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Research Article

ANTIBACTERIAL ACTIVITY OF GREEN BIOSYNTHESIS OF MAGNETIC IORN OXIDE NANOPARTICLE OF *MURRAYA EXOTICA* L. AQUEOUS EXTRACT AGAINST HUMAN PATHOGENS

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Abstract:

The green synthesis of magnetic iron oxide nanoparticles is a convenient, economical, rapid and eco-friendly method compared to physical and chemical synthesis methods. In the present study iron oxide nanoparticles synthesized by Murraya exotica L. leaves extract. The formation of iron oxide nanoparticles was confirmed by the colour change and further characterized by UV-Visible Spectroscopy, FT-IR analysis, DLS and XRD. The morphology and the size of nanoparticles were analyzed by SEM and HR - TEM analysis. The antibacterial efficacy of synthesized iron oxide nanoparticles exhibited considerable activity against the tested human pathogens. Our study shows that green synthesized iron oxide nanoparticles can be a good source for alternative therapy of bacterial diseases.

Key words: Green synthesis, iron oxide, nanoparticles, Murraya exotica, antibacterial efficacy

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INTRODUCTION:

In nanotechnology, magnetic iron oxide nanoparticles are microscopic particles and sized between 1 and 100 nanometers [1]. It has unique and most important property i.e larger surface area than superior particles which cause them to be more reactive to some other molecules. They are extensively synthesized by using physical and chemical methods. These synthesized methods are needed to use high energy, temperature, toxic chemicals and expensive. The primary goal of nanotechnology is to develop convenient, economical, rapid and eco-friendly green synthesis methods [2].

Magnetic iron oxide nanoparticles research is presently an area of passionate scientific interest due to a broad range of prospective applications. It is used as catalysis [3] high-density magnetic storage media [4] and chemical sensors [5]. It is an effective nano agent to remove a number of pollutants from water resources [6]. It has many important biomedical applications such as for targeted drug delivery in clinical trials [7], contrast agents in magnetic resonance imaging (MRI) [8], antibacterial activity [9] and anticancer activities [10].

Murrava exotica L. commonly known as Chinese box belongs to the family of Rutaceae. It is an evergreen shrub, habitually 2-3 m in height. It is traditionally used in India and China for treatment of diarrhea, dysentery, toothache and body pains from injury or trauma [11]. It was documented to exhibit antimicrobial [12] anti-inflammatory, antinociceptive [13], anti-oxidant [14] and larvicidal activities [15]. In addition, various bioactive compounds such as Colensenone and colensanone [16], cinnamic acid [17], coumarins [18] methoxylated flavonoids [19], alkaloids [20] and phytosterols [21] have been reported in M. exotica L leaves. Furthermore, a study by Lv et al. (2013) [22] revealed sesquiterpenes are the main constituents in essential oil of M. exotica. Considering the ethnomedicinal properties and reported activities of M. exotica L., in the present investigation was preferred for nanoparticles synthesis.

MATERIALS AND METHODS:

Materials: Collection and identification of plant

Fresh healthy leaves of *M. exotica* were collected from Thiruvanamalai Local Park (Figure 1) and were authentically identified by Prof. P. Jayaraman, Institute of Herbal Science, Plant Anatomy Research Centre, West Tambaram, Chennai, India as Rutaceae with voucher specimen number PARC/2015/3147.



Fig 1: Habit of Murraya exotica L.

Scientific classification of *M. exotica* L.

Class	:	Magnoliopsida – Dicotyledons
Subclass	:	Rosidae
Order	:	Sapindales
Family	:	Rutaceae
Genus	:	Murraya
Species	:	<i>Murraya exotica</i> L. – Chinese box
Synonymous	:	Chalcas exotica, Chalcas
		Paniculata, Murraya paniculata

Synthesis of iron oxide nanoparticles using *M. exotica* extract

About 100 g of fresh healthy leaves of *M. exotica* were washed thoroughly with running tap water and double distilled water, cut into fine pieces and shade dried for 10 days under dark condition. After drying the leaves were powdered using kitchen blender. The powdered leaves were soaked in the 200 ml of double distilled water for overnight in a fridge for 4° C and then the rinsed mixtures were boiled for 10 minutes. The extracts were cooled to room temperature and then filtered through Whatman filter paper.

Iron oxide nanoparticles were synthesized by taking FeCl₃.6H₂O and FeCl₂.4H₂O (1:2 molar ratios) and were dissolved in 100 ml of double distilled water in a 250 ml beaker and heated at 80°C with mild stirring using magnetic stirrer under atmospheric pressure. After 10 minutes, 20 ml of the aqueous solutions of *M. exotica* extract was added to the mixture, immediately the light green colour of the *M. exotica* extract of the mixture changed to dark brownish colour. After 10 minutes, 20 ml aqueous solution of sodium hydroxide was added to the mixtures with the rate of 3 ml per minutes for allowing the iron oxide

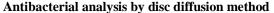
precipitations uniformly. From the first addition of sodium hydroxide the dark brown mixture changed to black suspended particles. The mixture was allowed to cool down to room temperature and the iron oxide nanoparticles were obtained by decantation to form magnetite. The magnetites formed were washed 3 times with double distilled water and 3 times with ethanol and air dried at room temperature.

Characterization

The surface Plasmon resonances (SPR) of synthesized iron oxide nanoparticles have been studied bv UV-Vis double-beam biospectrophotometer Elico-Bl-198 using the software Spectral Treats Version 2.37.4 Rel-1 in the range of 300 to 700 nm. The diffraction pattern was recorded by Seifert Rayflex Software which provides control modules for the complete range of diffractometer accessories together with the corresponding analysis software XRD with Cu-Ka radiation. Particle size of magnetic iron oxide nanoparticles was measured by laser diffractometry using a Nano Size Particle Analyzer in the range between 0.6 nm to 6.0 μ . Morphological analysis of nanoparticles was done using Vega 3 Tescan SEM machine. The morphology of magnetic iron oxide nanoparticles was viewed under a Transmission electron microscope (HR-TEM, Jeol model 3010, at 200 Kv and 104.1µA).

Test Bacteria

The Bacterial cultures employed in this study are Bacillus cereus, Bacillus subtilis, Enterococcus faecalis, Escherichia coli, Klebsiella pneumonia, Micrococcus luteus, Proteus mirabilis, Proteus vulgaris, Pseudomonas fluorescens, Staphylococcus aureus and Vibrio fluvialis.



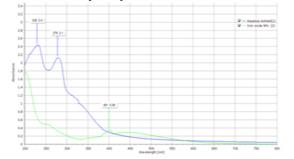


Figure 2. UV absorption spectrum of plant extract and synthesized iron oxide nanoparticles by *M. exotica* aqueous extract

The antibacterial activity of synthesized iron oxide nanoparticles were evaluated using disc diffusion method [23]. A set of sterile discs (6 mm, Hi-media) were impregnated with different concentrations of iron nanoparticles *i.e.* 10 µg/ disc (10µg/µl), 15 µg/ disc (15µg/µl), 20 µg/ disc (20µg/µl), 25 µg/ disc $(25\mu g/\mu l)$ 30 $\mu g/$ disc $(30\mu g/\mu l)$ respectively. Subsequently culture plates were prepared by pouring 20 mL of Mueller-Hinton agar (Hi-media) medium and bacterial suspension swabbed on the medium plates using sterile cotton swab and the plates were kept aside for few minutes. The discs were gently pressed and incubated in inverted position for 24 hours at 37°C. The discs with Norfloxacin (20 µg/ disc) were placed on the MHA plates maintained as positive control. After the incubation period, the susceptibility of the test organisms was determined by measuring the diameter of the zone of inhibition using Himedia zone scale and the obtained results were tabulated for evaluation.

RESULTS AND DISCUSSION:

UV-Visible spectroscopy analysis

In the present investigation, the formation and stability of synthesized iron oxide nanoparticles was further confirmed by UV-Vis spectral analysis. *M. exotica* aqueous extract has the absorption peaks at 230-279 nm regions and a synthesized iron oxide nanoparticle has the absorption peak at 401 nm (Figure 2). It might be due to the excitation of surface plasmon vibrations in the iron oxide nanoparticles, which are very similar to the characteristics UV – visible spectrum of β Fe₂O₃ [24]. Balamurugan *et al.* (2014) [25] reported UV-Vis spectrum of iron oxide nanoparticles synthesized by *Eucalyptus globulus* leaf extract showed absorption peak around 402 nm.

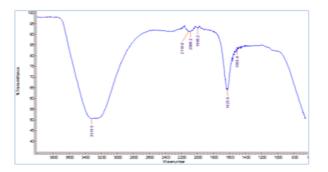


Figure 3. FT-IR spectrum of synthesized iron oxide nanoparticles by *M. exotica* aqueous extract

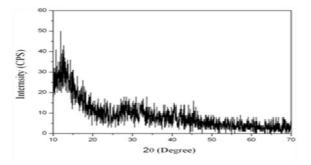


Figure 4. XRD patterns of synthesized iron oxide nanoparticles by *M. exotica* aqueous extract

FTIR analysis

Figure 3 shows the Fourier transform infrared (FTIR) spectra of magnetic nanoparticles. The strong absorption peaks at 3319, 2110, 2088, 1998, 1625 and 1506 cm⁻¹are assigned to O-H stretching, C=N stretching vibrations, aliphatic C-H stretching, C-C multiple bond stretching, conjugated carbonyl (– C=O) group stretching vibration, O-H deformed vibtration and C-O stretching vibrations of synthesized iron oxide nanoparticles respectively. These functional groups are harmony with previous FT- IR spectrum of iron oxide nanoparticles synthesized by various extracts such as *Sargassum muticum* [26], *Passiflora tripartita* var. *mollissima* [27] and *Caricaya papaya* [28].

XRD analysis

The X-ray diffraction (XRD) patterns of Fe₃O₄ by *M.* exotica aqueous extract is shown in Figure 4. In figure 4, weak diffraction peaks with 2θ at 30.0° , 35.6° , 48.3° , 57.2° and 62.5° are observed, which indicate that the Fe₃O₄ particles have an amorphous structure.

DLS analysis

The particle size distributions of green synthesized iron oxide nanoparticles are shown in figure 5. The

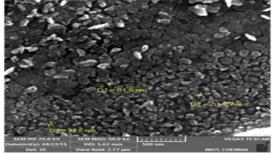


Figure 6. SEM image of synthesized iron oxide nanoparticles by *M. exotica* aqueous extract

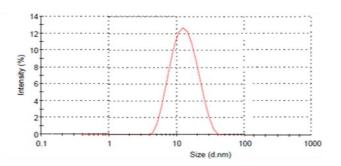


Figure 5. particle size analysis of synthesized iron oxide nanoparticles by *M. exotica* aqueous extract

average size of iron oxide nanoparticles is found to be below100 nm. Similar work was done by Kumar *et al.* (2014) [27] who reported the average particle size of spherical iron oxide nanoparticles synthesized by *Passiflora tripartita* var. *mollissima* fruit is 22.3 ± 3 nm by DLS analysis.

SEM analysis

To determine the morphology and the average size of Fe₃O₄ particles, scanning electron microscopy (SEM) is used. The SEM image shows that magnetite nanoparticles have a mean diameter of about 50nm and a nearly spherical shape. The SEM image of iron oxide nanoparticles synthesized by M. exotica aqueous extract was shown in figure 6. and the size of the iron oxide nanoparticles ranges from 44.5 to 61.9 nm. This is comparable to the findings of Wang et al. (2014) [29] who reported the size of iron nanoparticles by using Eucalyptus leaves was diameter ranging from 20 to 80 nm. On the contrary Latha and Gowri (2014) [28] analysed the SEM image of iron oxide nanoparticles synthesized by *Carica papaya* leaf extracts demonstrated uniformly distributed spherical shaped particles. The increase in the size of nanoparticles confirms the presence of iron oxide nanoparticles with agglomerated in its structures.

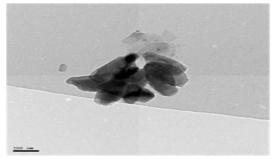


Figure 7. HR-TEM image of synthesized iron oxide nanoparticles by *M. exotica* aqueous extract

Name of the bacterial pathogens	Gree	Standard antibiotic					
	10 µg/disc	15 μg/disc	20 μg/disc	25 μg/disc	30 μg/disc	Norfloxacin 20 µg/ disc	
	Zone of inhibition (Diameter in mm)						
Bacillus cereus	10±1.0	12±1.0	15±1.7	16±1.0	18±2.0	20±0.0	
Bacillus subtilis	12±2.0	14±1.7	15±1.0	16±1.0	18±2.0	22±0.0	
Enterococcus faecalis	12±2.0	13±1.0	15±1.0	17±1.0	18±2.0	23±0.2	
Escherichia coli	10±2.0	12±1.7	13±2.6	14±2.0	18±2.0	10±0.0	
Klebsiella pneumoniae	8±1.0	9±1.0	10±2.0	12±2.0	14±1.0	16±0.1	
Micrococcus luteus	10±1.7	12±1.0	13±0.9	15±0.9	16±0.5	13±0.2	
Proteus mirabilis	9±1.1	10±1.0	8±0.7	11±0.6	13±1.0	14±0.0	
Proteus vulgaris	12±0.9	13±1.4	15±0.9	17±1.4	18±1.4	16±0.0	
Pseudomonas fluorescens	14±1.0	15±1.2	17±1.3	18±1.2	19±1.3	22±0.0	
Staphylococcus aureus	7±0.8	8±1.0	10±1.2	11±1.0	12±1.0	15±0.0	
Vibrio fluvialis	11±1.1	9±1.1	9±0.5	10±0.9	11±1.0	14±0.2	

Table 1: Antibacterial activity of iron oxide nanoparticles synthesized by M. exotica aqueous extract

HR-TEM analysis

The morphology and structure of the iron oxide nanoparticles were further investigated by HR-Transmission Electron Microscopy. Figure 7 shows the TEM image of iron oxide nanoparticles synthesized by aqueous leaves extract of *M. exotica*. TEM image also revealed the successful synthesis of nanosized iron oxide particles, the average core diameter of 100 nm and the nanoparticles are agglomerated and cluster. The aggregation might be due to a magnetic property of Iron oxide nanoparticles. Iron oxide nanoparticles have a large surface to volume ratio and possess high surface energies. Accordingly, they tend to aggregate so as to minimize the surface energies [30].

Antibacterial activity

In the present investigation, the different concentrations of green synthesized iron oxide nanoparticles were exhibited variable degrees of antibacterial activity against the tested bacterial pathogens (Table 1). The activity of the magnetite iron oxide nanoparticles was concentration dependent; with the increase in concentration the activity was also increased. The inhibition activity of the iron oxide nanoparticles were compared with standard antibiotic Norfloxacin. The iron oxide nanoparticles showed minimum zone of inhibition (ranging 7-14mm) against the tested pathogens at 10 µg/disc concentration. The maximum zone of inhibition (ranging 17-19mm) was observed at 30 µg/disc concentration of iron oxide nanoparticles. These findings are in agreement with the earlier research on the antibacterial activity of iron oxide nanoparticles synthesized by *Lawsonia inermis* and *Gardenia jasminoides* leaves extract against *E. coli*, *P. mirabilis* and *S. aureus* [31]. Likewise, in another study by Groiss *et al.* (2017) [32] who reportediron oxide nanoparticles synthesized by leaf extract of *Cynometra ramiflora* exhibited effective inhibition against *E. coli* and *S. epidermidis*.

CONCLUSION:

For the first time, biosynthesis of magnetic iron oxide nanoparticles by using *M. exotica* L aqueous extract is reported. Measurement of UV, IR, XRD, DLS, SEM, and TEM analysis confirmed the structures. The antibacterial activity of iron oxide nanoparticles showed potent activity against human pathogens. On the basis of this research work, green synthesized iron oxide nanoparticles can be a good source for alternative therapy of bacterial diseases. The study can be extended for nanomedicine application and preclinical studies in relevant animal models.

REFERENCES:

1. Thakkar KN, Mhatre SS, Parikh RY. Biological synthesis of metallic nanoparticles. Nanomedicine, 2010; 6(2) : 257–262.

2.Allouche J. Synthesis of organic and bioorganic nanoparticles: an overview of the preparation methods. In *Nanomaterials: A Danger or a Promise?* 2013; (pp. 27-74). Springer London.

3.Weiss W, Ranke W. Surface chemistry and catalysis on well-defined epitaxial iron-oxide layers. Prog. Surf. Sci, 2002; 70 : 1-15.

4.Zeng H, Li J, Liu JP, Wang ZP, Sun SH. Exchanged-coupled nanocomposite magnets via nanoparticle self-assembly. Nature, 2002; 420 : 395.

5.Raj VB, Nimal AT, Parmar Y, Sharma MU, Sreenivas K, Gupta V. Cross sensitivity and selectivity studies on ZnO surface acoustic wave ammonia sensor. Sens. Actuators B, 2010; 147 : 517.

6.Feng J, Hu X, Yue PL, Zhu HY, Lu GQ. Degradation of azo-dye orange II by a photoassisted Fenton reaction using a novel composite of iron oxide and silicate nanoparticles as a catalyst. Ind Eng Chem Res, 2003; 42 : 2058–2066.

7.Kim DK, Zhang Y, Voit W, Rao KV, Kehr J, Bjelke B, Muhammed M. Superparamagnetic iron oxide nanoparticles for bio-medical applications. Scripta Materialia, 2001; 44(8) : 1713-1717.

8.Modo MMJ, Bulte JWM. Molecular and Cellular MR Imaging; CRC Press: Boca Raton, FL, 2007.

9.Ismail RA, Sulaiman GM, Abdulrahman SA, Marzoog TR. Antibacterial activity of magnetic iron oxide nanoparticles synthesized by laser ablation in liquid. Materials Science and Engineering C, 2015; 53 : 286-297.

10.Nagajyothi PC, Pandurangan M, Kim DH, Sreekanth TVM, Shim J. Green synthesis of iron oxide nanoparticles and their catalytic and *in vitro* anticancer activities. Journal of Cluster Science, 2017; 28(1): 245-257.

11. China Pharmacopoeia Committee, Pharmacopoeia of the People's Republic of China. Beijing, Chemical Industry Press, 2005; p. 9.

12.EI-Sakhawy FS, EI-Tantawy ME, Ross SA, EI-Sohly MA. Composition and antimicrobial activity of the essential oil of *Murraya exotica* L. Flavour Frag J, 1998; 13 : 59–62.

13.Wu L, Li P, Wang X, Zhuang Z, Farzaneh F, Xu R. Evaluation of anti-inflammatory and antinociceptive activities of *Murraya exotica*. Pharmaceutical biology, 2010; 48(12) : 1344-1353.

14.Khatun A, Rahman M, Jahan S. Preliminary phytochemical, cytotoxic, thrombolytic and antioxidant activities of the methanol extract of *Murraya exotica* Linn. leaves. Oriental Pharmacy and Experimental Medicine, 2014; 14(3) : 223-229.

15.Krishnamoorthy S, Chandrasekaran M, Raj GA, Jayaraman M, Venkatesalu V. Identification of chemical constituents and larvicidal activity of essential oil from *Murraya exotica* L.(Rutaceae) against *Aedes aegypti, Anopheles stephensi* and *Culex quinquefasciatus* (Diptera: Culicidae). Parasitology Research, 2015; 114(5):1839-1845.

16.Ahmad ZA, Begum S. Colensenone and colensanone (non-diterpene oxide) from *Murraya exotica* Linn. Indian Drugs, 1987; 24 : 322.

17.Barik BR, Kundu AB. A cinnamic acid derivative and a coumarin from *Murraya exotica*. Phytochemistry, 1987; 26 : 3319- 3321.

18.Ito C, Furukawa H. Constituents of *Murraya exotica* L. structural elucidation of new coumarins. Chemical and Pharmaceutical Bulletin, 1987;35 : 4277-4285.

19.Bishay DW, El-Sayyad SM, Abd El-Hafiz MA, Achenbach H, Desoky EK. Phytochemical study of *Murraya exotica* L. 1: Methoxylated flavonoids of the leaves. Bulletin of Pharmaceutical Sciences, 1987; 10:55-70.

20.Desoky EK, Kamel MS, Bishay DW. Alkaloids of *Murraya exotica* L. (Rutaceae) cultivated in Egypt. Bulletin of Faculty of Pharmacy Cairo University, 1992; 30 : 235-238.

21.Desoky EK. Phytosterols from *Murraya exotica*. Phytochemistry, 1995; 40 : 1769-1772.

22.Lv HN, Guo XY, Tu PF, Jiang Y. Comparative analysis of the essential oil composition of *Murraya paniculata* and *M. exotica*. Natural product communications, 2013; 8(10) : 1473-1475.

23.Bauer AW, Kirby WM, Sherris JC, Turck M. Antibiotic susceptibility testing by a standardized single disk method. American Journal of Clinical Pathology, 1996; 45(4) : 493–496.

24.Cornell RM, Schwertmann U. The Iron Oxides-Structure, Properties, Reactions, Occurrences and Uses. Darmstadt: Wiley–VCH GmbH & Co. KGaA. 2003.

25.Balamurugan M, Saravanan S, Soga T, Synthesis of iron oxide nanoparticles by using *Eucalyptus globulus* plant extract. e-Journal of Surface Science and Nanotechnology, 2014; 12 : 363-367.

26. Mahdavi M, Namvar F, Ahmad MB, Mohamad R. Green biosynthesis and characterization of magnetic iron oxide (Fe₃O₄) nanoparticles using seaweed (*Sargassum muticum*) aqueous extract. Molecules, 2013; 18(5): 5954-5964.

27.Kumar B, Smita K, Cumbal L, Debut A. Biogenic synthesis of iron oxide nanoparticles for 2arylbenzimidazole fabrication. Journal of Saudi Chemical Society, 2014;18(4) : 364-369.

28.Latha N, Gowri M. Biosynthesis and characterization of Fe₃O₄ nanoparticles using *Carica*

papaya leaves extract. Int. J. Sci. Res, 2014; 3(11) : 1551-1556.

29.Wang T, Jin X, Chen Z, Megharaj M, Naidu R. Green synthesis of Fe nanoparticles using *Eucalyptus* leaf extracts for treatment of eutrophic wastewater. Science of the total environment, 2014; 466 : 210-213.

30.Wu W, He Q, Jiang C. Magnetic iron oxide nanoparticles: synthesis and surface functionalization strategies. Nanoscale research letters, 2008; 3(11) : 397.

31.Naseem T, Farrukh MA. Antibacterial activity of green synthesis of iron nanoparticles using *Lawsonia inermis* and *Gardenia jasminoides* leaves extract. Journal of Chemistry, 2015.

32.Groiss S, Selvaraj R, Varadavenkatesan T, Vinayagam R. Structural characterization, antibacterial and catalytic effect of iron oxide nanoparticles synthesized using the leaf extract of *Cynometra ramiflora*. Journal of Molecular Structure, 2017; 1128 : 572-578.