found very helpful.

Other immunohistochemical markers used, that can

guide the diagnosis are CD34 (with a much higher positivity in the stomach-80% then in the small intestine-35%) together with SMA Desmin, Vimentin. CD34 was done only in one of our cases, a small intestine GIST and it was negative so the only reliable marker was DOG1.

Most of the GISTs are reported in the stomach (50-60%), jejunum and ileum (30-35%), duodenum (5%), colorectal (4%), and rarely in the esophagus and appendix (<1%) [10, 20]. Primary tumors outside the GI tract have been reported in small numbers in omentum [21,22], mesenteries and retroperitoneum [23,24]. The location of c-kit negative GIST is more

frequently reported in the stomach [25, 26, 27] and less in the small intestine [28,29]. Two of our cases were located in the stomach but the other two located in the small intestine gave rise to diagnostic suspicion so we had to pay more attention at the differential diagnosis due to the rarity of the tumor site.

Location and size of the tumor influence the apparition of symptoms. Small tumors are usually silent and are being discovered incidentally during investigations or surgical procedures for other disease. However if a GIST becomes symptomatic, it usually cause non-specific symptoms like weight loss, fatigue, nausea, bowel obstruction or overt or occult gastrointestinal bleeding due to mucosal ulceration or tumor rupture[30,9]. Two of our patients presented with nonspecific symptoms like abdominal pain, nausea, fatigue, weight loss, and the other two with bleeding signs. Non-specific symptoms delays the diagnosis and we can see from Table 1 that those two cases have also the largest diameter (8.5 cm in the small intestine and 12 cm in the stomach).

The prognosis and predictive factors for survival of patients with C-kit negative GISTs are still unclear and difficult to assess because of insufficient data. The tumor diameter, metastatic disease, incomplete resection is associated in our cases with a shorter survival rate (Table1).

DOG1 especially clone K9 is an invaluable marker of the diagnosis but mutational status comes in hand in the treatment management. Complete resection and inhibitors of tyrosine kinase are considered for the treatment as in c-kit positive GIST [31].

However when we encounter a c-kit negative GIST we need to take into consideration:

- The antibody that we use, the clone and the laboratory technique
- The fixation methods of the surgical specimen may influence the reaction
- The CD117 negativity does not exclude a GIST

References:

1. Judson I, Demetri G. Advances in the treatment of gastrointestinal stromal tumors. *Ann Oncol* 2007; 18(Suppl 10): 20–4.

diagnosis and other markers like DOG1 are necessary to complete the immunohistochemical diagnostic panel; we also must think at GIST with other mutations like PDGFRA

- For a possible treatment with inhibitors of tyrosine kinase (imatinib) we must have in mind mutation analysis as well

- GIST wild type is a constantly changing concept, at first considered a KIT negative GIST, now is described as GIST with no identified gain of function mutation. Besides the KIT gene, GISTs may present mutations in another receptor tyrosine kinase - the PDGFRA which accounts for less than 10% of all GISTs [32].

According to most studies, activating mutations in KIT or PDGFRA are present in 85 - 90% and are mutually exclusive [33]. Other driver mutations studied, like BRAF, RAS, PIC3K, SDHA, NF1 need to have other genetic event in order to develop and progress [34,33, 35,36]

CONCLUSIONS

Kit negative GIST is a problematic entity which makes the diagnostic difficult.

In conclusion, we report four cases of CD117 negative, DOG1 positive GIST located in the stomach and small intestine, all being male patients.

It is important for the pathologists to beware of the fact that a CD117 negative in the context of a typical morphological appearance does not exclude a GIST tumor and also the oncologist must be aware not to exclude the therapy with imatinib based on the negativity of CD117.

Acknowledgement

We like to thank our lab technicians George Badicut and Mariana Tudor for their assistance.

Conflict of interests

The authors declare no conflict of interests.

2. Fletcher CD, Hogendoorn P, Mertens F, Bridge J. WHO Classification of Tumors of Soft Tissue and Bone. 4th ed. Lyon, France: IARC Press; 2013.

3. Cho MY, Sohn JH, Kim JM, Kim KM, Park YS, Kim WH, et al. Current trends in the epidemiological and pathological characteristics of gastrointestinal stromal tumors in Korea, 2003-2004. *J Korean Med Sci.* 2010;25:853–862.
4. Tryggvason G, Gíslason HG, Magnússon MK, Jónasson JG. Gastrointestinal stromal tumors in Iceland, 1990-2003: The Icelandic GIST study, a population-based incidence and pathologic risk stratification study. *Int J Cancer* 2005; 117(2):289–93.
5. Goettsch WG, Bos SD, Breekveldt-Postma N, Casparie M, Herings RM, Hogendoorn PC. Incidence of gastrointestinal stromal tumours is underestimated: results of a nation-wide study. *Eur J Cancer* 2005; 41(18):2868–72.
6. Nilsson B, Bümming P, Meis-Kindblom JM, Odèn A, Dortok A, Gustavsson B, et al. Gastrointestinal stromal tumors: the incidence, prevalence, clinical course, and prognostication in the preimatinib mesylate era: a population-based study in western Sweden. *Cancer*. 2005;103:821–829.
7. Ma GL, Murphy JD, Martinez ME, Sicklick JK. Epidemiology of Gastrointestinal stromal tumors in the era of histology codes: Results of a population-based study. *Cancer Epidemiol Biomarkers Prev* 2015; 24(1):298–302.
8. Joensuu H. Gastrointestinal stromal tumor (GIST). *Ann Oncol* 2006; 17(suppl 10):x280–6.
9. Scarpa M, Bertin M, Ruffolo C, Polese L, D'Amico DF, Angriman I. A systematic review on the clinical diagnosis of gastrointestinal stromal tumors. *J Surg Oncol* 2008; 98(5):384–92.
10. Joensuu H, Hohenberger P, Corless CL. Gastrointestinal stromal tumour. *Lancet* 2013;382:973-983
11. Corless CL, Barnett CM, Heinrich MC: Gastrointestinal stromal tumours: origin and molecular oncology. *Nature reviews*. *Cancer* 2011; 11:865-78.
12. Hsu SM, Raine L, and Fanger H. Use of avidin-biotin peroxidase complex (ABC) in immunoperoxidase techniques: a comparison between ABC and unlabeled antibody (PAP) procedures. *J Histochem Cytochem* 1981;29:577-80.
13. Iorgescu A.C., Herlea V., Mustafa E.S., Pechianu C., Sajin M., Dumitru A., Tampa M., Georgescu S.R., Popescu I. Morphological, immunohistochemical and prognostic analysis of gastrointestinal stromal tumors with gastric and small intestine origin. *Medicine in Evolution*. 2016 Dec, (4) :498-508
14. Miettinen M, Lasota J. Gastrointestinal stromal tumors. *Gastroenterol Clin North Am* 2013; 42(2):399–415.
15. Novelli M, Rossi S, Rodriguez-Justo M, et al. DOG1 and CD117 are the antibodies of choice in the diagnosis of gastrointestinal stromal tumours. *Histopathology* 2010; 57(2):259–70.
16. Espinosa I, Lee CH, Kim MK, Rouse BT, Subramanian S, Montgomery K, et al. A novel monoclonal antibody against DOG1 is a sensitive and specific marker for gastrointestinal stromal tumors. *Am J Surg Pathol*. 2008;32:210–8.
17. Miettinen M, Wang ZF, Lasota J. DOG1 antibody in the differential diagnosis of gastrointestinal stromal tumors: a study of 1840 cases. *Am J Surg Pathol* 2009;33:1401–1408.
18. Liegl B, Hornick JL, Corless CL, Fletcher CD. Monoclonal antibody DOG1.1 shows higher sensitivity than KIT in the diagnosis of gastrointestinal stromal tumors, including unusual subtypes. *Am J Surg Pathol.* 2009;33:437–46.
19. Lopes LF, West RB, Bacchi LM, et al. DOG1 for the diagnosis of gastrointestinal stromal tumor (GIST): comparison between 2 different antibodies. *Appl Immunohistochem Mol Morphol* 2010;18:333–337.
20. Miettinen M, Lasota J. Gastrointestinal stromal tumors: review on morphology, molecular pathology, prognosis, and differential diagnosis [review] *Arch Pathol Lab Med*. 2006;130:1466–78.
21. Yamamoto H, Kojima A, Nagata S, Tomita Y, Takahashi S, Oda Y. KIT-negative gastrointestinal stromal tumor of the abdominal soft tissue: a clinicopathologic and genetic study of 10 cases. *Am J Surg Pathol*. 2011 Sep;35(9):1287-95
22. Lee HE1, Kim MA, Lee HS, Lee BL, Kim WH. Characteristics of KIT-negative gastrointestinal stromal tumours and diagnostic utility of protein kinase C theta immunostaining. *J Clin Pathol*. 2008 Jun;61(6):722-9.
23. Reith JD, Goldblum JR, Lyles RH, et al. Extragastric (soft tissue) stromal tumors: an analysis of 48 cases with emphasis on histologic predictors of outcome. *Mod Pathol* 2000;13:577-85.
24. Miettinen M, Monihan JM, Sarlomo-Rikala M, et al. Gastrointestinal stromal tumors/smooth muscle tumors (GISTs) primary in the omentum and mesentery: clinicopathologic and immunohistochemical study of 26 cases. *Am J Surg Pathol* 1999;23:1109-18.
25. Seo HS, Hyeon JY, Shin OR, et al. C-kit-negative gastrointestinal stromal tumor in the stomach. *J Gastric Cancer*. 2015;15(4):290–4.
26. Wada T, Tanabe S, Ishido K, Higuchi K, Sasaki T, Katada C, et al. DOG1 is useful for diagnosis of KIT-negative gastrointestinal stromal tumor of stomach. *World J Gastroenterol*. 2013;19(47):9133–6.
27. Kang GH, Srivastava A, Kim YE, Park HJ, Park CK, Sohn TS, et al. DOG1 and PKC-θ are useful in the diagnosis of KIT negative gastrointestinal stromal tumors. *Mod Pathol*. 2011;24:866–875
28. Tzen CY, Mau BL. Analysis of CD117-negative gastrointestinal stromal tumors. *World J Gastroenterol*. 2005;11:1052–1055.
29. Kontogianni-Katsarou K, Lariou C, Tsompanaki E, et al: KIT-negative gastrointestinal stromal tumors with a long term follow-up: A new subgroup does exist. *World J Gastroenterol* 13:1098-1102, 2007
30. Agaimy, A; Wunsch, PH; Hofstaedler, F; et al. Minute gastric Sclerosing stromal tumours (GIST tumourlets) are common in adults and frequently show c-KIT mutations. *Am J Surg Pathol*, 2007, 31,113.
31. Medeiros F, Corless CL, Duensing A, Hornick JL, Oliveira AM, Heinrich MC, et al. KIT-negative gastrointestinal stromal tumors: proof of concept and therapeutic implications. *Am J*

Surg Pathol.2004;28:889–894.

32. Heinrich MC, Corless CL, Duensing A, et al. PDGFRA activating mutations in gastrointestinal stromal tumors. *Science*, 2003.299(5607):708-710.

33. Minarik G, Plank L, Lasabová Z, et al. Spectrum of mutations in gastrointestinal stromal tumor patients - a population-based study from Slovakia. *APMIS*; 2013. 121(6):539-48.

34. Doyle LA, Nelson D, Heinrich MC, Corless CL, Hornick JL. Loss of succinate dehydrogenase subunit B (SDHB) expression is limited to a distinctive subset of gastric wild-

type gastrointestinal stromal tumours: a comprehensive genotype-phenotype correlation study. *Histopathology* 2012; 61(5):801-9.

35. Agaram NP, Wong GC, Guo T, et al. Novel V600E BRAF mutations in imatinib-naive and imatinib-resistant gastrointestinal stromal tumors. *Genes Chromosomes Cancer* 2008; 47(10):853-9.

36. Yamamoto H, Oda Y. Gastrointestinal stromal tumor: Recent advances in pathology and genetics. *Pathol Int* 2015; 65(1):9-18.